# Continuous preparation and use of dibromoformaldoxime as a reactive intermediate for the synthesis of 3-bromoisoxazolines

Claudio Battilocchio,<sup>1\*</sup> Francesco Bosica,<sup>1</sup> Sam M. Rowe,<sup>1</sup> Bruna L. Abreu,<sup>1</sup> Edouard

Godineau,<sup>2\*</sup> Matthias Lehmann,<sup>2</sup> Steven V. Ley<sup>1</sup>

<sup>1</sup>Innovative Technology Centre, Department of Chemistry, University of Cambridge, Lensfield Road, CB2 1EW Cambridge, UK;

<sup>2</sup>Syngenta Crop Protection AG, Crop Protection Research, Schaffhauserstrasse 101, CH-4332 Stein, Switzerland

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#### **1.** General information

All batch reactions were performed using oven-dried glassware (200 °C) under an atmosphere of argon unless otherwise stated. All flow reactions were performed using a Vapourtec R2+R4 system,<sup>1</sup> a Vapourtec E-Series system,<sup>2</sup> HiTech Zang SyrDos syringe pumps<sup>3</sup> and Knauer pumps K120.<sup>4</sup>

Unless stated otherwise, reagents were obtained from commercial sources and used without further purification.

Flash column chromatography was performed using high-purity grade silica gel (Merck grade 9385) with a pore size 60 Å and 230–400 mesh particle size under air pressure. Analytical thin layer chromatography (TLC) was performed using silica gel 60 F254 precoated glass backed plates and visualized by ultraviolet radiation (254 nm) and/or potassium permanganate solution as appropriate.

<sup>1</sup>H NMR spectra were recorded on Bruker Avance DPX-600 (600 MHz), with the residual solvent peak as the internal reference (CDCl<sub>3</sub> = 7.26 ppm). <sup>1</sup>H resonances are reported to the nearest 0.01 ppm. <sup>13</sup>C-NMR spectra were recorded on the same spectrometer with proton decoupling, with the solvent peak as the internal reference (CDCl<sub>3</sub> = 77.00 ppm). All <sup>13</sup>C resonances are reported to the nearest 0.01 ppm. The multiplicity of <sup>1</sup>H signals are indicated as: s =singlet, d = doublet, dd = doublet of doublets, ddd = doublet of doublets of doublets, t = triplet, q = quadruplet, sext = sextet, m = multiplet, br = broad, or combinations of thereof. Coupling constants (*J*) are quoted in Hz and reported to the nearest 0.1 Hz. Where appropriate, measures of the same coupling constant are averaged. The removal of solvent under reduced pressure was carried out on a standard rotary evaporator.

High resolution mass spectrometry (HRMS) was performed using positive electrospray ionisation (ESI+), on either a Waters Micromass LCT Premier spectrometer or performed by the Mass Spectrometry Service for the Chemistry Department at the University of Cambridge. All m/z values are reported to 4 decimal places and are within  $\pm$  5 ppm of theoretical values.

Infrared spectra were recorded on a Perkin-Elmer Spectrum RX One FT-IR ATR (Attenuated Total Reflectance) spectrometer. The samples were prepared as thin films deposited on the ATR, unless otherwise specified. Only structurally important absorptions are quoted. Absorption maxima (vmax) are reported in wavenumbers (cm<sup>-1</sup>).

#### 2. Synthetic procedures for DFBO and 3-bromoisoxazolines



#### Step 1: synthesis of hydroximinoacetic acid 6

An aqueous stream of glyoxylic acid (5 M in H<sub>2</sub>O, 3 mL min<sup>-1</sup>) was combined at a T-piece with a stream of hydroxylamine aqueous solution (5 M in H<sub>2</sub>O, 3 mL min<sup>-1</sup>). The resulting reaction stream was then directed towards a 10mL politetrafluoroetilene (PTFE) coil reactor (o.d. 1/16'',  $\tau = 130$  s), operating at room temperature, to ensure full conversion to hydroxyiminoacetic acid **6.** The reaction solution was directed through a 75 psi backpressure regulator (BPR) and the reactor output was collected in a reservoir.

