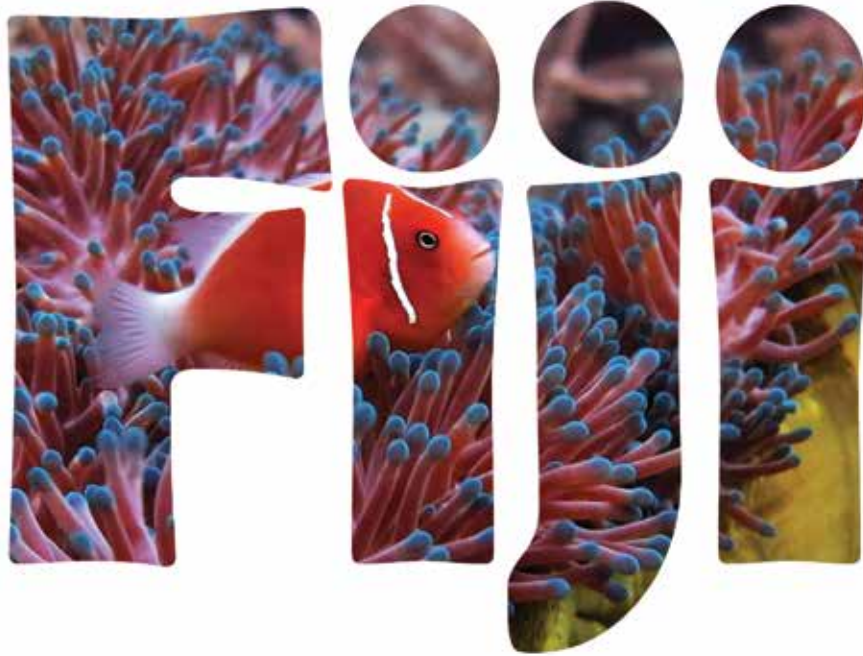


Lau Province



Global Reef Expedition Final Report

June 2-28, 2013

Andrew W. Bruckner, Alexandra Dempsey, Georgia Coward,
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Front cover: Clownfish in an anemone by Derek Manzello
Back cover: Coral reefs of Fiji by Derek Manzello

**Khaled bin Sultan Living Oceans Foundation
Global Reef Expedition**

Lau Province, Fiji

June 2-28, 2013

FINAL REPORT

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**Support for the research conducted during June 2013
as part of the Global Reef Expedition (GRE),
provided by His Royal Highness Prince Khaled bin Sultan**

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Executive Summary

The Khaled bin Sultan Living Oceans Foundation is conducting the Global Reef Expedition – the largest coral reef survey and mapping expedition in history. The Global Reef Expedition is a five-year mission to survey the health and resiliency of coral reefs around the world. In June of 2013, the Global Reef Expedition came to Fiji to assess the state of their coral reefs and create detailed habitat maps of their shallow water marine habitats.

Using standardized methods, scientists on the Global Reef Expedition mapped and surveyed the coral reefs of Fiji over the course of four weeks, from June 2-28. The research focused on coral reefs surrounding 11 islands in Lau Province, Fiji: Cicia, Fulaga, Kabara, Mago, Matuka, Moala, Nayau, Totoya, Tuvuca, Vanua Balavu, and Vanua Vatu. These small, remote islands are far from the tourist center of Fiji, and home to small villages of Pacific Islanders run by traditional leaders. These communities rely heavily on the coral reef ecosystem for their livelihood, mainly through engaging in artisanal and substance fishing.



Fijian fishing vessel

The research objectives of the mission were to:

1. *Develop detailed habitat maps of shallow marine habitats*
2. *Evaluate the composition, structure, and health of coral reefs*
3. *Assess the diversity and abundance of reef fish and algae, including commercially valuable species*
4. *Document impacts of broad scale disturbances and patterns of recovery with emphasis on storm damage and crown of thorns predation impacts*
5. *Evaluate the effects of environmental stressors on coral health*



Live coral cover on Fiji's reefs was exceptional

In addition, research fellows and partners on the Global Reef Expedition studied ocean acidification and its effects on coral growth, surveyed the symbiotic community inhabiting coral colonies and studied how environmental factors influenced their composition and photosynthetic efficiency, and surveyed the population of sea cucumbers and the impact of the sea cucumber fishery.



The research vessel, M/Y Golden Shadow, in Fiji

The Expedition was conducted in partnership with local experts from the Wildlife Conservation Society of Fiji, Lau Provincial Office and Fiji Department of Fisheries, and scientists from the University of the South Pacific, Department of Fisheries. The Foundation also welcomed researchers from Nova Southeastern University, University of the Azores, University of the Philippines, NOAA/University of Miami, National Museum of Marine Biology and Aquarium of Taiwan, Reef Environmental Education Foundation (REEF), and Victoria University to join us on the expedition. Roko Sau Josefa Cinavilakeba from the Pacific Blue Foundation and High Chief of Yasayasa Moala Group and Roko Laitia Raloa from the Lau Provincial Office, joined the Expedition to provide assistance with traditional protocols, meetings and deliberations with the Chiefs, Council and elders on each island, and education activities.

Habitat Mapping

Detailed shallow water marine habitat maps were produced for all 11 islands surveyed on this Expedition. Using a combination of satellite imagery and surveys in the field, over 2,000 square kilometers of high-resolution habitat and bathymetric maps were created that extended from the shoreline to 25 m depth. Twenty one different types of shallow water marine habitats were identified in this region within lagoonal, back reef and fore reef locations. Seven intertidal, coastal and nearshore terrestrial habitats were also identified and mapped. Coral reefs covered 285 km² in this region, with an additional 781 km² of coral habitat present in shallow lagoonal habitats.

Coral Reef Surveys

Scientists conducted comprehensive coral reef surveys at 70 dive sites throughout Lau Province, Fiji that included a representative sample of fore reef, back reef, and lagoonal habitats on each island when possible. Scientific dive teams conducted transect surveys to determine substrate composition live cover (including live coral cover), and an assessment of reef fish abundance, diversity, and population structure at each of the dive sites at multiple depths from 5-30m.

Survey	Depth (m)					TOTAL
	<8	8-13	14-18	19-25	>25	
Benthic	57	74	102	48	1	282
Coral	15	34	25	19	1	94
Photo-transects	75	124	120	106	0	425
Fish	14	161	183	1	0	359

Table 1. Total number of benthic, coral, photo-transect and fish transects completed in Lau Province.

The reefs surveyed were generally healthy with high coral cover, diverse soft corals, and well developed deep-water coral communities. There was also a high diversity of motile invertebrates including octopus, sea cucumbers and giant clams. Overall, 85% of the substrate surveyed was covered in living organisms, and the remainder was either uncolonized sand, pavement, or rubble.



A clownfish peeks out from its anemone home. Invertebrate diversity on Fiji's reefs was high.

Live coral cover was high on all of the islands surveyed, averaging 36%. On several islands including Cicia, Fulaga, Mago and Nayau, average coral cover exceeded 45%. Coral cover was dominated by species of *Acropora*, with strong healthy communities of faviids, *Pocillopora*, *Porites*, and *Montipora* coral species present as well. The island of Vanua Balavu had the most diverse coral community, with over 24% of the coral community composed of uncommon species. At most sites, macroalgae cover was low (4%), and cover of crustose coralline algae was high (21.6%), signs of a healthy reef system, composed of a fair number of herbivores and plenty of prime settlement opportunities for new coral recruits.



Reef fish in Fiji were small but diverse

Fish communities on Fiji's reefs varied significantly in species assemblages and biomass between islands, but the fish were small with few large fish remaining. An unusually high proportion of fish on the reef were smaller than 20cm (92%), and less than 1% of the fish were large fish (>40cm). In particular, large individuals of important functional groups such as herbivores and piscivores were few and far between. This population structure suggests the current level of fishing pressure may be too high, as fishermen take out the large and commercially valuable species first, and the populations of these fish are not given time to recover.

Despite their small size, the reef fish communities were fairly diverse, with a total of 482 species recorded on our surveys. However, they were not

very dense, averaging about 2 fish per square meter of reef. Fish biomass was low, averaging 0.1 kg / m² across all islands. Biomass was highest on Toyoya and Cicia islands, where it reached 0.2 kg/ m². The low biomass of fish on Fiji's reefs can be attributed to the small size of the fish and their relatively low abundance on the reefs.



Fiji's reefs recovered well from a coral bleaching event and crown of thorns starfish outbreak

The most spectacular find of the Expedition was to see how well the reefs recovered from periods of stress, which is a great sign of hope for their future. Fiji's reefs proved to be exceptionally resilient. Both coral and fish communities rebounded from a mass bleaching event in 2002 and subsequent crown of thorns starfish (COTS) outbreak. Crown of thorns starfish are a voracious coral predator, a few individuals on a reef is normal and healthy, but an outbreak can decimate a reef in a matter of weeks. We found COTS on the reefs of Cicia Island in Fiji, but no lingering signs of the outbreak. The great news was that reefs damaged by bleaching or COTS years ago were now covered in new colonies of acroporid corals.

One of the few disconcerting observations of threats to reef health was the presence of cyanobacterial mats. This microorganism was found on many the islands surveyed and usually covered areas of pavement, dead coral, or sandy areas in the lagoon. Cyanobacteria was particularly abundant on the island of Tuvuca where it covered 14% of the seafloor. The development of these mats is likely the result of intense fishing pressure for sea cucumbers, which filter the sand and are a vital component of a healthy coral reef ecosystem. Scientists observed hundreds of sea cucumbers drying on racks on shore, but not many in the marine ecosystem. The absence of sea cucumbers is extremely alarming and could cause a phase shift, or transition in the health and community structure of the reef. Increased regulation of this fishery may be warranted to ensure the health and resiliency of Fiji's coral reef ecosystems.



Additional regulation of the sea cucumber fishery may be needed to maintain the health of Fiji's reefs

Education & Outreach

In addition to conducting scientific surveys, the Khaled bin Sultan Living Oceans Foundation conducted a series of educational seminars with students, teachers, stakeholders, and community leaders in Lau Province, Fiji. A coral reef education curriculum was presented at 12 schools and 8 community groups that covered coral reefs, their

benefits to the community, and what can be done to protect these important marine ecosystems. The Foundation also discussed the state of the coral reefs in Fiji with government officials, community leaders, and stakeholders at a series of seminars on the island of Suva. Together, these outreach and education seminars reached over 1,500 people. This was the first time the Foundation conducted such a large and extensive education and outreach on the Global Reef Expedition.



Children learn about coral reefs on Totoya Island

Overall, the Global Reef Expedition found Fiji's reefs to be healthy and resilient, and on-par with other remote island reefs in the region. The amount of live coral cover on Fiji's reefs was particularly high, the fish and coral communities were diverse even though the individual fish were small, and the ecosystem recovered quickly. The Foundation hopes the habitat maps and scientific information collected on the health of Fiji's coral reefs on the Global Reef Expedition will help Fiji's leaders in their efforts to manage their marine resources, and continue to be a valuable resource on the health and resiliency of these coral reefs for many years to come.



A fishing village on the island of Totoya

Introduction

The Lau Islands form one of 14 provinces in Fiji. They are located in the South Pacific Ocean, due east of the Koro Sea and separated from the Fiji Platform by the Nanuku Channel. Lau Province covers an area of 114,000 sq km and includes a chain of about 57 islands and 43 islets at the eastern edge of Fiji on the top of the north-south running Lau-Colville Ridge. While most of the northern group of islands are high and volcanic in origin, those in the south are a mix of extinct oceanic volcanoes and low-lying carbonate islands. These include a number of atolls and extensive barrier reef systems that enclose a number of smaller islands. Together, they have a total land area of 487 sq km. The approximate land area, reef length and lagoonal area is shown in Table 2.

Approximately 30 islands in the Lau Province are sparsely populated. The total population of Lau Province recorded in 2007 is 10,700 with residents inhabiting 13 districts and 72 villages. The population is reported to have declined substantially between 1996- 2006 (Turner et al. 2007) and it continues to decline. Best available population estimates for the 11 villages examined in this study are shown in Table 2. Although human population is relatively low, a number of human stressors are still reported to have contributed to the decline of these reef systems. The most disconcerting human threats affecting these remote reef systems are unsustainable fishing pressure, destructive fishing, sedimentation and nutrients associated with runoff from agriculture and waste disposal, and removal of mangroves.

Previous surveys of Fiji's coral reefs indicate they have high biodiversity, including 219 species of scleractinian corals, over 2,030 species of reef fishes, 478 species of marine molluscs and 422 species of algae. Shallow marine habitats also include mangrove forests composed of nine species and sea grass beds consisting of five different species. Fiji's reefs have an average live coral cover of 45%, although there has been considerable variability between sites and over time (Wilkinson 2008).

The most significant regional changes to Fiji's coral reefs over the last 15 years have been attributed to the 2000 coral bleaching event, with localized damage from crown of thorns starfish (COTS) and *Drupella* gastropods, cyclones, and coral disease (Wilkinson 2011). Archipelago-wide mortality from bleaching was reported to range from 40% to 80% (Lovell and Sikes 2008). The acroporids were the most severely impacted corals, with near total losses of these species noted in some locations. Fortunately, the rapid recolonization and growth of acroporid corals promoted rapid increases in coral cover with some locations reaching pre-disturbance levels by 2007.

While less historic data is available on the condition of coral reefs in Lau Province, these reefs have also experienced large declines in coral cover since 1998. For instance, at Totoya and Matuka, coral cover declined from 50-70% in 2000 to 20% in 2006, with a shift from a dominance of acroporids to *Pocillopora spp.* and *Porites spp.* (Wilson et al. 2008). Our coral reef surveys in 2013 indicate that these reefs are starting to rebound with coral cover at Matuka and Totoya islands rising to 30% and 32% respectively.

Cyclones, bleaching events and outbreaks of coral predators are thought to be the main factors responsible for the observed changes in the composition and structure of benthic communities in Lau Province (Cumming et al. 2002, Dulvy et al. 2004). Mass bleaching was reported in 2000, with localized bleaching in 2002 and 2006. Large aggregations of COTS were first reported in 1998. Interestingly, Kabara had recovered from a COTS outbreak by 2000 and coral cover increased from 1-18% by 2006 (Wilson et al. 2008). Cicia and Vanua Vatu experienced localized damage from Cyclone Ami in 2003.

Changes in fish community structure has also been reported for several islands in the Lau Province between 2000-2006. These predominantly included declines in small bodied fishes, and not the larger species that are targeted for subsistence and export fisheries. One of the primary reasons for the

Island	Land area (km ²)	Elevation (m)	Lagoonal area (km ²)	Reef area (km ²)	Reef length (km)	# of villages	Approx. Population
Cicia	34.9	?	9.2	9.3	25.8	5	1047
Fulaga	17.5	79	43.0	14.1	31.2	3	400
Kabara	31.8	?	10.4	7.0	24.8	4	700
Mago	22.0	?	11.5	8.8	22.4	1	Private island
Matuka	29.1	385	30.6	19.2	30.3	7	800
Moala	64.7	468	115.6	46.0	59.0	8	1596*
Nayau	19.2	?	6.8	5.6	20.8	3	500
Totoya	30.9	366	109.0	26.9	43.2	4	800
Tuvuca	13.3	243	14.3	8.8	20.8	1	180
Vanua Balavu	69.1	283	730.1	116.8	155.4	17	1200
Vanua Vatu	4.2	79	7.3	5.3	14.5	1	89

Table 2. Total areal extent of the major island groups, lagoons and reef, linear distance of the outer reef system, maximum elevation, number of villages and estimated population for each island/atoll examined in 2013. Land area and reef length were calculated from satellite imagery. Reef length is linear extent of reef surrounding major island group. Lagoonal area was calculated from the sum of all back reef and lagoonal habitats. Reef area was estimated by adding up the total area of all fore reef, lagoonal and back reef habitat classes, excluding rubble and soft bottom classes. Population data is from the 1996 census as reported in Dulvy et al 2004; these were updated using 2014 census data from Wikipedia when available.

decline was thought to be the loss of key habitat areas associated with the demise of acroporid communities from past bleaching events and COTS outbreaks (Wilson et al. 2008, 2010). Because fishing on these islands is predominantly restricted to subsistence fishing by local residents and most areas are still managed using traditional practices, stressors are likely to be minimal. Further, a reduction in human populations and shifts to other occupations is thought to have caused a decline in fishing pressure between 1996-2006 (Turner et al. 2007).

The current study was undertaken to obtain updated information on the condition of coral reef communities off islands that were previously examined, and also to fill gaps and obtain baseline data for reef systems in unstudied areas. The surveys were designed to provide relevant information on the changes to coral reef communities that occurred since the 2000-2007 studies undertaken by Dulvy et al. (2004), Turner et al. (2007) and Wilson et al. (2008). This

includes information on the patterns of recovery from bleaching events in 2000 and 2002, COTS outbreaks, and other large scale stressors, as well as the response of fish populations to these changes and to changing human pressure. Data were also collected on current stressors impacting these ecosystems. Targeted research was conducted to evaluate the health of corals, symbiont communities harbored by *Pocillopora spp.*, ocean chemistry, and potential effects of ocean acidification on massive reef building corals. A second component of the study involved the characterization of different shallow marine habitats and mapping of their distribution and bathymetry. These datasets are designed to aid in the development of spatially-based management tools. A third component, conducted by partners from the Wildlife Conservation Society, involved an assessment of the status of invertebrate taxa targeted by fisheries, with emphasis on sea cucumber populations.

Date	Survey Location
1 June	Arrival of scientists
2 June	Depart NADI, Overnight passage
3-5 June	Totoya
6-7 June	Matuka
8-10 June	Moala
11-12 June	Fulaga
13-14 June </td <td>Kabara</td>	Kabara
15 June	Vanua Vatu
16 June	Nayau
17 June	Tuvuca
18-19 June	Cicia
20 June	Mago
21-26 June	Vanua Balavu
27 June	Mago, overnight transit
28 June	Science team disembarks

Table 3. Schedule of the Lau Province, Fiji research mission.

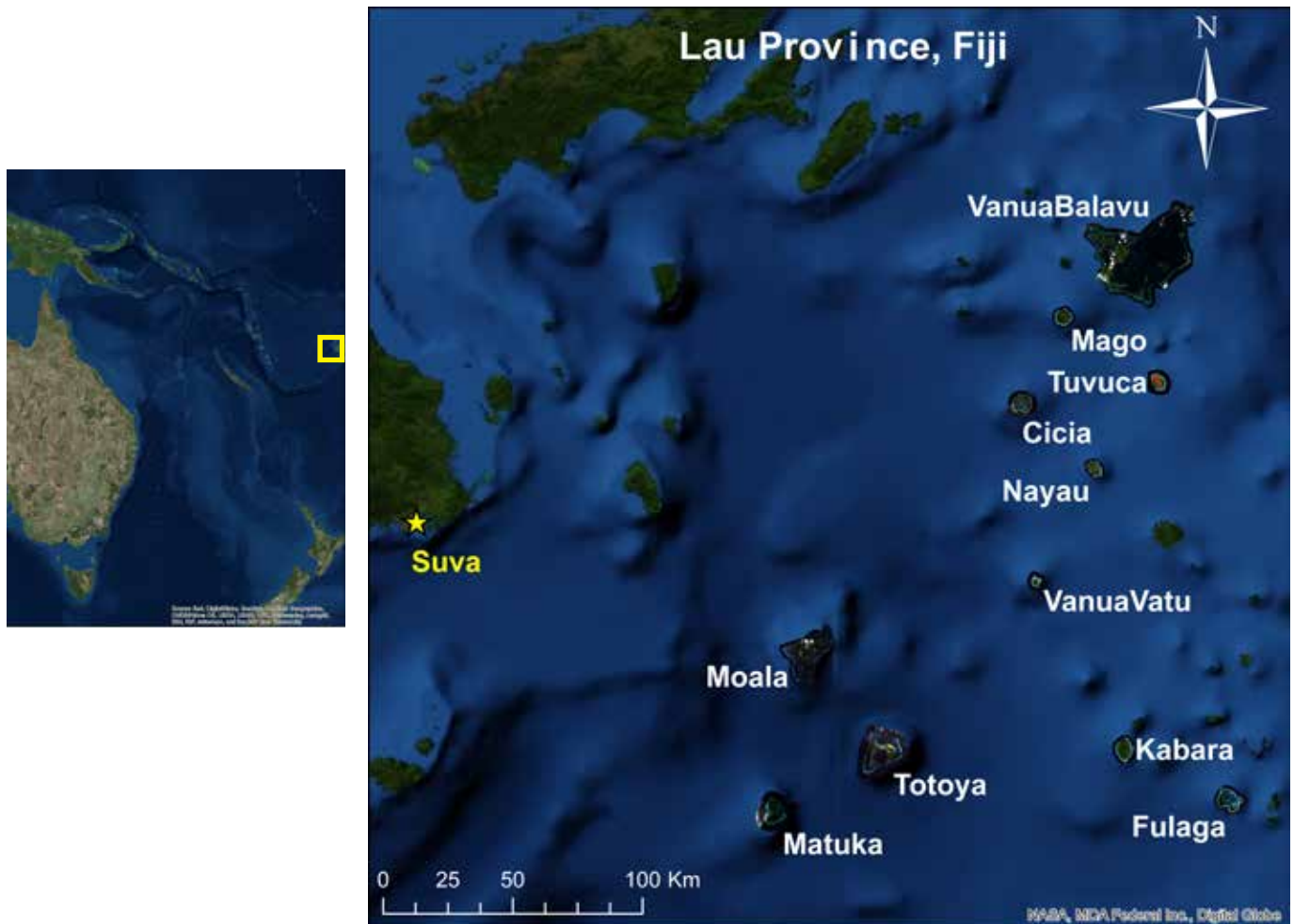


Fig. 1. Locations examined during the research mission to the Lau Province, Fiji.

Habitat Mapping

Using multispectral WorldView-2 satellite imagery obtained from DigitalGlobe Inc., in combination with data obtained from groundtruthing (Fig. 5-7), high resolution bathymetric maps and habitat maps were created for shallow marine environments within the lagoon and fore reef. The maps extend from the shoreline to approximately 25 m depth. Groundtruthing efforts included continuous bathymetry measures, drop camera analysis, characterization of sediment and hard substrates and habitat features using two acoustic sub-bottom profiling equipment (Stratabox and Hydrobox), snorkel and dive assessments, and fine scale photo-transect surveys.



Fig. 2. Acoustic sub-bottom profiling equipment. Stratabox (left) and Hydrobox (right).

Satellite Imagery

A total of 1,105 sq. km of WorldView-2 (8 band) satellite imagery was acquired for the eight islands targeted for mapping (Table 4). The satellite images had a spatial resolution of 2 m by 2 m (i.e., each pixel covers a 4 m² area) enabling real-time navigation in the field to locate features of interest and to avoid dangerous features (e.g., emergent reefs). In order to navigate, the team used the scenes in conjunction with a differential GPS device (dGPS). The imagery was used in conjunction with groundtruth data to create bathymetric and benthic habitat maps.

Benthic Video

An underwater video camera attached to a cable, called a drop-cam, was used to gather video on the benthic composition at each survey site. At each

point, the drop-cam was held from the survey boat enabling it to ‘fly’ along the sea floor as it records video for 15 to 60 seconds. During this time, the laptop operator watched the video in real-time and guided the drop-cam operator to raise or lower the camera. In this manner, we were able to prevent damage to marine life. The video was recorded on a ruggedized laptop, and the geographic position, time, date, boat heading, and boat speed were stamped into the video. Drop-cam deployment was limited to depths above 40 m due to the limited length of the tether cable (50 m). The acquired videos are used to create the benthic habitat maps by providing the necessary information for the development of a habitat classification scheme and training of classification models. A minimum of 30 drop-cam videos were gathered per day (Table 4). The locations of drop camera deployments are shown in Fig. 5-7.



Fig. 3. SeaView Underwater Video System.

Acoustic Depth Soundings

Depth soundings were gathered along transects shown in Fig. 5-7 using Hydrobox, a single-beam acoustic transducer, developed by Syqwest. The instrument emits 3 pings per second. Depths were estimated based on the time the return-pulse’s reaches the sounder’s head. Geopositional data were simultaneously acquired by the dGPS unit. The estimated depth values and their geographic location were recorded in the ruggedized laptop, with an average of 100,000 acoustic depth soundings gathered during a full work day (Table 4). The soundings were used to train a water-depth derivation model, which is based on the spectral attenuation of light in the water column. The final bathymetric maps have the same spatial resolution as the satellite imagery.

Site	Imagery (sq km)	# of drop-cams	# of depth soundings	Track length (km)
Cicia	91	38	96,931	32.159
Fulaga	102	58	250,518	71.533
Kabara	76	31	156,591	40.175
Mago	57	29	102,444	25.169
Matuka	168	54	181,378	45.706
Moala	268	157	524,732	124.100
Nayau	45	24	82,152	25.427
Totoya	311	105	412,190	130.740
Tuvuca	78	53	181,574	42.533
Vanua Balavu	1,036	215	983,605	243.290
Vanua Vatu	41	23	65,708	17.392
Total	2,273	787	3,037,823	798.224

Table 4. Summary of groundtruthing data sets. The total area of satellite imagery acquired, number of deployments of the drop camera, number of depth soundings and total distance covered by the groundtruthing team are shown for the 11 islands.

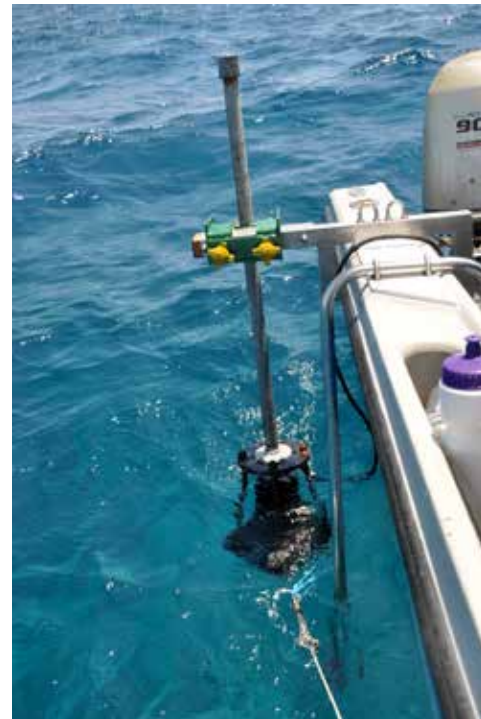


Fig. 4. The groundtruthing team deploying the drop camera (top left). The Hydrobox acoustic sounder recording water depths (top right). Recording and examining the video image collected of the benthos by the drop camera (bottom left).

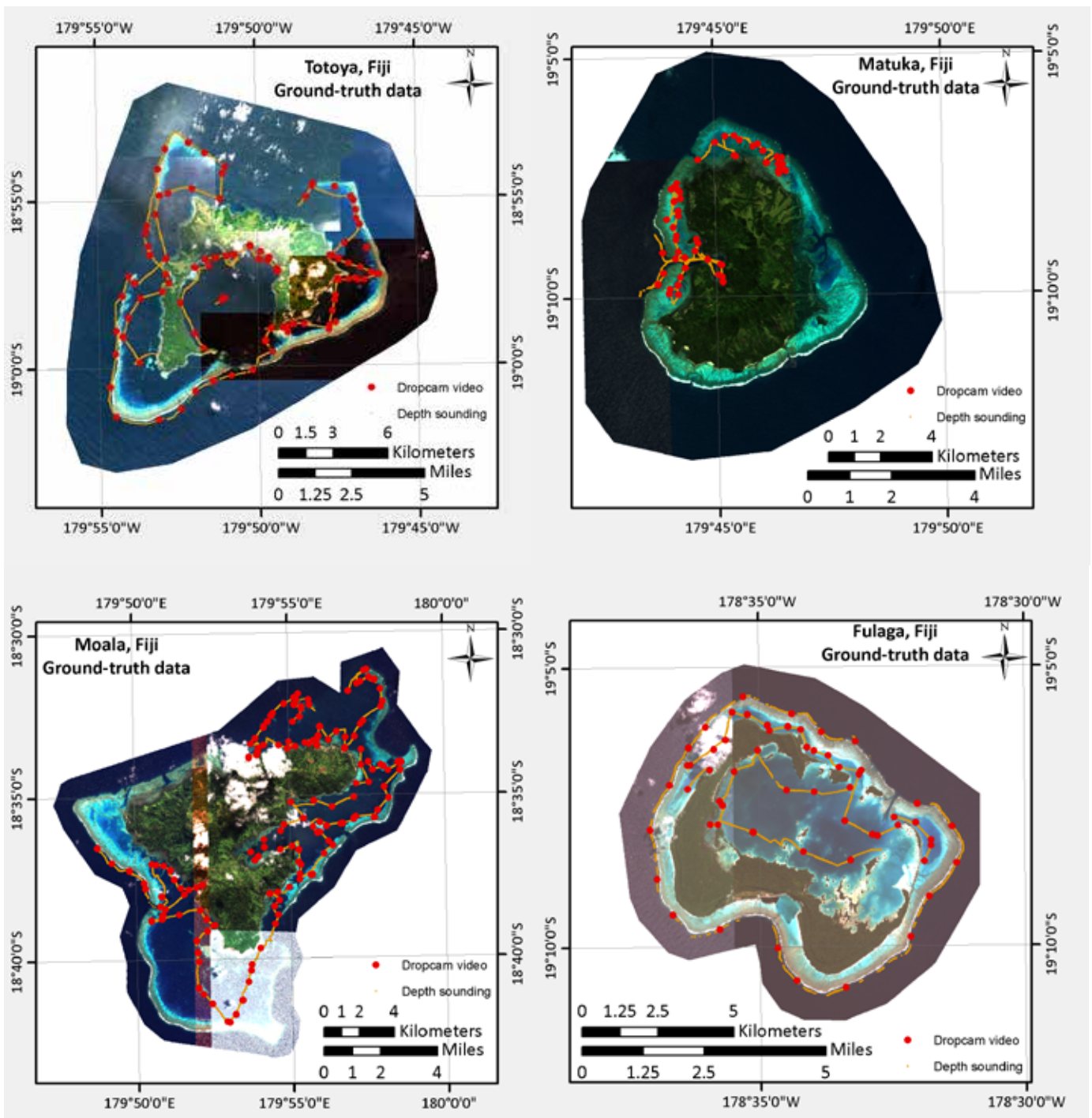


Fig. 5. Groundtruthing track (orange line) and location of drop camera videos (red dots) for Totoya, Matuka, Moala and Fulaga, Fiji.

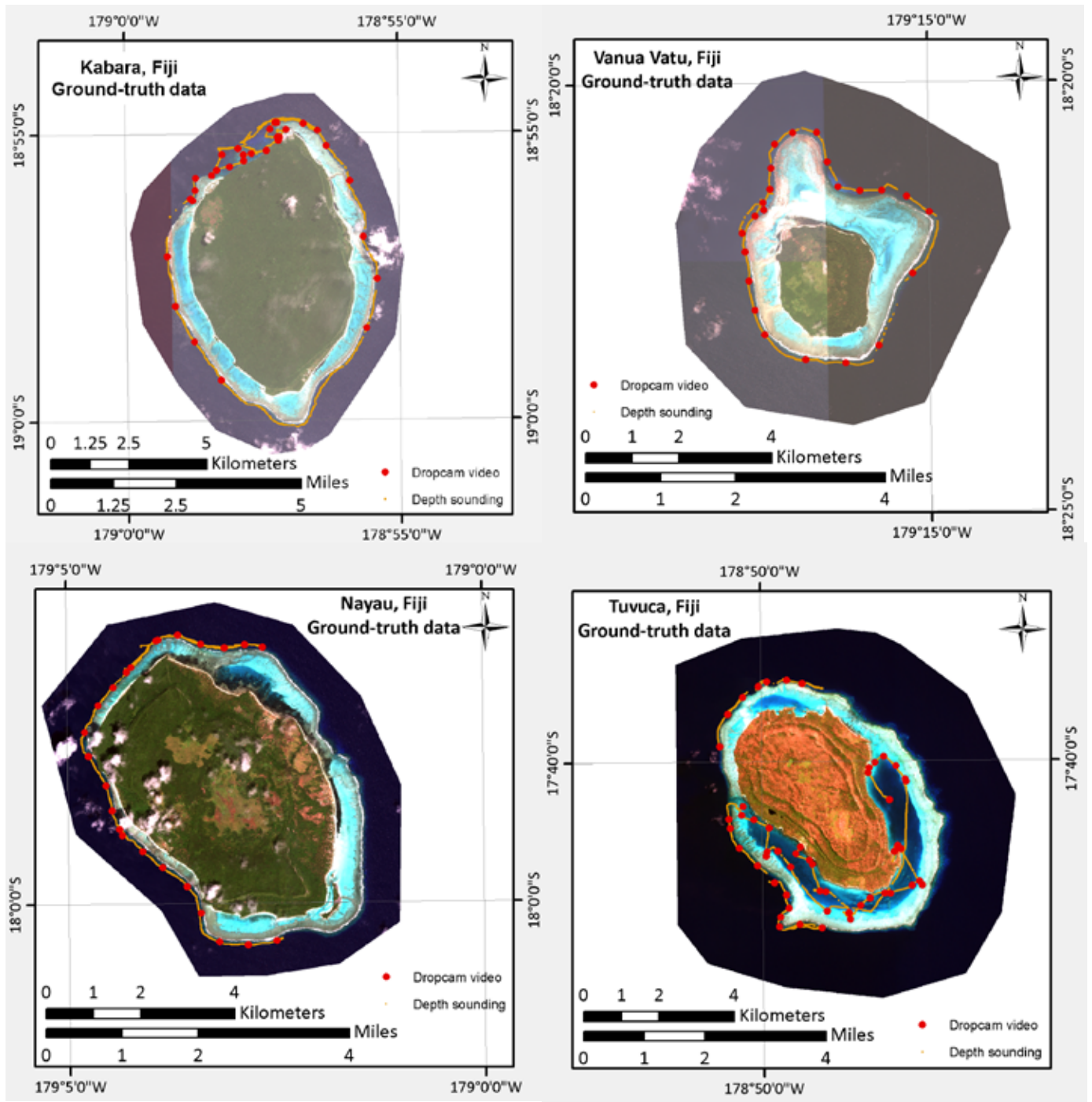


Fig.6. Groundtruthing track (orange line) and location of drop camera videos (red dots) for Kabara, Vanua Vatu, Nayau, and Tuvuca, Fiji.

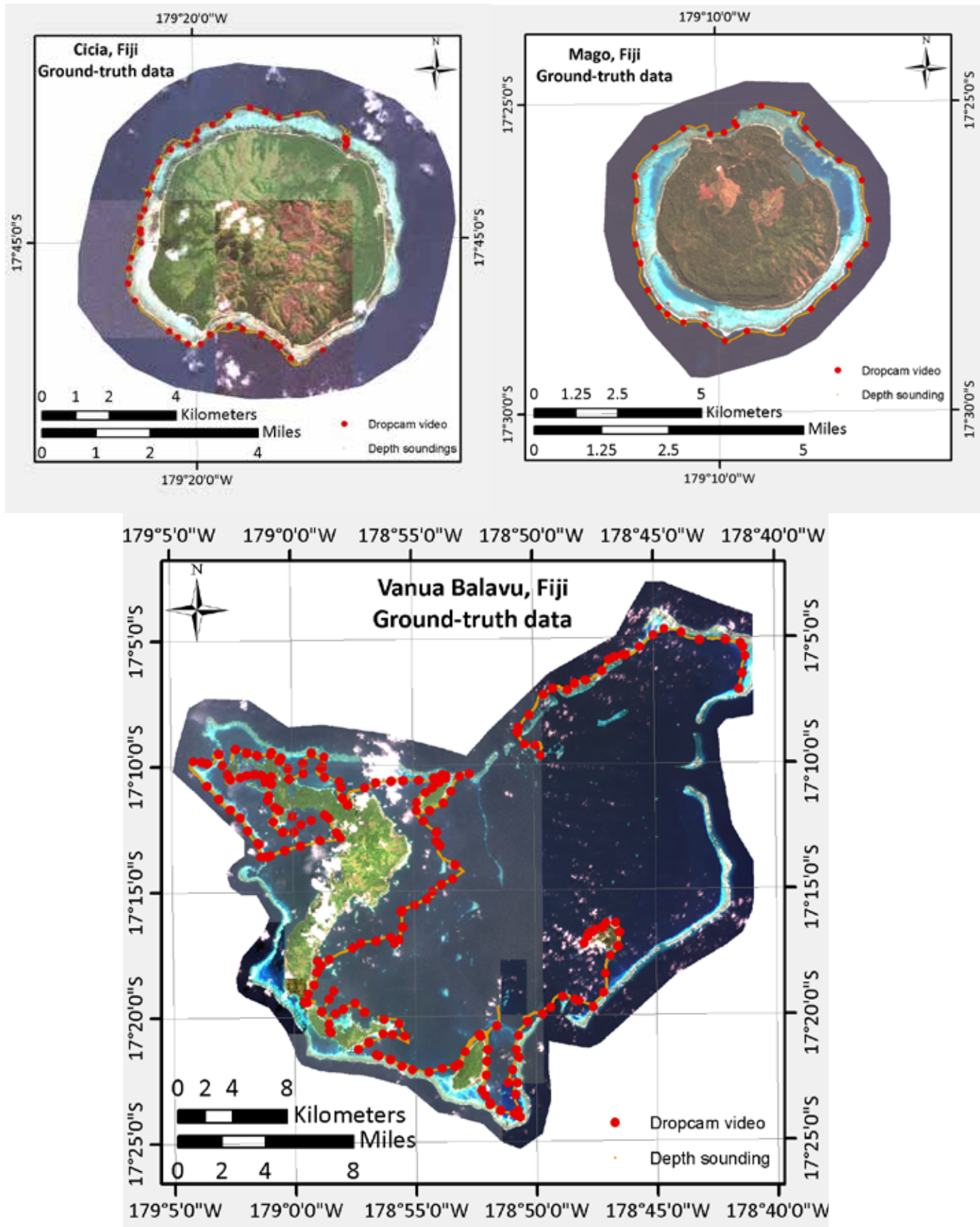


Fig. 7. Groundtruthing track (orange line) and location of drop camera videos (red dots) for Cicia, Mago and Vanua Balavu, Fiji.

Habitat Key – Lau Province, Fiji

A description of all the marine and terrestrial habitat types identified and characterized in Lau Province, Fiji are provided along with representative photographs of each of these habitats taken during the mission. A total of 21 shallow marine habitats, deep (unmapped) fore reef and lagoonal environments and four terrestrial habitats were identified. The total area encompassed by each of the habitats is shown in Table 5.

Habitat Class	Total Area (km ²)
Shallow fore reef terrace	20.9
Shallow fore reef slope	22.6
Deep fore reef slope	44.2
Fore reef sand flats	7.9
Coralline algal ridge	14.0
Back reef rubble dominated	47.8
Back reef sediment dominated	10.3
Back reef pavement	104.3
Back reef coral framework	6.7
Back reef coral bommies	1.1
Lagoonal sediment apron sediment dominated	10.7
Lagoonal floor barren	695.9
Lagoonal floor macroalgae on sediment	3.8
Lagoonal pinnacle reefs massive coral dominated	15.8
Lagoonal patch reefs	2.4
Lagoonal floor coral bommies	0.8
Lagoonal fringing reefs	35.0
Lagoonal bommie field	1.9
Dense seagrass meadows	13.3
Dense macroalgae on sediment	1.3
Deep lagoonal water	134.4
Mangroves	3.8
Mud flats	0.3
Terrestrial vegetation	308.9
Beach sand	2.6
Inland waters	0.2
Urban	0.6
Unvegetated Terrestrial	24.3
Deep ocean water	2029.7
TOTAL	3565.6

Table 5. Total area mapped for each habitat class for 11 islands/atolls within Lau Province, Fiji.

Terrestrial Habitats

For terrestrial (coastal/nearshore) habitats, the following classes were discriminated: Terrestrial-Vegetated, Terrestrial - Unvegetated, Beach Sand and Urban (human-made structures).



Fig. 8. Examples of coastal habitats. Terrestrial-vegetated, Mago (top left). Terrestrial-Unvegetated, Cicia (top right). A mix of Terrestrial -Unvegetated and Vegetated habitats at Fulaga (center left). A hillside with dense vegetation, Moala (center right). Beach Sand, Nayau (lower left). Urban, Vanua Balavu (lower right).