#### Step 2: synthesis of DBFO 2



menna maging camera

A solution of bromine (2.0 M, in  $CH_2CI_2$ ) was loaded in a 50 mL perfluoroalkoxy alkane (PFA) pre-loading coil (o.d. 1/8") before being mixed at a T-piece with a solution of **6** (1.25 M, in H<sub>2</sub>O). The resulting mixture was pumped (flow rate 1.20 mL min<sup>-1</sup> each channel) through the

packed bed column mixing element and then directed into 5 mL PFA coil reactor (o.d. 1/16'',  $\tau = 126$  s). The output stream of DBFO **2** was directed through a 100 psi BPR and then towards a multi-port valve. After reaching steady-state, the valve was switched to collection and the reaction mixture directed to a liquid-liquid separator, equipped with a turbulence settler. An IR thermal imaging camera<sup>5</sup> was used to monitor the reaction exotherm.



#### Step 3: synthesis of 3-bromoisoxazolines

The solution of DBFO **2** was pumped (1 mL min<sup>-1</sup>) to meet a stream of the appropriate alkene (0.5 M, in CH<sub>2</sub>Cl<sub>2</sub>, flow rate of 1 mL min<sup>-1</sup>), at a T-piece, combined with a  $KH_2PO_4/K_2HPO_4$  buffer (1.0 M, in H<sub>2</sub>O, pH = 6.8) delivered at a flow rate of 2 mL min<sup>-1</sup>, allowed to mix through the packed bed column mixing element, and then reacted in a 5 mL PFA coil reactor (o.d. 1/16"). The reactor output was directed towards a multi-port valve. After reaching steady-state, the valve was switched to collection and the reaction mixture directed to a liquid-liquid separator, where the organic layer was recovered and the solvent evaporated *in vacuo*.

The crude mixture was purified by flash chromatography (eluent: Hexane/EtOAc 100:0 to 0:100) to afford the desired products **7b-n**.

# Large scale, telescoped synthesis of *N*-(3-Bromo-4,5-dihydroisoxazol-5-yl)benzamide (7a)

An aqueous stream of glyoxylic acid (5 M in H<sub>2</sub>O, 3 mL min<sup>-1</sup>) was combined at a T-piece with a stream of hydroxylamine aqueous solution (5 M in H<sub>2</sub>O, 3 mL min<sup>-1</sup>). The resulting reaction stream was then directed towards a 10mL PTFE coil reactor (o.d. 1/16'',  $\tau = 130$  s), operating at room temperature, to ensure full conversion to hydroxyiminoacetic acid **6.** The reaction solution was directed through a 75 psi back-pressure regulator (BPR) and the reactor output was combined with a solution of potassium monohydrogen phosphate (2.5 M, 6 mL min<sup>-1</sup>) and then passed through a mixing element prior to going being directed though a 100 psi bpr and hydroxyiminoacetic acid **6** is collected in a reservoir. The freshly made solution of **6** was then combined with a solution of bromine (2.0 M, in CH<sub>2</sub>Cl<sub>2</sub>) at a T-piece (flow rate 11 mL min<sup>-1</sup> each channel) into a 20 mL Vapourtec "static mixer" coil reactor<sup>6</sup> (o.d. 1/8"). An IR thermal imaging camera<sup>5</sup> was used to monitor the reaction exotherm (tested over a period of 4 h, Figure S1).