Shallow Fore Reef Terrace

BIOLOGY: Benthic community structure varies due to the local disturbance regime and resilience. Spur tops may host a complex community of short, stout branched acroporids and pocilloporids, small encrusting and submassive faviids; on the deeper ends, *Montipora*, *Astreopora* and other corals take on a plating morphology. Live coral cover in these cases may be very high (60% to 80%). In other cases, the spur top is a scoured hardground with isolated scleractinians and abundant calcareous coralline algae (CCA). Coral density on spur walls ranges from sparse to a dense assemblage of plating and encrusting colonies. Similarly, grooves may be uncolonized coral rock, host CCA accumulations, or support dense coral assemblages.

HYDROGRAPHY: Oceanic and wind-driven waves strongly influence this habitat. Large waves begin to break over spurs, and may generate currents off the reef as water circulation is channeled down the grooves.

SEDIMENTOLOGY: Little sediment accumulates on spurs. Coral rubble and sand accumulates in the grooves. Fine grained sediments are exported by wave generated water motion and currents.

TOPOGRAPHY: Spur tops and grooves have low rugosity. Grooves are highly variable in width and height with some examples being narrow (<1 m) and deep (>2 m) and others being wide (>2 m) and shallow (<50 cm). Grooves may be shallow or deep near the reef crest and similarly shallow or deep near their termination on the reef slope.

DEPTH RANGE: Emergent to 8 m.

*Fig. 9. Shallow fore reef terrace communities. Coral community is dominated by digitate and tabular acroporids, Kabara, 3 m depth (top). Large *Diploastrea* colony with table acroporids, foliose *Turbinaria* and other species, Cicia, 8 m depth (center). Shallow fore reef terrace at Vanua Balavu, 5 m depth (bottom).*



Shallow Fore Reef Slope

BIOLOGY: The benthic community is highly diverse and coral-dominated often with a substantial macroalgae component. Environment and disturbance regime govern benthic community structure. Scleractinian diversity is very high and often dominated by acroporids, pocilloporids, poritids, and faviids. The edge of the shallow slope is often dominated by large *Pocillopora eydouxi* colonies, thick-branched and elkhorn-like *Acropora* (e.g. *A. cuneata*, *A. palifera*), and unusually large *Favia stelligera* and/or *Pavona clavus* stands. Common macroalgae include the genera *Halimeda*, *Turbinaria*, and *Sargassum*.

HYDROGRAPHY: Water motion is driven by wind and oceanic waves and alongshore currents. Wave influence on the zone abates as depth increases.

SEDIMENTOLOGY: Coral dominated. When present, calcareous macroalgae contribute substantial amounts of sediment. Terraces on the reef slope may capture patches of sand and coral rubble.

TOPOGRAPHY: Gently sloping with some scattered mounds and spurs of coral framework running perpendicular to the reef crest. At the transition from this zone to the “Fore reef deep slope” (near 15 m), a build-up of corals often forming a ridge or mound perforated by shore-perpendicular channels often occurs and runs parallel to the reef crest.

DEPTH RANGE: 8 m to 15 m.

Fig. 10. Shallow fore reef slope at 8-12 m depth, Reefs often had high cover of small pocilloporids, acroporids, Montipora and faviids such as that seen at Kabara (top), Cicia (center), and Nayau (bottom). In most cases the branching corals were fairly small (>1 m) and generally less than 10 years old which suggests they have recruited and regrown since the 2002 bleaching event.



Deep Fore Reef Slope

BIOLOGY: A diverse benthic community with the dominant scleractinian community differing by location. Shallower areas may be dominated by branching acroporids and (sub)massive poritids with encrusting and plating corals also present. Small mounds of hosting diverse mixed coral assemblages may be present. At greater depths, colonies take on plating morphologies and may form a series of shingle-like structures. Large monospecific stands of foliaceous and plating *Pachyseris*, *Leptoseris*, *Montipora* and *Merulina* occur on more gradual slopes while encrusting and laminar morphologies become more common when the slope becomes close to vertical. Prominent macroalgae include *Halimeda*, *Caulerpa*, and *Dictyota*.

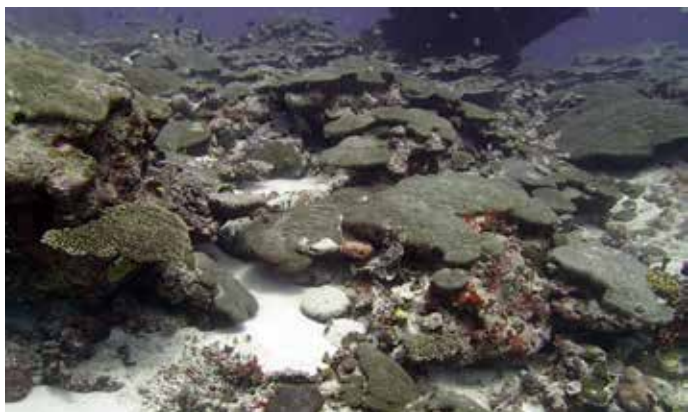
HYDROGRAPHY: Deep fore reef slope lies below the fair-weather wave base but is episodically exposed to high energy during severe storm events and tsunamis. Alongshore currents may be present. Differences in storms, surge, wind and wave exposure influence the benthic community.

SEDIMENTOLOGY: Coral dominated, however *Halimeda* algae may also contribute substantial volumes of sediment. Terraces on the reef slope may capture patches of sand and coral rubble. Water motion tends to be lower than the shallow reef slope resulting in greater rates of deposition.

TOPOGRAPHY: This zone occurs where the shallow fore reef slope begins to steepen (typically 12 – 15 m) and continues into the ocean depths.

DEPTH RANGE: 15 m to 40 m.

Fig. 11. Deeper fore reef off slope off Vanua Vatu (top) and Vanua Balavu (bottom). The deeper fore reef often had small pinnacles surrounded by sand and rubble such as that seen in Vanua Balavu. Porites lobata colonies exhibited a plating morphology on the deeper fore reef environments such as that seen here from Totoya (bottom).



Fore Reef Sand Flats

BIOLOGY: Interstitial fauna account for the majority of biomass. Benthic cover is usually very low because the substrate is mobile and largely unconsolidated thus inhibiting colonization by benthic organisms. In some cases, large *Porites lobata* colonies and tabular acroporids occur within the sand flats, especially in deeper areas (30-50 m). Macroalgae become more common with depth due to the abatement of wave energy with increasing depth.

HYDROGRAPHY: Water motion is driven by wind and oceanic waves and alongshore currents. Wave influence on the zone abates as depth increases.

SEDIMENTOLOGY: Coral dominated. When present, calcareous macroalgae contributes.

TOPOGRAPHY: Generally low relief. Large areas may have higher relief as they follow the antecedent topography's slope into deeper waters.

DEPTH RANGE: 10 m to 40 m.

*Fig. 12. The deeper fore reef habitats at the base of the reef were generally low relief sand flats, mostly uncolonized. Often there were sparse assemblages of cyanobacteria and small patches of macroalgae, along with isolated larger corals. Examples of deeper sand flats from Vanua Balavu are shown in the top and center. A coral trout, *Plectropomus leopardus* is swimming across the sand flat (top). Closer to the reef system, sandy areas typically had more rubble (bottom).*



Coralline Algal Ridge (Reef Crest)

BIOLOGY: Crustose coralline algae (CCA) accounts for the majority of biotic cover. Coral growth is limited due to wave action and episodic aerial exposure and is limited to species tolerant of the chronic disturbances.

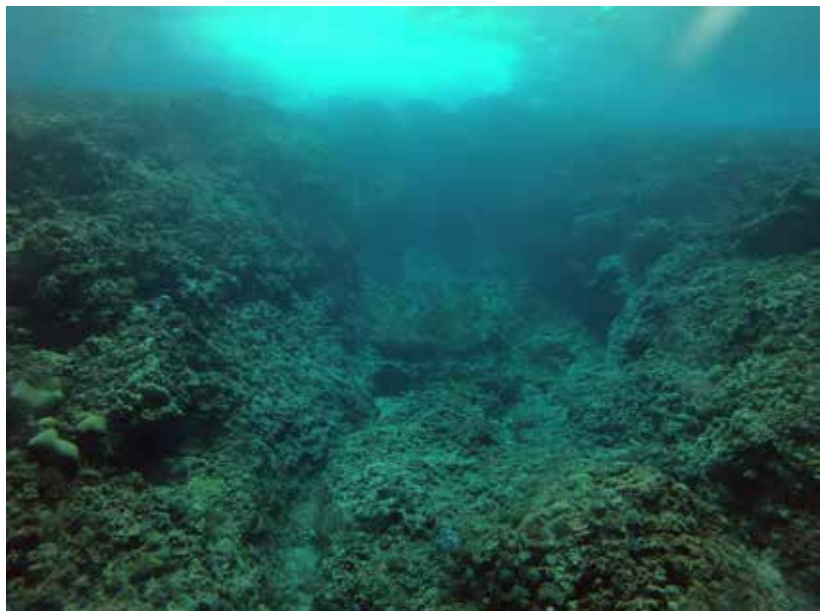
HYDROGRAPHY: A high energy environment exposed to large (often > 1 m) waves on a regular basis. The reef crest may be aerially exposed during low tides. The continuity of the algal ridge is broken at irregular intervals by shore-perpendicular grooves cut by waves and currents. These channels provide paths of tidal water exchange between the open ocean and the lagoon or back reef.

SEDIMENTOLOGY: The reef crest is an important source of carbonate detritus. Continuous wave energy generates and transports sediment, shoreward, or down-slope.

TOPOGRAPHY: The reef crest's ocean-face is a cement-like barrier with irregularly spaced ridges and grooves. Ridges are exposed at low-tide and between waves, and grooves allow for seaward transport of water.

DEPTH RANGE: +2 m to -1 m.

Fig. 13. Reef crest habitats in Lau Province. The reef crest at Fulaga had small encrusting corals and coralline algae (top). Above water view of the emergent reef crest off the west side of Vanua Balavu at low tide (center). Windward reef crest off Vanua Vatu (bottom). There are prominent channels that transport sediments to the fore reef.



Back Reef – Rubble Dominated

BIOLOGY: Biotic cover is low in this zone because the substrate is mobile and largely unconsolidated. Algae dominate, and isolated coral colonies are present, but at low density and of a small size. Calcareous red algae and turf algae are prominent, and rhodoliths may accumulate. Coral colonies are typically encrusting, submassive and free living (e.g. *Psammocora* and fungiids) and hardier than those found in other zones.

HYDROGRAPHY: Water motion in this zone is driven by waves, tides, and wind. Breaking waves transport water and sediment over the reef crest into this zone. Tidal pools form during low tide when water is trapped in topographic lows.

SEDIMENTOLOGY: Sediment is composed primarily of accumulated dead coral fragments ("rubble") of varying sizes. Biotic and chemical processes cement the rubble in place over time. The degree of cementation varies.

TOPOGRAPHY: The back reef is the shoreward extension of the reef crest. It is fully aggraded to sea level and has low-relief. Large boulders of reef rock moved up over the reef crest by severe storms and tsunamis may be present and mix with outcrops of stranded fossil reef deposited during geological periods of higher sea level.

DEPTH RANGE: Emergent to 1 m.

Fig. 14. Shallow back reef environments with accumulations of rubble. These habitats generally have very few live corals. Unconsolidated, poorly sorted rubble at Matuka (top). Back reef environment in Vanua Balavu (center). Shallow back reef rubble field in Totoya (bottom).



Back Reef – Sediment Dominated

BIOLOGY: Biotic cover is low due to high disturbance rates. Sparse turf and macroalgae may stabilize the accumulated sediments.

HYDROGRAPHY: Water motion in this zone is driven by waves, tides, and wind. Breaking waves transport water and sediment over the reef crest into this zone. Tidal pools form during low tide when water is trapped in topographic lows.

SEDIMENTOLOGY: Where wave action is more moderate, the back reef tends to be less rugose and composed of compacted carbonate sands deposited periodically by large storm waves.

TOPOGRAPHY: The back reef is the shoreward extension of the reef crest. It is fully aggraded to sea level and has low-relief. Large boulders of reef rock moved up over the reef crest by severe storms and tsunamis may be present and mix with outcrops of stranded fossil reef deposited during geological periods of higher sea level.

DEPTH RANGE: 0 m to 1 m.

Fig. 15. Sediment-dominated shallow back reef environments were relatively uncommon except at Totoya, Tuvuca, Cicia and Vanua Balavu. These soft bottom communities had sparse cover of benthic organisms, consisting of small free living corals and occasional patches of macroalgae, cyanobacteria or turf algae. Algae were more common in calm, protected areas. Caulastrea colonies were found in sandy areas in the back reef at Tuvuca (top). Small Porites lobata colonies were found close to the reef crest. Some shallow back reef environments had high numbers of Holothuria edulis sea cucumbers (bottom).



Back Reef – Pavement

BIOLOGY: A planation hardground colonized by turf algae and isolated coral colonies (<1% live cover). Scleractinians found in this habitat include stout branched *Pocillopora*, *Stylophora*, and submassive *Platygyra*, *Favites*, and *Favia*.

HYDROGRAPHY: Water motion in this zone is driven by waves, tides, and wind. Breaking waves transport water and sediment over the reef crest into this zone. Tidal pools form during low tide when water is trapped in topographic lows.

SEDIMENTOLOGY: Very little sediment, except in depressions between pavement. In protected areas the surface of the pavement may have a fine layer of sediment or turf with sediment.

TOPOGRAPHY: The back reef is the shoreward extension of the reef crest. It is fully aggraded to sea level and has low-relief. Large boulders of reef rock moved up over the reef crest by severe storms and tsunamis may be present and mix with outcrops of stranded fossil reef deposited during geological periods of higher sea level.

DEPTH RANGE: 0 m to 1 m.

Fig. 16. The shallow Back Reef - Pavement community at Vanua Balavu had turf algae and small corals, with scattered sand channels (top). At Kabara, the extensive Back Reef - Pavement habitat was colonized by turf algae (center). In some areas off Matuka, crustose coralline algae and turf were common colonizers and sea urchins had bioeroded small holes into the pavement (bottom).



Back Reef – Coral Framework

BIOLOGY: Coral framework with variable benthic community composition. The coral community may be composed of dense acroporid thickets or massive poritids and agariciids forming microatolls whose tops are colonized by submassive, branching, and foliose coral colonies.

HYDROGRAPHY: The back reef is a low energy environment as wave energy dissipates across the reef crest and back reef. Tides and wind are the dominant drivers of water motion.

SEDIMENTOLOGY: The back-reef has little sedimentation. Topographic depressions serve as collection points for coarse carbonate sands, *Halimeda* plates, and coral rubble.

TOPOGRAPHY: The back reef lies shoreward of the back reef and displays varying degrees of reef framework aggradation depending on the width of the back reef and hydrodynamics.

DEPTH RANGE: 0 m to 2 m.

Fig. 17. The shallow back reef community often had high cover of small massive corals including Porites, faviids, digitate acroporids and other species in exposed areas (top). More sediment tolerant corals such as Pavona clavus were seen in protected environments (bottom).



Back Reef – Coral Bommies

BIOLOGY: Isolated or composite coral heads with a plan-form area <200 sq. m growing to near sea level. The coral community may be composed of dense acroporid thickets or massive poritids and agariciids forming microatolls whose tops are colonized by submassive, branching, and foliose coral colonies.

HYDROGRAPHY: The back reef is a low energy environment as wave energy dissipates across the reef crest and back reef. Tides and wind are the dominant drivers of water motion.

SEDIMENTOLOGY: The back-reef has little sedimentation. Topographic depressions serve as collection points for coarse carbonate sands, *Halimeda* plates, and coral rubble.

TOPOGRAPHY: The back reef lies shoreward of the back reef and displays varying degrees of reef framework aggradation depending on the width of the back reef and hydrodynamics.

DEPTH RANGE: 1 m to 2 m.

*Fig. 18. Examples of Back Reef-Coral Bommies. Shallow back reef environments in sandy areas often had small coral heads scattered over the substrate. These typically consisted of small clumps of staghorn corals and table acroporids (top), colonies of *Porites lobata* (center) and various faviid corals (bottom).*



Dense Seagrass Meadows

BIOLOGY: An expanse of dense seagrass in sediment.

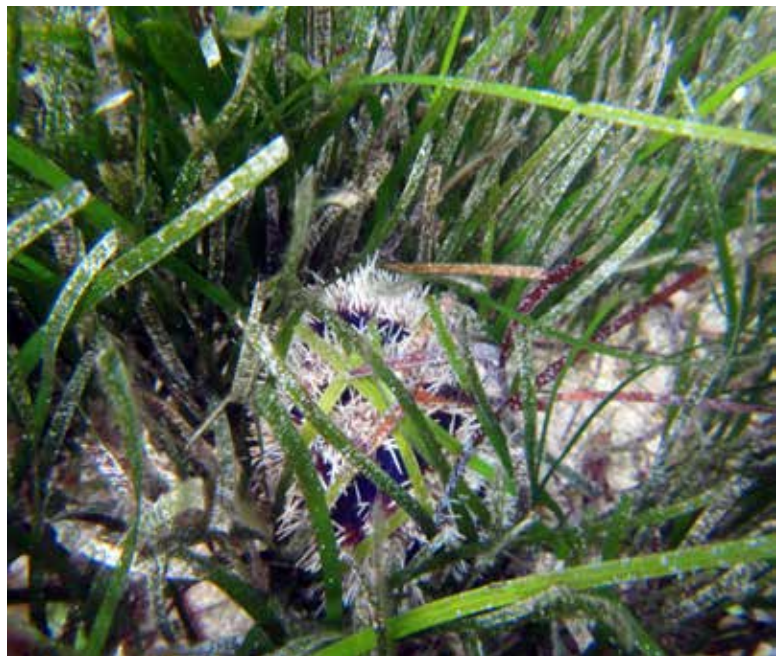
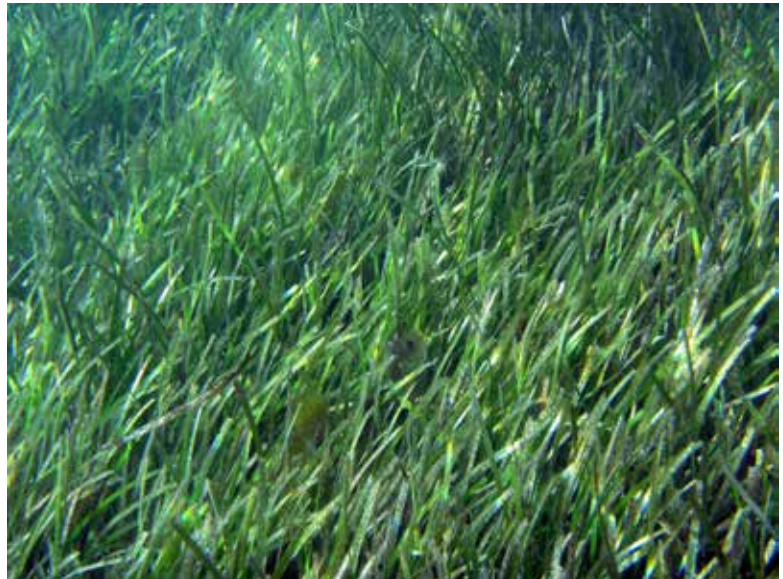
HYDROGRAPHY: Restricted water motion with little influence from waves due to sheltering by reefs and islands. Water restriction leads to long resident times for water masses, and the suspension of detritus creates moderate turbidity.

SEDIMENTOLOGY: Predominantly fine-grained sediments trapped by the seagrasses. Most sediments are biogenic, resulting from the breakdown of the marine vegetation and associated calcareous epibionts.

TOPOGRAPHY: Low relief.

DEPTH RANGE: From 1 m to 15 m.

*Fig. 19. Dense seagrass beds were seen in intertidal and shallow subtidal lagoonal habitats off Matuka, Tuvuca, Kabara and Cicia, with sparse seagrass coverage in deeper habitats. There are five species and one subspecies reported from Fiji. *Syringodium isoetifolium* was found from 1-6 m depth (top right). *Thalassia hemprichii* was the most common species (center right). Shallow areas also had a high abundance of *Halophila ovalis* and *Halodule uninervis*, but beds were not as dense as *Thalassia* beds. Seagrass beds are extremely productive, efficient recyclers of nutrients, critical nursery areas, and they support a high biomass of consumers including sea urchins (bottom right). *Halophila teneded* to occur in deeper lagoonal areas, rarely forming dense assemblages (bottom left).*



Lagoon – Sediment Apron

BIOLOGY: The lagoonal sediment apron supports a shallow and narrow terrace of unconsolidated sand, lagoonward of the back reef that spans a range of depths from the intertidal and shallow subtidal to 5 m depth.

HYDROGRAPHY: A low energy environment dominated by tides and wind-driven waves. Oceanic influence is minimal because long-swell waves break on the reef crest, and their energy is greatly attenuated before reaching this zone.

SEDIMENTOLOGY: The active face of lagoon in-filling by an accumulation of well-sorted sand-sized carbonate detritus. Most sediments in this zone are transported from the reef crest and fore reef during storms and their size distribution is largely the result of differential transport by high bottom-water velocities during those events. Sandy islands or shingle banks may develop in between the back reef and deep lagoon.

TOPOGRAPHY: Generally low relief.

DEPTH RANGE: 1 m to 5 m.

*Fig. 20. Unconsolidated sand and fine silt was common in shallow lagoonal areas adjacent to the shoreline (top). In areas with higher wave action, sand waves and small pieces of rubble were observed (center). Sandy lagoonal environments often contained erect coralline algae such as *Halimeda* and supported a number of small burrowing fish such as the tile fish *Malacanthus brevirostris* (bottom).*



Lagoonal Floor – Barren

BIOLOGY: Few benthic organisms with exception of motile invertebrates. Isolated colonies of sediment-tolerant and low-light adapted *Acropora* colonies. Sparse community of fleshy macroalgae may grow in the sediment.

HYDROGRAPHY: Restricted water motion with little influence from waves due to sheltering by the reef. Oceanic wave energy dissipates before reaching this zone, and the orbitals of wind generated waves attenuate before reaching the seafloor. Tidal cycles flush the lagoon, however restricted flow leads to long resident times for water masses and high turbidity.

SEDIMENTOLOGY: Sediment entirely skeletal in origin and tends to have a very poorly sorted, coral gravel-rich base which grades up into poorly sorted muddy sands with infaunal molluscan gravel. Intense bioturbation by callianassid shrimps.

TOPOGRAPHY: Generally has low relief. Local highs created by coral colonies.

DEPTH RANGE: 15 m to 35 m.

Fig. 21. Deeper lagoonal environments were dominated by sand with small pieces of rubble and scattered, sediment tolerant corals. Often the sediment was colonized by cyanobacteria and motile invertebrates were seen, such as the helmet snail (top right) and sea cucumber (bottom left). Branching acroporids were common in many areas (bottom right).



Lagoonal Floor – Macroalgae on Sediment

BIOLOGY: An expanse of dense macroalgae in which thalli are interspersed by unconsolidated sediment.

HYDROGRAPHY: Restricted water motion with little influence from waves due to sheltering by the reef. Oceanic wave energy dissipates before reaching this zone, and the orbitals of wind generated waves attenuate before reaching the seafloor. Tidal cycles flush the lagoon, however restricted flow leads to long resident times for water masses and high turbidity.

SEDIMENTOLOGY: Predominantly fine-grained carbonate sediments trapped by the macroalgae thalli.

TOPOGRAPHY: Low relief.

DEPTH RANGE: 2 m to 30 m.

Fig. 22. Soft bottom habitats in the lagoon often had dense assemblages of macroalgae, including a number of taxa of erect coralline algae. Common taxa included Halimeda (top). Turf algae was also found in lagoonal soft bottom areas, especially in areas where the sand was mixed with coral rubble (center). In some areas the sediment was stabilized by the rhizomes of Caulerpa (bottom).



Lagoonal Pinnacle Reefs – Massive Coral Dominated

BIOLOGY: Coral framework with a plan-form area >400 sq. m aggraded from the lagoon floor to the water surface with steep, sediment dominated flanks. Coral community composition is determined by the development stage of each patch. In the early stage, the coral community is dominated by massive species (e.g., *Porites* sp.) which build to form a pinnacle. Accommodation space becomes limited as the pinnacle aggrades to sea level, leading to the formation of microatolls from the massive colonies. Branching, submassive, and encrusting coral species colonize the tops of the microatolls. Over time, skeletal debris and sediment collect in the center of the microatoll leading to the formation of cays. A relative sea level drop may expose the top of the pinnacle to form a lagoonal island, which, with time, can then be colonized by vegetation. Simultaneously, the coral community may back-step to form a Lagoonal Island Fringing Reef.

HYDROGRAPHY: Tides and wind-driven waves affect the benthic community and spatial distribution of sediments. Water motion in the lagoon is restricted which can lead to highly turbid waters. Water in the center of the patches is restricted further causing water masses to warm from solar radiation.

SEDIMENTOLOGY: The tops of the pinnacles are coral-dominated and characterized by reef derived debris in the sand and rubble size fractions. Sediments become muddier with increasing depth.

TOPOGRAPHY: Isolated coral pinnacles rise from the lagoon-floor into the zone of maximum carbonate production. Pinnacles are usually circular in shape, but may elongate along hydrodynamic gradients. Pinnacle-tops flatten as they near sea level

DEPTH RANGE: 1 m to 30 m.



*Fig. 23. Lagoonal pinnacle reef in Vanua Balavu, 15 m depth (top). Large *Diploastrea* colonies often colonized the sides and tops of the pinnacle reefs (center). Many of the lagoonal pinnacle reefs had flourishing acroporid communities on their top surfaces, especially in shallow water (bottom).*

Lagoonal Patch Reefs

BIOLOGY: Coral framework with a plan-form area >100 sq. m and <400 sq. m aggraded from the lagoon floor to sea level. Coral community composition is determined by the framework's development stage. The coral community may be composed of dense acroporid thickets, massive poritids and agariciids forming microatolls whose tops are colonized by submassive, branching, and foliose coral colonies, or a mixture of the two.

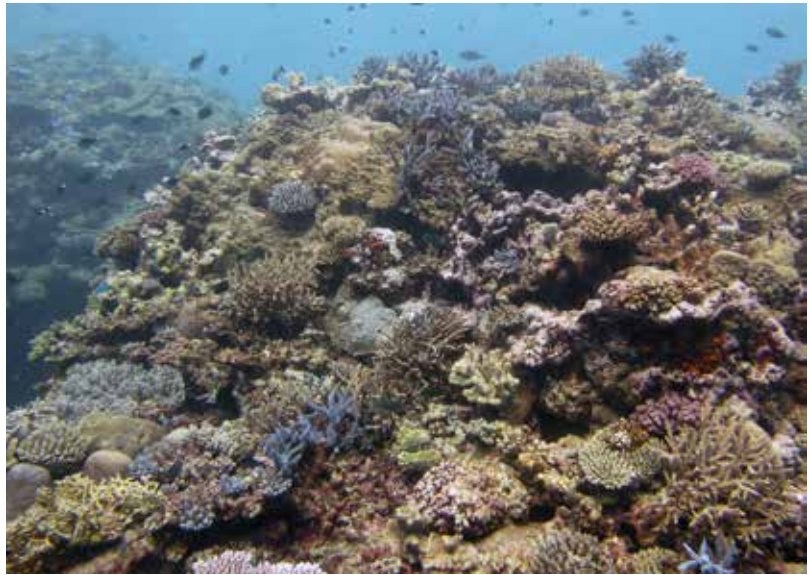
HYDROGRAPHY: Tides and wind-driven waves affect the benthic community and spatial distribution of sediments. Water motion in the lagoon is restricted which can lead to highly turbid waters. Water in the center of the patches is restricted further causing water masses to warm from solar radiation.

SEDIMENTOLOGY: Upper reaches of the patches are coral dominated and characterized by reef derived debris in the sand and rubble size fractions.

TOPOGRAPHY: Coral framework rises from the lagoon floor. Patches are usually circular in shape, but may elongate along hydrodynamic gradients, and their tops flatten as they near sea level.

DEPTH RANGE: 2 m to 15 m.

Fig. 24. Vanua Balavu had an extensive lagoonal habitat with numerous patch reefs such as this reef (top). The top and sides of a large patch reef at Totoya colonized primarily by staghorn and digitate acroporids and small massive corals. An unusual field of branching Galaxea at the eastern end of Vanua Balavu lagoon.



Lagoonal Floor – Coral Bommies

BIOLOGY: Isolated or composite coral heads with a plan-form area <100 sq. m growing up from the lagoon floor to near sea level. The coral community may be composed of dense acroporid thickets or massive poritids and agariciids forming microatolls whose tops are colonized by submassive, branching, and foliose coral colonies.

HYDROGRAPHY: Restricted water motion with little influence from waves due to sheltering by the reef. Oceanic wave energy dissipates before reaching this zone, and the orbitals of wind generated waves attenuate before reaching the seafloor. Tidal cycles flush the lagoon, however restricted flow leads to long resident times for water masses and high turbidity.

SEDIMENTOLOGY: Sediment entirely skeletal in origin and tends to have a very poorly sorted, coral gravel-rich base which grades up into poorly sorted muddy sands with infaunal molluscan gravel. Intense bioturbation by callianassid shrimps.

TOPOGRAPHY: Generally has low relief. Local highs created by coral colonies.

DEPTH RANGE: 2 m to 10 m.

Fig. 25. Scattered coral bommies were seen throughout lagoonal environments in shallow water. These included mixed species assemblages with Porites, Favia, Lobophyllia, Acropora dominating. In most cases they were only a few meters diameter. A small coral bommie with Acropora, Lobophyllia and Porites at Vanua Balavu, 3 m depth (top). Coral bommies constructed of large massive Porites lobata colonies off Kabara at 12 m depth (center). Larger colonies of Lobophyllia in the lagoon off Totoya at 5 m depth (bottom).



Lagoonal Floor – Bommie Field

BIOLOGY: A high density of small patch reefs, so called “bommies,” grouped together, and regularly spaced on the lagoon floor. These reefs, tend to be circular with plan-form areas always <100 sq. m, but typically <10 sq. m and do not reach sea level. While the individual bommies are constructed from massive corals, such as poritids and agariciids, portions of the tops may be inhabited by dense acroporid thickets.

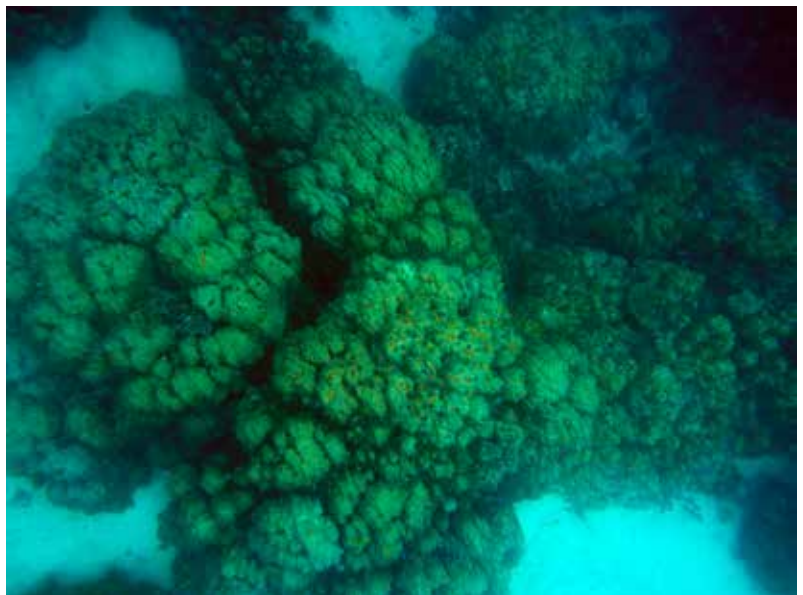
HYDROGRAPHY: Tides and wind-driven waves affect the benthic community and spatial distribution of sediments. Water motion in the lagoon is restricted which can lead to highly turbid waters. Water in the center of the patches is restricted further causing water masses to warm from solar radiation.

SEDIMENTOLOGY: The tops of the pinnacles are coral-dominated and characterized by reef derived debris in the sand and rubble size fractions. Sediments become muddier with increasing depth.

TOPOGRAPHY: The bommies rise a few meters from the lagoon floor and are spaced less than 10 m from one another with channels of sand or fine grained lagoon floor mud, located between them as described in “Lagoon floor - barren”.

DEPTH RANGE: 5 m to 35 m.

*Fig. 26. Examples of bommie fields within lagoonal habitats constructed primarily of *Porites lobata* colonies. *Porites* bommies at 5 m depth in Vanua Balavu (top). Coral bommies at Moala. A deeper lagoon floor coral bommie field dominated by very large colonies of *Porites* at Vanua Balavu, 20 m depth (bottom).*



Lagoonal Fringing Reefs

BIOLOGY: Coral reef frameworks fringing islands. Benthic community composition is similar to that of lagoonal patch reefs.

HYDROGRAPHY: Tides and wind-driven waves affect the benthic community and spatial distribution of sediments. Water motion in the lagoon is restricted which can lead to highly turbid waters. Water in the center of the patches is restricted further causing water masses to warm from solar radiation.

SEDIMENTOLOGY: Similar to that of lagoonal pinnacle reefs. Lagoonal islands may be circular to elongate in shape and develop through two mechanisms. Lagoonal pinnacle reefs may fill the accommodation space, or the islands may be remnants of fossil reef stranded from episodes of higher-than-present sea level. The most mature lagoonal islands possess a water table and are heavily vegetated.

TOPOGRAPHY: Moderate to steeply sloping reef framework rising from the lagoon-floor into the zone of maximum carbonate production. Usually elongate and narrow, extending along the coastline. The reef top flattens as it approaches sea level.

DEPTH RANGE: 1 m to 30 m.

Fig. 27. Fringing reefs were seen along several of the islands examined in Lau Province. A shallow lagoonal fringing reef in Vanua Balavu exposed at low tide. Image by Stacy Jupiter (top). Close-up of the coral community of Vanua Balavu exposed at low tide (center). A shallow submerged fringing reef in Totoya (bottom).



Dense Macroalgae on Sediment

BIOLOGY: An expanse of dense macroalgae in which thalli are interspersed by unconsolidated sediment.

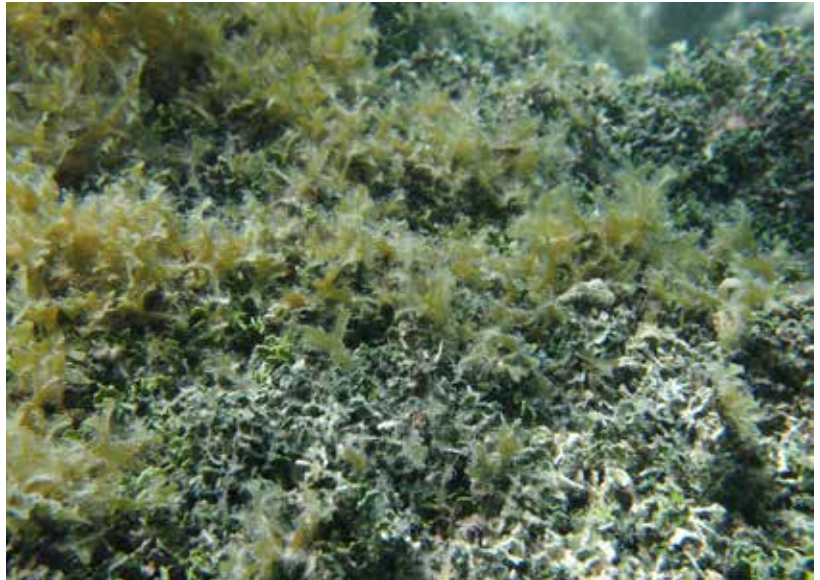
HYDROGRAPHY: Restricted water motion with little influence from waves due to sheltering by reefs and islands. Moderate turbidity results from the typically restricted hydrodynamic conditions and the suspension of detritus.

SEDIMENTOLOGY: Predominantly fine-grained carbonate sediments trapped by the macroalgae thalli.

TOPOGRAPHY: Low relief.

DEPTH RANGE: 1 m to 15 m.

Fig. 28. Soft bottom habitats occasionally had dense assemblages of Halimeda and Dictyota (top right). In some area a variety of red and brown macroalgae were common including Padina (center). A variety of green algae also occurred with erect red coralline algae and red macroalgae (bottom right). Large stands of Caulerpa were seen in many locations. Some species primarily occurred on vertical surfaces and at the base of the reef on rubble and dead coral such as Caulerpa racemosa (bottom left), while others carpeted sandy areas.



Mud Flats

BIOLOGY: Benthic community is primarily microbial. Benthic invertebrates are isolated or absent due to aerial exposure during low tides. Interstitial fauna are common. Sparse seagrass (e.g. *Halodule*) may be present.

HYDROGRAPHY: Restricted water motion with little influence from waves due to sheltering by reefs and islands. Tidal cycles flush the area. The water column has high turbidity due to suspension of detritus.

SEDIMENTOLOGY: Predominantly fine-grained sediments trapped by mangrove prop roots and seagrasses. Sediments are biogenic, resulting from the breakdown of the marine vegetation.

TOPOGRAPHY: Low relief. Seaward from shore, the substrate may slope to the base of the lagoon, with accumulations of debris (leaf litter, macroalgae) and scattered patches of rubble.

DEPTH RANGE: 0 to 2 m.

Fig. 29. Fine grained sediments and silt in a protected nearshore lagoonal habitat in Mago. Adjacent to the mud flat is a well developed mangrove community (top). An intertidal mudflat exposed at low tide. The flat is colonized by sparse seagrass (center). Submerged mudflat adjacent to a mangrove community at Vanua Balavu (bottom).



Mangroves

BIOLOGY: Coastal area dominated by mangroves. Diverse filter-feeding invertebrate communities including sponges, tunicates, bryozoans and certain cnidarians may occur among prop roots

HYDROGRAPHY: Restricted water motion with little influence from waves due to sheltering by reefs and islands. Tidal cycles flush the area. The water column has high turbidity due to suspension of detritus.

SEDIMENTOLOGY: Predominantly fine-grained sediments trapped by mangrove prop roots and seagrasses. Sediments are biogenic, resulting from the breakdown of the marine vegetation.

TOPOGRAPHY: Mud flats around the roots are low relief. Root system provides relief, substrate for invertebrate colonization, and habitat for fishes.

DEPTH RANGE: Dependent on tidal range. From 0 m to 2 m.

*Fig. 30. Fiji supports nine different species of mangroves and one hybrid. Three most abundant taxa include two red mangroves, *Rhizophora mangle* (red mangrove) and *Rhizophora stylosa* (spotted mangrove), and the black mangrove *Bruguiera gymnorhiza*. The red mangroves are found at the edge of the water, while black mangroves are generally found landward of these species. Close-up of a red mangrove community at Mago (top). Small mangrove along the shoreline at Moala (center). Red mangroves can be identified by their prominent prop roots which help anchor them into soft substrates (bottom).*



Deep Ocean Water

Submerged areas seaward of the fore reef that are too deep for observation via satellite. These deeper fore reef environments may be sand-bottom with scattered coral heads and rubble. In some locations deep water *Porites lobata* plates and tabular acroporids are dispersed over the bottom.

DEPTH RANGE: below 30 m

Deep Lagoonal Water

Submerged areas seaward of the fore reef that are too deep for observation via satellite. These are typically sand bottom, but may have some rock/coral rubble patches and often have high cover of macroalgae.

DEPTH RANGE: Below 30 m.

Fig. 31. Examples of deep water. The lagoon at Moala (top). Deep Ocean Water at Fulaga. The reef crest and lagoon are visible in the center of the image (center). Deep Ocean Water off Matuka. Image by Stacy Jupiter.



Habitat Maps

Totoya Island - Satellite Map

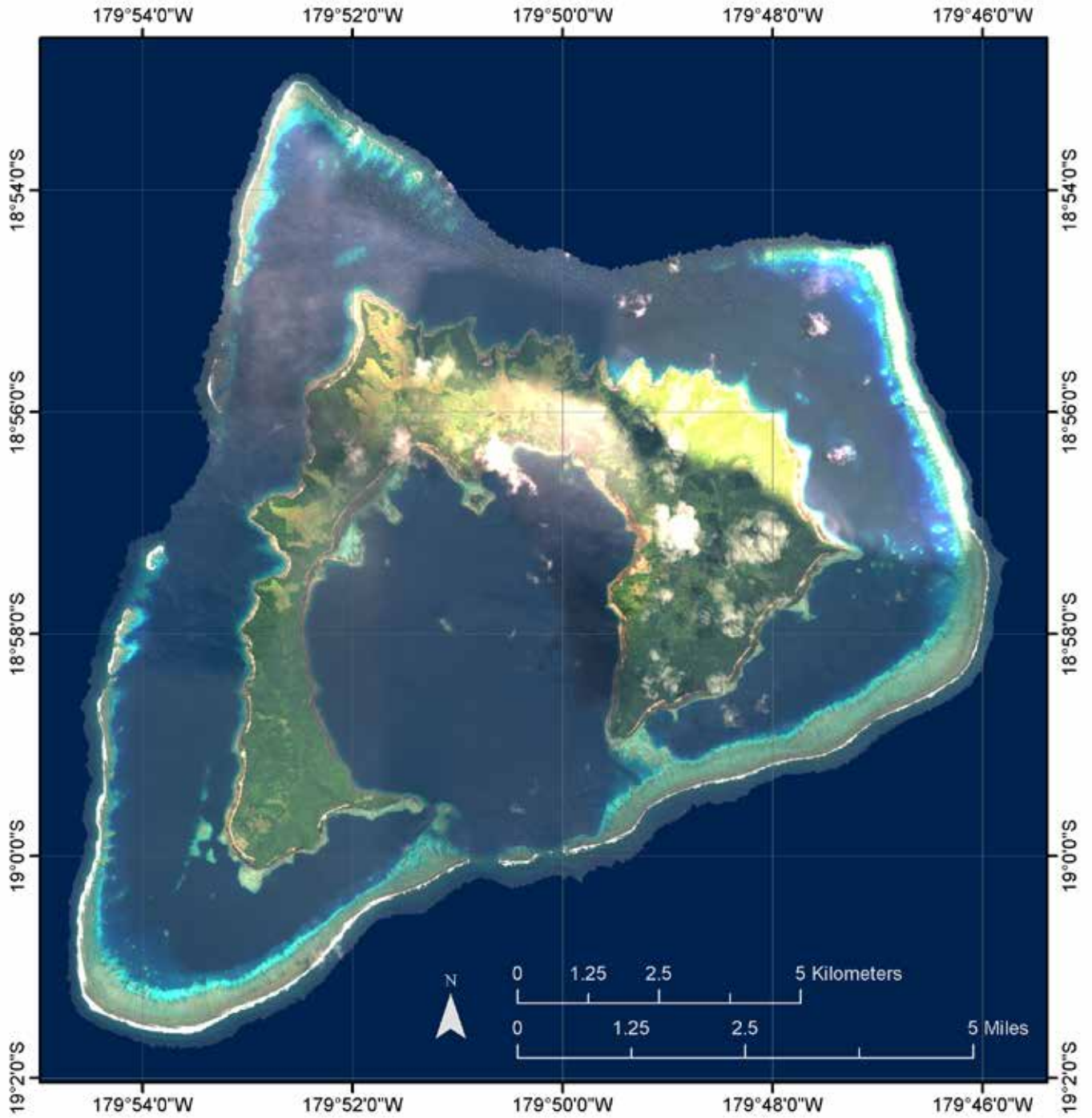
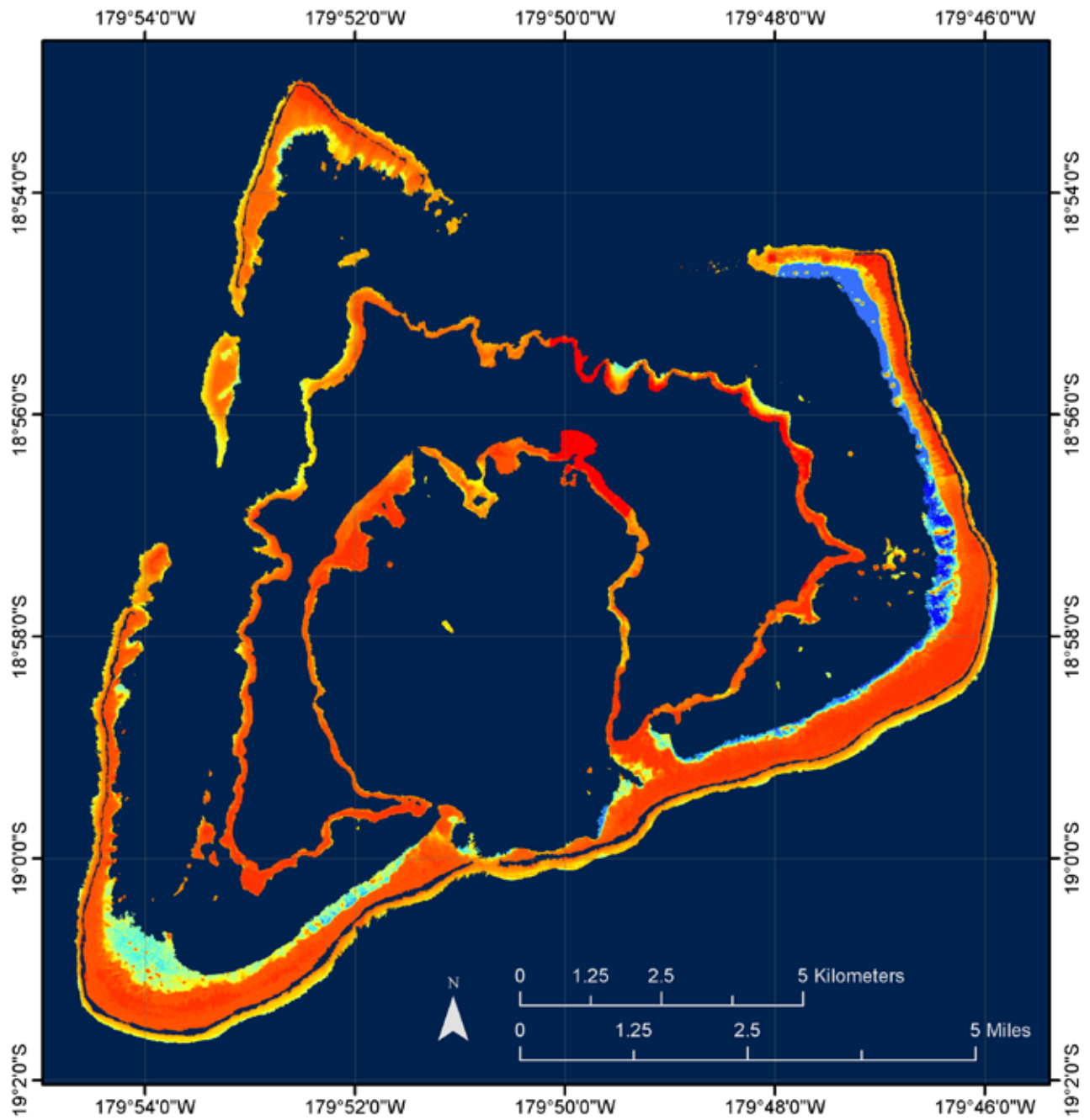


Fig. 32. WorldView-2 Satellite imagery for Totoya.

Totoya Island - Bathymetric Map

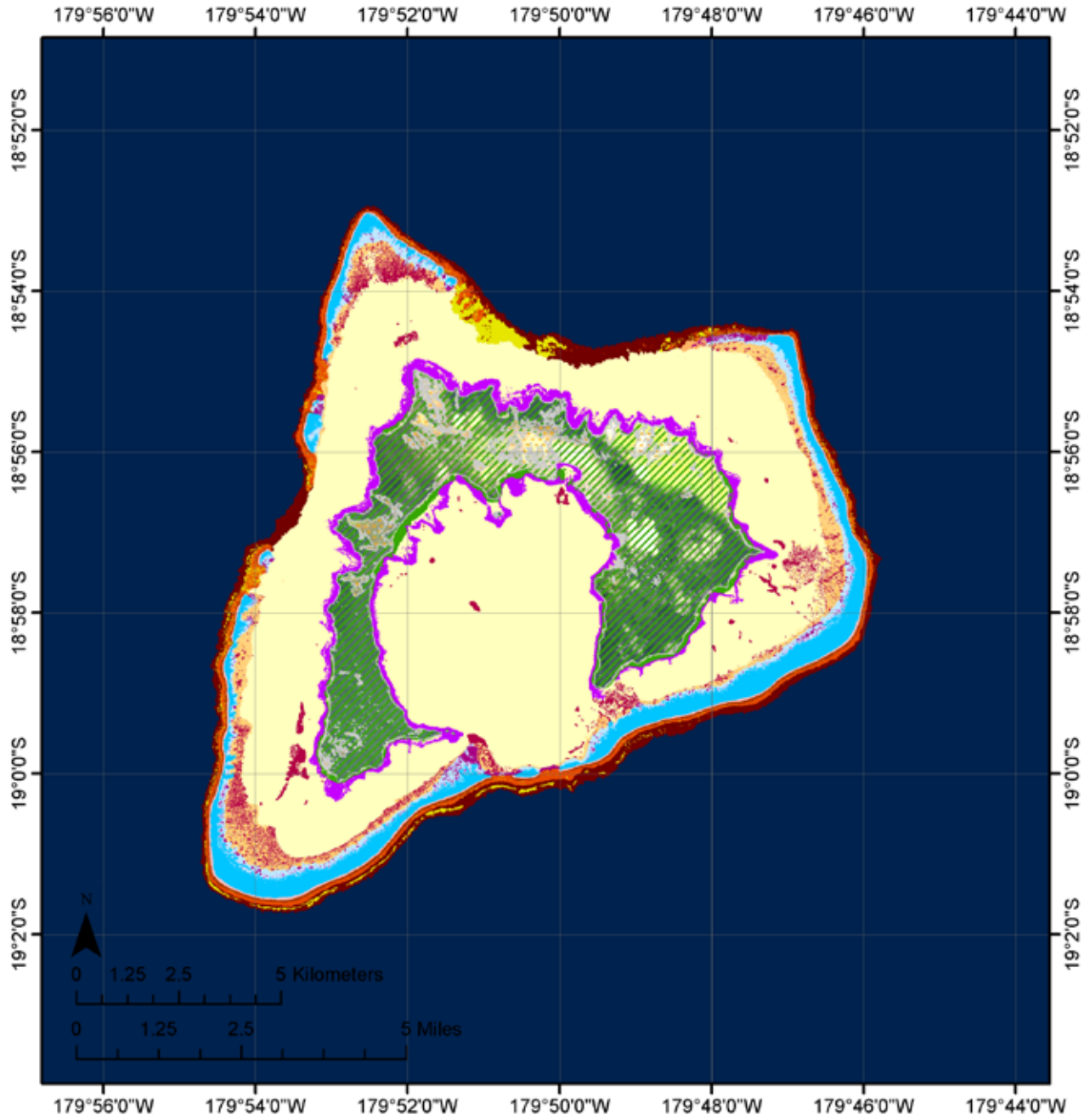


Water Depth (m)



Fig. 33. Bathymetric map for Totoya.

Totoya Island - Habitat Map



Habitats

- | | | |
|------------------------------|---|-------------------------|
| Deep fore reef slope | Back reef coral framework | Dense seagrass meadows |
| Shallow fore reef slope | Back reef coral bommies | Mud flats |
| Shallow fore reef terrace | Lagoonal sediment apron sediment dominated | Deep ocean water |
| Fore reef sand flats | Lagoonal floor barren | Terrestrial vegetation |
| Coralline algal ridge | Lagoonal pinnacle reefs massive coral dominated | Beach sand |
| Back reef rubble dominated | Lagoonal patch reefs | Urban |
| Back reef sediment dominated | Lagoonal floor coral bommies | Unvegetated terrestrial |
| Back reef pavement | Lagoonal fringing reefs | |

Fig. 34. Habitat map for Totoya.

Matuka Island - Satellite Map

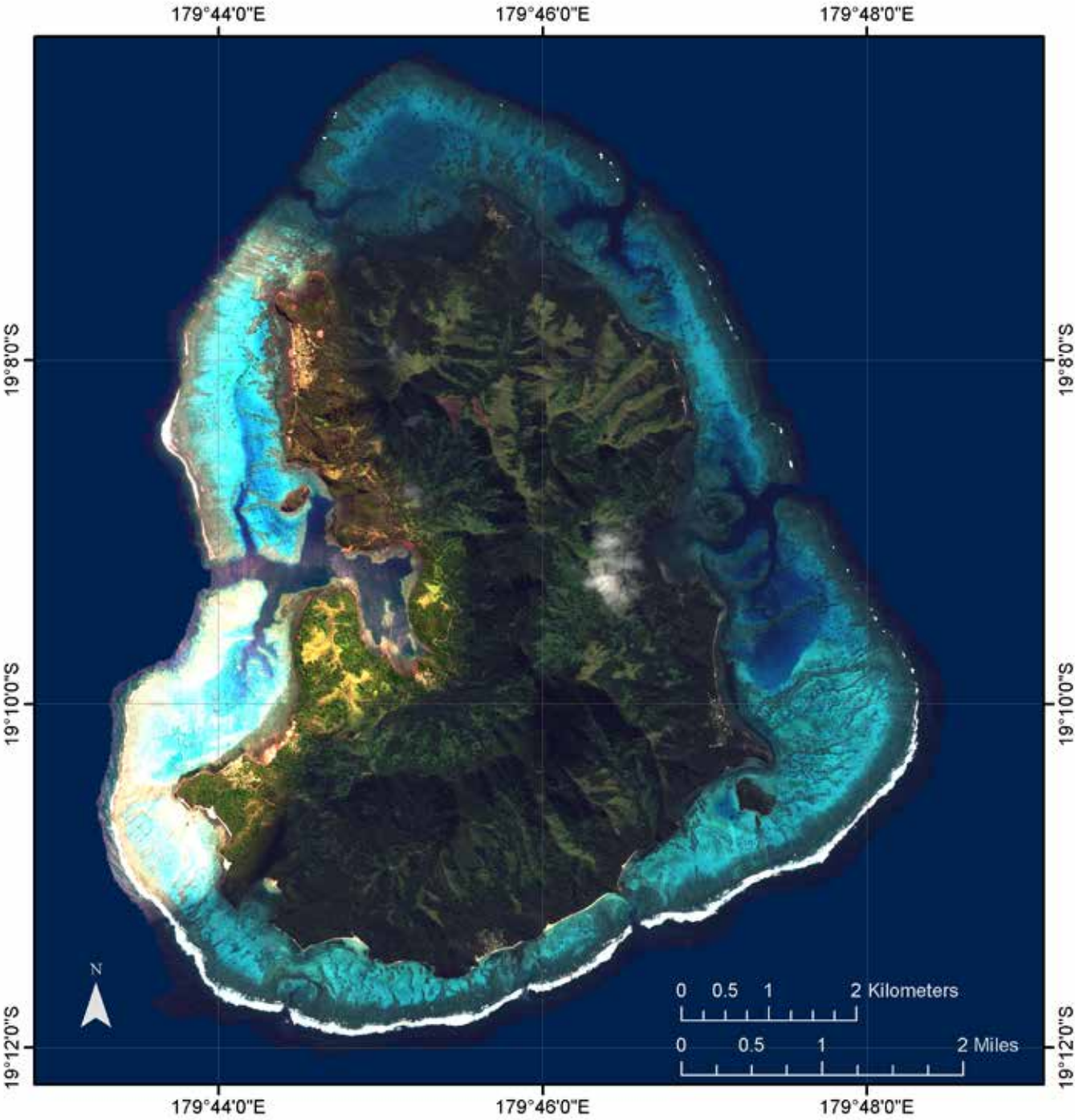
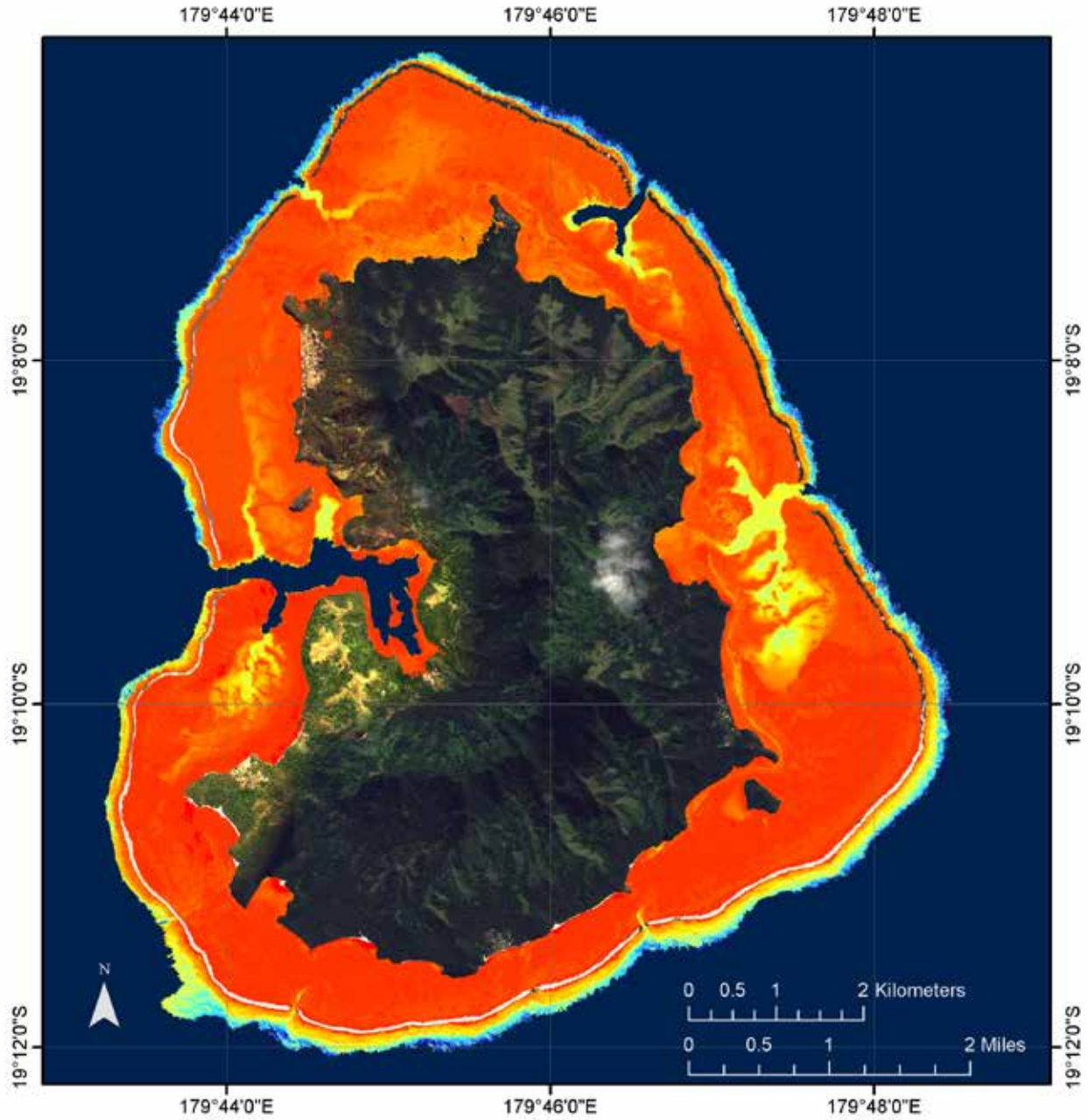


Fig. 35. WorldView-2 Satellite imagery for Matuka.

Matuka Island - Bathymetric Map

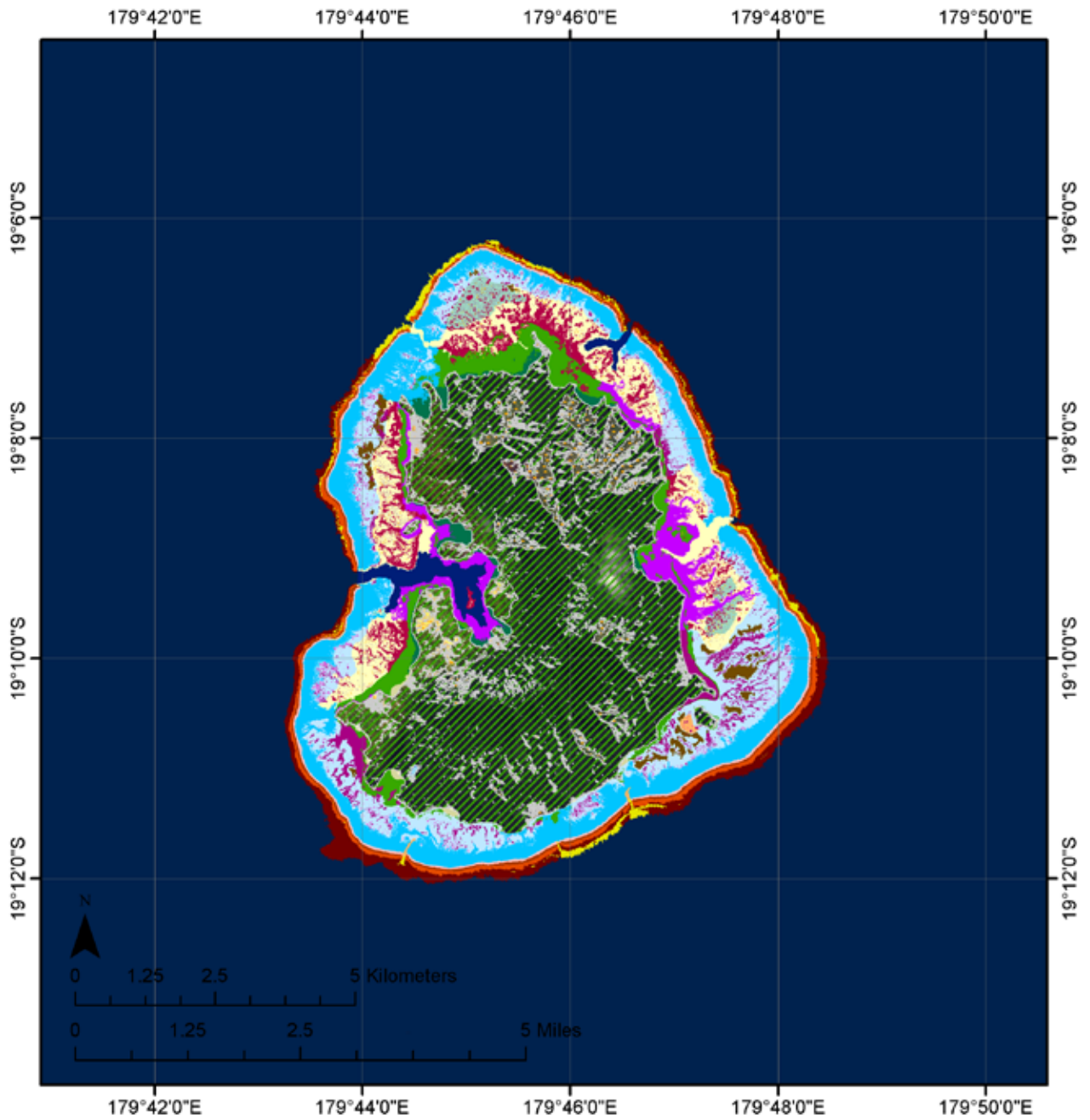


Deep Water (m)

0.10 - 1.00	8.01 - 9.00	16.01 - 17.00	24.01 - 25.00
1.01 - 2.00	9.01 - 10.00	17.01 - 18.00	25.01 - 26.00
2.01 - 3.00	10.01 - 11.00	18.01 - 19.00	26.01 - 27.00
3.01 - 4.00	11.01 - 12.00	19.01 - 20.00	27.01 - 28.00
4.01 - 5.00	12.01 - 13.00	20.01 - 21.00	28.01 - 29.00
5.01 - 6.00	13.01 - 14.00	21.01 - 22.00	29.01 - 30.00
6.01 - 7.00	14.01 - 15.00	22.01 - 23.00	30.01 - 31.00
7.01 - 8.00	15.01 - 16.00	23.01 - 24.00	31.01 - 32.00
			Deep water

Fig. 36. Bathymetric map for Matuka.

Matuka Island - Habitat Map



Habitats

- | | | |
|---|--|---|
| Deep fore reef slope | Back reef coral bommies | Mangroves |
| Shallow fore reef slope | Lagoonal floor barren | Mud flats |
| Shallow fore reef terrace | Lagoonal floor bommie field | Deep lagoonal water |
| Fore reef sand flats | Lagoonal pinnacle reefs massive coral dominated | Deep ocean water |
| Coralline algal ridge | Lagoonal patch reefs | Terrestrial vegetation |
| Back reef rubble dominated | Lagoonal floor coral bommies | Beach sand |
| Back reef sediment dominated | Lagoonal fringing reefs | Inland waters |
| Back reef pavement | Dense seagrass meadows | Urban |
| Back reef coral framework | Dense macroalgae on sediment | Unvegetated terrestrial |

Fig. 37. Habitat map for Matuka.

Moala Island - Satellite Map

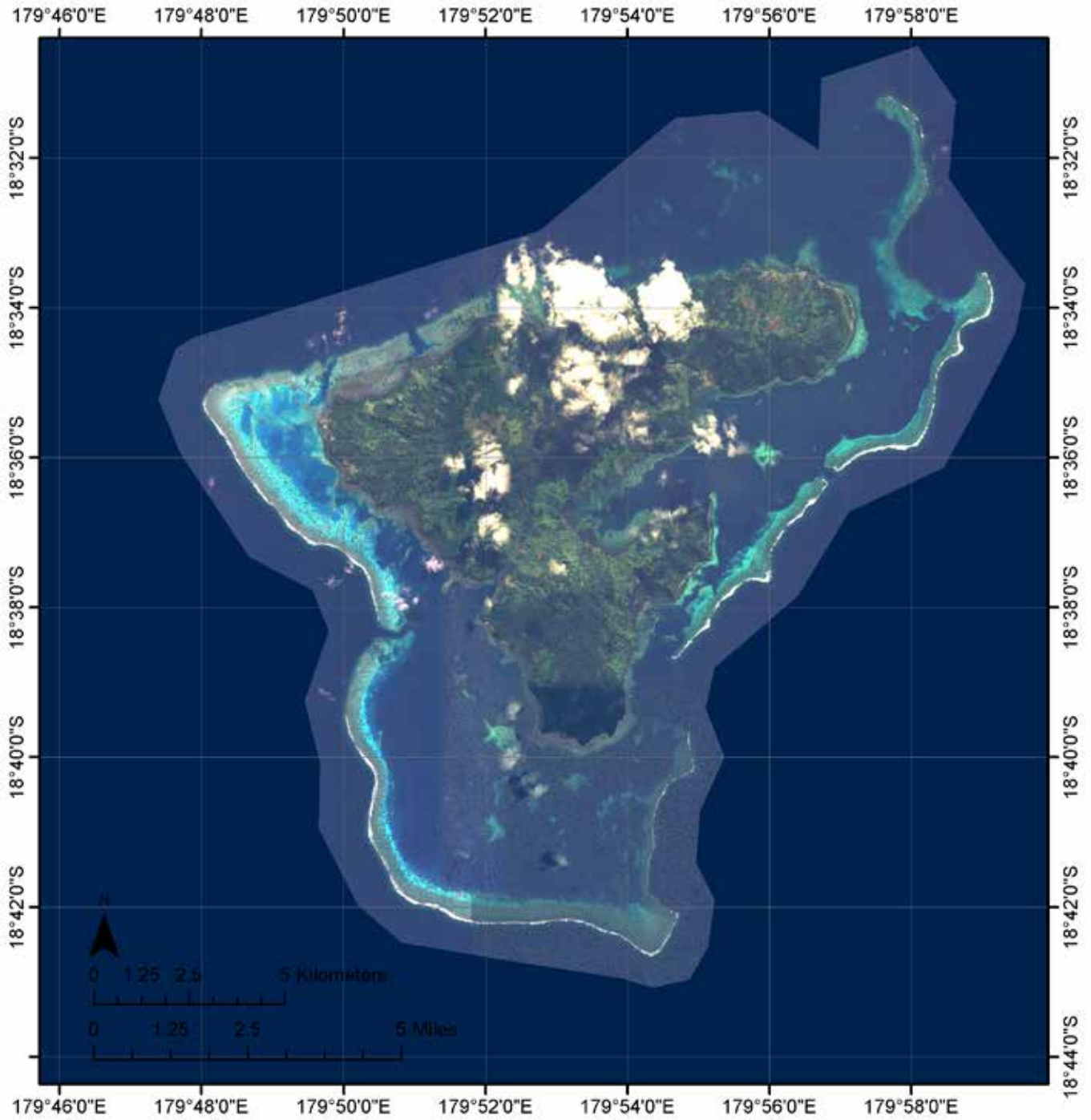


Fig. 38. WorldView-2 Satellite imagery for Moala.

Moala Island - Bathymetric Map

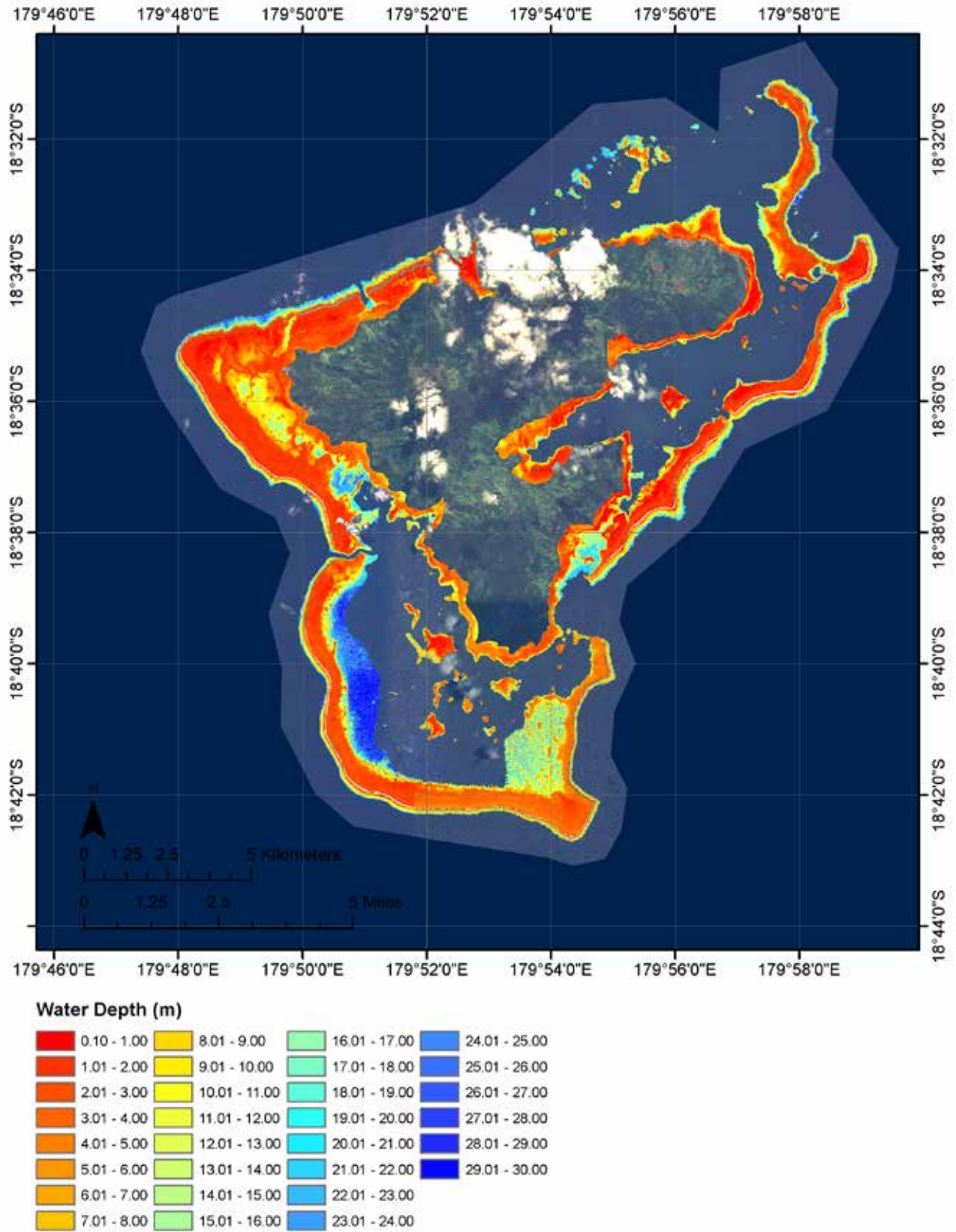


Fig. 39. Bathymetric map for Moala.

Moala Island - Habitat Map

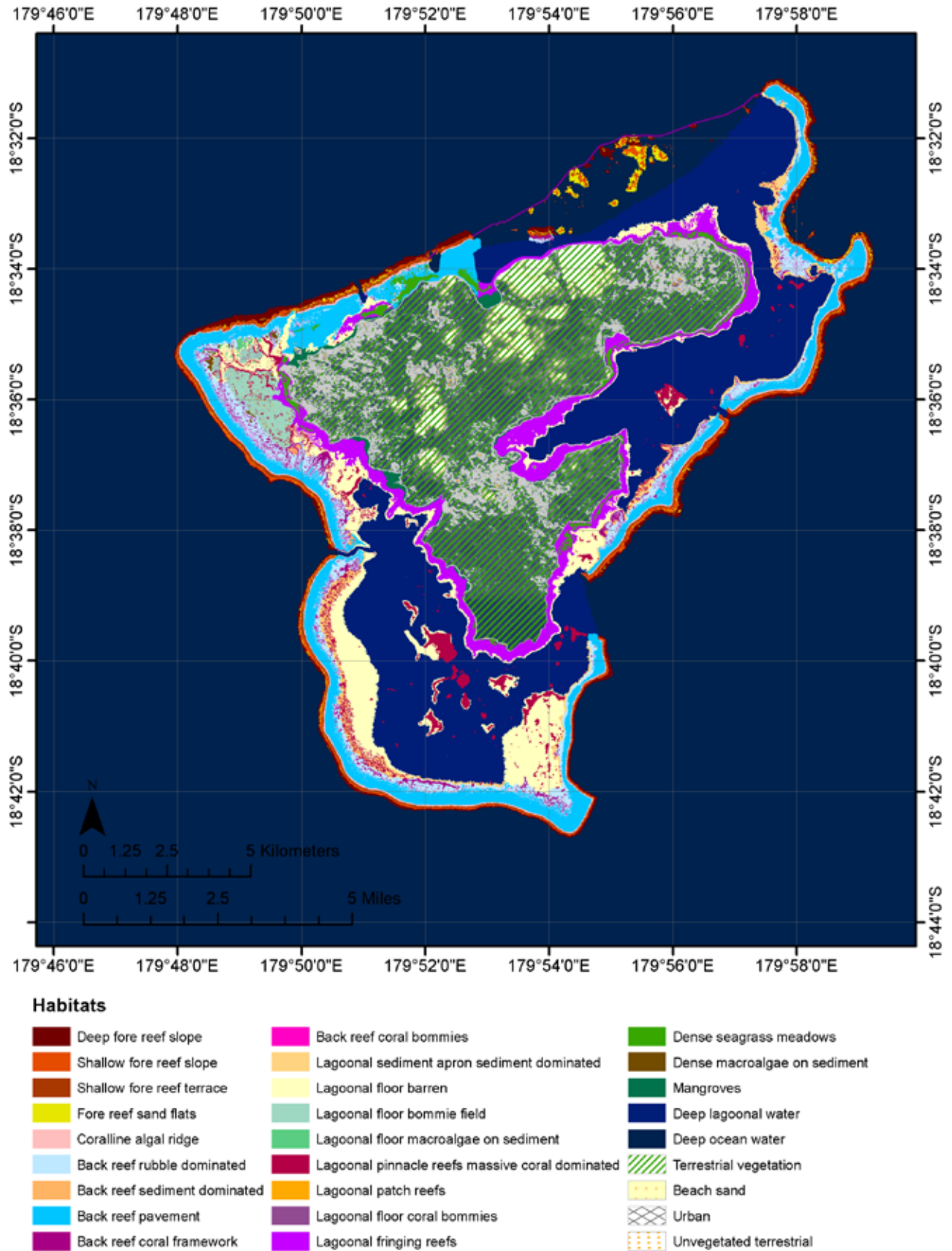


Fig. 40. Habitat map for Moala.

Fulaga Island - Satellite Map

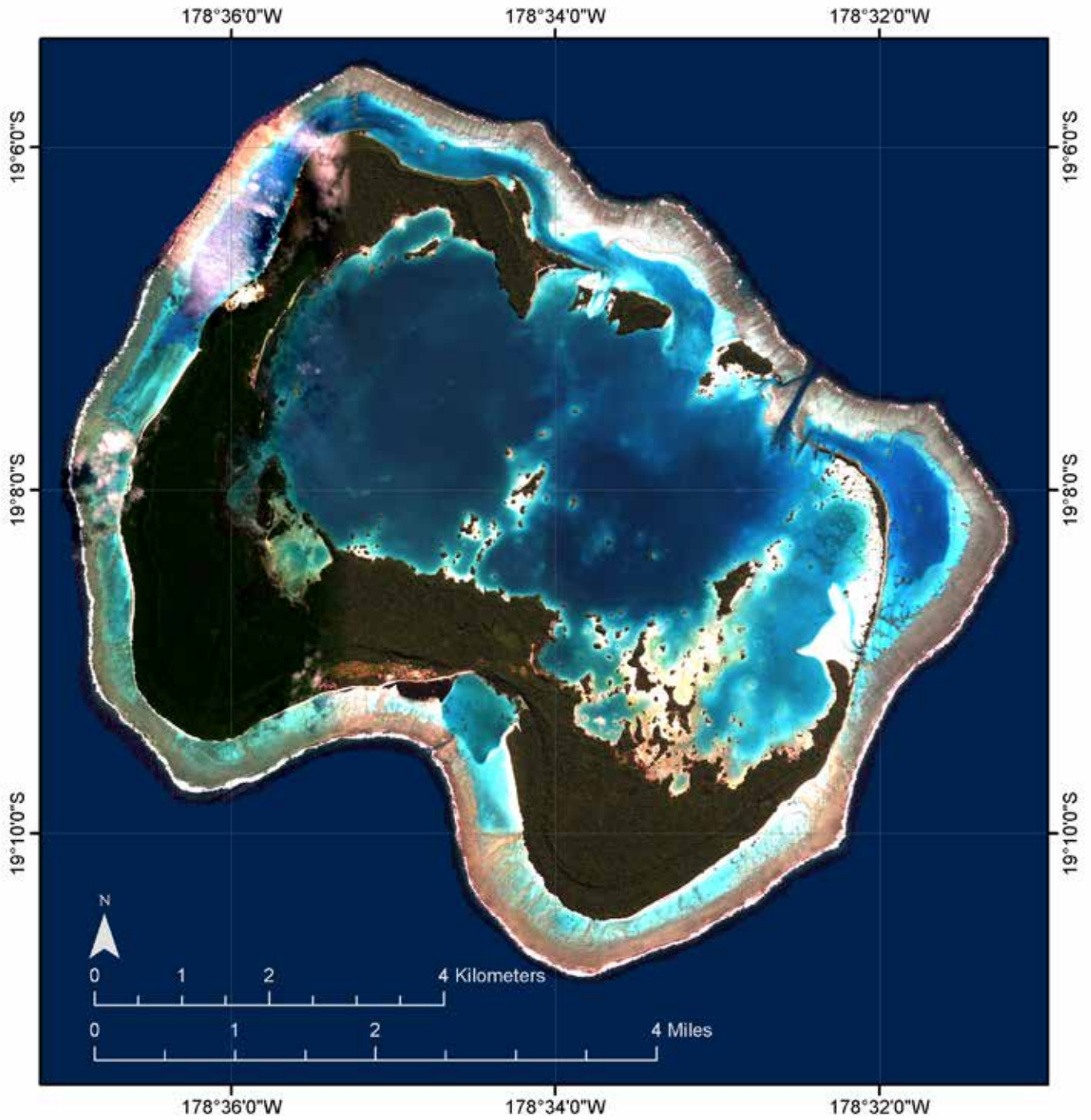
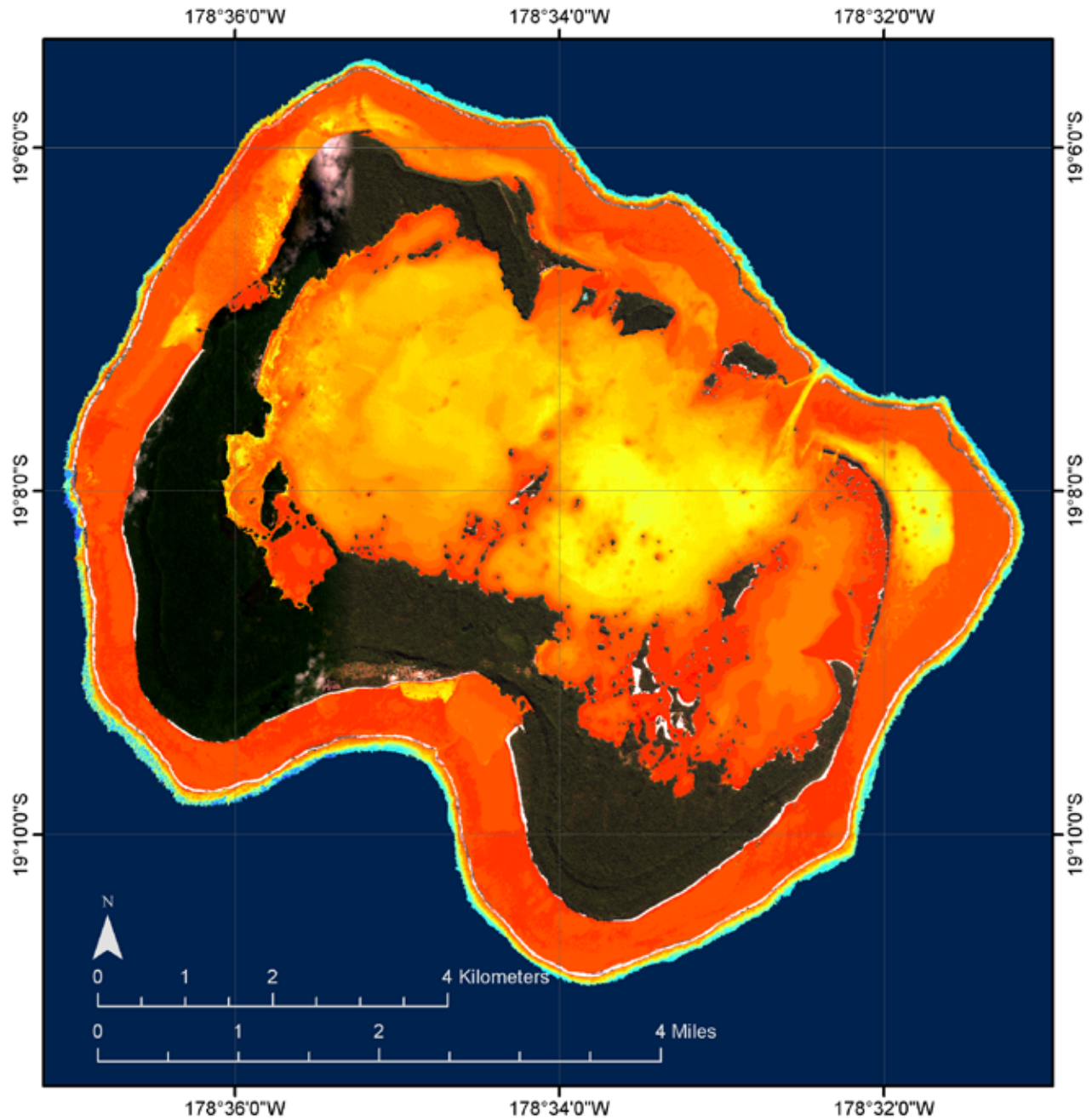


Fig. 41. WorldView-2 Satellite imagery for Fulaga.