**Figure S1.** Yield of DBFO **2** and temperature detection at the mixing element during a reaction period of 4 h.

The output stream of DBFO **2** was directed through a 100 psi BPR and then directed towards a multiport valve. After reaching steady-state, the valve was switched and the reaction

mixture directed to a liquid-liquid separator, equipped with a turbulence settler (see Figure S5). The solution of DBFO **2** was then pumped (10 mL min<sup>-1</sup>) to meet a stream of N-vinylbenzamide **8a** (0.75 M, in CH<sub>2</sub>Cl<sub>2</sub>, flow rate of 10 mL min<sup>-1</sup>), at a T-piece, combined with a KH<sub>2</sub>PO<sub>4</sub>/K<sub>2</sub>HPO<sub>4</sub> buffer (1.0 M, in H<sub>2</sub>O, pH = 6.8) delivered at a flow rate of 20 mL min<sup>-1</sup>, and then reacted through a 20 mL Vapourtec "static mixer" coil reactor<sup>6</sup> (o.d. 1/8"). The resulting biphasic mixture was collected, the organic layer recovered and the solvent evaporated *in vacuo* to give **7a** (97% yield) (Figure S2).



Figure S2. Reaction set up for the telescoped synthesis of 7a.



3. Mixing elements and liquid-liquid separator

Figure S3. Packed bed column mixing element, Step 2.



Figure S4. Static mixer coil (20 mL).



Figure S5. Turbulence settler and liquid-liquid separator.

#### 4. Safety data

#### Hydroxyiminoacetic acid 6

Differential Scanning Calorimetry (DSC)

Methodenname: 20-350°C 4° Min. Modul: DSC1/700/ID2570, 03.07.2014 15:45:29 Bemerkungen: weisser Feststoff in Goldkapsel



DSC, isotherm 110°C, 15h



#### DSC, isotherm 100°C, 15h



9

#### DSC, isotherm 80°C, 48h



RADEX



#### RA-734; Radex up to 220°C; open system



#### **Dibromoformaldoxime 2**

#### Thermal stability

Hydroxylamine and its derivatives tend to be thermally unstable and some of these compounds exhibit explosive properties. The decomposition of this type of material is accelerated by the presence of metals but the corresponding salts tend to be relatively thermally stable.

An initial thermal stability screening, using DSC showed evidence of endothermic activity with a minimum at 65°C, likely to be associated to melting (literature: mp. 65-71°C; Rohloff, J. C.; Robinson III, J.; Gardner J. O. *Tetrahedron Lett.* **1992**, *33*, 3113-3116). There is evidence of a rather sharp exotherm with onset temperature at 122°C, reaching a peak at 131°C (peak height 66.5 mW), with an associated heat output of *ca*. 950 J/g. There is further exothermic activity above 156°C. The magnitude of the sharp exotherm is sufficient to result in a temperature rise under low heat loss conditions of >550 K, and due to the structure of the material this would be expected to be accompanied by the generation of significant quantities of gas. Testing using RADEX provided additional information regarding the thermal stability and the potential of gas release of the material. There is evidence of pressure rise which can be attributed to a gas evolution from around 78°C, whilst at *ca*. 123°C and 131°C there is evidence of two separate, extremely rapid pressure rises. The shape of the pressure curve confirms the autocatalytic nature of the decomposition and, based upon the structure, it is likely that the gas evolved as the material starts to

decompose. The total volume of gas generated is equivalent to *ca.* 57.5 L/kg at ambient temperature.

The thermal stability experiments confirmed the initial analysis which indicated that the material could undergo a violent exothermic decomposition. The results of the initial screening tests pointed out that the necessary analyses are to be carried out on this material, in accordance with the UN Transport Regulations (Orange book), to determine whether the compound is to be classified as an explosive, should it need to be prepared on a large scale.

Based upon this data, it is not possible to define a safe temperature threshold which can guarantee that no violent decomposition would occur; however, additional experimental work allowed defining parameters to safely store this material, for a limited period.

#### Storage stability

The exothermic event shown in the DSC analysis and the pressure curve in the RADEX test are of concern, as they exhibit the classic profile of an autocatalytic decomposition process. This means that safe storage of the material cannot be simply based upon temperature considerations, as the storage period might have a significant impact upon the stability.

To investigate the autocatalytic nature of decomposition, a series of isothermal DSC experiments were carried out with particular focus on the storage period (induction time) which would induce decomposition.