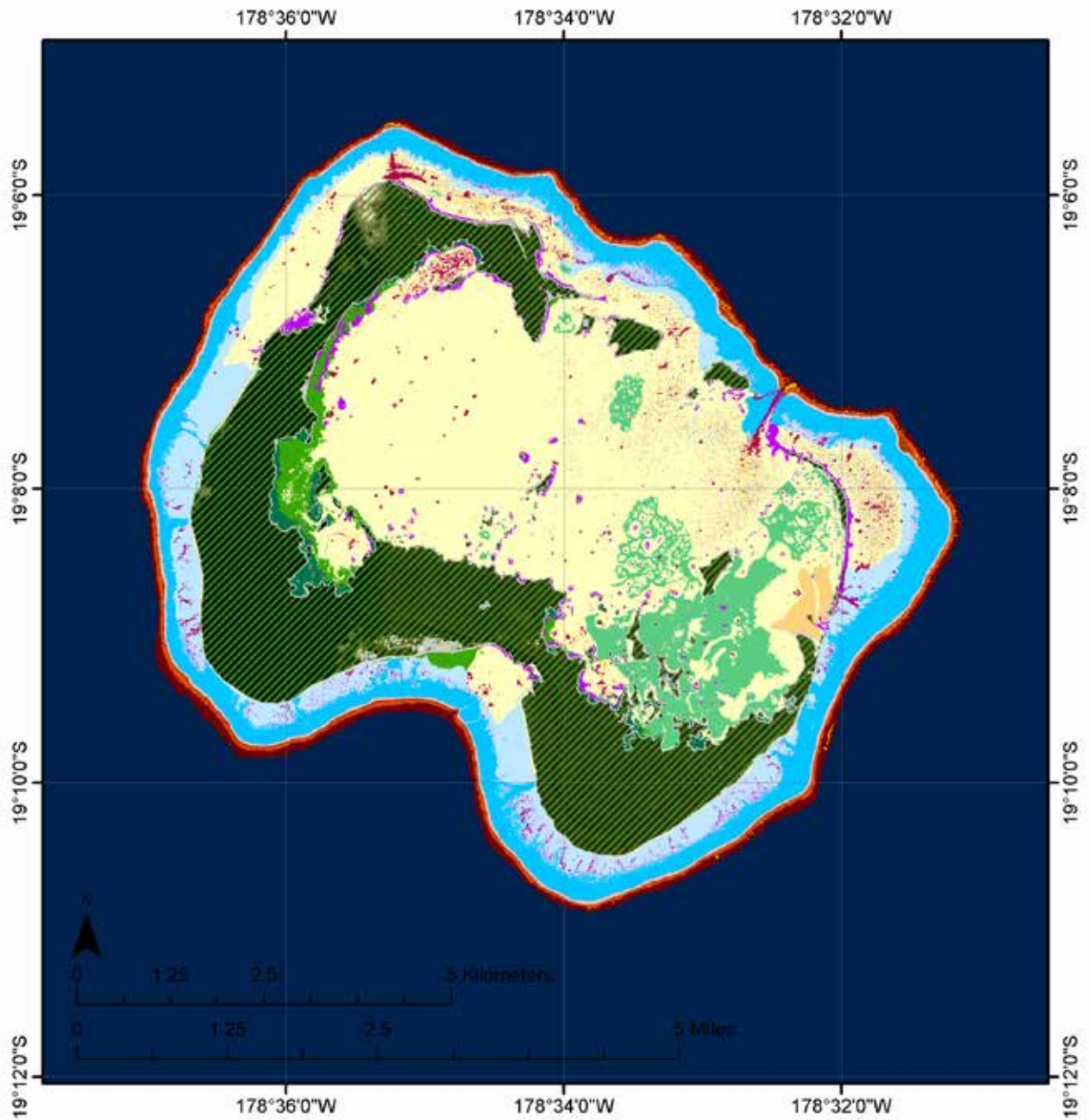
Fulaga Island - Bathymetric Map



Water Depth (m)			
0.10 - 1.00	8.01 - 9.00	16.01 - 17.00	24.01 - 25.00
1.01 - 2.00	9.01 - 10.00	17.01 - 18.00	25.01 - 26.00
2.01 - 3.00	10.01 - 11.00	18.01 - 19.00	26.01 - 27.00
3.01 - 4.00	11.01 - 12.00	19.01 - 20.00	27.01 - 28.00
4.01 - 5.00	12.01 - 13.00	20.01 - 21.00	28.01 - 29.00
5.01 - 6.00	13.01 - 14.00	21.01 - 22.00	29.01 - 30.00
6.01 - 7.00	14.01 - 15.00	22.01 - 23.00	Deep water
7.01 - 8.00	15.01 - 16.00	23.01 - 24.00	

Fig. 42. Bathymetric map for Fulaga.

Fulaga Island - Habitat Map



Habitats

- | | | |
|----------------------------|---|------------------------------|
| Deep fore reef slope | Reef-top algal mats | Dense macroalgae on sediment |
| Shallow fore reef slope | Lagoonal sediment apron sediment dominated | Mangroves |
| Shallow fore reef terrace | Lagoonal floor barren | Mud flats |
| Fore reef sand flats | Lagoonal floor macroalgae on sediment | Deep ocean water |
| Coralline algal ridge | Lagoonal pinnacle reefs massive coral dominated | Terrestrial vegetation |
| Back reef rubble dominated | Lagoonal patch reefs | Beach sand |
| Back reef pavement | Lagoonal floor coral bommies | Inland waters |
| Back reef coral framework | Lagoonal fringing reefs | Urban |
| Back reef coral bommies | Dense seagrass meadows | Unvegetated terrestrial |

Fig. 43. Habitat map for Fulaga.

Kabara Island - Satellite Map

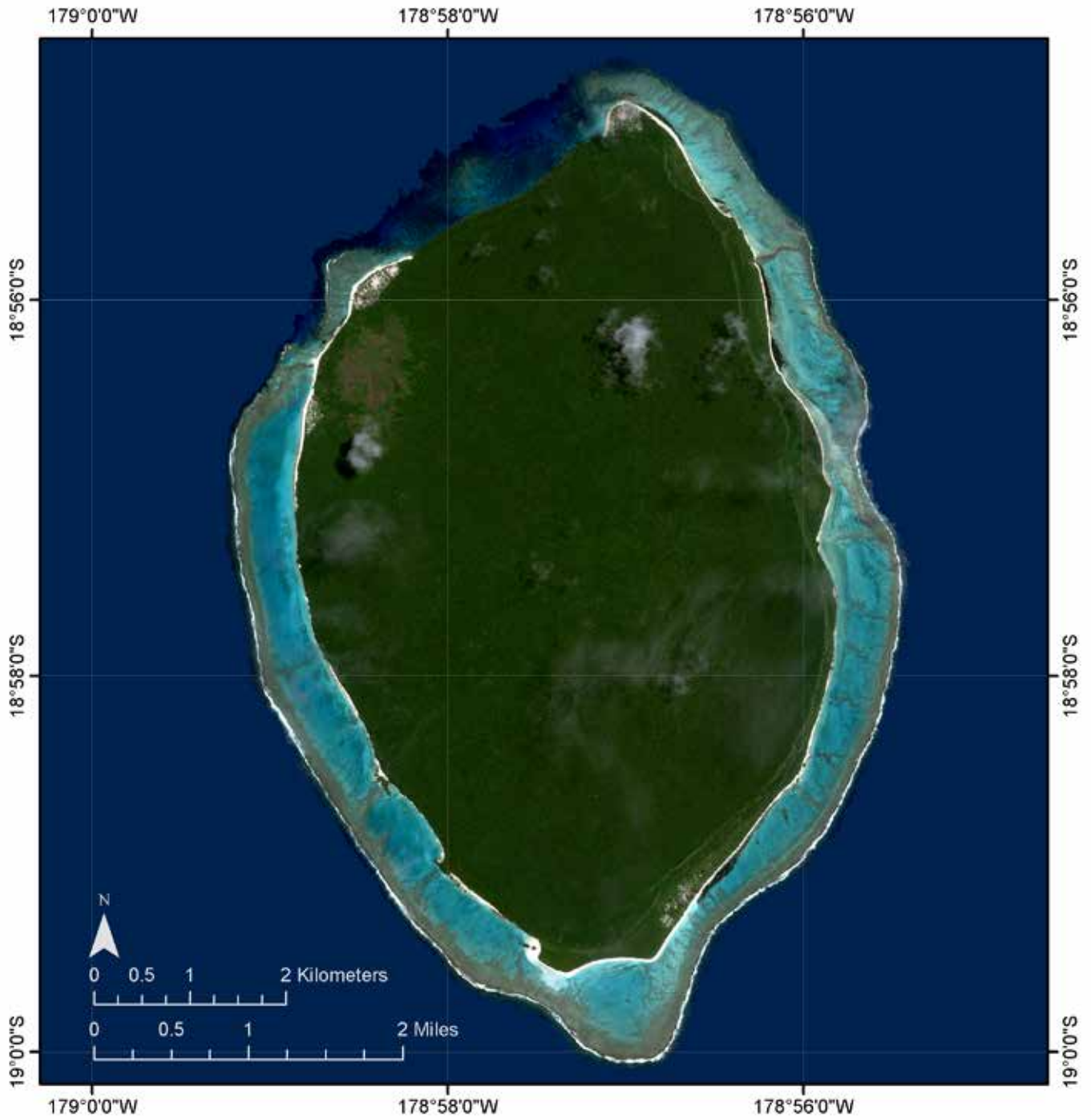


Fig. 44. WorldView-2 Satellite imagery for Kabara.

Kabara Island - Bathymetric Map

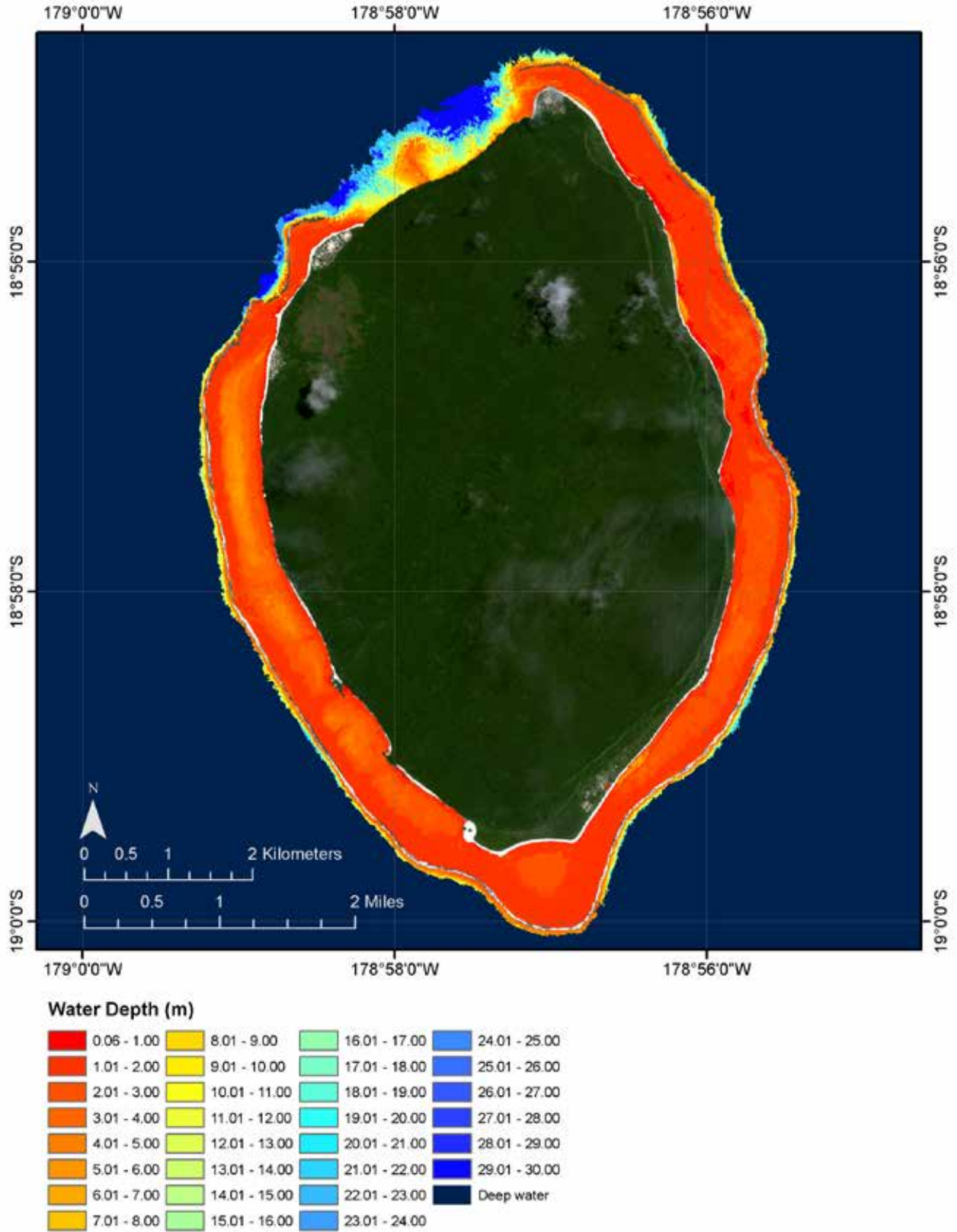


Fig. 45. Bathymetric map for Kabara.

Kabara Island - Habitat Map

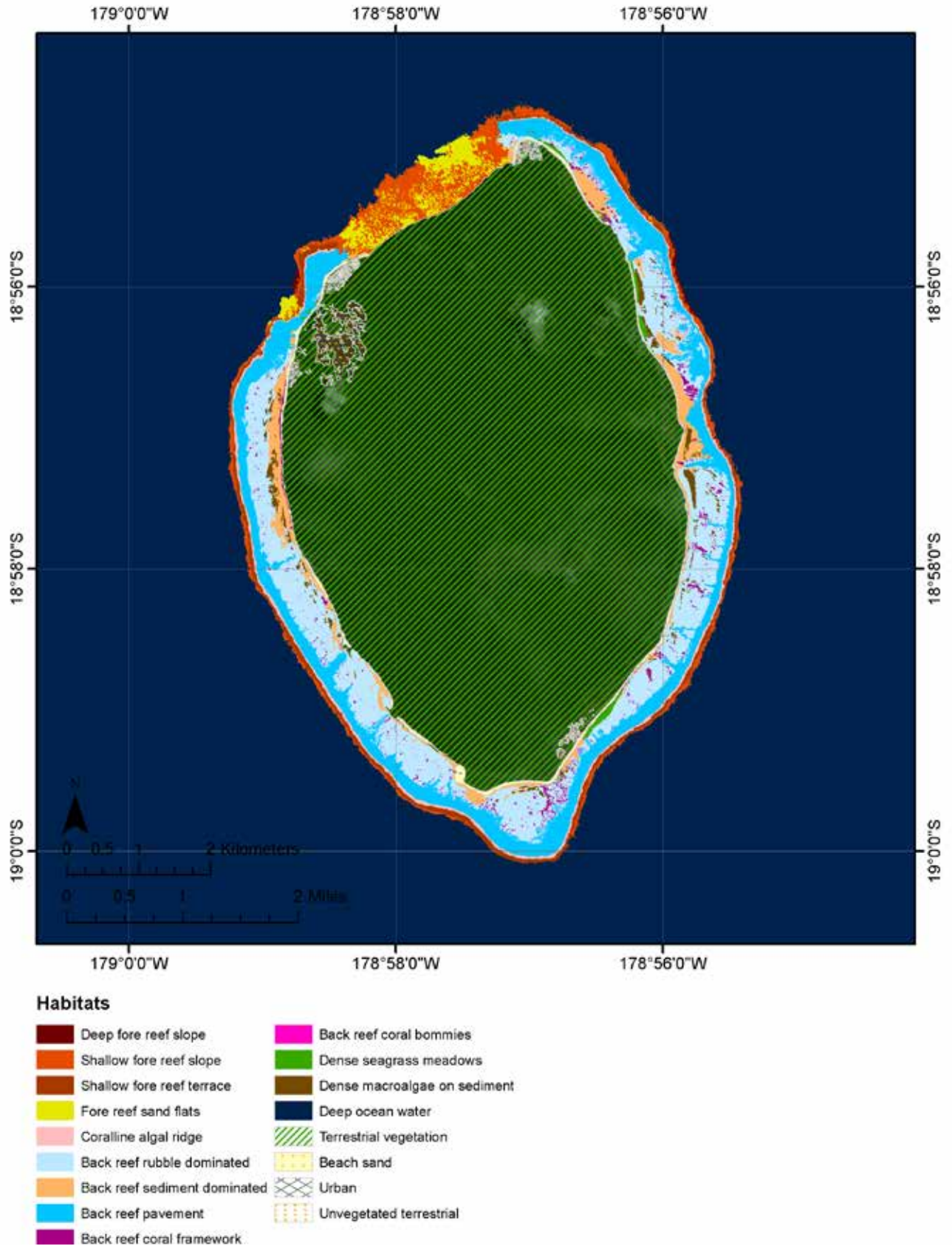


Fig. 46. Habitat map for Kabara.

Vanua Vatu Island - Satellite Map

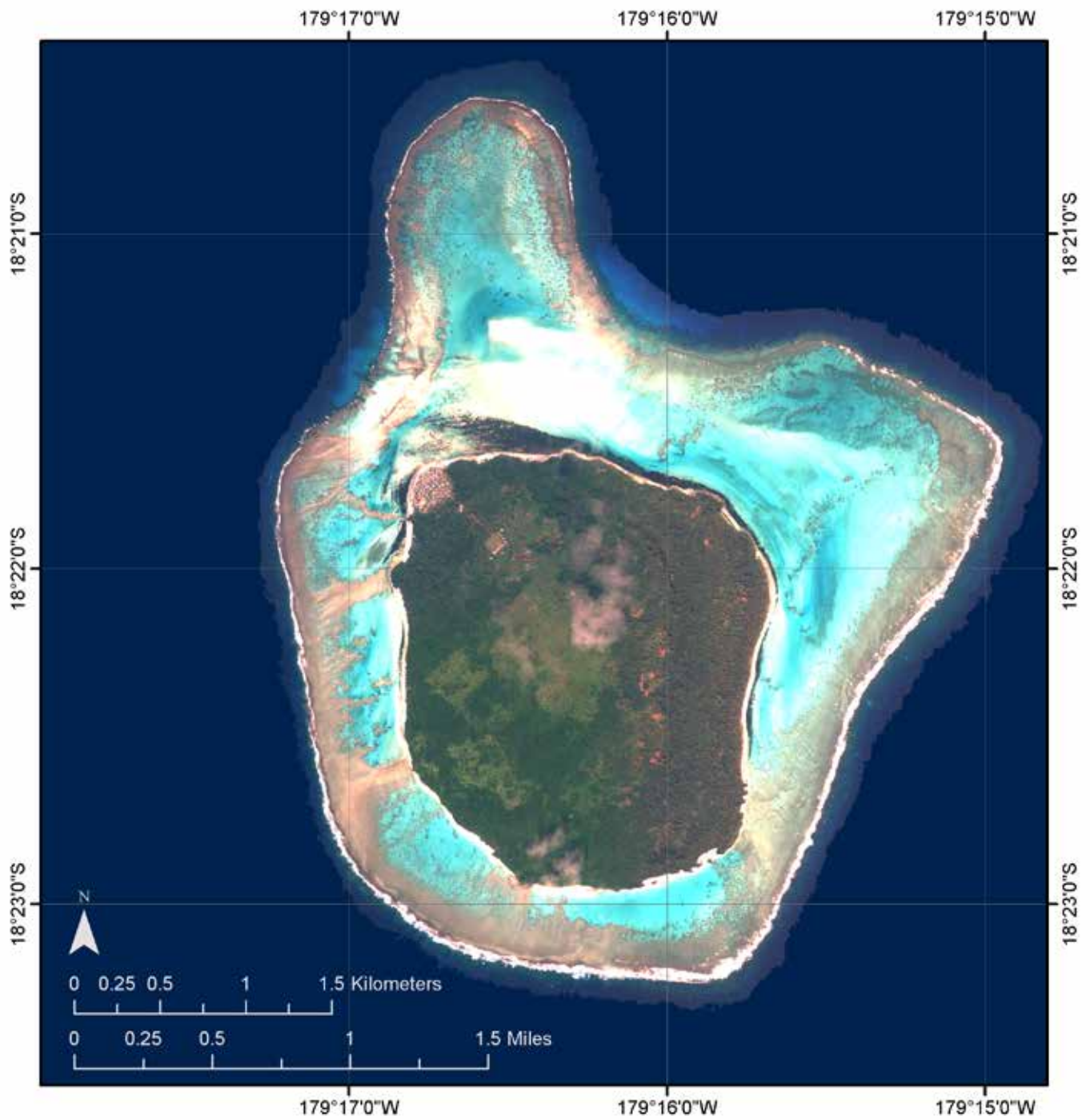


Fig. 47. WorldView-2 Satellite imagery for Vanua Vatu.

Vanua Vatu Island - Bathymetric Map

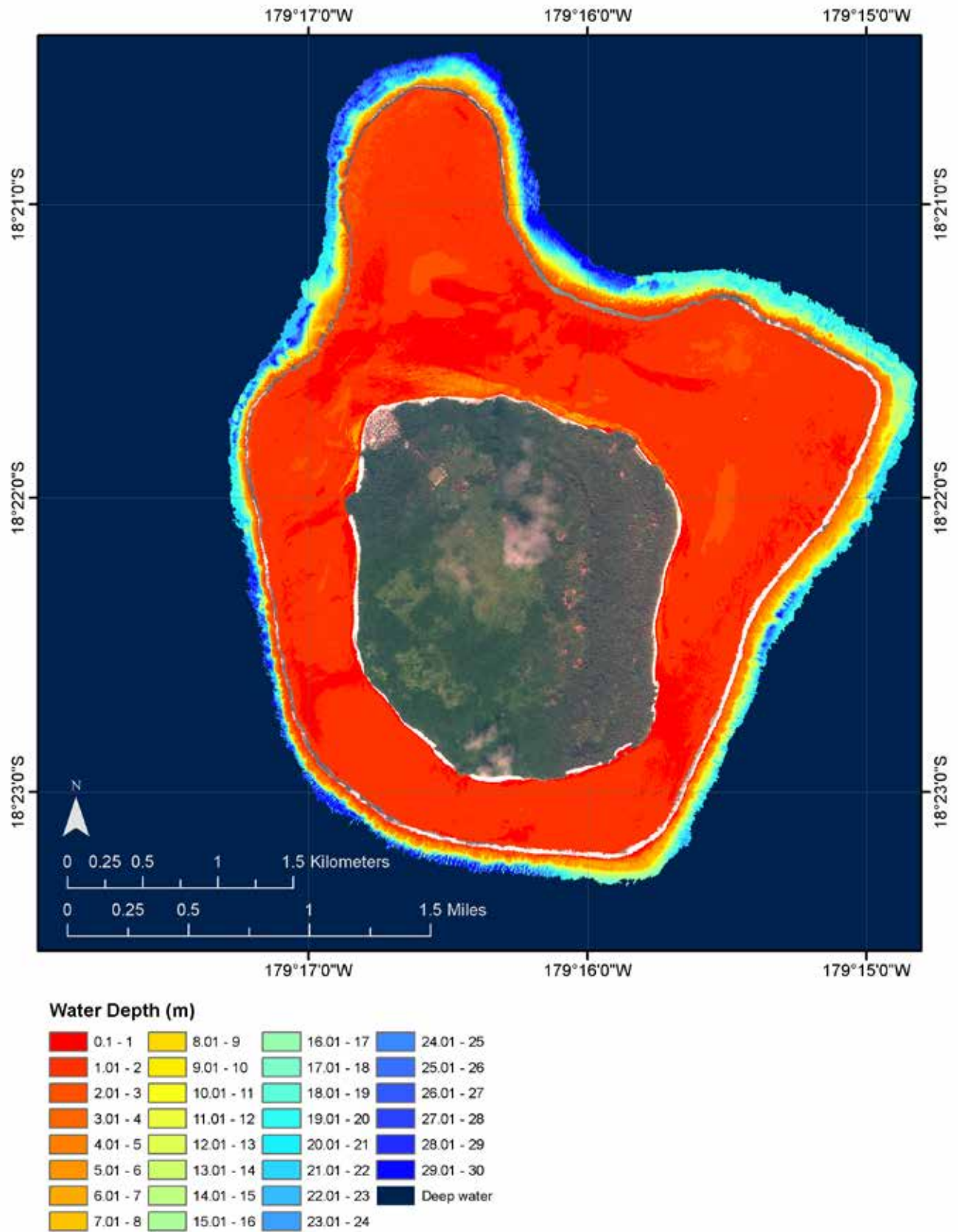


Fig. 48. Bathymetric map for Vanua Vatu.

Vanua Vatu Island - Habitat Map

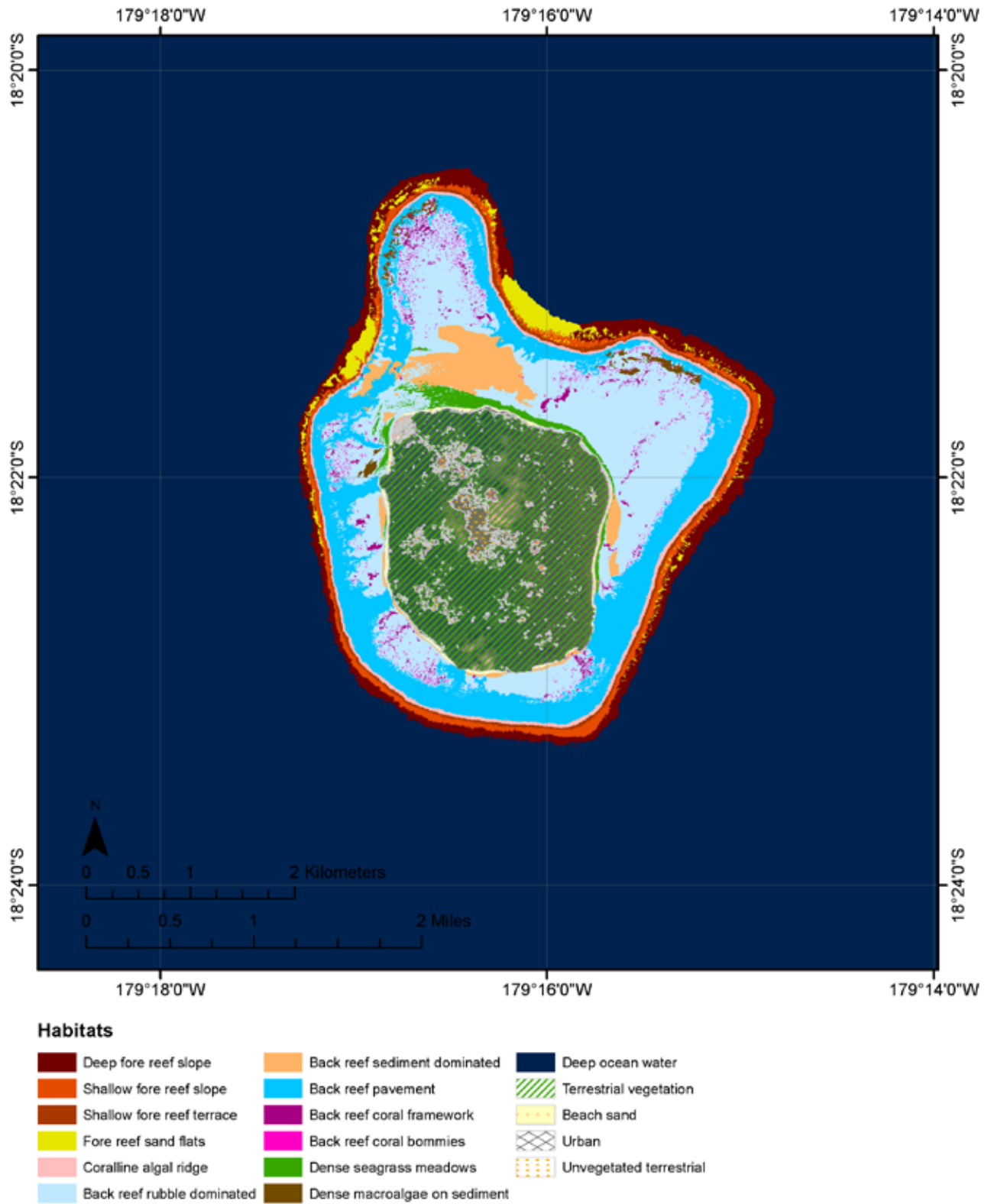


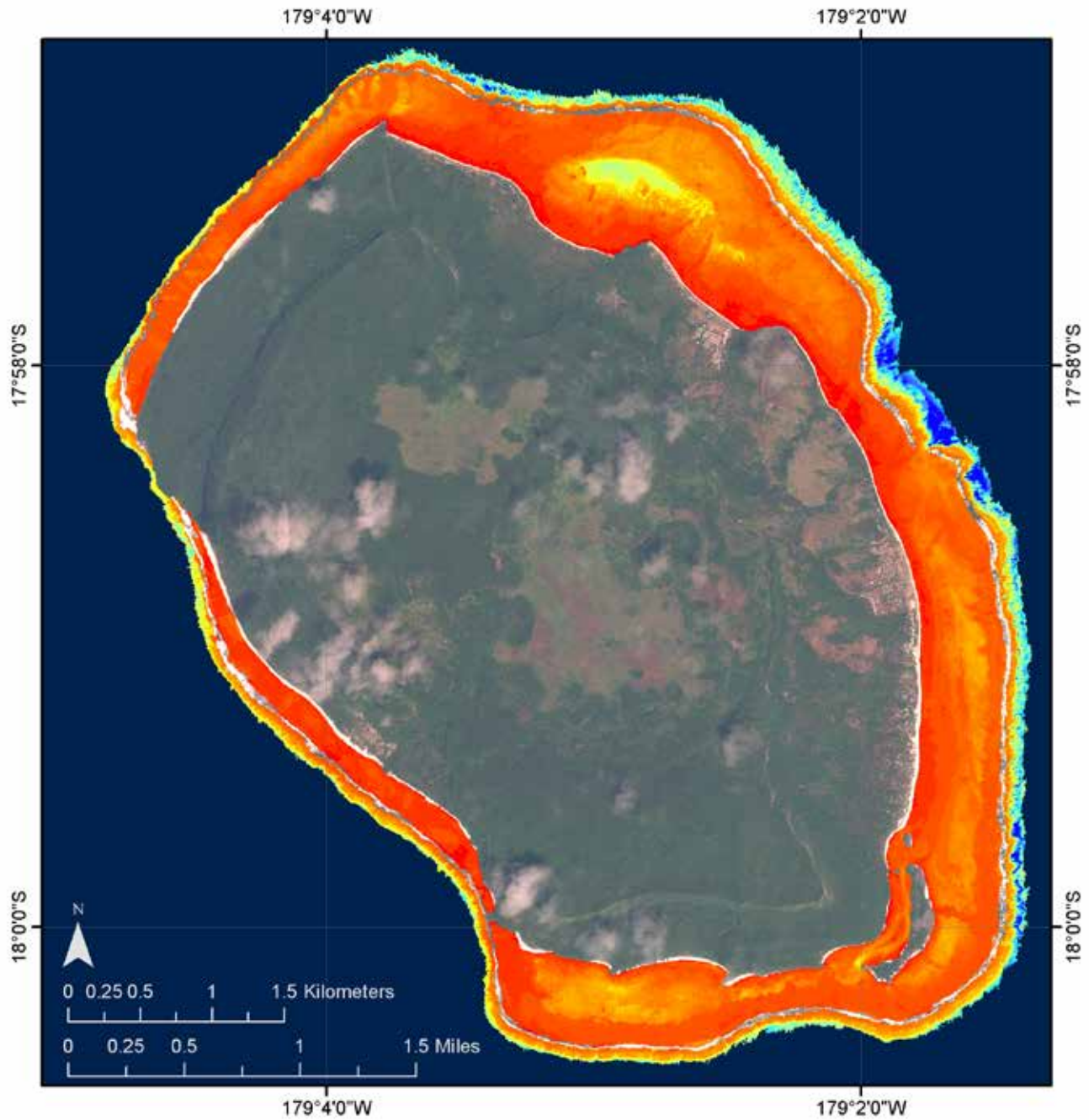
Fig. 49. Habitat map for Vanua Vatu.

Nayau Island - Satellite Map



Fig. 50. WorldView-2 Satellite imagery for Nayau.

Nayau Island - Bathymetric Map



Water Depth (m)

0.10 - 1.00	8.01 - 9.00	16.01 - 17.00	24.01 - 25.00
1.01 - 2.00	9.01 - 10.00	17.01 - 18.00	25.01 - 26.00
2.01 - 3.00	10.01 - 11.00	18.01 - 19.00	26.01 - 27.00
3.01 - 4.00	11.01 - 12.00	19.01 - 20.00	27.01 - 28.00
4.01 - 5.00	12.01 - 13.00	20.01 - 21.00	28.01 - 29.00
5.01 - 6.00	13.01 - 14.00	21.01 - 22.00	29.01 - 30.00
6.01 - 7.00	14.01 - 15.00	22.01 - 23.00	Deep water
7.01 - 8.00	15.01 - 16.00	23.01 - 24.00	

Fig. 51. Bathymetric map for Nayau.

Nayau Island - Habitat Map

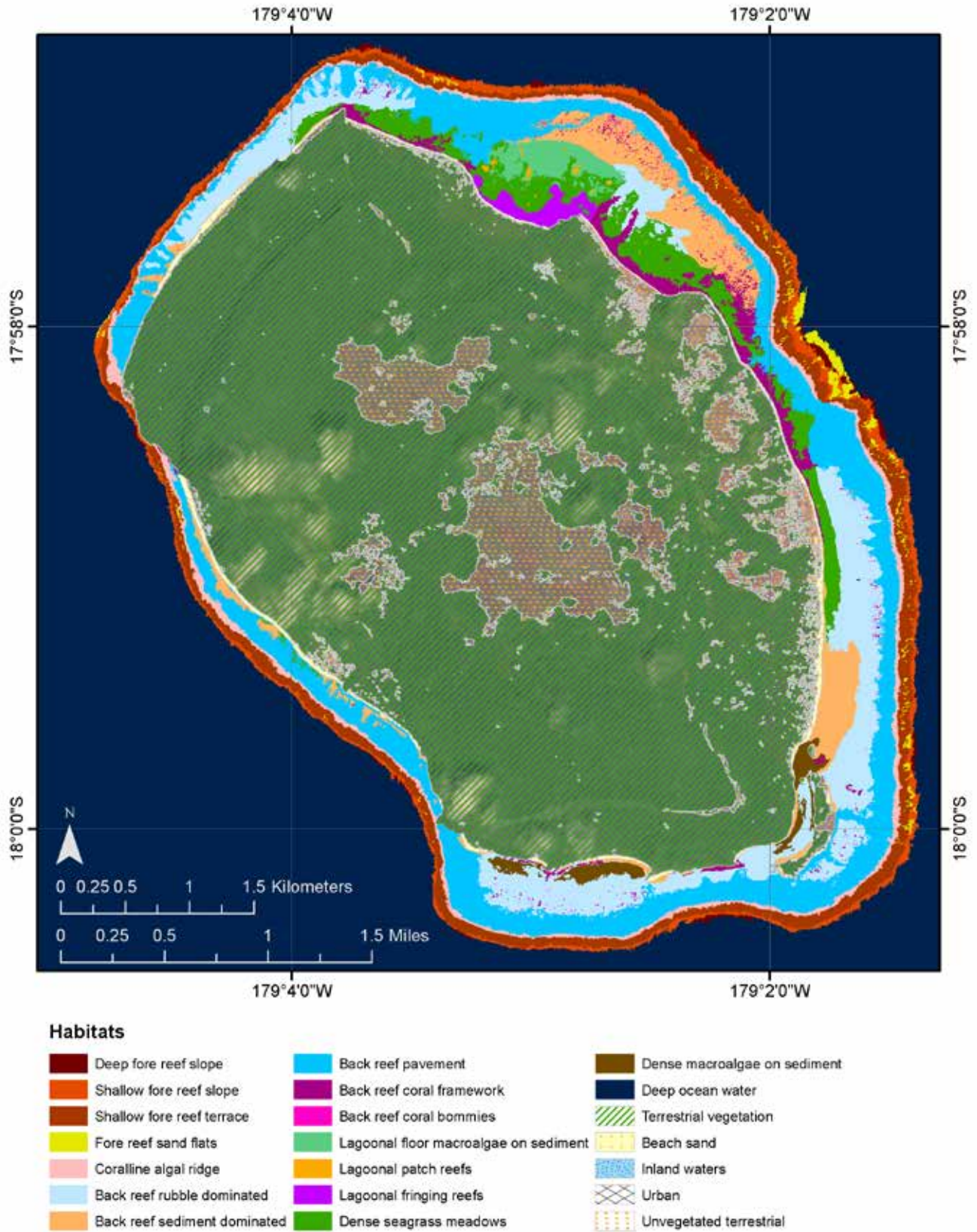


Fig. 52. Habitat map for Nayau.

Tuvuca Island - Satellite Map

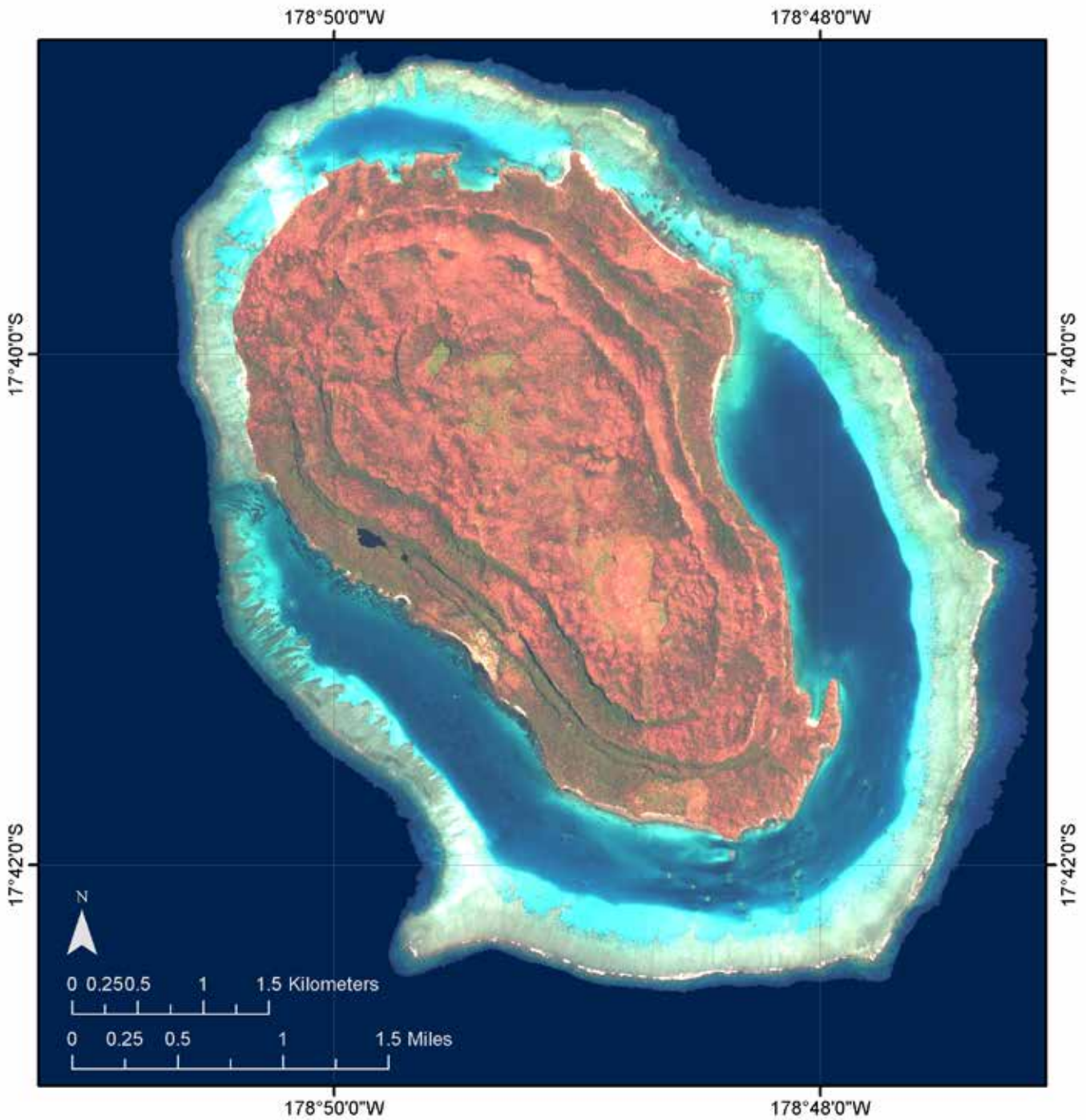
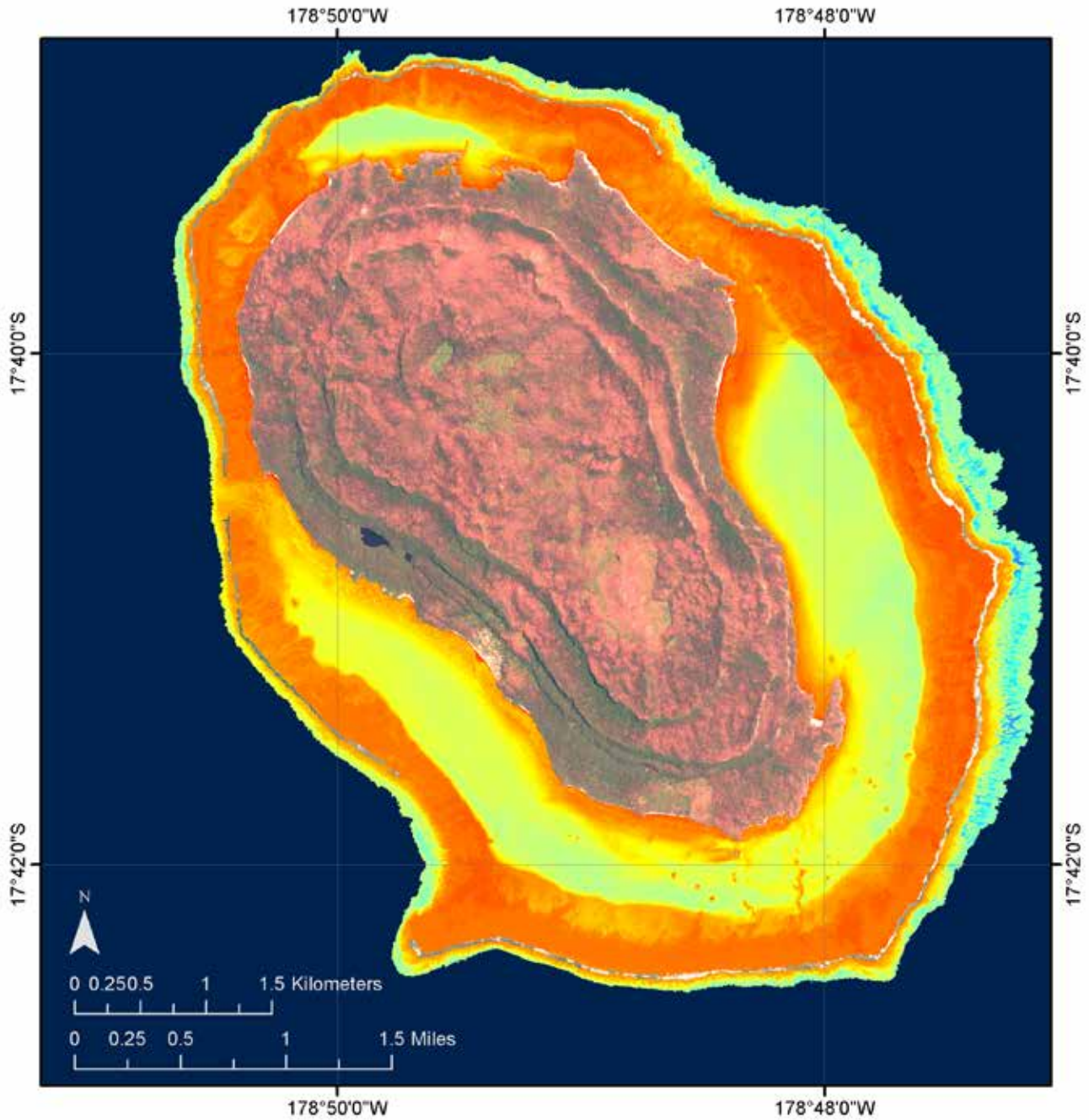


Fig. 53. WorldView-2 Satellite imagery for Tuvuca.

Tuvuca Island - Bathymetric Map



Water Depth (m)

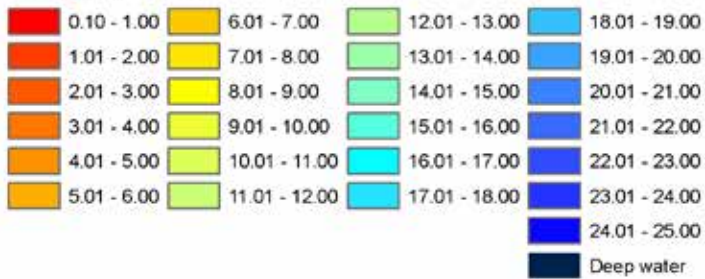
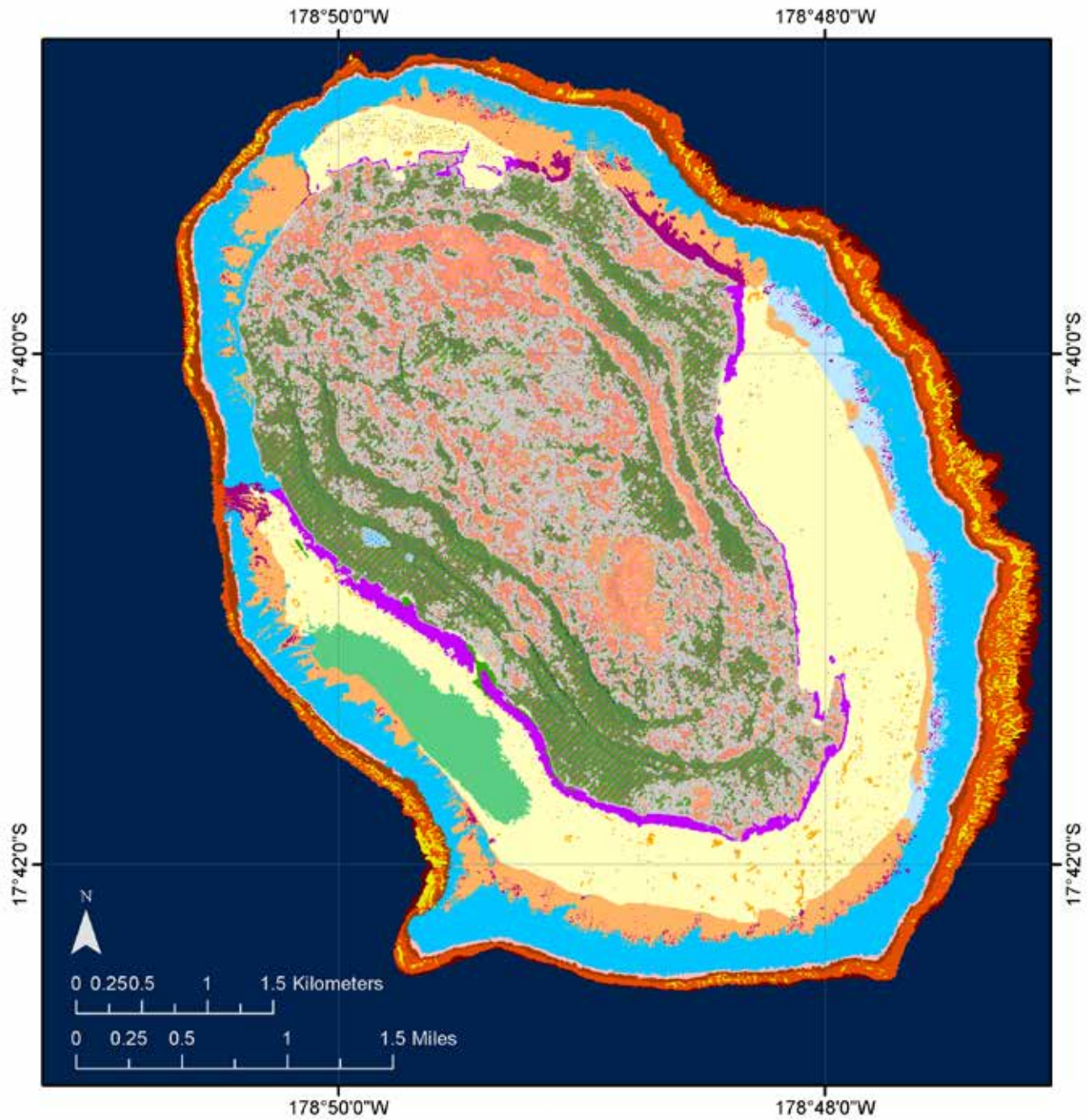


Fig. 54. Bathymetric map for Tuvuca.

Tuvuca Island - Habitat Map



Habitats

- | | | |
|------------------------------|---------------------------------------|-------------------------|
| Deep fore reef slope | Back reef coral framework | Deep ocean water |
| Shallow fore reef slope | Back reef coral bommies | Terrestrial vegetation |
| Shallow fore reef terrace | Lagoonal floor barren | Beach sand |
| Fore reef sand flats | Lagoonal floor macroalgae on sediment | Inland waters |
| Coralline algal ridge | Lagoonal patch reefs | Urban |
| Back reef rubble dominated | Lagoonal floor coral bommies | Unvegetated terrestrial |
| Back reef sediment dominated | Lagoonal fringing reefs | |
| Back reef pavement | Dense seagrass meadows | |

Fig. 55. Habitat map for Tuvuca.

Cicia Island - Satellite Map

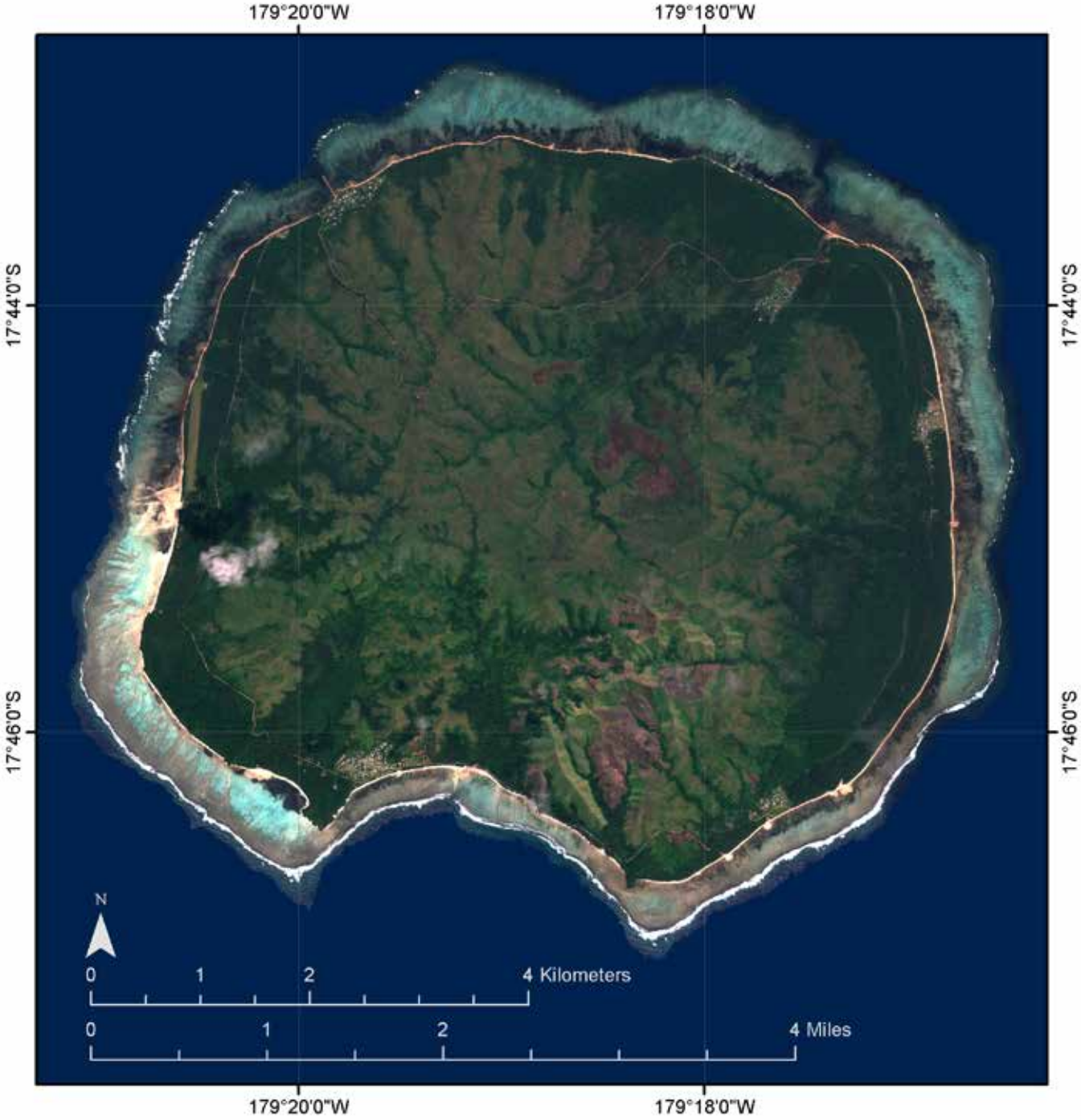
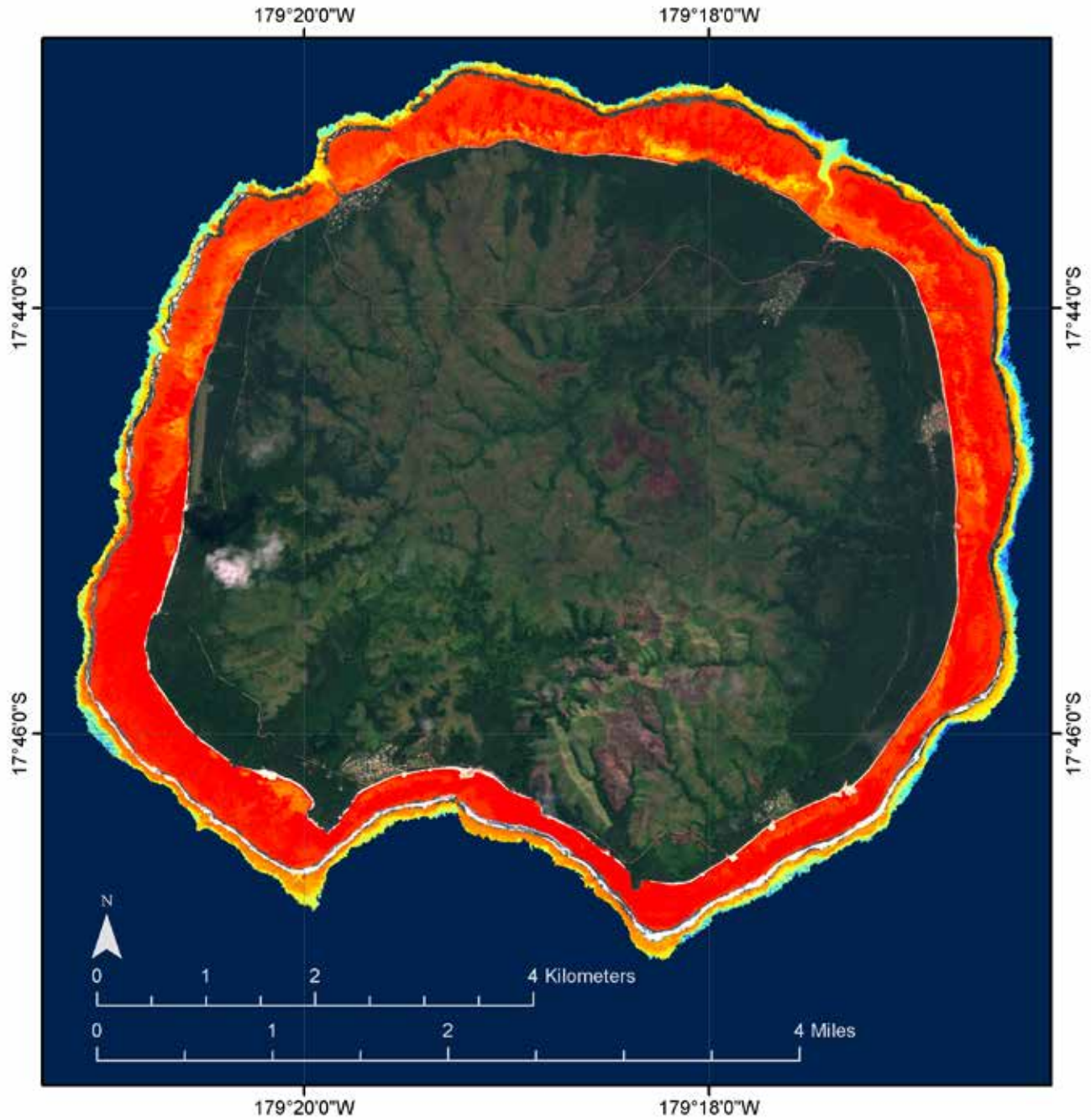


Fig. 56. WorldView-2 Satellite imagery for Cicia.

Cicia Island - Bathymetric Map

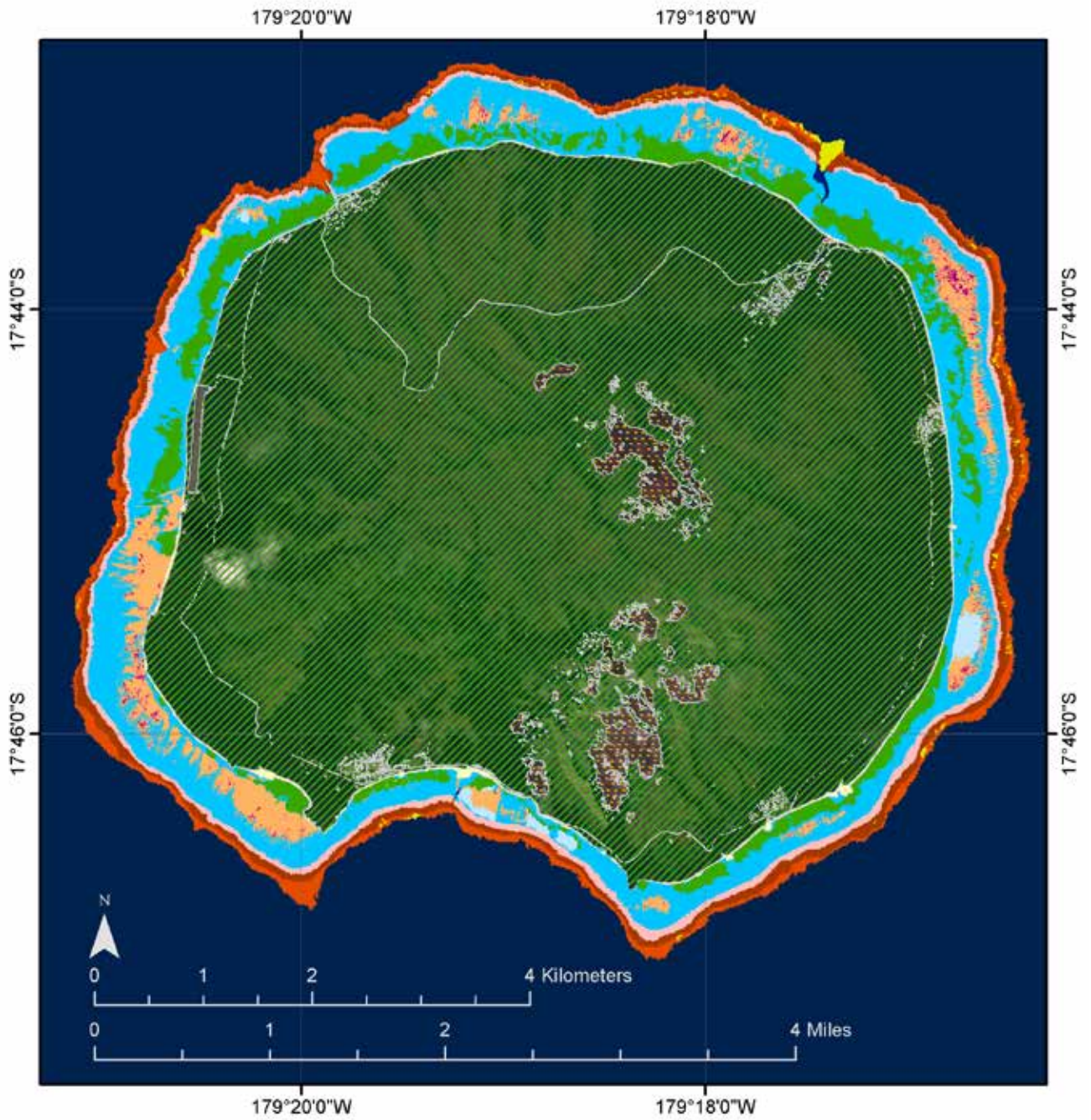


Water Depth (m)

0.10 - 1.00	8.01 - 9.00	16.01 - 17.00	24.01 - 25.00
1.01 - 2.00	9.01 - 10.00	17.01 - 18.00	25.01 - 26.00
2.01 - 3.00	10.01 - 11.00	18.01 - 19.00	26.01 - 27.00
3.01 - 4.00	11.01 - 12.00	19.01 - 20.00	27.01 - 28.00
4.01 - 5.00	12.01 - 13.00	20.01 - 21.00	28.01 - 29.00
5.01 - 6.00	13.01 - 14.00	21.01 - 22.00	29.01 - 30.00
6.01 - 7.00	14.01 - 15.00	22.01 - 23.00	Deep water
7.01 - 8.00	15.01 - 16.00	23.01 - 24.00	

Fig. 57. Bathymetric map for Cicia.

Cicia Island - Habitat Map



Habitats

- | | | |
|------------------------------|---------------------------|-------------------------|
| Shallow fore reef slope | Back reef pavement | Terrestrial vegetation |
| Shallow fore reef terrace | Back reef coral framework | Beach sand |
| Fore reef sand flats | Back reef coral bommies | Urban |
| Coralline algal ridge | Dense seagrass meadows | Unvegetated terrestrial |
| Back reef rubble dominated | Deep lagoonal water | |
| Back reef sediment dominated | Deep ocean water | |

Fig. 58. Habitat map for Cicia.

Mago Island - Satellite Map

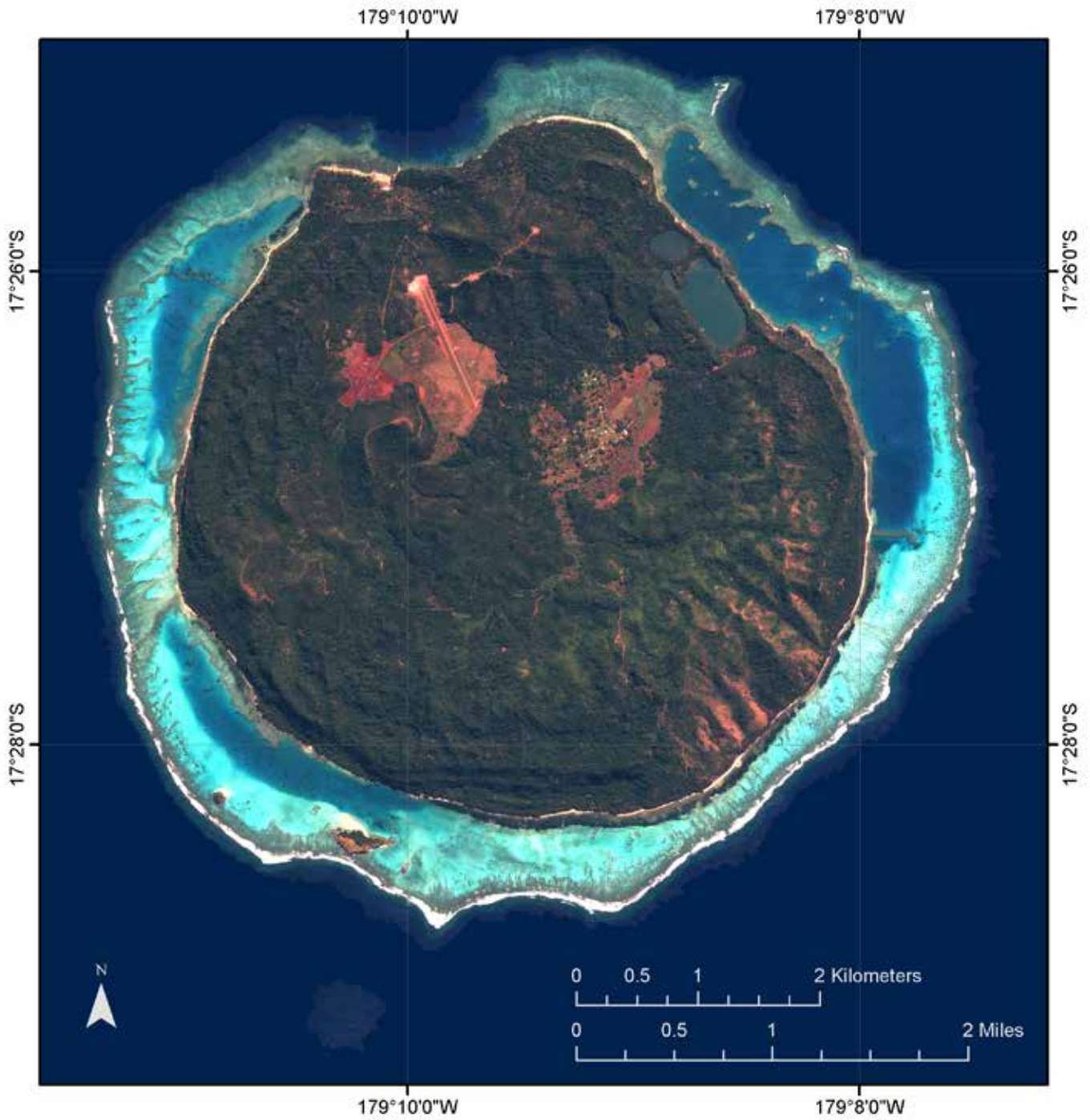
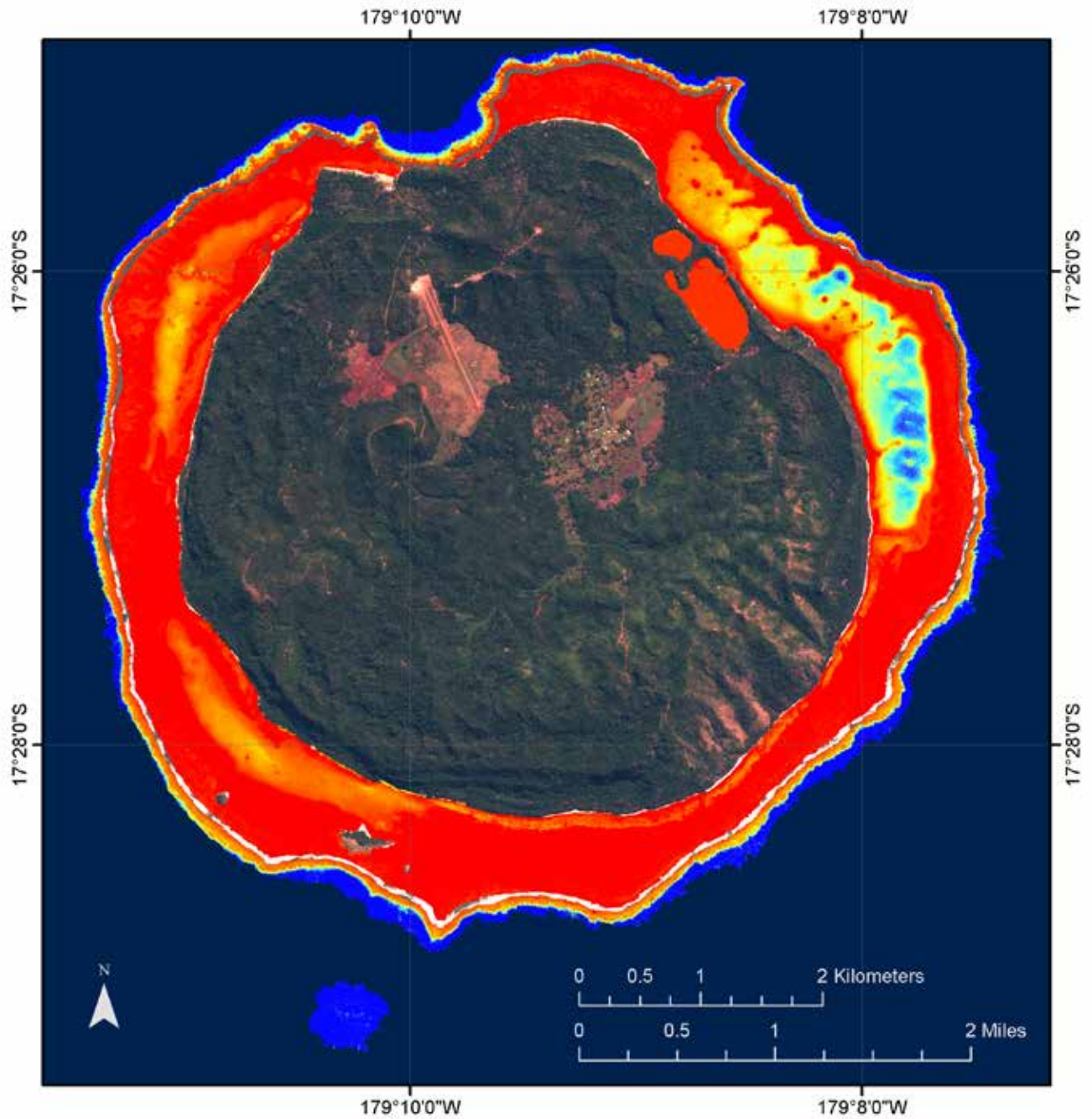


Fig. 59. WorldView-2 Satellite imagery for Mago.

Mago Island - Bathymetric Map

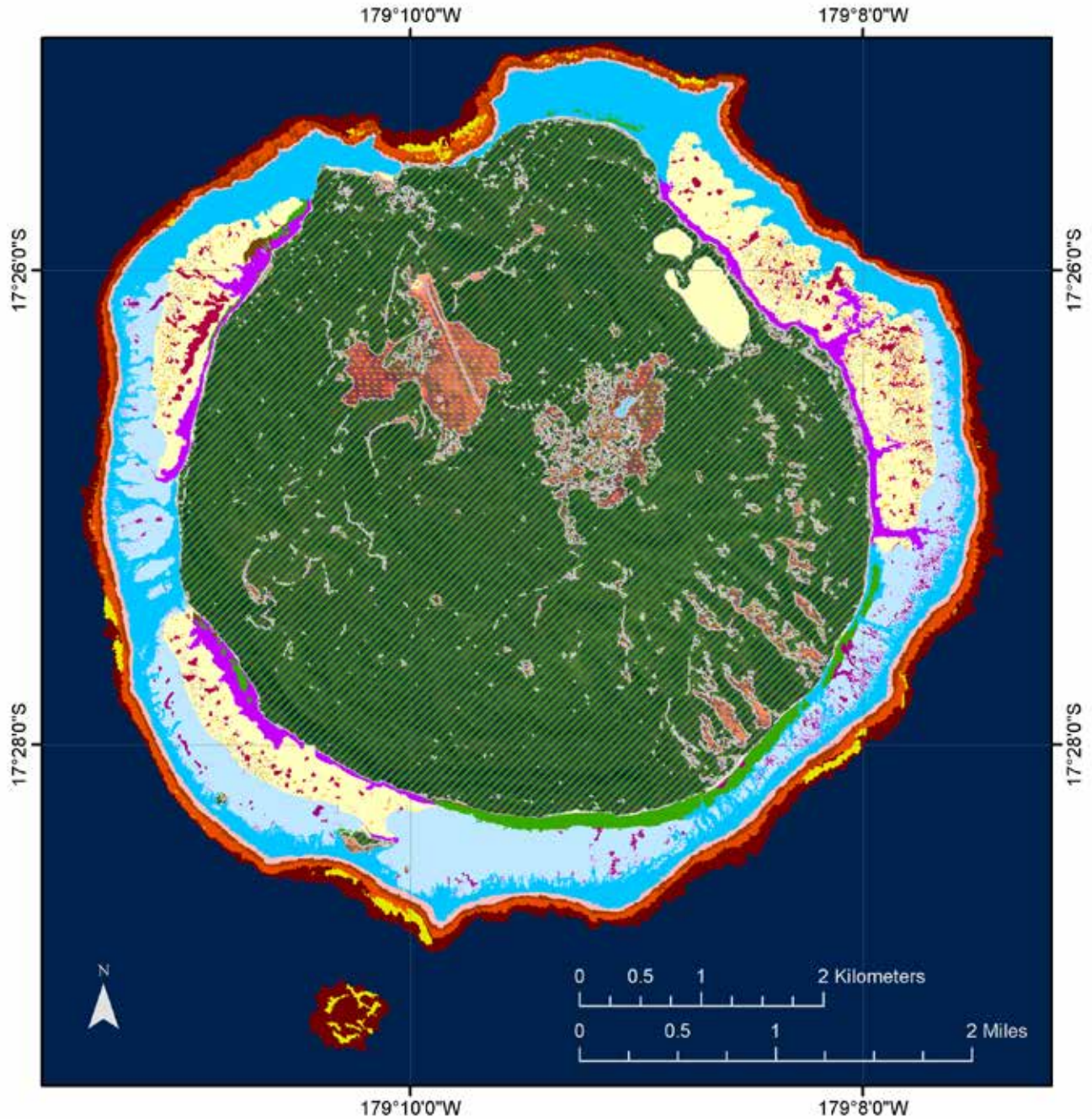


Water Depth (m)

0.30 - 1.00	8.01 - 9.00	16.01 - 17.00	24.01 - 25.00
1.01 - 2.00	9.01 - 10.00	17.01 - 18.00	25.01 - 26.00
2.01 - 3.00	10.01 - 11.00	18.01 - 19.00	26.01 - 27.00
3.01 - 4.00	11.01 - 12.00	19.01 - 20.00	27.01 - 28.00
4.01 - 5.00	12.01 - 13.00	20.01 - 21.00	28.01 - 29.00
5.01 - 6.00	13.01 - 14.00	21.01 - 22.00	29.01 - 30.00
6.01 - 7.00	14.01 - 15.00	22.01 - 23.00	Deep water
7.01 - 8.00	15.01 - 16.00	23.01 - 24.00	

Fig. 60. Bathymetric map for Mago.

Mago Island - Habitat Map



Habitats

- | | | |
|------------------------------|---|------------------------------|
| Deep fore reef slope | Back reef coral framework | Dense macroalgae on sediment |
| Shallow fore reef slope | Back reef coral bommies | Deep ocean water |
| Shallow fore reef terrace | Lagoonal floor barren | Terrestrial vegetation |
| Fore reef sand flats | Lagoonal pinnacle reefs massive coral dominated | Beach sand |
| Coralline algal ridge | Lagoonal patch reefs | Inland waters |
| Back reef rubble dominated | Lagoonal floor coral bommies | Urban |
| Back reef sediment dominated | Lagoonal fringing reefs | Unvegetated terrestrial |
| Back reef pavement | Dense seagrass meadows | |

Fig. 61. Habitat map for Mago.