The isothermal DSC experiments carried out at 80, 90 and 100°C showed excellent agreement with both the 'induction time' (*i.e.*, time before heat output was detected) and the "time to peak" rate of decomposition; this was used to calculate the data given in the table below:

| Temperature (°C) | Time to start of decomposition (hours) | Time to peak rate of decomposition (hours) | Time from "start of<br>decomposition to peak"<br>rate (hours) |
|------------------|--|--|---|
| -40              | 6000                                   | 60330                                      | 54330   |
| -35              | 3390                                   | 30100                                      | 26710   |
| -30              | 1960                                   | 15450                                      | 13490   |
| -25              | 1160                                   | 8145                                       | 6985  |
| -23              | 945                                    | 6350                                       | 5405  |
| -20              | 700                                    | 4405                                       | 3705  |
| -15              | 432                                    | 2440                                       | 2008  |
| -10              | 271                                    | 1382                                       | 1111  |
| -5               | 173                                    | 800  | 627   |
| 0                | 112                                    | 472  | 360   |
| 5                | 74                                     | 284  | 210   |
| 10               | 49.5                                   | 174  | 124.5   |
| 15               | 33.5                                   | 108  | 74.5  |
| 20               | 23                                     | 68.6                                       | 45.6  |
| 25               | 16                                     | 44.1                                       | 28.1  |

#### Table S1.

| 30  | 11.3 | 28.8  | 17.5  |
|-----|------|-------|-------|
| 35  | 8    | 19    | 11    |
| 40  | 5.8  | 12.75 | 6.95  |
| 45  | 4.2  | 8.7   | 4.5   |
| 50  | 3.1  | 5.95  | 2.85  |
| 55  | 2.3  | 4.13  | 1.83  |
| 60  | 1.7  | 2.9   | 1.2   |
| 65  | 1.3  | 2.06  | 0.76  |
| 70  | 1    | 1.48  | 0.48  |
| 75  | 0.76 | 1.07  | 0.31  |
| 80  | 0.59 | 0.78  | 0.19  |
| 85  | 0.46 | 0.575 | 0.115 |
| 90  | 0.35 | 0.43  | 0.08  |
| 95  | 0.28 | 0.32  | 0.04  |
| 100 | 0.22 | 0.24  | 0.02  |

The sample tested had been obtained from a third party and it is not known how old the material were, and how it had been stored. However, given the decomposition potential of this material the values shown in the second column should be used as the basis for determining appropriate safe storage period for this material.

Due to the thermal instability of the material, its use as a synthetic reagent would need to be carefully considered, especially on scale.

Generation of the reagent in-situ may be a more likely prospect for larger scale operation or indeed safer laboratory scale work.

#### Differential Scanning Calorimetry (DSC)

Experimental investigations were carried out in the PHS laboratory at Muenchewilen (Switzerland). Gold plated stainless steel high pressure crucibles were used with a heating rate of 4 K/min over the range -20 to 350°C for the initial dynamic test and 20 – 350°C for the tests carried out after isothermal experiments.

#### Initial dynamic test





Isothermal at 80°C



Isothermal at 90°C



Isothermal at 100°C

WMU/Stein/150505-470 05.05.2015 15:42:46 ^exo WMU/Stein/150505-470, 05.05.2015 13:48:18 Methodenname: Iso 100°C/12h + Restwärme 20-350°C CSAA115064; 1, 1-Dibromoformaldoxime, 2.0000 mg Modul: DSC1/700/ID2570, 03.07.2014 15:45:29 Bemerkungen: Hellgelber Feststoff in Goldkapsel Auftraggeber: M.EL Qacemi Integral normalisiert 1655.62 mJ 827.81 Jg^-1 Peakhöhe 42.91 m¥ =21455 W/kg Peak 101.76 °C Extrapolierter 101.69 °C Peakweite Linke Grenze 679.02e-06 °C 100.00 °C Rechte Grenze 100.02 °C 50 10 15 20 25 35 0 30 40 m in mW Bemerkungen: Rückstand aus Lauf150505-470 in Goldkapsel 485.94 mJ 242.97 Jg^-1 Integral normalisiert 103.11 mJ Integral normalisiert 51.55 Ja^-1 Peakhöhe 2.00 mW Peakhöhe 0.20 mW 228.62 °C Peak Peak 136 77 °C Extrapolierter 231.73 °C Extrapolierter Peak 148.21 °C Peakweite 13.68 °C eakweite 33.81 °C Linke Grenze 214.19 °C 105.95 °C Linke Grenze Rechte Grenze 255.31 °C Rechte Grenze 168.56 °C TTTTI AUTOLICE -WMU/WST/150505-470-1, 05.05.2015 15:19:15 CSAA115064; 1, 1-Dibromoformaldoxime, 2.0000 mg Methodenname: 20-350°C 4°Min. Modul: DSC1/700/ID2570, 03.07.2014 15:45:29 °C 20 40 60 80 100 120 140 160 180 200 220 240 260 280 300 320

#### Radex

#### Radex; pressure/power output relation



#### 5. Characterization of compounds 7a-n



*N*-(3-Bromo-4,5-dihydroisoxazol-5-yl)benzamide (7a): yellowish solid (97%). <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>):  $\delta$  8.40 (s, 1H), 7.58 (d, J = 7.5 Hz, 2H), 7.36 (t, J = 7.6 Hz, 2H), 7.17 (t, J = 7.6 Hz, 2H), 5.17 (dd, J = 7.6, 9.8 Hz, 1H), 3.68 (dd, J = 9.8, 18Hz, 1H), 3.61 (dd, J = 7.6, 18 Hz, 1H) ppm. <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>):  $\delta$  167.7, 139.3, 136.4, 129.1, 125.1, 119.9, 79.1, 45.9 ppm.<sup>7</sup>



**3-Bromo-5-(4-bromophenyl)-isoxazoline (7b):** yellow solid (98%). **FTIR** (ν<sub>max</sub>, cm<sup>-1</sup>) 1593. <sup>1</sup>**H NMR** (600 MHz, CDCl<sub>3</sub>) δ 7.61 – 7.44 (m, 2H), 7.25 – 7.21 (m, 2H), 5.64 (dd, *J* = 10.9, 8.8 Hz, 1H), 3.64 (dd, *J* = 17.3, 11.0 Hz, 1H), 3.17 (ddd, *J* = 17.3, 8.7, 3.9 Hz, 1H) ppm. <sup>13</sup>**C NMR** (151 MHz, CDCl<sub>3</sub>) δ 138.4, 136.8, 132.1, 127.7, 122.8, 82.5, 49.2 ppm. **HRMS** (ESI) calculated for C<sub>9</sub>H<sub>7</sub>Br<sub>2</sub>NO [M+H]<sup>+</sup> 303.8967, found 303.8960.



**3-Bromo-5-(o-tolyl)-isoxazoline (7c):** yellowish oil (98%). **FTIR** ( $\nu_{max}$ , cm<sup>-1</sup>) 1607. <sup>1</sup>**H NMR** (600 MHz, CDCl<sub>3</sub>)  $\delta$  7.35 – 7.29 (m, 1H), 7.18 – 7.12 (m, 2H), 7.11 – 7.06 (m, 1H), 5.74 (dd, *J* = 11.0, 9.0 Hz, 1H), 3.54 (dd, *J* = 17.2, 11.1 Hz, 1H), 2.98 (dd, *J* = 17.2, 9.0 Hz, 1H), 2.21 (s, 3H) ppm. <sup>13</sup>**C NMR** (151 MHz, CDCl<sub>3</sub>)  $\delta$  137.5, 136.7, 134.4, 130.7, 128.4, 126.6, 125.2, 80.8, 48.3, 19.3 ppm. **HRMS** (ESI) calculated for C<sub>10</sub>H<sub>10</sub>BrNO [M+H]<sup>+</sup> 240.0019, found 240.0015.