Vanua Balavu Island Group - Satellite Map

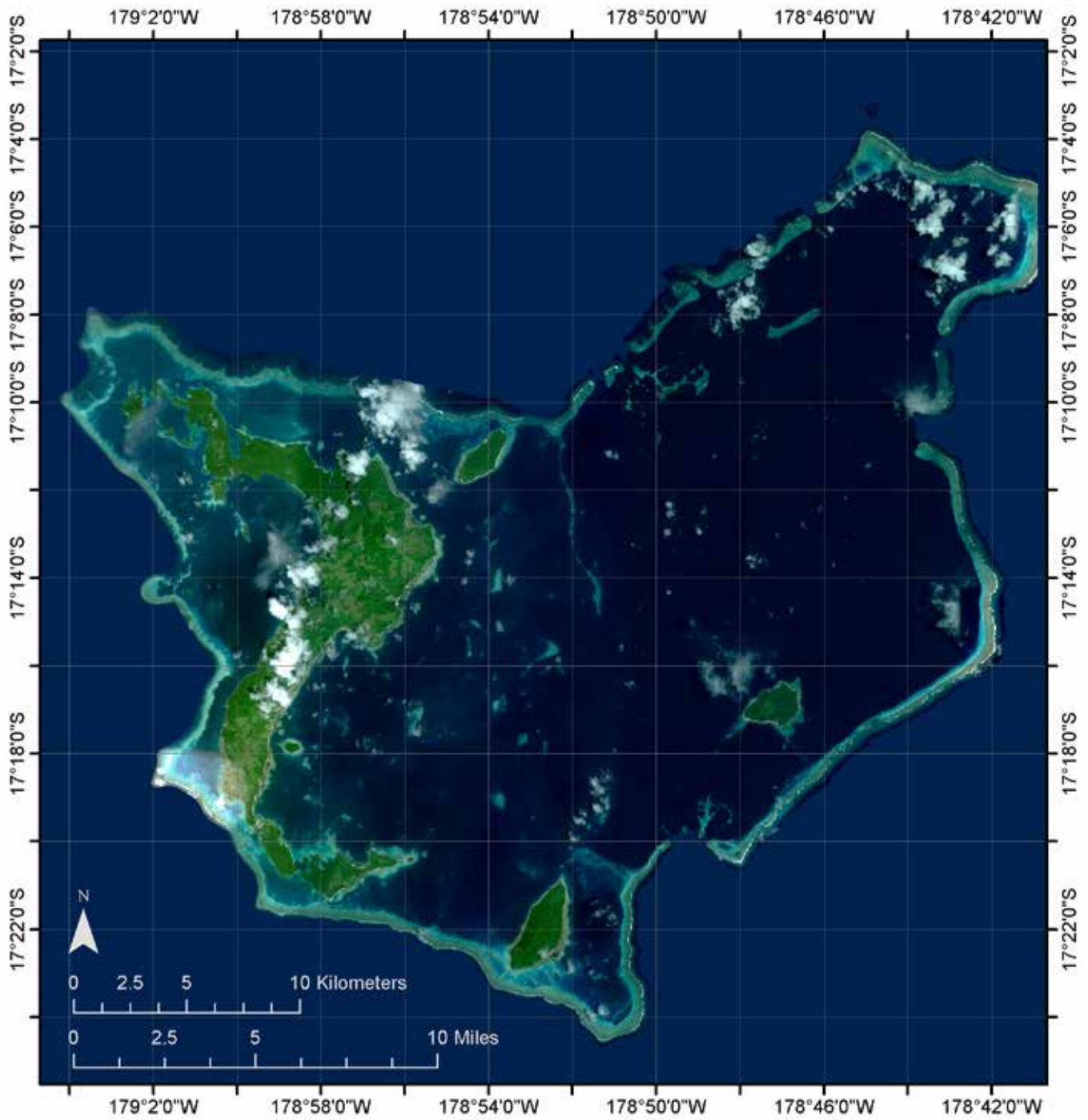
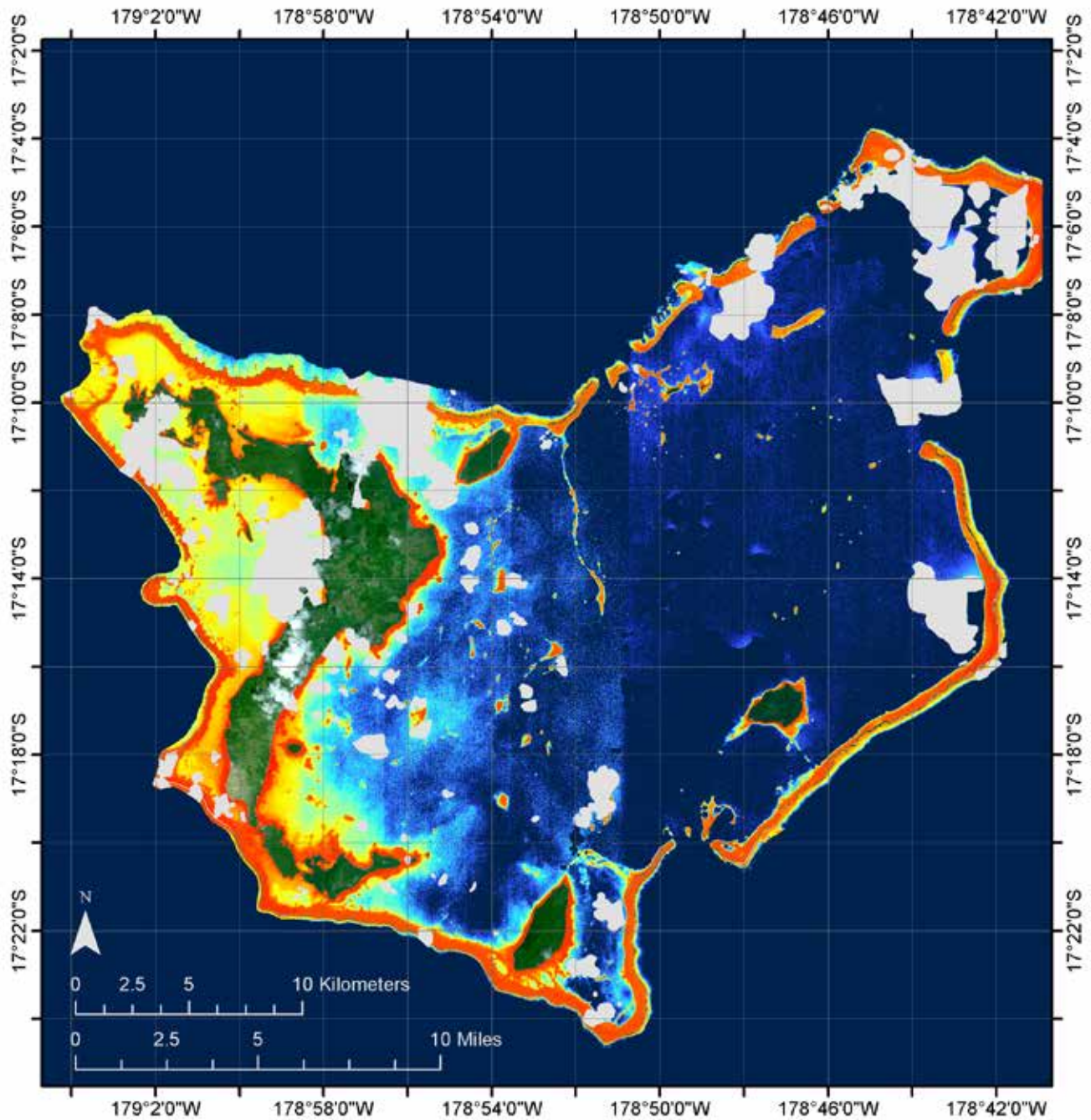


Fig. 62. WorldView-2 Satellite imagery for Vanua Balavu.

Vanua Balavu Island Group - Bathymetric Map

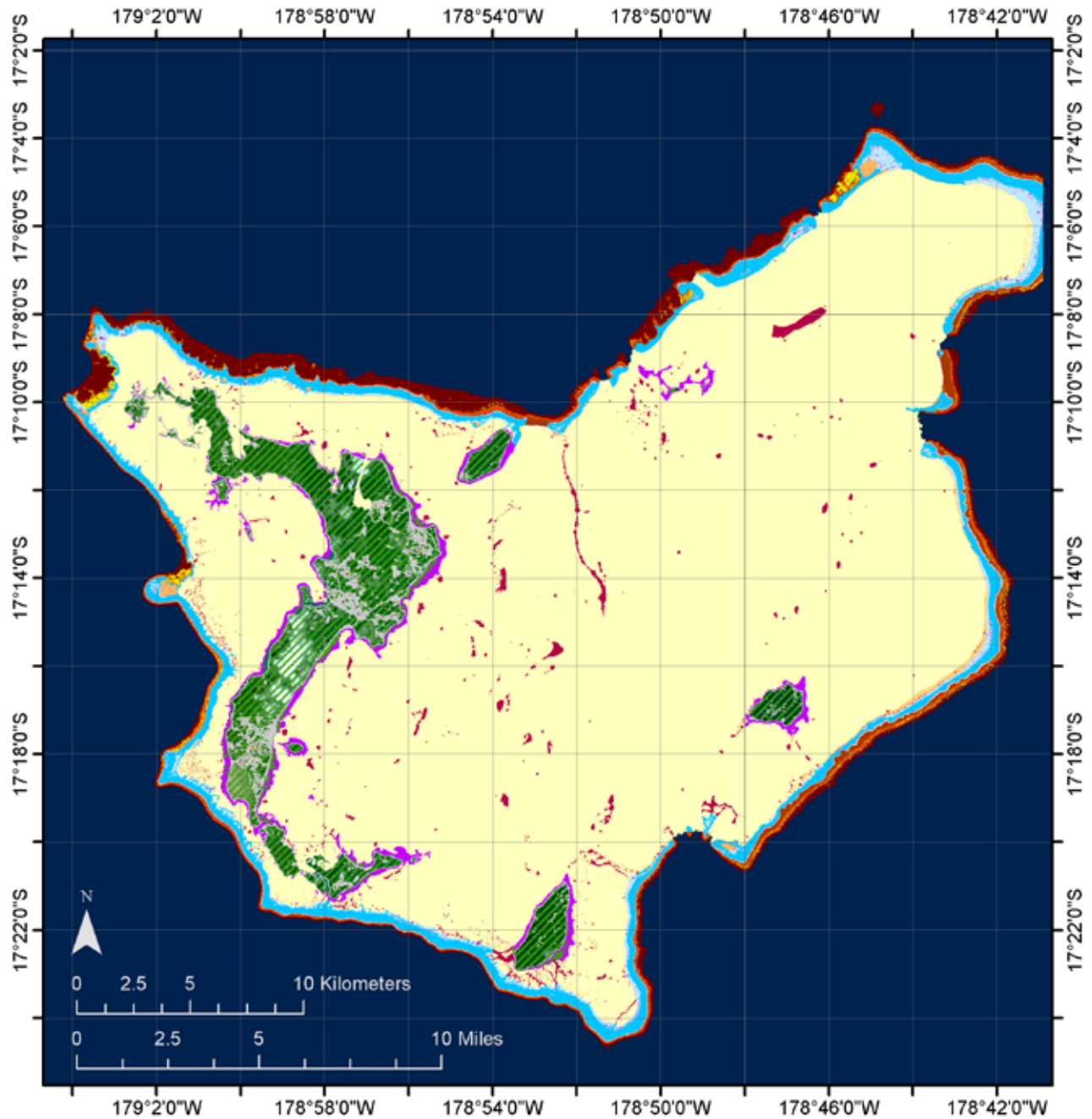


Water Depth (m)



Fig. 63. Bathymetric map for Vanua Balavu.

Vanua Balavu Island Group - Habitat Map



Habitats

- | | | |
|------------------------------|---|-------------------------|
| Deep fore reef slope | Back reef coral bommies | Mangroves |
| Shallow fore reef slope | Lagoonal sediment apron sediment dominated | Mud flats |
| Shallow fore reef terrace | Lagoonal floor barren | Deep ocean water |
| Fore reef sand flats | Lagoonal floor macroalgae on sediment | Terrestrial vegetation |
| Coralline algal ridge | Lagoonal pinnacle reefs massive coral dominated | Beach sand |
| Back reef rubble dominated | Lagoonal patch reefs | Inland waters |
| Back reef sediment dominated | Lagoonal floor coral bommies | Urban |
| Back reef pavement | Lagoonal fringing reefs | Unvegetated terrestrial |
| Back reef coral framework | Dense seagrass meadows | |

Fig. 64. Habitat map for Vanua Balavu.

Coral Reef Surveys

A combination of quantitative methods including belt transects, point intercept transects, radial plots and quadrats were used to assess corals, fish, and benthic cover of reefs in Lau Province, Fiji.

Benthic Cover

Cover of major functional groups (corals identified to genus, other invertebrates identified to phylum or class, and six groups of algae including macroalgae, crustose coralline algae, erect coralline algae, fine turfs, turf algae with sediment, and cyanobacteria) and substrate type (hardground, sand, loose rubble, fused rubble, recently dead coral, bleached coral, and live coral) were assessed along 10 m transects using recorded observations and/or photographic assessments. Recorded observations involved a point intercept method, whereas the organism and substrate was identified every 10 cm along a 10 m transect (total 100 points/transect), with a minimum of six transects examined per location. When possible, surveys were completed at 30, 25, 20, 15, 10 and 5 m depth.



Fig. 65. A diver conducting a benthic assessment. A ten meter lead line, subdivided into 10 cm intervals is deployed along the bottom. The diver records the substrate type and organism under each 10 cm point.

Coral Assessments

Five measurements were recorded for corals: 1) benthic cover (point intercept, see above); 2) coral diversity and abundance (by genus, except certain common species); 3) coral size class distributions; 4) recruitment; and 5) coral condition. Additional

information was collected on causes of recent mortality, including signs of coral disease and predation.



Fig. 66. A diver conducting a coral assessment. A 10 m lead line is deployed and all corals within 1 m of the line are identified and measured. A 1 m PVC pipe, is used to determine the proper width of the line and to aid in measurement of the corals.

Assessment of corals smaller than 4 cm was achieved by using a minimum of five 0.25 m² quadrats per transect, with each quadrat located at fixed, predetermined intervals, alternating between the right and left side of the transect line. Recruits were identified in both point intercept surveys and belt transects. Recruits were divided into two categories: corals up to 2 cm in diameter and larger corals, 2-4 cm in diameter.



Fig. 67. Assessing recruits within a 25 X 25 cm quadrat.
©Keith Ellenbogen/iLCP

Coral population structure and condition was assessed within belt transects (each 10 m x 1 m), with a minimum of two transects completed per depth. Each coral 4 cm or larger was identified (to genus at minimum) and its growth form was recorded. Visual estimates of tissue loss were recorded for each colony over 4 cm in diameter using a 1 m bar marked in 1 cm increments for scale. If the coral exhibited tissue loss, estimates of the amount of remaining tissue, percent that recently died and percent that died long ago were made based on the entire colony surface. Tissue loss was categorized as recent mortality (occurring within the last 1-5 days), transitional mortality (filamentous green algae and diatom colonization, 6-30 days) and old mortality (>30 days).



Fig. 68. A small section of reef substrate (approximately 15 cm x 20 cm) illustrating patterns of colonization. The substrate is colonized by turf algae, crustose coralline algae, macroalgae, cyanobacteria and four coral recruits (red circles).

For each coral with partial or whole colony mortality, the cause of mortality is identified when possible. The diagnosis included an assessment of the type of disease, extent of bleaching, predation, competition, overgrowth or other cause of mortality. Each coral was first carefully examined to identify cryptic predators. Lesions were initially diagnosed into four categories: recent tissue loss, skeletal damage, color change, and unusual growth patterns; an individual colony could have multiple characteristics (e.g. color change and recent tissue loss). The location (apical, basal, and medial) and pattern of tissue loss (linear, annular,

focal, multifocal, and coalescing) was recorded. If an outbreak of coral disease was documented, sampling of the affected corals was undertaken to further characterize the disease (see Figure 69).

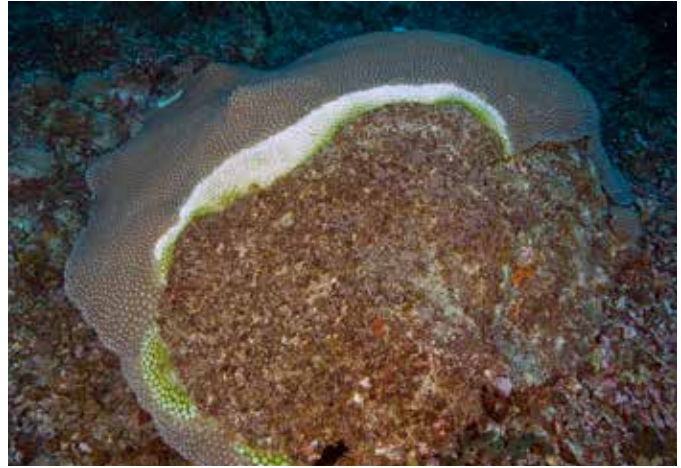


Fig. 69. A colony of *Diploastrea* with white syndrome. The coral is about 30% live, with a narrow band of exposed skeleton (recently mortality) and a larger patch of long dead algal colonized skeleton.

Fish Assessments

Fish abundance and size structure was collected for over 450 species of fishes (Appendix 1), with an emphasis on species that have a major functional role on reefs or are major fisheries targets. Reef fishes were assessed along 4 m X 30 m belt transects. A T-square marked in 5 cm increments was used to gauge fish size. A minimum of two transects per depth category and six transects per site were conducted. A roving survey was also completed to document additional species that were not seen within belt transects.



Fig. 70. A diver conducting a fish survey. The transect tape is attached to the divers BC and the end is secured to the bottom to gauge 30 m distance.

Biomass was determined by estimating the body mass of each individual fish using length-weight relationships ($W=aL^b$). Constants (a & b) were obtained from Fishbase (www.fishbase.org). Each fish species was grouped into one of seven functional guilds (benthic carnivores, corallivores, herbivores, piscivores, planktivores, omnivores or other), a fisheries category (targeted, target, or indicator species), and species of major or minor ecological significance using published literature (Rotjan and Lewis, 2008; Barneche et al., 2009; Bruckner and Bruckner, 2015; Fishbase). Mean values for fish biomass and density were calculated for each island, genus, and functional guild. To examine the size structure of coral reef fish communities, fish were pooled into eight size class bins.

Photographic Assessment

Belt transects, 1 m x 10 m were photographed along depth gradients to supplement in situ recorded benthic and coral data. One scientist extended a 10 m long lead line along depth contours at 30, 20, 15, 10 and 5 m depth. The line was marked with cable ties at 10 cm intervals. Continuous digital still photographs were taken of the reef substrate from a height of approximately 0.6-0.75 meters above the substrate, using a one meter bar divided into 5 cm increments placed perpendicular to the transect tape as a scale bar. Approximately 20 photographs were taken per transect to allow for overlap between adjacent images with two photo transects (each 10 m in length) per depth. A second scientist used a 1 m x 1 m quadrat, flipping over the quadrat a total of 10 times per transect.



Fig. 71. A diver photographing a 1 m X 1 m quadrat.

Images were downloaded and analyzed using Coral Point Count (CPCe) software developed by the National Coral Reef Institute (NCRI). To determine benthic community composition, coral cover, algae cover, and cover of different substrate types, a total of 50 points were randomly placed on each 1 m x 1 m image and the attribute directly below the point was recorded. The coral community structure was also assessed within the same transects to determine the size (planar surface area), amount of partial mortality and condition. Planar surface area was measured by tracing the outline of individual corals.

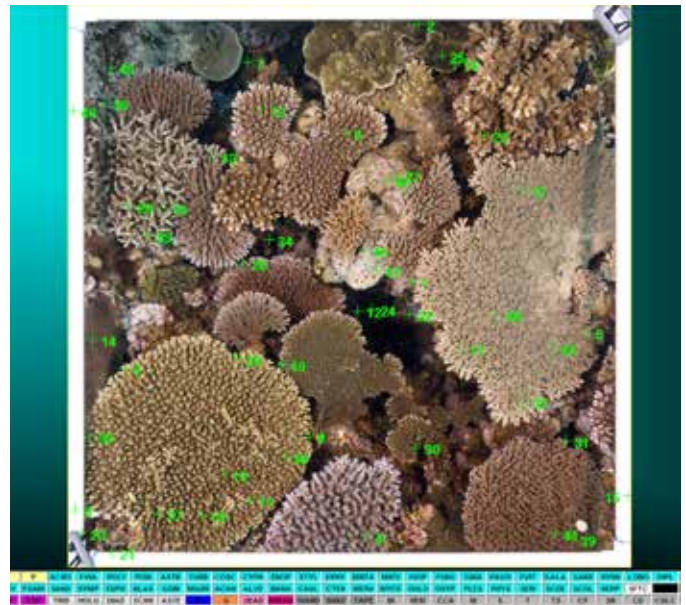


Fig. 72. Example of one 1 m X 1 m quadrat photographed on a reef in Fiji and imported into CPCe. 50 random points have been placed on the image.

Date	Lat	Long	Site Name	Island	Reef Zone	Reef Type	Exposure
3-Jun-13	-18.996	-179.9032	FJTO01	Totoya	back reef	barrier reef	leeward
3-Jun-13	-18.9728	-179.9068	FJTO02	Totoya	fore reef	barrier reef	leeward
3-Jun-13	-18.9273	-179.8907	FJTO03	Totoya	fore reef	barrier reef	leeward
4-Jun-13	-18.8886	-179.8677	FJTO04	Totoya	fore reef	barrier reef	windward
4-Jun-13	-18.8981	-179.8836	FJTO05	Totoya	fore reef	barrier reef	leeward
4-Jun-13	-18.9976	-179.8473	FJTO06	Totoya	channel	patch reef	currents/swell
5-Jun-13	-19.0032	-179.8485	FJTO07	Totoya	fore reef	barrier reef	leeward
5-Jun-13	-19.023	-179.8808	FJTO08	Totoya	fore reef	barrier reef	leeward
5-Jun-13	-18.9082	-179.7864	FJTO09	Totoya	fore reef	barrier reef	windward
6-Jun-13	-19.1178	179.7382	FJMT10	Matuka	fore reef	barrier reef	leeward
6-Jun-13	-19.1585	179.7304	FJMT11	Matuka	fore reef	barrier reef	leeward
6-Jun-13	-19.1534	179.7401	FJMT12	Matuka	back reef	patch reef	leeward
7-Jun-13	-19.1172	179.7783	FJMT13	Matuka	fore reef	barrier reef	windward
7-Jun-13	-19.129	179.7866	FJMT14	Matuka	fore reef	barrier reef	windward
8-Jun-13	-18.5919	179.97308	FJML15	Moala	fore reef	barrier reef	windward
8-Jun-13	-18.5204	179.96562	FJML16	Moala	fore reef	barrier reef	leeward
8-Jun-13	-18.5325	179.92	FJML17	Moala	fore reef	patch reef	leeward
9-Jun-13	-18.5461	179.9013	FJML18	Moala	fore reef	patch reef	leeward
9-Jun-13	-18.5794	179.8201	FJML19	Moala	fore reef	barrier reef	leeward
9-Jun-13	-18.5577	179.8785	FJML20	Moala	fore reef	barrier reef	leeward
10-Jun-13	-18.5972	179.9337	FJML21	Moala	back reef	patch reef	leeward
10-Jun-13	-18.6168	179.9389	FJML22	Moala	fore reef	barrier reef	windward
10-Jun-13	-18.5575	179.9851	FJML23	Moala	fore reef	barrier reef	windward
11-Jun-13	-19.124	-178.548	FJFU24	Fulaga	lagoonal	back reef	leeward
11-Jun-13	-19.094	-178.5809	FJFU25	Fulaga	fore reef	barrier reef	windward
11-Jun-13	-19.1011	-178.6011	FJFU26	Fulaga	fore reef	barrier reef	leeward
12-Jun-13	-19.1299	-178.6174	FJFU27	Fulaga	fore reef	barrier reef	leeward
12-Jun-13	-19.1411	-178.5706	FJFU28	Fulaga	lagoonal	patch reef	leeward
12-Jun-13	-19.1184	-178.5918	FJFU29	Fulaga	lagoonal	patch reef	leeward
13-Jun-13	-18.9414	-178.9847	FJKA30	Kabara	fore reef	barrier reef	leeward
13-Jun-13	-18.9194	-178.9577	FJKA31	Kabara	fore reef	pinnacles	leeward
13-Jun-13	-18.9136	-178.9455	FJKA32	Kabara	fore reef	barrier reef	windward
14-Jun-13	-18.9545	-178.9874	FJKA33	Kabara	fore reef	barrier reef	leeward
14-Jun-13	-18.9228	-178.9363	FJKA34	Kabara	fore reef	barrier reef	windward
15-Jun-13	-18.3864	-179.2786	FJVV35	Vanua Vatu	fore reef	barrier reef	swellward
15-Jun-13	-18.3439	-179.2803	FJVV36	Vanua Vatu	fore reef	barrier reef	leeward
15-Jun-13	-18.3584	-179.2847	FJVV37	Vanua Vatu	fore reef	barrier reef	leeward
16-Jun-13	-17.9512	-179.067	FJNA38	Nayau	fore reef	barrier reef	leeward
16-Jun-13	-17.9569	-179.0723	FJNA38SN	Nayau	fore reef	reef flat	leeward
16-Jun-13	-17.9759	-179.0767	FJNA39	Nayau	fore reef	fringing reef	leeward
16-Jun-13	-17.9651	-179.0789	FJNA40	Nayau	fore reef	barrier reef	leeward
17-Jun-13	-17.6498	-178.8354	FJTV41	Tuvuca	fore reef	barrier reef	leeward
17-Jun-13	-17.7041	-178.8291	FJTV42	Tuvuca	fore reef	barrier reef	leeward
17-Jun-13	-17.6935	-178.8325	FJTV43	Tuvuca	fore reef	barrier reef	leeward
18-Jun-13	-17.7167	-179.3243	FJCC44	Cicia	fore reef	barrier reef	leeward
18-Jun-13	-17.7238	-179.3386	FJCC45	Cicia	fore reef	barrier reef	leeward
18-Jun-13	-17.7265	-179.3408	FJCC46	Cicia	fore reef	barrier reef	leeward

Table 6. Coordinates, reef zone, reef type and exposure for survey locations examined in Lau Province, Fiji.

Date	Lat	Long	Site Name	Island	Reef Zone	Reef Type	Exposure
19-Jun-13	-17.7671	-179.3491	FJCC47	Cicia	fore reef	barrier reef	windward
19-Jun-13	-17.7498	-179.3841	FJCC48	Cicia	fore reef	barrier reef	leeward
20-Jun-13	-17.4785	-179.1672	FJMG49	Mago	fore reef	barrier reef	windward
20-Jun-13	-17.4639	-179.1877	FJMG50	Mago	fore reef	barrier reef	leeward
20-Jun-13	-17.4249	-179.1655	FJMG51	Mago	fore reef	fringing reef	leeward
21-Jun-13	-17.3028	-179.0309	FJVB52	Vanua Balavu	fore reef	barrier reef	leeward
21-Jun-13	-17.1394	-179.0600	FJVB53	Vanua Balavu	fore reef	barrier reef	leeward
21-Jun-13	-17.2376	-179.0386	FJVB54	Vanua Balavu	fore reef	barrier reef	leeward
22-Jun-13	-17.1534	-179.0049	FJVB55	Vanua Balavu	fore reef	barrier reef	leeward
22-Jun-13	-17.1395	-179.0599	FJVB56	Vanua Balavu	fore reef	barrier reef	leeward
22-Jun-13	-17.1715	-178.8871	FJVB57	Vanua Balavu	fore reef	barrier reef	leeward
22-Jun-13	-17.196	-178.8707	FJVB58	Vanua Balavu	patch reef	lagoonal	leeward
22-Jun-13	-17.2419	-178.8556	FJVB59	Vanua Balavu	patch reef	lagoonal	leeward
23-Jun-13	-17.1206	-178.8265	FJVB60	Vanua Balavu	fore reef	barrier reef	leeward
23-Jun-13	-17.1518	-178.8512	FJVB61	Vanua Balavu	fore reef	barrier reef	leeward
23-Jun-13	-17.2824	-178.9267	FJVB62	Vanua Balavu	patch reef	lagoonal	leeward
24-Jun-13	-17.2923	-178.8856	FJVB63	Vanua Balavu	patch reef	lagoonal	leeward
24-Jun-13	-17.2877	-178.9281	FJVB62B	Vanua Balavu	patch reef	lagoonal	leeward
24-Jun-13	-17.3364	-178.86	FJVB64	Vanua Balavu	linear reef	lagoonal	leeward
25-Jun-13	-17.3355	-178.8337	FJVB65	Vanua Balavu	back reef	barrier reef	windward
25-Jun-13	-17.3234	-178.8167	FJVB66	Vanua Balavu	patch reef	lagoonal	leeward
25-Jun-13	-17.2709	-178.7774	FJVB67	Vanua Balavu	back reef	fringing reef	windward
26-Jun-13	-17.1066	-178.6897	FJVB68	Vanua Balavu	patch reef	lagoonal	leeward
26-Jun-13	-17.0895	-178.7692	FJVB69	Vanua Balavu	fore reef	barrier reef	windward
26-Jun-13	-17.1352	-178.7775	FJVB70	Vanua Balava	patch reef	lagoonal	leeward

Table 6. Coordinates, reef zone, reef type and exposure for survey locations examined in Lau Province, Fiji (cont.).

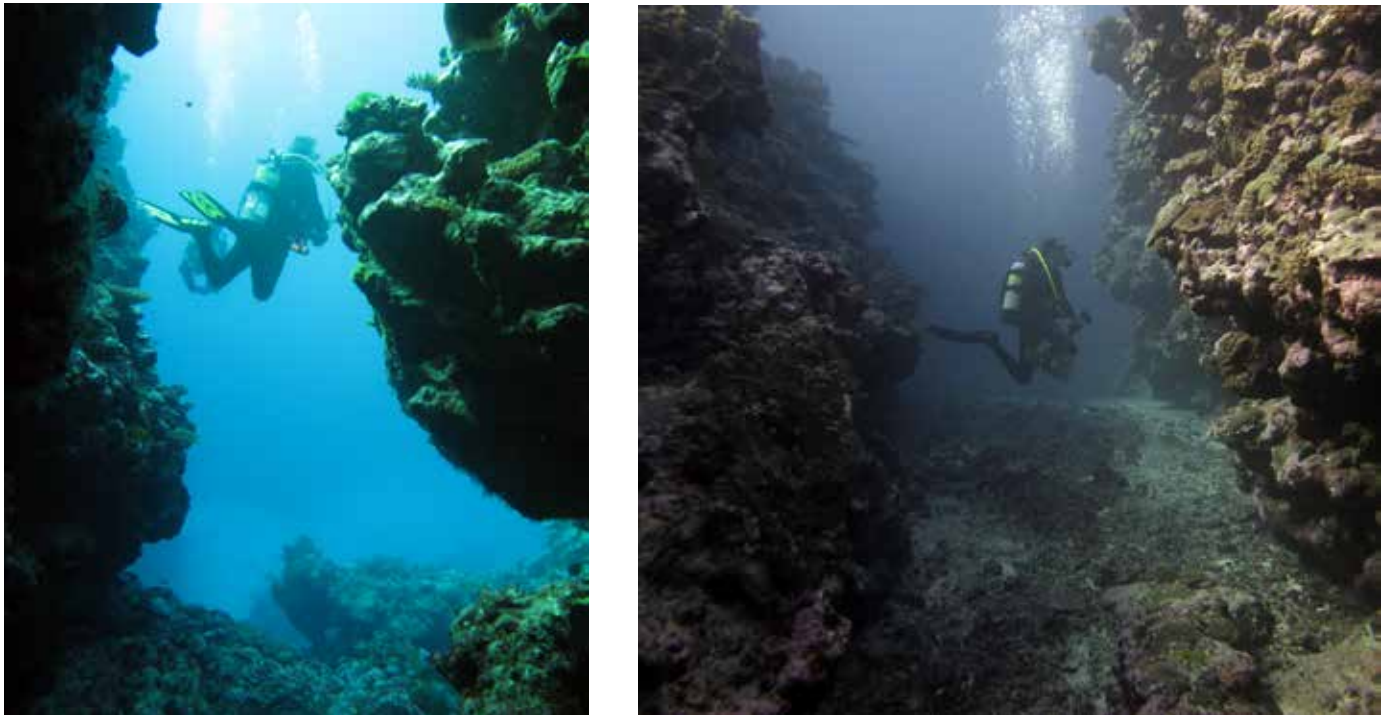


Fig. 73. A number of reefs had vertical drop-offs, overhangs, caves and canyons such as the two reefs shown here. Surveys were not conducted on vertical surfaces. Images by Anderson Mayfield.

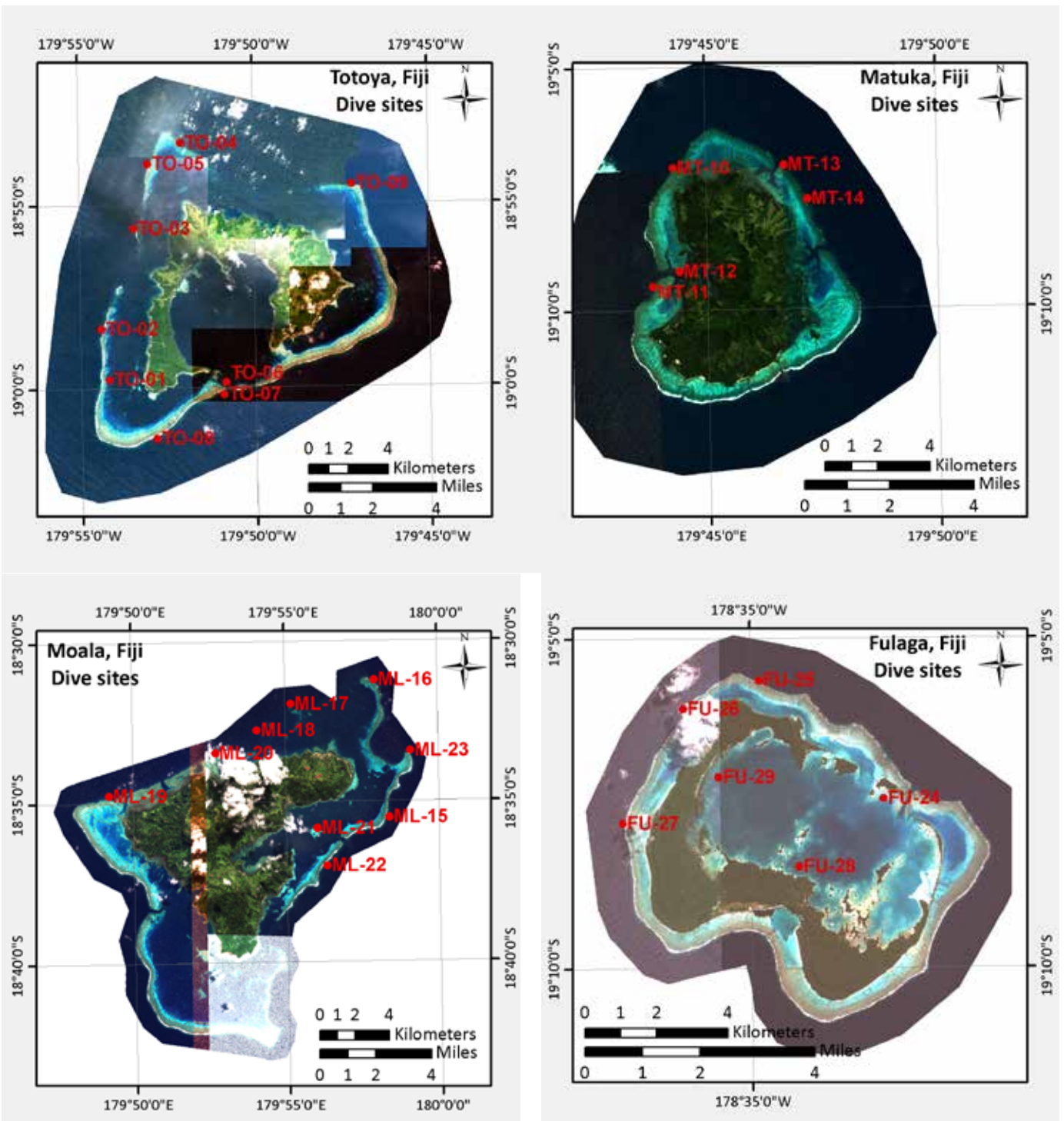


Fig. 74. Location of SCUBA assessments off Totoya, Matuka, Moala and Fulaga, Fiji.

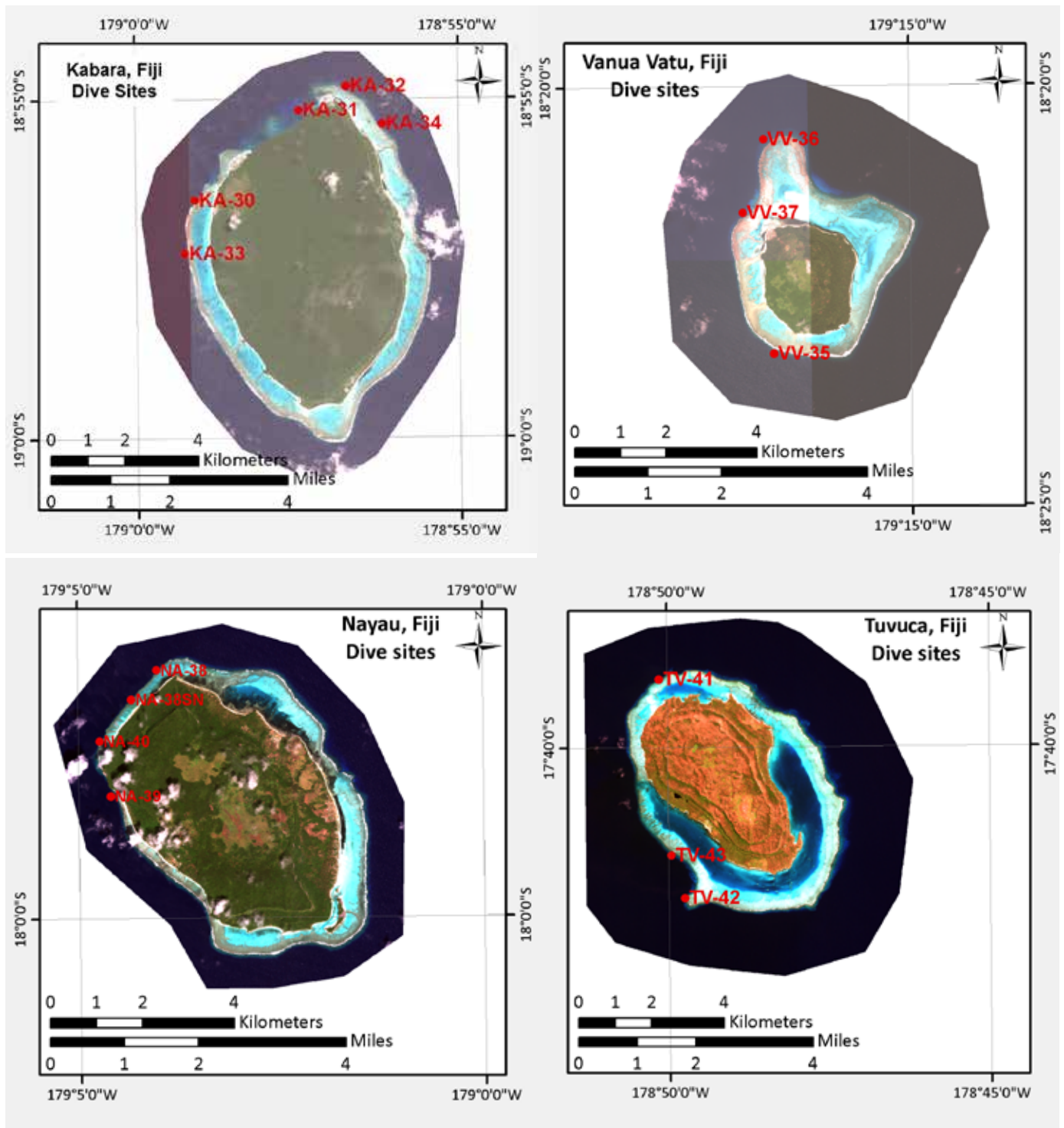


Fig. 75. Location of SCUBA assessments for Kabara, Vanua Vatu, Nayau and Tuvuca, Fiji.

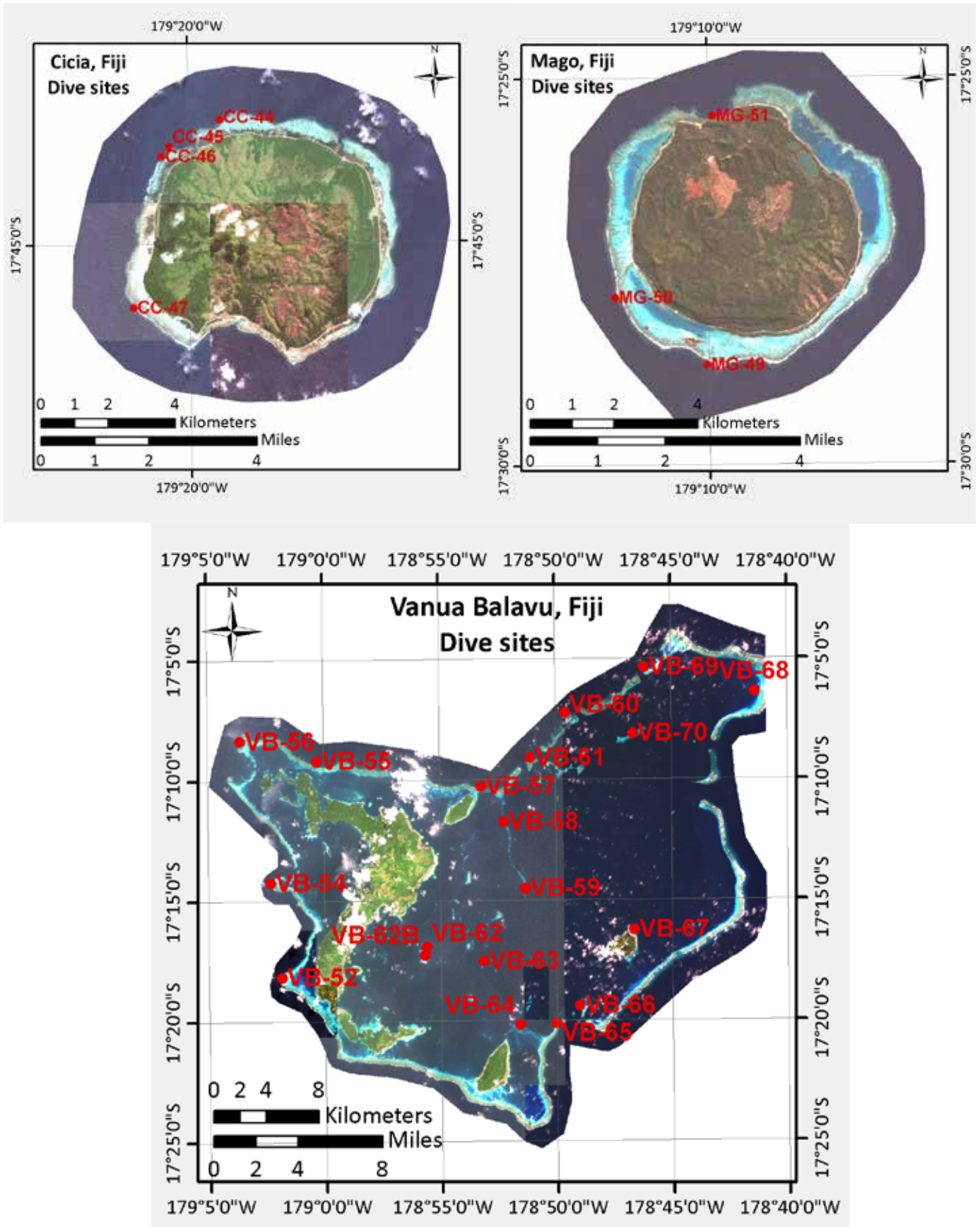


Fig. 76. Location of SCUBA assessments for Cicia, Mago and Vanua Balavu, Fiji.

Substrate Composition and Cover

During benthic surveys, substrates in coral reef environments were classified as pavement (hard bottom), live coral, dead coral, fused rubble, loose rubble and sand. The majority of the substrate was pavement (41-61% cover) and live coral (22-50% cover). Benthic cover of rubble (predominantly loose, unconsolidated rubble) ranged from 2.8-11% with the highest amount of rubble at Tuvuca (11.2%), Vanua Balavu (8.6%), Matuka (6.8%) and Totoya (5.9%). Dead coral covered approximately 5% of the substrate with the lowest cover at Vanua

Vatu (1.4%) and Kabara (2.3%) (Fig. 77).

An examination of substrate data, pooled by depth for all sites and islands suggests that the percent cover of different substrate types was very similar among depths (Fig. 78). However, closer examination by island suggests that there is variation between depths, and also among sites within an island and between depths at an individual site (Fig 79).

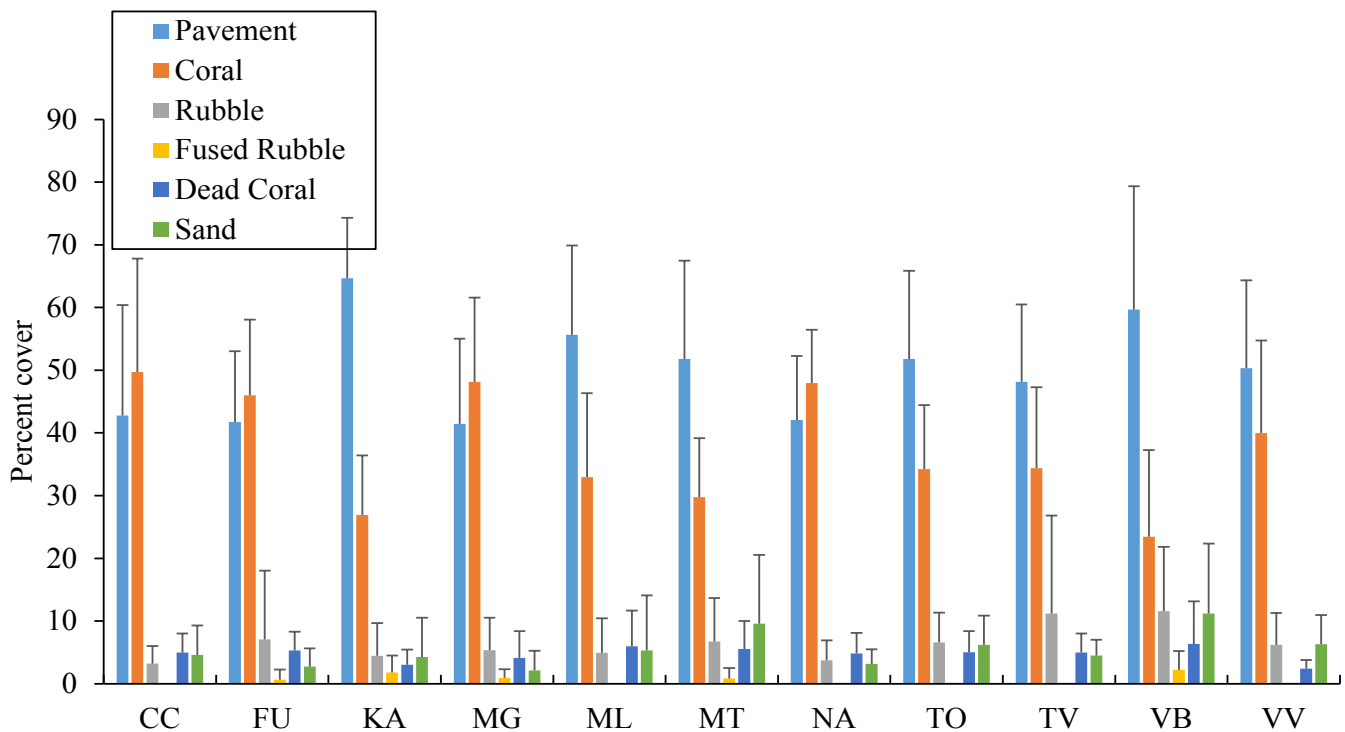


Fig. 77 Cover of different benthic substrates within reef habitats off 11 islands in Lau Province, Fiji. Islands include: Cicia (CC), Fulaga (FU), Kabara (KA), Mago (MG), Moala (ML), Matuka (MT), Nayau (NA), Totoya (TO) Tuvuca (TV), Vanua Balavu (VB) and Vanua Vatu (VV). Substrate types were subdivided into pavement (hard bottom), loose rubble, fused rubble, sand, live coral and dead coral. Data are pooled by island for all depths and reefs.

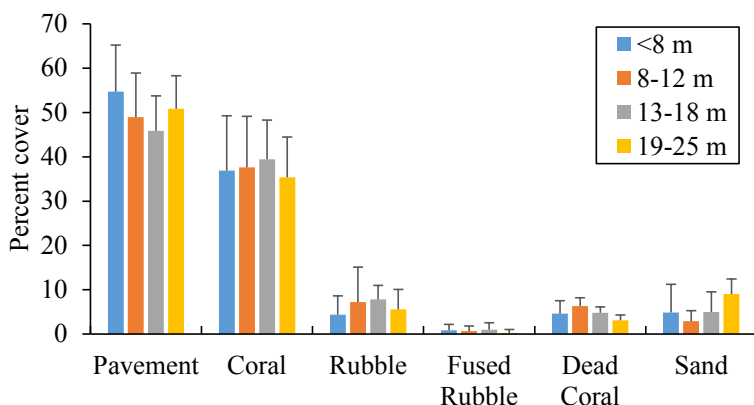
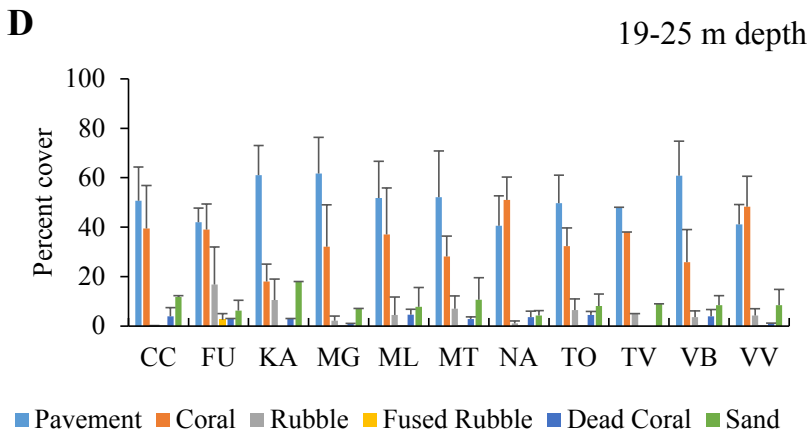
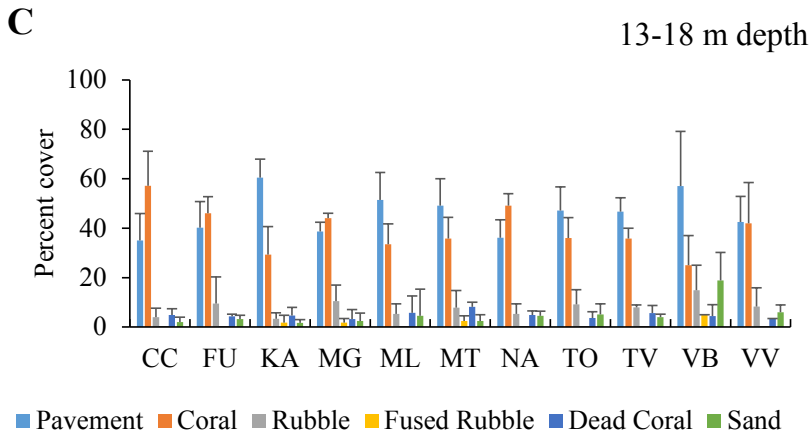
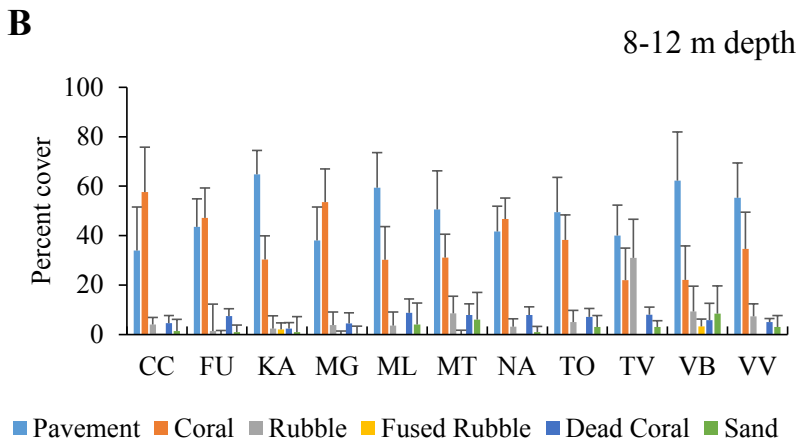
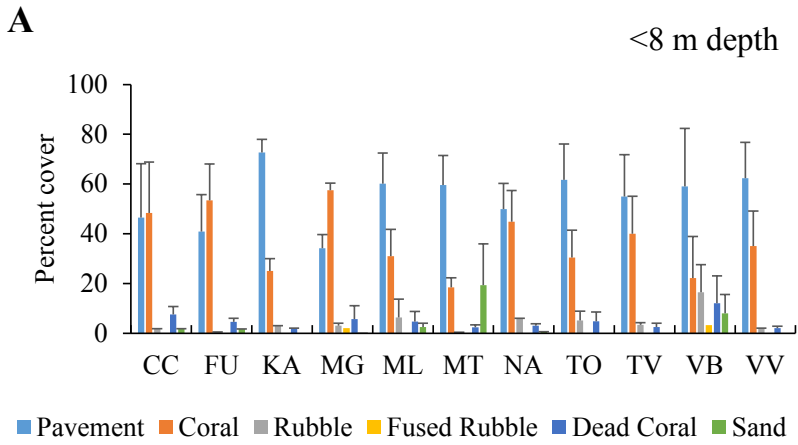


Fig. 78. Cover of different benthic substrates within reef habitats pooled for the 11 islands in Lau Province, Fiji into four depth classes, <8 m, 8-12 m, 13-18 m and 19-25 m.



Of the six main substrate categories, pavement and live coral cover varied most among islands. The lowest mean coral cover and highest cover of pavement was recorded at Kabara (27% and 65% respectively) and Vanua Balavu (23% and 60%). Coral cover was highest at the shallowest survey sites (<8 m) at three islands (Fulaga, Mago, and Tuvuca), highest at mid depths (8-18 m) at Totoya, Matuka, Kabara and Cicia, and highest at the deepest depths (19-25 m) at Vanua Balavu, Vanua Vatu, Nayau, and Moala (Fig. 79).

Fig. 79 (graphs A-D on the left). Cover of different benthic substrates within reef habitats off 11 islands in Lau Province, Fiji broken down by the four depth classes, <8 m, 8-12 m, 13-18 m and 19-25 m. Islands listed in the figures include: Cicia (CC), Fulaga (FU), Kabara (KA), Mago (MG), Moala (ML), Nayau (NA), Totoya (TO) Tuvuca (TV), Vanua Balavu (VB) and Vanua Vatu (VV).



Fig. 80. Many reefs had high cover of corals dominated by small to medium-sized table acroporids, especially on the tops of pinnacles as seen in the above photograph

Live Cover

Knowing what organisms live on a reef (and their relative proportions) is a reliable indicator of the health of a coral reef. Four main categories of substrate cover were examined on reefs in Lau Province: coral, algae, other invertebrate and uncolonized substrate. Nearly 85% of the bottom was colonized by live organisms, while uncolonized substrate (15.8%) was predominantly small patches of sand between colonized pavement. Coral cover (36%) was slightly higher than algal cover on four islands, Cicia, Fulaga, Moala and Nayau. Algal cover ranged from 37-86%, with highest coverage at Vanua Balavu, Kabara, Vanua Vatu, Moala and Tuvuca, respectively. Other invertebrates made up a low portion of the total cover except at Matuka (8.7%) and Vanua Balavu (4.7%) (Fig. 82).

Algal cover varied considerably among sites, depths and habitats. Some reefs had very dense patches of macroalgae, especially *Halimeda spp.* and *Caulerpa spp.*, but this tended to be on vertical slopes and at the base of the reef. Algal cover was much less common on horizontal reef surfaces. The highest cover of macroalgae was recorded at Vanua Balavu (8.6%), Kabara (4.7%) and Fulaga (4.3%). All other sites had less than 3% cover of macroalgae.



Fig. 81. Macroalgae often covered dead coral and rubble at the base of the reef.

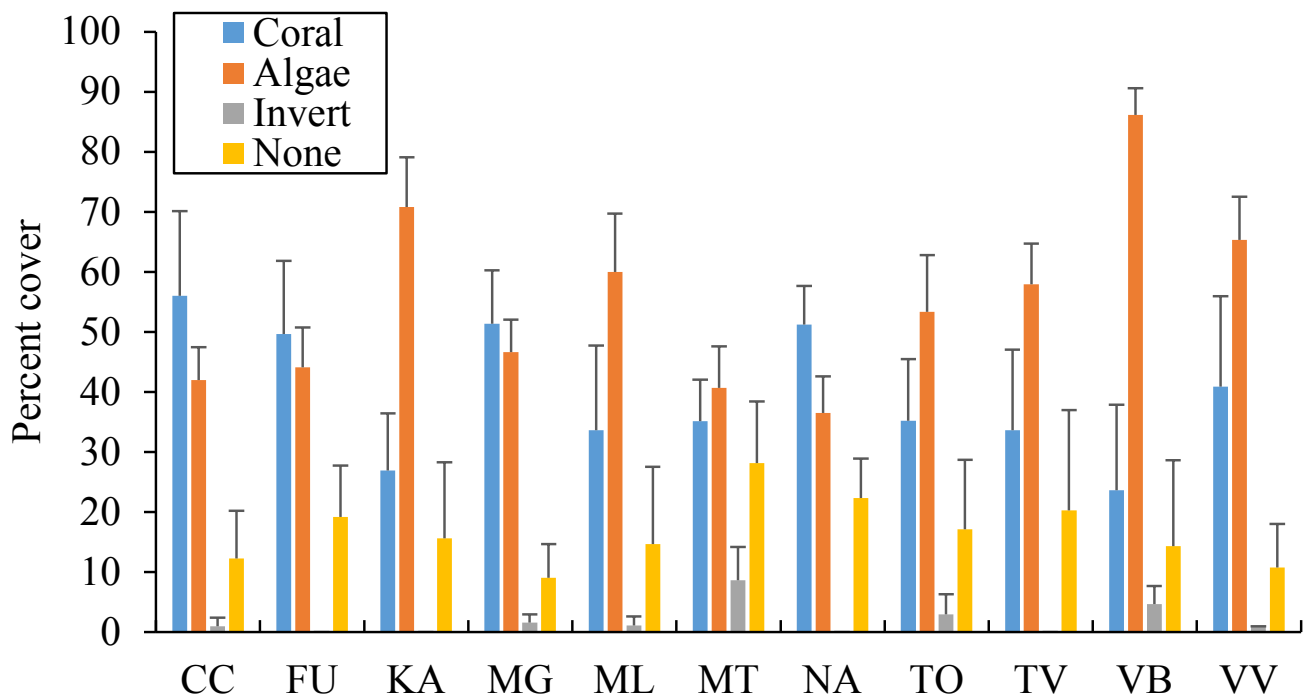
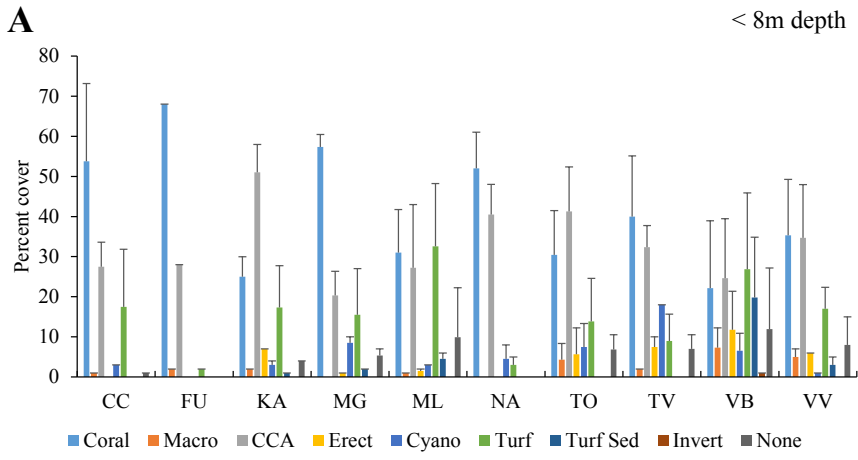
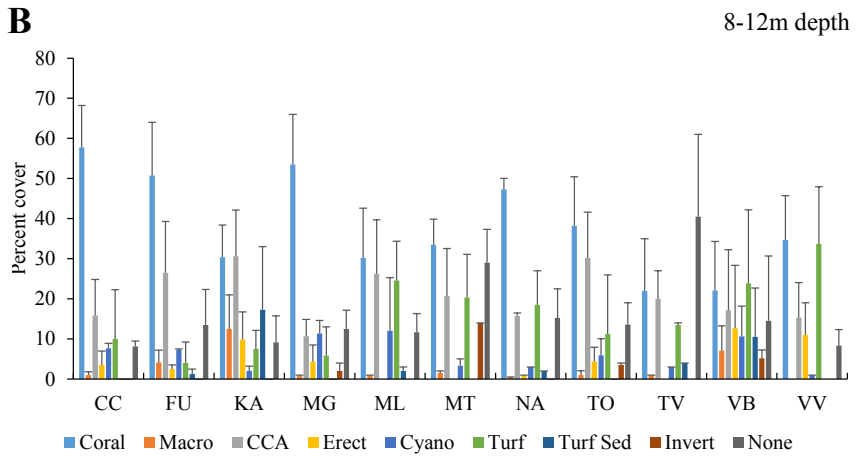


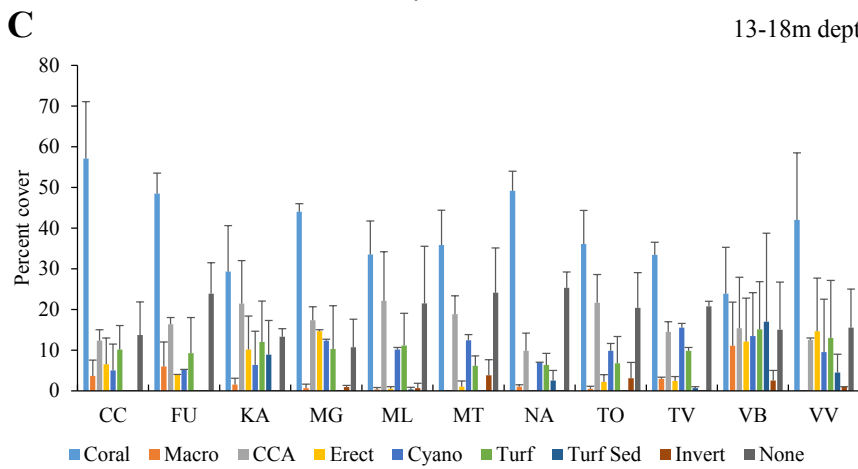
Fig. 82 Cover of living organisms and bare substrate at each of the 11 islands surveyed in Lau Province, Fiji.



Turf algae ranged from 3-22% cover, with the highest cover at Moala, Vanua Balavu and Vanua Vatu. High cover of turf algae with sediment was recorded at Kabara (10%) and Vanua Balavu (15%), possibly due to the large number of lagoonal sites examined in this area. Most reefs had high cover of CCA, with >25% cover at Kabara, Totoya and Moala, the lowest at Ciccia (16%) and Mago (15%).



Cyanobacterial mats were also a common feature (mean cover, all islands = 9.3%) with over 10% cover at Tuvuca, Mago, Moala and Vanua Balavu and over 5% at all other islands. Vanua Balavu, and Vanua Vatu also had a substantial amount of erect coralline algae (e.g. *Halimeda*) with >10% cover.



Within individual islands, the cover of macroalgae did not differ substantially between depths. In contrast, CCA was highest in shallow water, declining with depth. The cover of turf algae was also highest in shallow water, except at Fulaga. In contrast, the cover of cyanobacteria increased with depth in most locations (Fig 83).

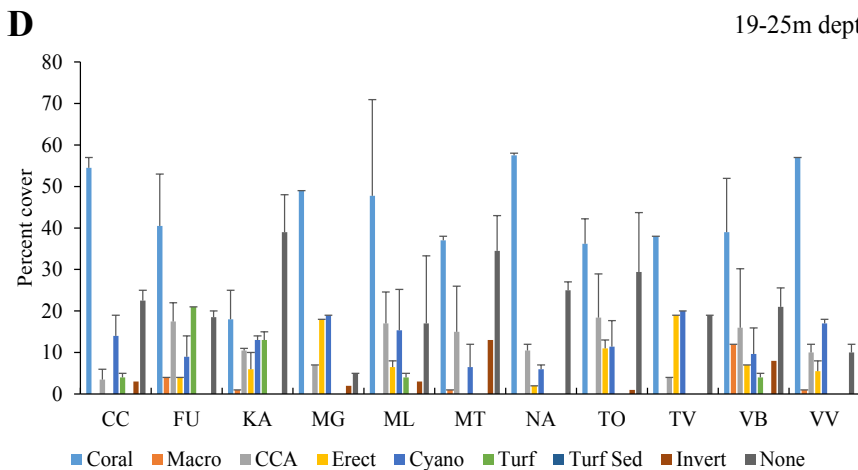


Fig. 83. Substrate cover at each of the 11 islands surveyed in the Lau Province, Fiji at different depth categories.

Coral Cover

A total of 45 genera of scleractinian corals and one hydrozoan coral were recorded within benthic transects. Overall, *Acropora* was the dominant coral on these reefs, making up 35% of the living coral cover.

Five functional groups of corals, *Acropora*, *Pocillopora*, *Porites*, *Montipora* and faviids (*Goniastrea*, *Montastraea*, *Favia*, *Favites* and

Cyphastrea) made up over 80% of the total live coral cover in Lau Province. Hydrozoan corals (*Millepora*) were also an important component of the live cover, occupying 5% of the sea floor, on average. Of the faviid corals, *Goniastrea* was the most common (3.9%) followed by *Favia* (3.5%), while other submassive and massive faviids contributed about 2% of the total coral cover.

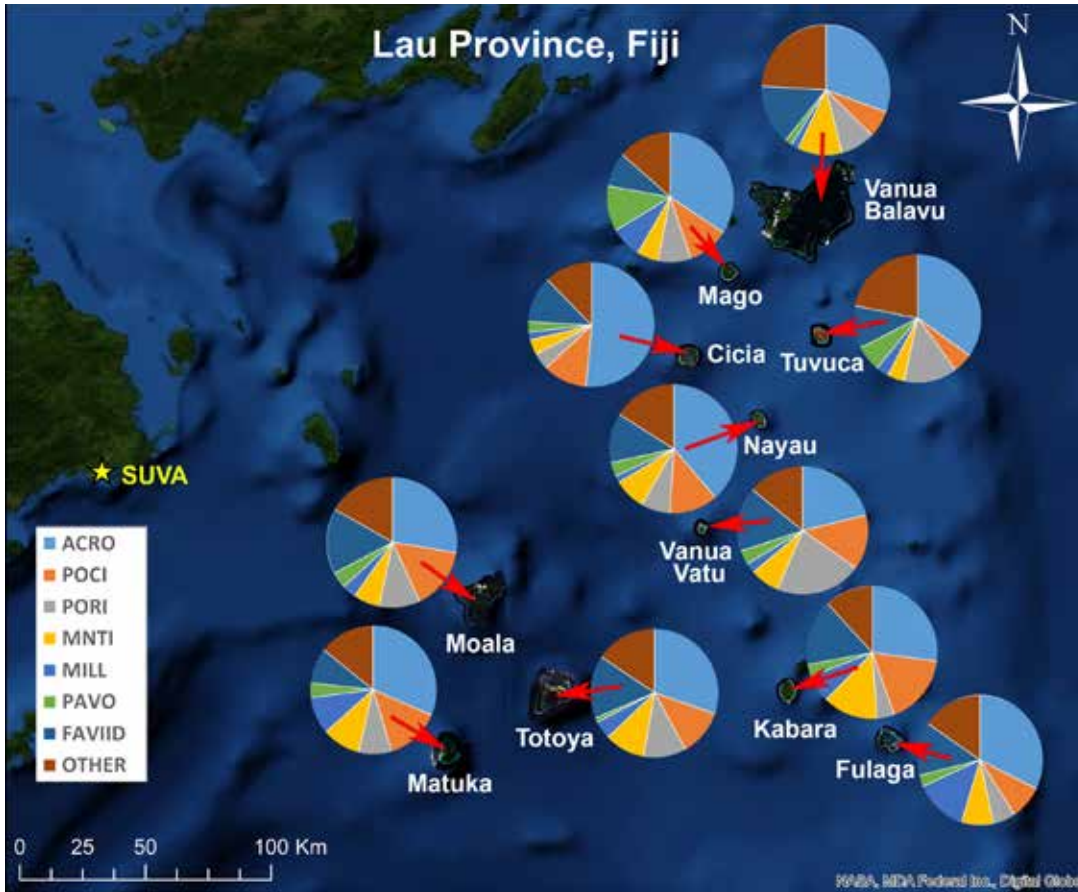


Fig. 84. Cover of the dominant coral taxa by island. Coral cover was dominated by *Acropora* (light blue), *Pocillopora* (orange), *Porites* (gray), *Montipora* (yellow), *Millepora* (blue), *Pavona* (green), faviids (navy blue), and 28 other genera (brown).

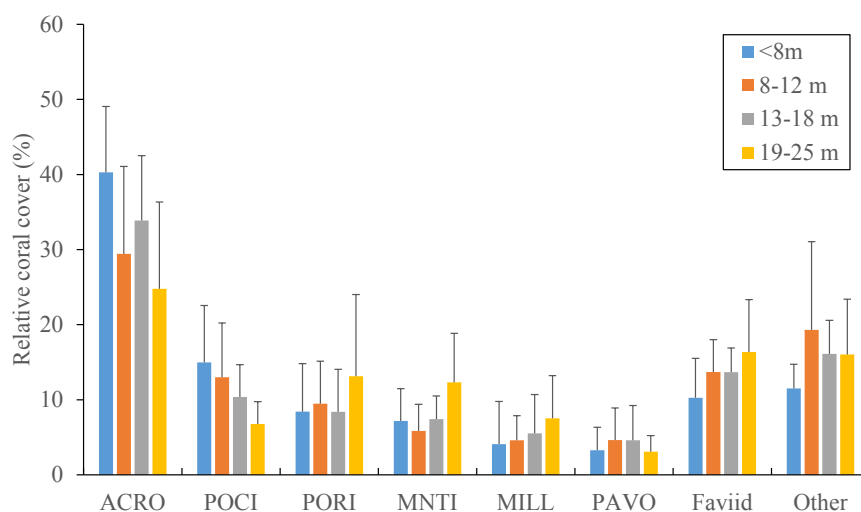
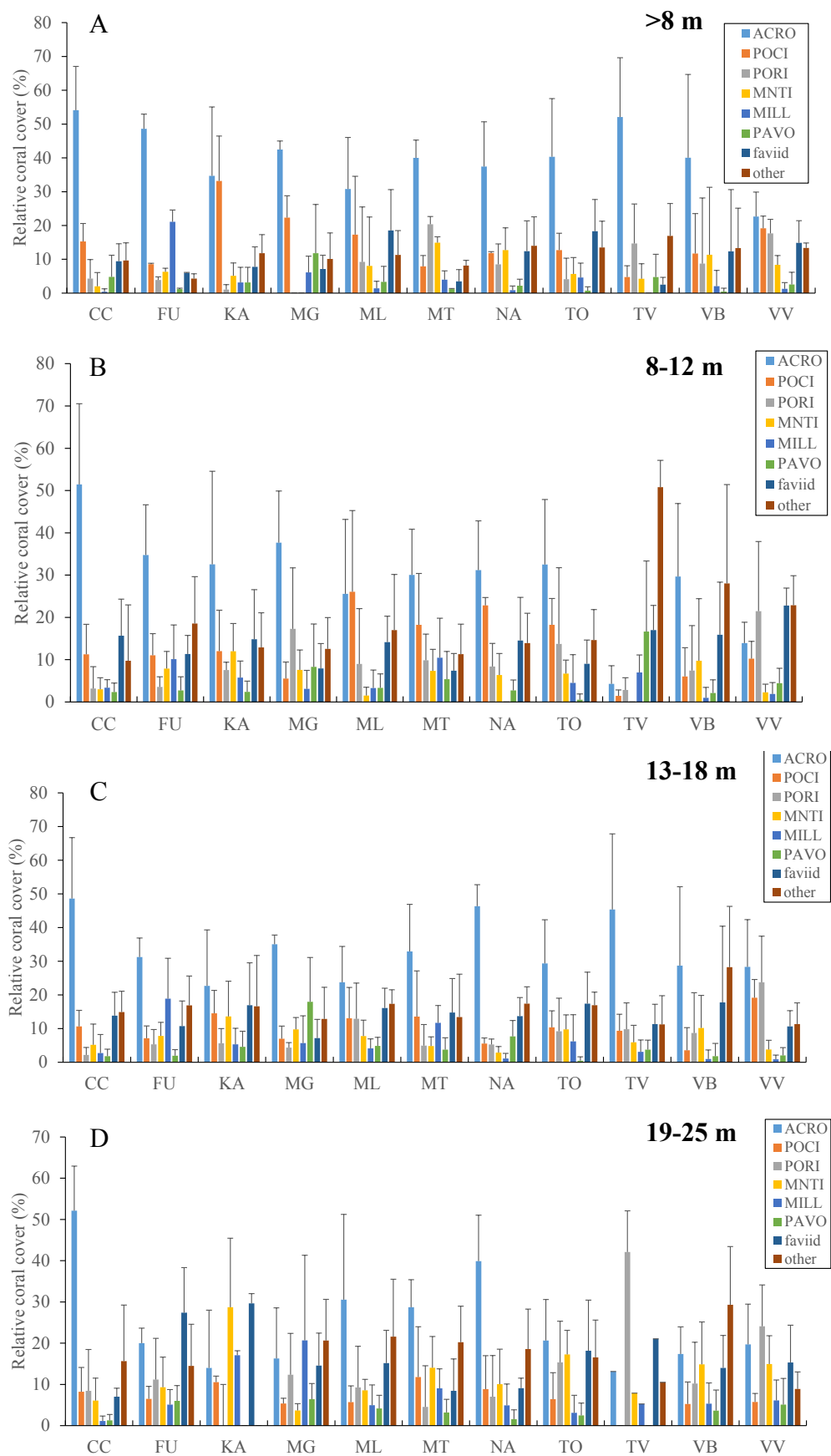


Fig. 85. Cover of the dominant coral taxa pooled for all islands into four depth categories.



The relative cover of different genera varied between islands. Cicia had the highest cover of *Acropora*, making up over half of all coral, followed by Nayau (39%) and Tuvuca (35%). The lowest cover of *Acropora* was recorded at Vanua Vatu (21% of the coral cover). *Pocillopora* cover was highest at Kabara (17.5%) and lowest (5%). The cover of *Porites* was greatest overall at Vanua Vatu (22%) and lowest at Cicia (4.5%). Vanua Balavu had the most diverse coral assemblages, with over 24% of the coral cover consisting of less common taxa (Fig. 84).

The live cover of *Acropora* and *Pocillopora* declined with depth, while *Porites*, *Montipora* and the five faviid genera was greatest at 18-25 m depth. In addition, the highest cover overall for *Porites* was noted at Tuvuca at 18-25 m. *Pavona* varied from 0-17% cover, with some reefs having large stands of *Pavona clavus*, especially Tuvuca at 8-12 m depth (Fig 86).

Some of the less common corals, especially *Diploastrea*, *Leptastrea*, *Galaxea*, *Merulina*, and *Lobophora* were locally abundant, making up 5-30% of the total live cover on certain reefs.

Fig. 86. Cover of the dominant coral taxa pooled on each island. A. cover at < 8 m depth. B. Cover at 8-12 m. C. Cover at 13-18 m. D. Cover at 19-25 m.

Reef Fish Assessments

Over the four-week research mission in Fiji, 359 belt transects were conducted to assess the structure of reef fish communities. The majority of transects were performed between 8 and 18 m depth (Table 7).

Diversity

A total of 482 species (172 genera, 51 families) were recorded within transects. Over 60% of recorded species were observed in over five locations and 101 species were seen on a single reef. Of the 51 families recorded during the mission, the 20 most common species belonged to just 10 families (Table 8). Of these 20 species, the Labridae and Pomacentridae families contained the highest numbers of species (4), followed by Chaetodontidae (3) and Acanthuridae and Scaridae (2). Six species were observed on every reef surveyed. The species observed less frequently belonged to a variety of families, feeding guilds and size classes. Diversity was fairly consistent between the 11 islands, with a total average of 250 species (Fig. 87). Reef fish diversity was highest at Vanua Balavu (375 species), followed by Totoya (278 species) and the lowest diversity was observed at Mago (211 species).

Overall, 32 species of Acanthurids, 23 species of Scarids and 8 Siganids were recorded (Fig. 88). Of these three key herbivorous families, diversity was fairly consistent for Acanthurids and Scarids. The highest diversity of Acanthurids and Siganids was at Vanua Balavu. Scarids were most diverse at both Moala and Totoya (Fig. 89). Diversity of Lutjanids (23) and members of the subfamily Epinephelinae (10) were also generally consistent among islands (Fig. 90). This excludes Vanua Balavu, where Epinephelinae diversity

Island	<8 m	8-13 m	14-18 m	19-25 m	Total
Cicia	0	10	14	0	24
Fulaga	0	10	12	0	22
Kabara	0	17	8	0	25
Mago	0	8	9	0	17
Matuka	1	12	14	0	27
Moala	2	24	25	1	52
Nayau	0	9	9	0	18
Totoya	2	20	26	0	48
Tuvuca	0	10	6	0	16
Vanua Balavu	9	35	49	0	93
Vanua Vatu	0	6	11	0	17
Total	14	161	183	1	359

Table 7. Total number of belt transects, each 30 m X 4 m, completed in each depth category on reefs associated with 11 islands/atolls in Lau Province, Fiji.

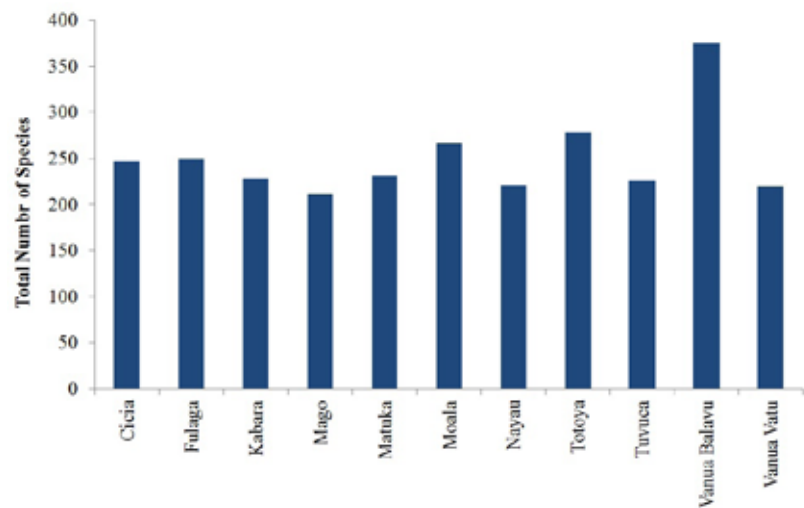


Fig. 87. The total number of species recorded in each island.



Fig. 88. Reef fish communities were dominated by a high diversity of small bodied fishes.

was almost double that of most of the other islands (18 species). Although 10 species of Carangids were recorded overall, this was the least diverse of the key predatory families between the islands (range:

9-0 species). The highest diversity of Carangid species was recorded in Tuvuca, where diversity of both Lutjanids and Epinephelinae was lower in comparison to many of the other islands.

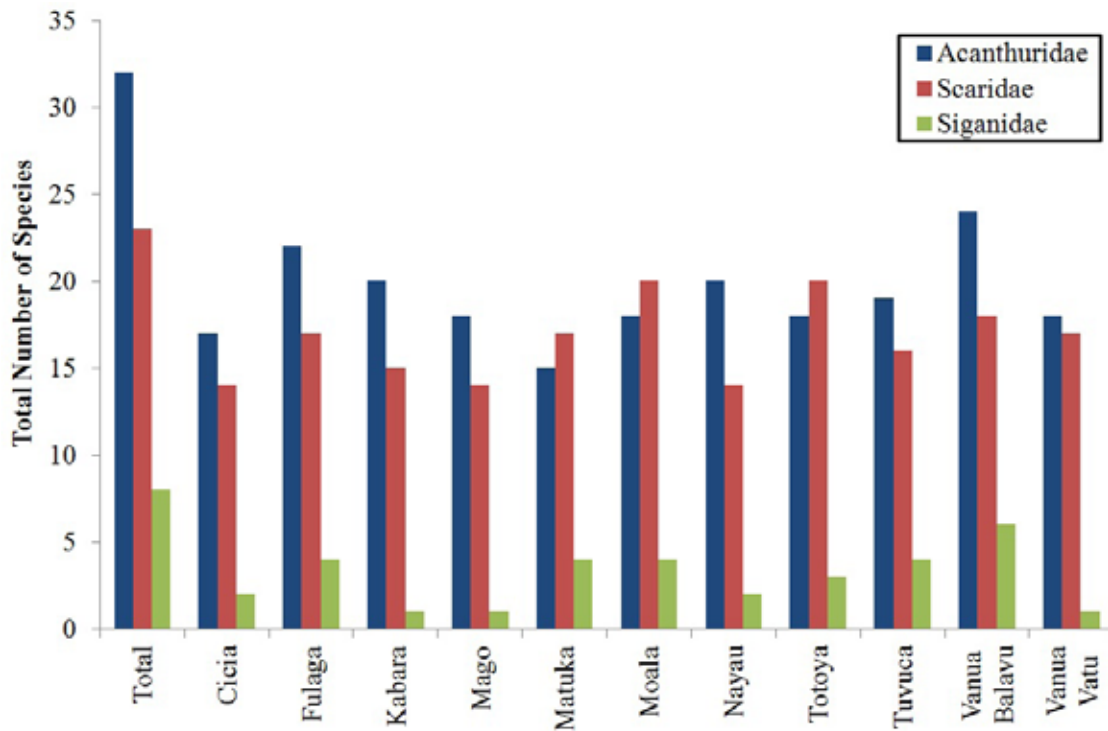


Fig. 89. The total number of species recorded for Acanthuridae, Scaridae and Siganidae families in each island.