**3-Bromo-5-(p-tolyl)-isoxazoline (7d):** colourless oil (98%). IR ( $\nu_{max}$ , cm<sup>-1</sup>) 1607, 2925. <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  7.13 (d, *J* = 8.1 Hz, 2H), 7.10 (d, *J* = 8.2 Hz, 2H) 5.52 (dt, *J* = 20.2, 10.1 Hz, 1H), 3.48 (dd, *J* = 17.3, 10.9 Hz, 1H), 3.09 (dd, *J* = 17.3, 9.2 Hz, 1H), 2.27 (s, 3H) ppm. <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>)  $\delta$  138.7, 136.8, 136.2, 129.6, 126.1, 83.3, 49.1, 21.2 ppm. HRMS (ESI) calculated for C<sub>10</sub>H<sub>10</sub>BrNO [M+H]<sup>+</sup> 240.0019, found 240.0008.



**3-Bromo-5-mesityl-isoxazoline (7e):** yellowish solid (20%). **IR** ( $v_{max}$  cm<sup>-1</sup>) 1608, 2967. <sup>1</sup>**H NMR** (600 MHz, CDCl<sub>3</sub>)  $\delta$  6.88 (s, 2H,), 6.07 (t, J = 12.2 Hz, 1H), 3.44 (dd, J = 17.5, 11.9 Hz, 1H), 3.25 (dd, J = 17.5, 12.5 Hz, 1H), 2.33 (s, 6H), 2.28 (s, 3H) ppm. <sup>13</sup>**C NMR** (151 MHz, CDCl<sub>3</sub>)  $\delta$  138.3, 137.0, 136.5, 130.3, 130.3, 80.7, 46.7, 20.8, 20.1 ppm. **HRMS** (ESI) calculated for C<sub>10</sub>H<sub>10</sub>BrNO [M+H]<sup>+</sup> 268.0332, found 268.0326.



**3-Bromo-5-(4-methoxyphenyl)-isoxazoline (7f):** amber solid (98%). **IR** ( $v_{max}$ , cm<sup>-1</sup>) 1609, 2960. <sup>1</sup>**H NMR** (600 MHz, CDCl<sub>3</sub>)  $\delta$  7.27 (d, *J* = 8.6 Hz, 2H), 6.91 (d, *J* = 8.7 Hz, 2H), 5.62 (dd, *J* = 10.5, 9.7 Hz, 1H), 3.81 (s, 3H), 3.56 (dd, *J* = 17.3, 10.8 Hz, 1H), 3.20 (dd, *J* = 17.3, 9.4 Hz, 1H) ppm. <sup>13</sup>**C NMR** (151 MHz, CDCl<sub>3</sub>)  $\delta$  160.1, 136.8, 131.1, 127.7, 114.4, 83.4, 55.5, 49.1 ppm. **HRMS** (ESI) calculated for C10H10BrNO2 [M+H]<sup>+</sup> 255.9968, found 255.9964.



**3-Bromo-5-(4-(chloromethyl)phenyl)-isoxazoline (7g):** brown solid (98%). **FTIR** ( $v_{max}$ , cm<sup>-1</sup>) 1609, 2965. <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  7.40 (d, *J* = 8.2 Hz, 2H), 7.33 (d, *J* = 8.2 Hz, 2H, H1-H5), 5.66 (dd, *J* = 10.9, 8.9 Hz, 1H), 4.58 (s, 2H), 3.62 (dd, *J* = 17.3, 11.0 Hz, 1H), 3.17 (dd, *J* = 17.3, 8.8 Hz, 1H) ppm. <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>)  $\delta$  139.5, 138.0, 136.9, 129.1, 126.3, 82.7, 49.1, 45.7 ppm. HRMS (ESI) calculated for C<sub>10</sub>H<sub>9</sub>BrCINO [M+H]<sup>+</sup> 273.9629, found 273.9624.



**3-Bromo-5-(4-methyl benzoyl)-isoxazoline (7h):** white powder (96%). **IR** ( $\nu_{max}$ , cm<sup>-1</sup>) 1612, 1709. <sup>1</sup>**H NMR** (600 MHz, CDCl<sub>3</sub>)  $\delta$  8.13 – 8.01 (m, 2H), 7.41 (d, *J* = 8.3 Hz, 2H), 5.73 (dd, *J* = 11.0, 8.7 Hz, 1H), 3.92 (s, 3H), 3.67 (dd, *J* = 17.3, 11.1 Hz, 1H), 3.18 (dd, *J* = 17.3, 8.6 Hz, 1H) ppm. <sup>13</sup>**C NMR** (151 MHz, CDCl<sub>3</sub>)  $\delta$  166.5, 144.2, 136.6, 130.5, 130.2, 125.8, 82.5, 52.3, 49.3 ppm. **HRMS** (ESI) calculated for C<sub>11</sub>H<sub>10</sub>BrNO<sub>3</sub> [M+H]<sup>+</sup> 283.9917, found 283.9912.



**3-bromo-5-(naphthalen-1-yl)-isoxazoline (7i):** orange oil (98%). **IR** ( $v_{max}$ , cm<sup>-1</sup>) 1600, 3056. <sup>1</sup>**H NMR** (600 MHz, CDCl<sub>3</sub>)  $\delta$  7.92 (d, J = 7.5 Hz, 1H), 7.85 (d, J = 8.2 Hz, 1H), 7.77 (d, J = 8.0 Hz, 1H), 7.66 (d, J = 7.1 Hz, 1H), 7.58 – 7.47 (m, 3H), 6.36 (dd, J = 11.1, 8.5 Hz, 1H), 3.84 (dd, J = 17.2, 11.2 Hz, 1H), 3.25 (dd, J = 17.2, 8.5 Hz, 1H) ppm. <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>)  $\delta$  137.3, 134.7, 134.1, 129.7, 129.4, 129.2, 126.8, 126.1, 125.6, 123.2, 122.6, 81.1, 49.2 ppm. **HRMS** (ESI) calculated for C<sub>13</sub>H<sub>10</sub>BrNO [M+H]<sup>+</sup> 276.0019, found 276.0015.



**3-bromo-isoxazolin-5-yl 4-(tert-butyl)benzoate (7j):** white solid (50%). **IR** (ν<sub>max</sub>, cm<sup>-1</sup>) 1640, 1728, 2915. <sup>1</sup>**H NMR** (600 MHz, CDCl<sub>3</sub>) δ 7.95 (d, *J* = 8.6 Hz, 2H), 7.46 (d, *J* = 8.6 Hz, 2H), 6.92 (dd, *J* = 7.0, 1.6 Hz, 1H), 3.64 (dd, *J* = 18.4, 7.0 Hz, 1H), 3.29 (dd, *J* = 18.4, 1.6 Hz, 1H), 1.33 (s, 9H) ppm. <sup>13</sup>**C NMR** (151 MHz, CDCl<sub>3</sub>) δ 165.0, 157.8, 138.3, 130.0, 126.0, 125.6, 96.7, 47.8, 35.3, 31.2 ppm. **HRMS** (ESI) calculated for C<sub>14</sub>H<sub>16</sub>BrNO<sub>3</sub> [M+Na]<sup>+</sup> 348.0206, found 348.0199.



**3-Bromo-3a,8b-dihydro-4H-indeno[2,1-d]isoxazole (7k**): white solid (38%). <sup>1</sup>**H** NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  7.49 (d, *J* = 7.5 Hz, 1H), 7.36-7.29 (m, 3H), 6.10 (d, *J* = 8.9 Hz, 1H), 4.14 (dt, *J* = 8.9, 5.3 Hz, 1H), 3.32 (d, *J* = 5.3 Hz, 2H) ppm. <sup>13</sup>**C** NMR (151 MHz, CDCl<sub>3</sub>)  $\delta$  141.9, 139.8, 139.6, 130.1, 128.1, 125.9, 124.9, 89.0, 55.7, 35.2 ppm. HRMS (ESI) calculated for C<sub>10</sub>H<sub>8</sub>BrNO [M+H]<sup>+</sup> 237.9862, found 237.9858.



**3-Bromo-3a,4,5,6,7,7a-hexahydrobenzo[d]isoxazole (7l):** colourless oil (48%). <sup>1</sup>**H NMR** (600 MHz, CDCl<sub>3</sub>) δ 4.60 (dt, *J* = 8.4, 5.0 Hz, 1H), 3.13-3.09 (m, 1H), 1.95 – 1.71 (m, 3H), 1.60 - 1.29 (m, 5H) ppm. <sup>13</sup>**C NMR** (151 MHz, CDCl<sub>3</sub>) δ 145.7, 80.4, 50.0, 25.8, 24.4, 21.2, 20.0 ppm. **HRMS** (ESI) calculated for C<sub>7</sub>H<sub>10</sub>BrNO [M+H]<sup>+</sup> 204.0019, found 204.0010.<sup>8</sup>



Ethyl 3-bromo-5-(trifluoromethyl)-dihydroisoxazoline-4-carboxylate, ethyl 3-bromo-4-(trifluoromethyl)-dihydroisoxazoline-5-carboxylate (7m): white solid (90%). IR ( $v_{max}$ , cm<sup>-1</sup>) 1605, 1742. <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  5.29 (m, *J* = 6.5 Hz, 1H), 5.16 (d, *J* = 5.8 Hz, 1H), 4.42

(dd, J = 8.0, 5.8 Hz, 1H), 4.37 - 4.22 (m, 5H), 1.34 (dd, J = 14.0, 7.0 Hz, 6H) ppm. <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>) δ 166.65, 164.60, 136.94, 134.13, 125.81, 125.31, 123.96, 123.45, 122.10, 121.59, 120.24, 119.73, 81.04, 80.81, 80.58, 80.36, 79.60, 79.58, 63.70, 63.40, 59.85, 59.65, 59.44, 59.23, 59.01, 59.00, 29.76, 14.13, 14.04, 14.02 ppm. **HRMS** (ESI) calculated for C<sub>7</sub>H<sub>7</sub>BrF<sub>3</sub>NO<sub>3</sub> [M+H]<sup>+</sup> 289.9634, found 289.9622.



**1,4-bis-(3-bromo-isoxazolin-5-yl)benzene (7n):** yellowish solid (98%). **FTIR** ( $v_{max}$ , cm<sup>-1</sup>) 1600, 2992, 3215. <sup>1</sup>H NMR (600 MHz, CDCl<sub>3</sub>)  $\delta$  7.37 (d, J = 4.3 Hz, 4H), 5.68 (dd, J = 10.9, 8.8 Hz, 2H), 3.64 (dd, J = 17.3, 11.0 Hz, 2H), 3.18 (ddd, J = 17.3, 8.8, 1.0 Hz, 2H) ppm. <sup>13</sup>C NMR (151 MHz, CDCl<sub>3</sub>)  $\delta$  140.0, 136.9, 126.6, 82.8, 49.3 ppm. **HRMS** (ESI) calculated for C<sub>12</sub>H<sub>10</sub>Br<sub>2</sub>N<sub>2</sub>O<sub>2</sub> [M+H]<sup>+</sup> 372.9182, found 372.9178.

6. Copies of <sup>1</sup>H- and <sup>13</sup>C-NMR spectra





# 3-Bromo-5-(o-tolyl)-isoxazoline (7c)



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# 3-Bromo-5-(p-tolyl)-isoxazoline (7d)



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## 3-Bromo-5-mesityl-isoxazoline (7e)





# 3-Bromo-5-(4-methoxyphenyl)-isoxazoline (7f)





# 3-Bromo-5-(4-methyl benzoyl)-isoxazoline (7h)



## 3-Bromo-5-(naphthalen-1-yl)-isoxazoline (7i)





#### 3-Bromo-3a,8b-dihydro-4H-indeno[2,1-d]isoxazole (7k)



Ethyl 3-bromo-5-(trifluoromethyl)-dihydroisoxazoline-4-carboxylate, ethyl 3-bromo-4-(trifluoromethyl)-dihydroisoxazoline-5-carboxylate (7m)





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