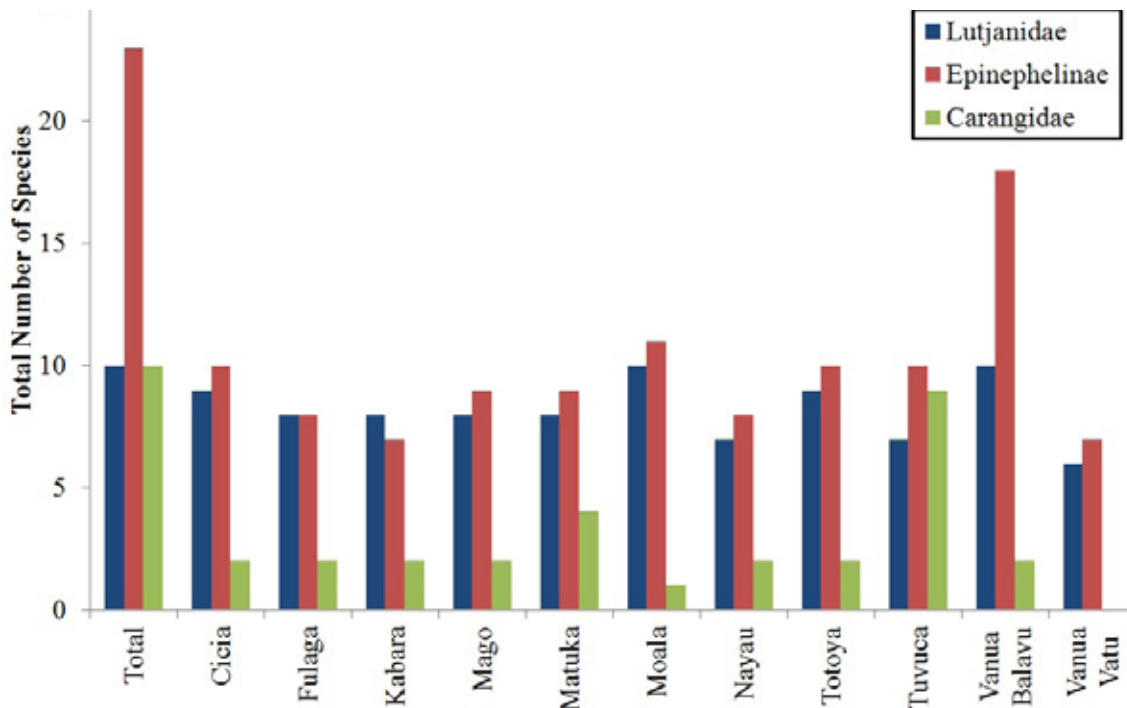


Fig. 90. The total number of species recorded for Carangidae, Lutjanidae and Epinephelinae families in each island.

Density

Overall mean density was 218 individuals/100 m² (Fig. 92). Fulaga recorded the greatest average density (297 individuals/100 m²), followed by Cicia (279 individuals/100 m²) and Matuka the lowest (162 individuals/100 m²).

Generally, the smaller-bodied and/or schooling genera contributed the most towards overall mean density in Fiji. Two Pomacentrid genera were the greatest contributors; *Chromis* (28%) and *Pomacentrus* (13%), followed by *Pterocaesio* (6%) and *Chaetodon* (4%). The majority of the other genera recorded during surveys had densities between 2-12 individuals/100 m².

Planktivores contained the highest mean density of all functional guilds with a 60% difference between this guild and the next highest guild, herbivores (101 and 40 individuals/100 m² respectively; Fig. 93). “Others”, piscivores and corallivores contained much lower densities (7 to 13 individuals/100 m² respectively). Carnivore density was fairly high, with an average total of 21 individuals/100 m².



Fig. 91 The Longnose Filefish, *Oxymonacanthus longirostris* was a common carnivore.

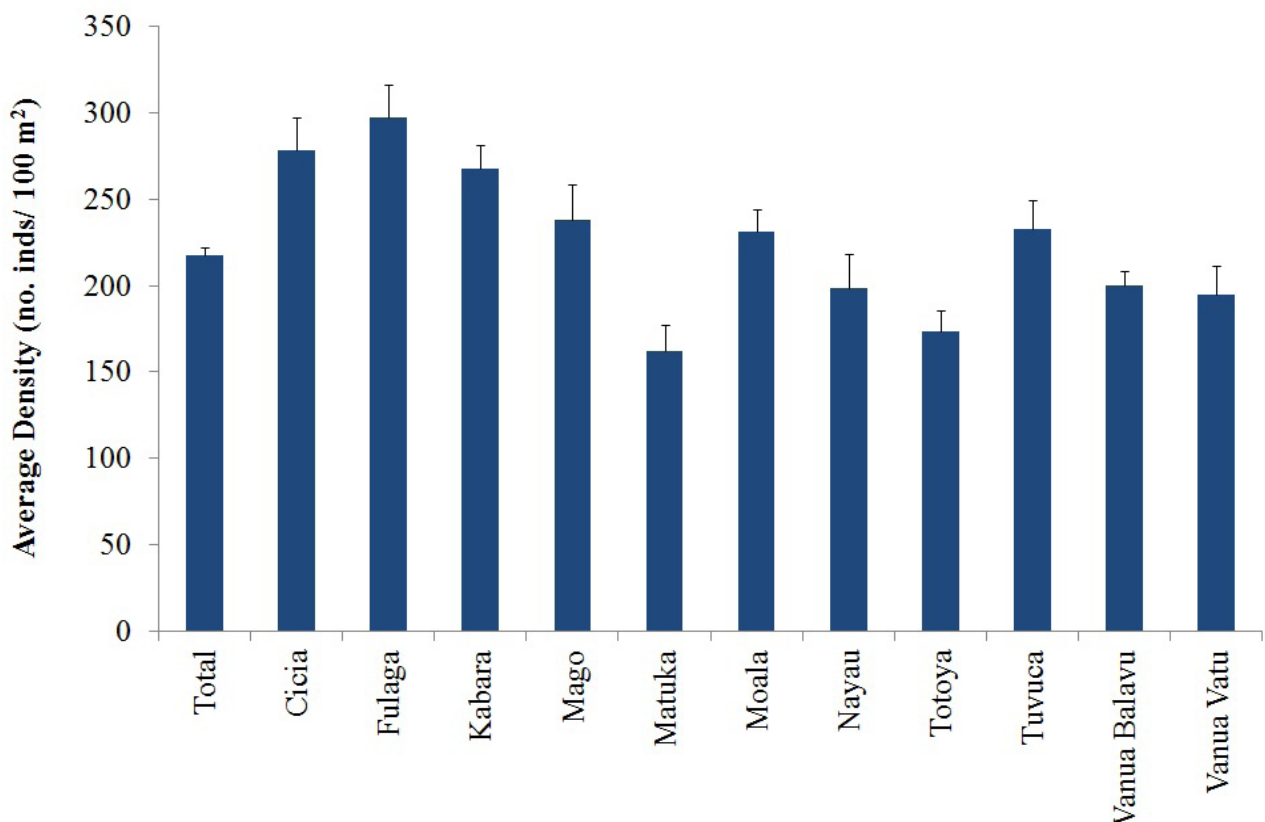


Fig. 92. The average total density (number of individuals/ 100 m² ±SE) recorded by island.

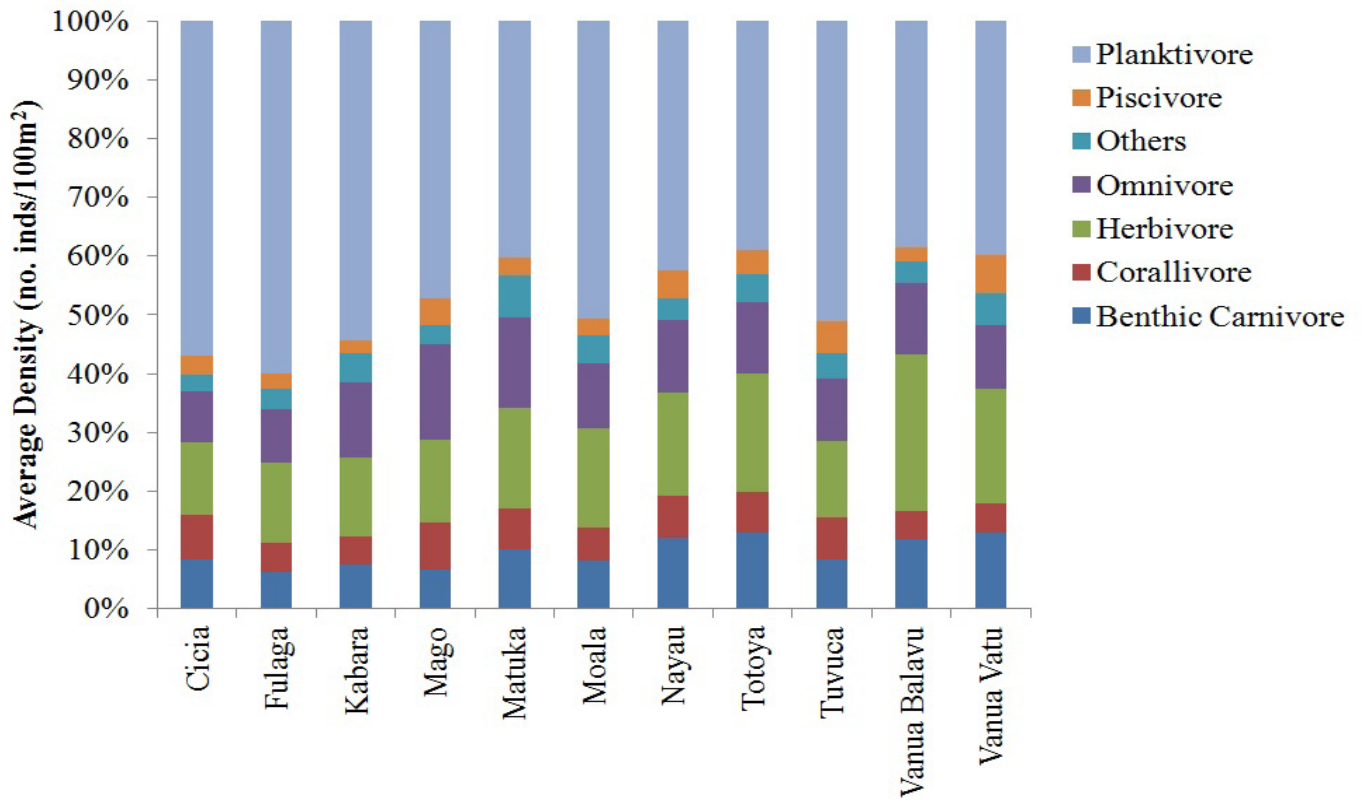


Fig. 93. The average total density (number of individuals/ 100 m² ±SE) by functional guild for each island.

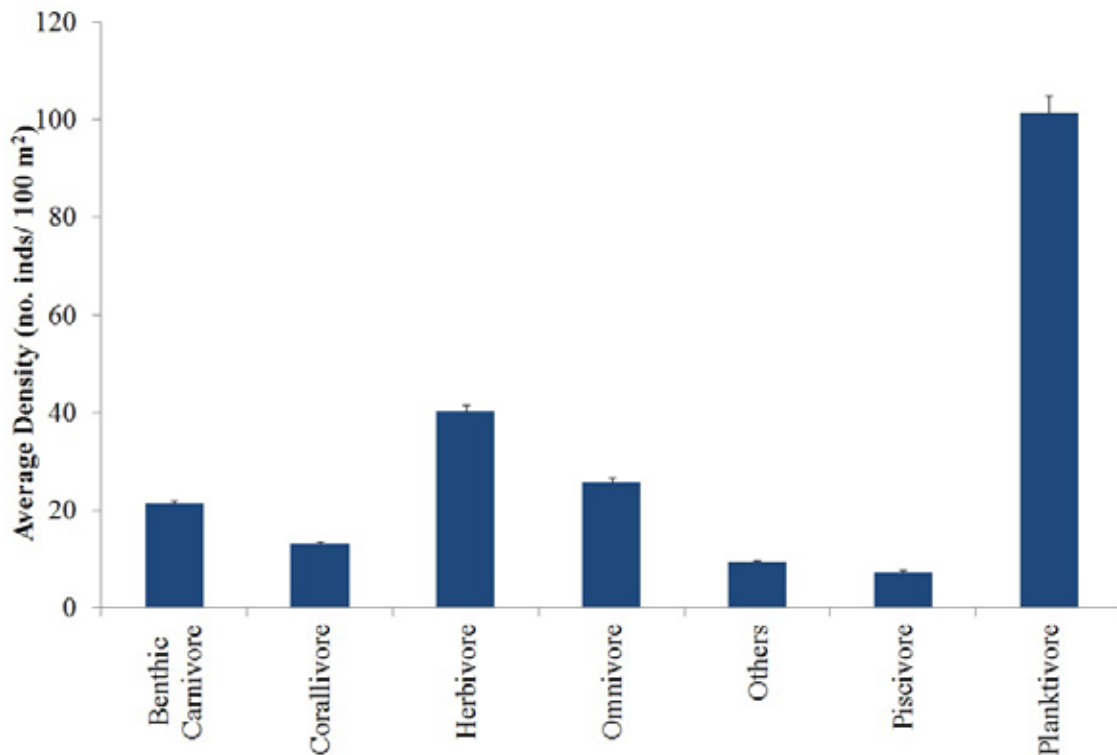


Fig. 94. The average total density (number of individuals/ 100 m² ±SE) recorded by feeding guild.

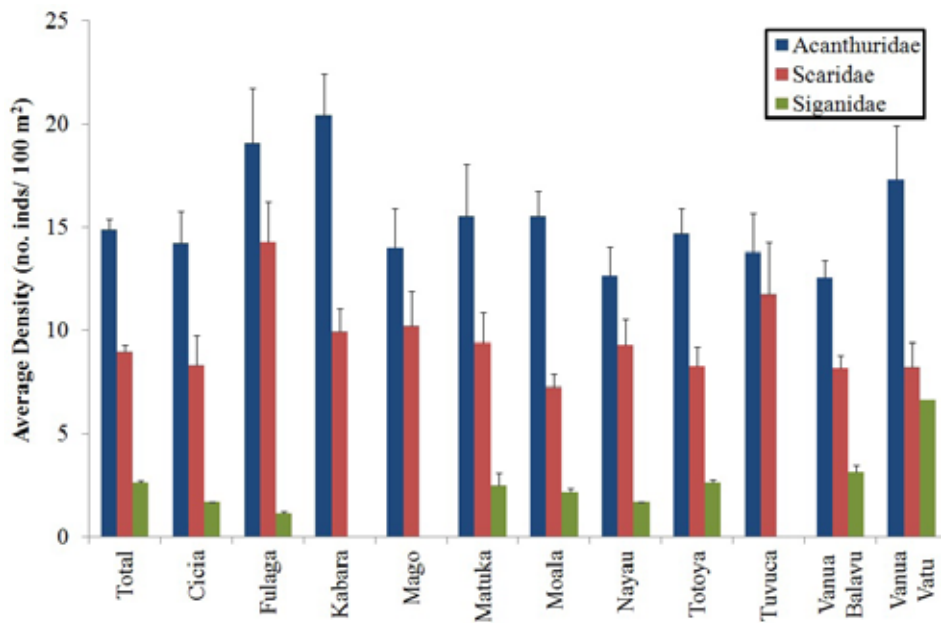


Fig. 95. The total average density (number of individuals/ 100 m² ±SE) recorded for Acanthuridae, Scaridae and Siganidae families at each island.

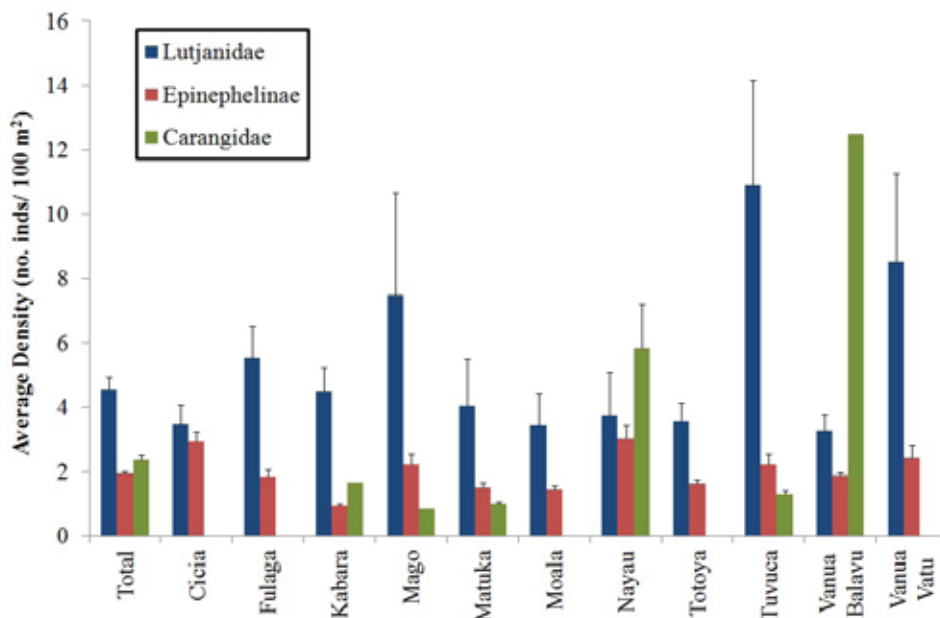


Fig. 96. The total average density (number of individuals/ 100 m² ±SE) recorded for Lutjanidae, Epinephelinae and Carangidae families at each island.

Lutjanid density peaked at three islands; Tuvuca, Vanua Vatu and Mago (11, 9 and 8 individuals/100 m² respectively), with 3-6 individuals/100 m² off other islands. Epinephelinae density ranged from 1- 3 individuals/100 m² (Nayau and Kabara, respectively). Carangids were recorded on transects at just 6 of the islands, with the greatest densities at Vanua Balavu (12.5 individuals/100 m²) and Nayau (6 individuals/100 m²). The remaining 4 islands recorded less than 2 individuals/100 m².

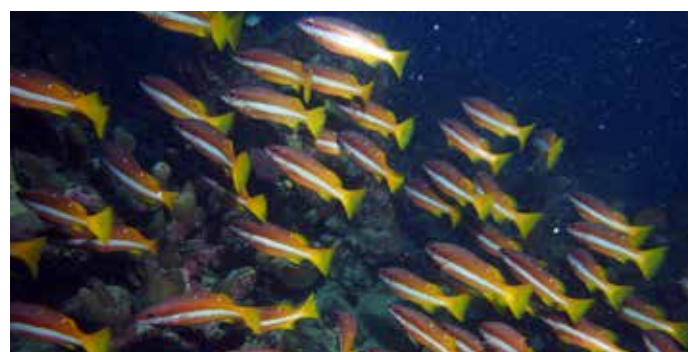


Fig. 97. A school of two-spot banded snapper, *Lutjanus biguttatus*.

Acanthurids were common and fairly consistent at most sites, dominating the herbivore assemblage at all of the islands (15 individuals/100 m²) (Fig. 95). The density of Scarids was approximately half that of Acanthurids (9 individuals/100 m²) and Siganid density was low throughout all islands (3 individuals/100 m²). Acanthurid density peaked at Kabara (20 individuals/100 m²) and Fulaga (19 individuals/100 m²). Scarid density was also high at Fulaga (14 individuals/100 m²) and lowest in Moala (7 individuals/100 m²). Siganid density also peaked at Vanua Vatu (7 individuals/100 m²).

As expected, predatory density was far lower than the herbivores (Fig. 96). Lutjanids were recorded more frequently than the other two predatory families, with an average of 5 individuals/100 m². The average density of the subfamily Epinephelinae was 60% lower than Lutjanids. Carangids had the lowest average density of these three families, with 2 individuals/100 m².

Size Class

Overall, the fish assemblage was dominated by fish less than 20 cm total length (92% of fish; Fig. 98). The larger size classes (i.e. above 41 cm) represented a small component of the total percentage of fish size (0.5%) and overall 7% of fish were between 21 and 40 cm.

Several islands were dominated by fish assemblages where individuals were less than 5 cm (Cicia 51%, Kabara 48% and Mago 51%). Vanua Vatu and Totoya had more fish within the 31 to 40 cm class than the other islands. This was probably due to the presence of large, schooling species at these islands (*C. teres*, *M. niger* and *C. lunaris*). Relative abundance of fish rarely exceeded 100 cm, with several islands recording no individuals within this class. Totoya had a slightly higher representation of fish over 100 cm, due to a higher abundance of sharks at this site.

Over 95% of omnivorous fish were under 10 cm in length and none exceeded 40 cm (Fig. 99). Of all feeding guilds, the largest proportion of fish greater than 51 cm in length belonged to the piscivores (2%). Almost half of the planktivores were between 6 and 20 cm, with 78% under 10 cm. Benthic carnivores were not particularly large with 89% under 20 cm and just 0.3% of fish recorded exceeding 41 cm in length. As expected, almost all corallivores were under 20 cm length. Just over a quarter of herbivorous fish were between 11 and 30 cm, with the majority of herbivores between 6 and 10 cm in length.

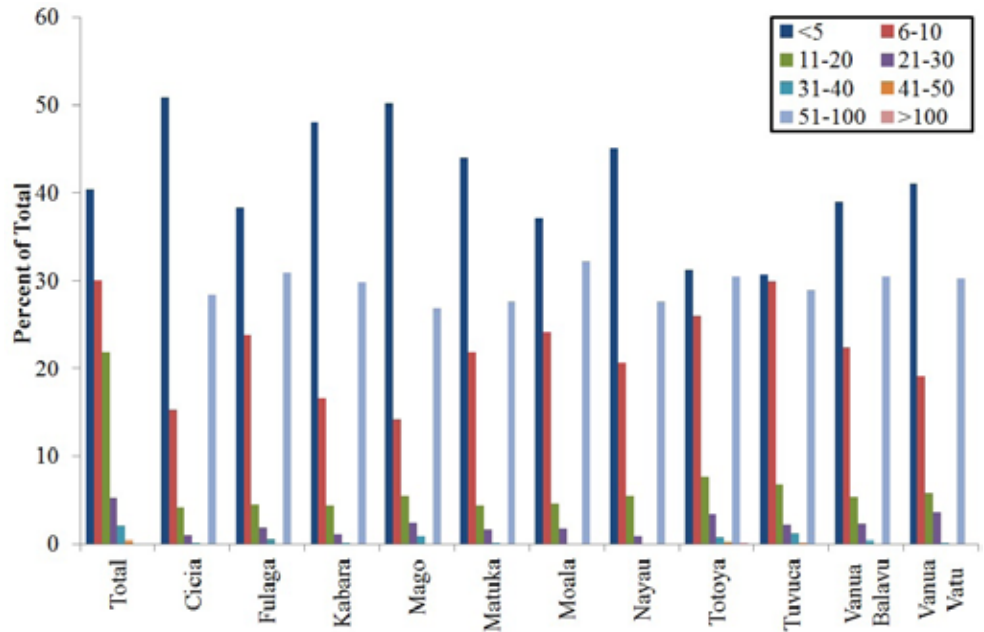


Fig. 98. The relative abundance (percentage of total) of all reef fish size classes recorded in each island.

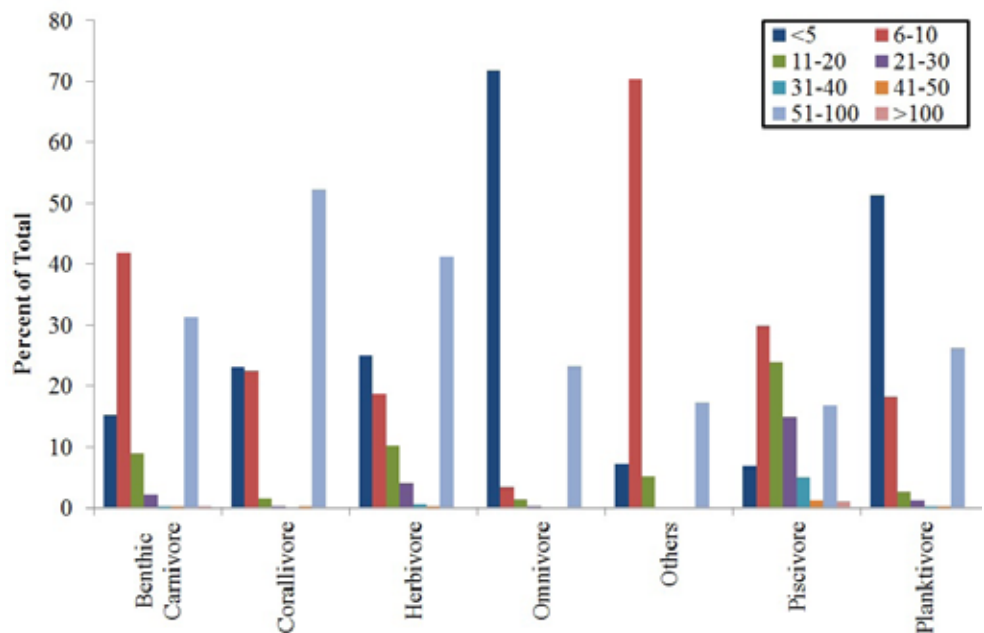


Fig. 99. The relative abundance (percentage of total) of reef fish size classes by feeding guild.



Fig. 100. Most groupers were 11-30 cm total length.

The three main groups of herbivores were mostly <30 cm in length. Scarids tended to fit within the larger size classes, with 92% ranging from 11 to 40 cm, and 3% exceeding 41 cm (Fig. 102). Overall, less than 1% of Scarids recorded were under 5 cm in length. The larger Scarids (e.g., *Chlorurus microhinos*, *Cetoscarus bicolor*, *Scarus forsteni* and *Hipposcarus longiceps*) were observed more frequently in Mago and Totoya. Acanthurids tended to be mid-sized, with the majority between 11 and 30 cm in length and less than 1% under 5 cm (Fig. 103). Fulaga's reefs comprised the greatest relative abundance of Acanthurids over 41 cm (6%). This was due to several larger unicornfish, *Naso hexacanthus* and *Naso lopezi*. Siganids tended to be primarily within the 11 to 20 cm size class, however at several islands, including Ciccia, most or all individuals were larger in size (Fig. 104). Only in Vanua Balavu were individuals under 10 cm in length.



Fig. 101. A small school of acanthurids.

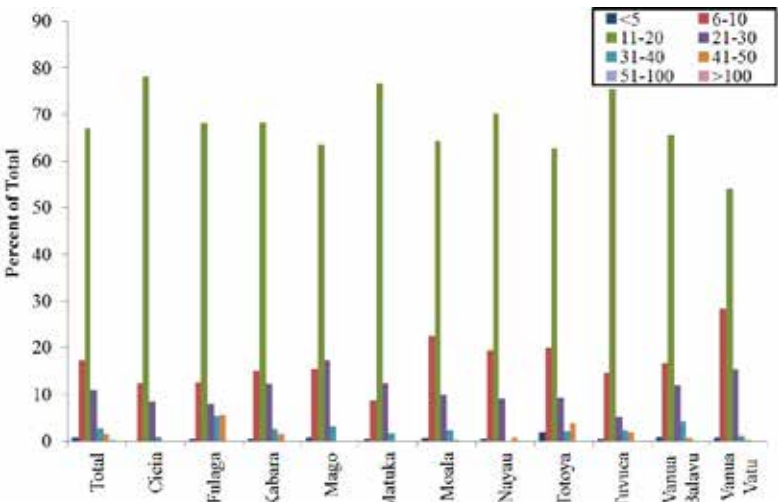


Fig. 102. The relative abundance (percentage of total) of reef fish size classes for Scaridae.

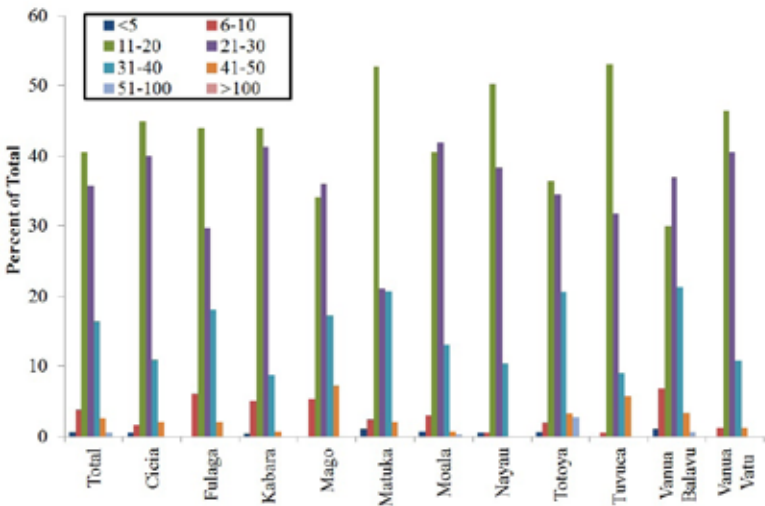


Fig. 103. The relative abundance (percentage of total) of reef fish size classes for Acanthuridae.

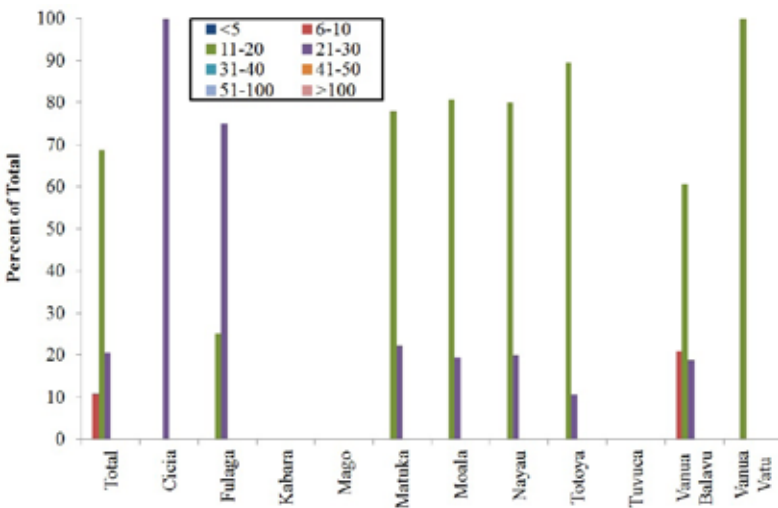


Fig. 104 The relative abundance (percentage of total) of reef fish size classes for Siganidae.

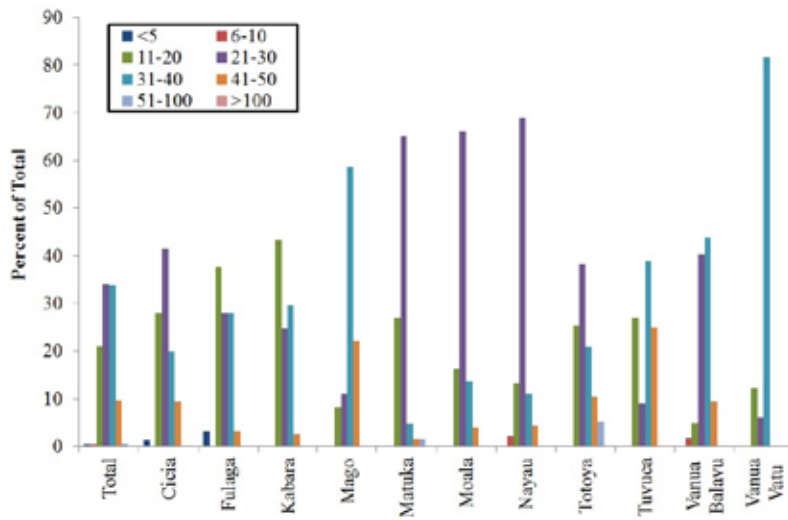


Fig. 105. The relative abundance (percentage of total) of reef fish size classes for Lutjanidae.

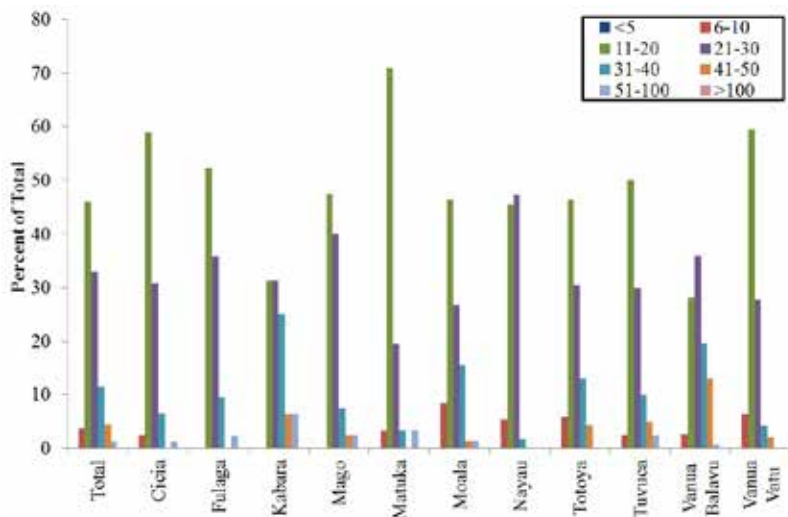


Fig. 106. The relative abundance (percentage of total) of reef fish size classes for Epinephelinae

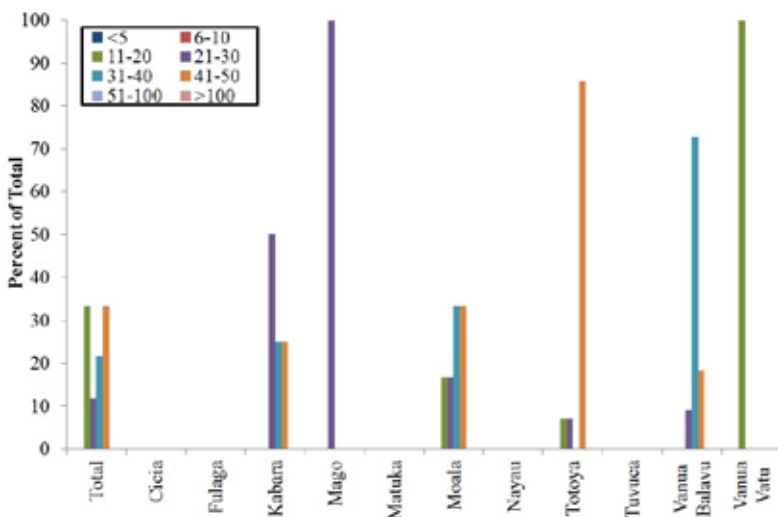


Fig. 107. The relative abundance (percentage of total) of reef fish size classes for Carangidae.

Lutjanids were mid to large body sizes, with two thirds of this family between 21 and 40 cm length (Fig. 105). Less than 1% of Lutjanids were under 10 cm, and no individuals exceeded 100 cm. A quarter of Lutjanids in Tuvuca were between 41 and 50 cm length, and in Totoya over 5% were larger than 51 cm.

Over 90% of all individuals from the subfamily Epinephelinae were within the middle of the size distribution and approximately 4% of individuals were at either end of the scale (Fig. 106). Groupers between 41 and 50 cm were most commonly observed at Vanua Balavu and Mago. No groupers under 5 cm were recorded at any island. All Carangids recorded throughout the islands were between 11 and 50 cm (Fig. 107). A third of all Carangids were between both 11 and 20 cm and 41 and 50 cm. The greatest variability of sizes was observed in Moala, with many other islands recording Carangids within just one or two size categories.

Sharks

Three species of sharks were recorded throughout the mission; *Carcharhinus amblyrhynchos*, *Triaenodon obesus* and *Carcharhinus melanopterus*. *T. obesus* was the most commonly observed species. Totoya was the only site where all three species were observed and where individuals were recorded most frequently. Individuals recorded on transects ranged from 31 cm to over 100 cm, with the majority exceeding 100 cm.



Fig. 108. A white tip shark from Totoya.

Biomass

Average biomass at these islands was 11.27 kg/100 m² (Fig. 109). Biomass was fairly consistent among islands, ranging between 8-12 kg/100 m². The only exception was Totoya, where biomass was far higher (19.4 kg/100 m²). The lowest average biomass was recorded at Matuka (8 kg/100 m²).

Overall, the larger-bodied genera contributed the most towards total biomass in Fiji. *Triaenodon* was the largest contributor of total biomass (18%), followed by two parrotfish genera; *Scarus* (13%) and *Chlorurus* (7%).

Piscivores and herbivores also contributed towards large proportions of overall biomass in each island. In Totoya, piscivores had the greatest biomass of all guilds which was due to the greater abundance of sharks at this location. *Triaenodon* and *Carcharhinus* contributed 85% of piscivore biomass in Totoya and over 55% of total biomass at this island.

Benthic carnivores contributed more towards total biomass in Vanua Vatu and Nayau than at other islands. In Vanua Vatu, *Chelinus* contributed almost 60% of the biomass of benthic carnivores. The proportion of corallivores was greater in Mago and Nayau and very low in Totoya and Kabara.

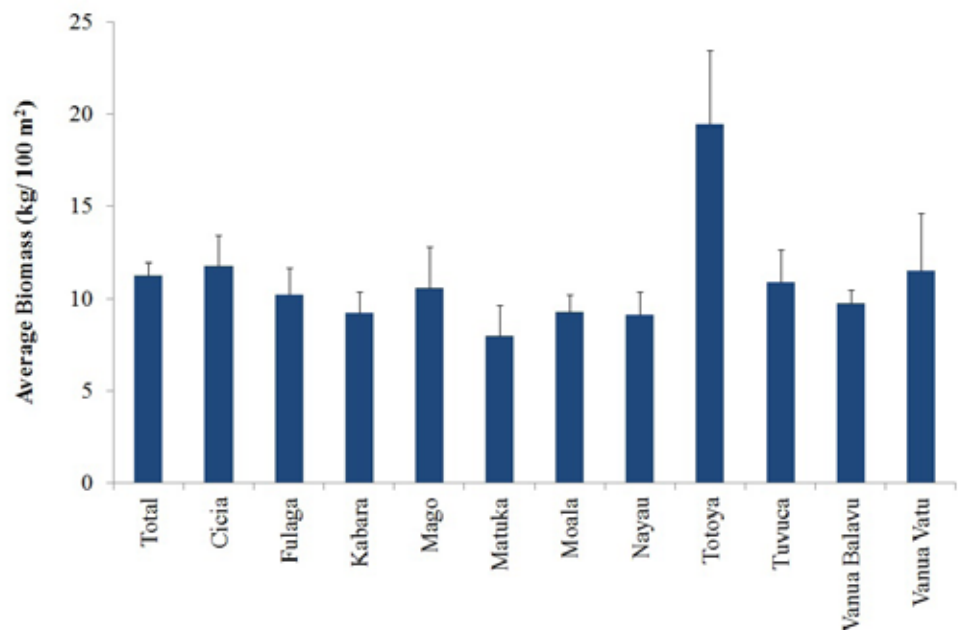


Fig. 109. The average total biomass (kg/ 100 m² ±SE) recorded by island.

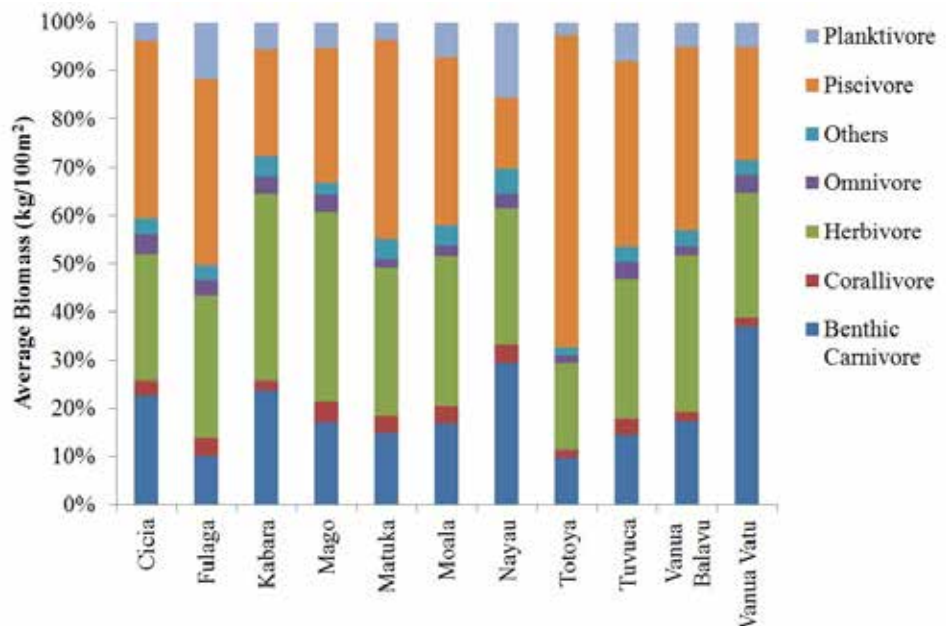


Fig. 110. The average total biomass (kg/ 100 m² ±SE) recorded by functional group for each island.

Fig. 111. Goatfish are important benthic carnivores that often occurred in schools of 20-50 fish, but most were small in size.



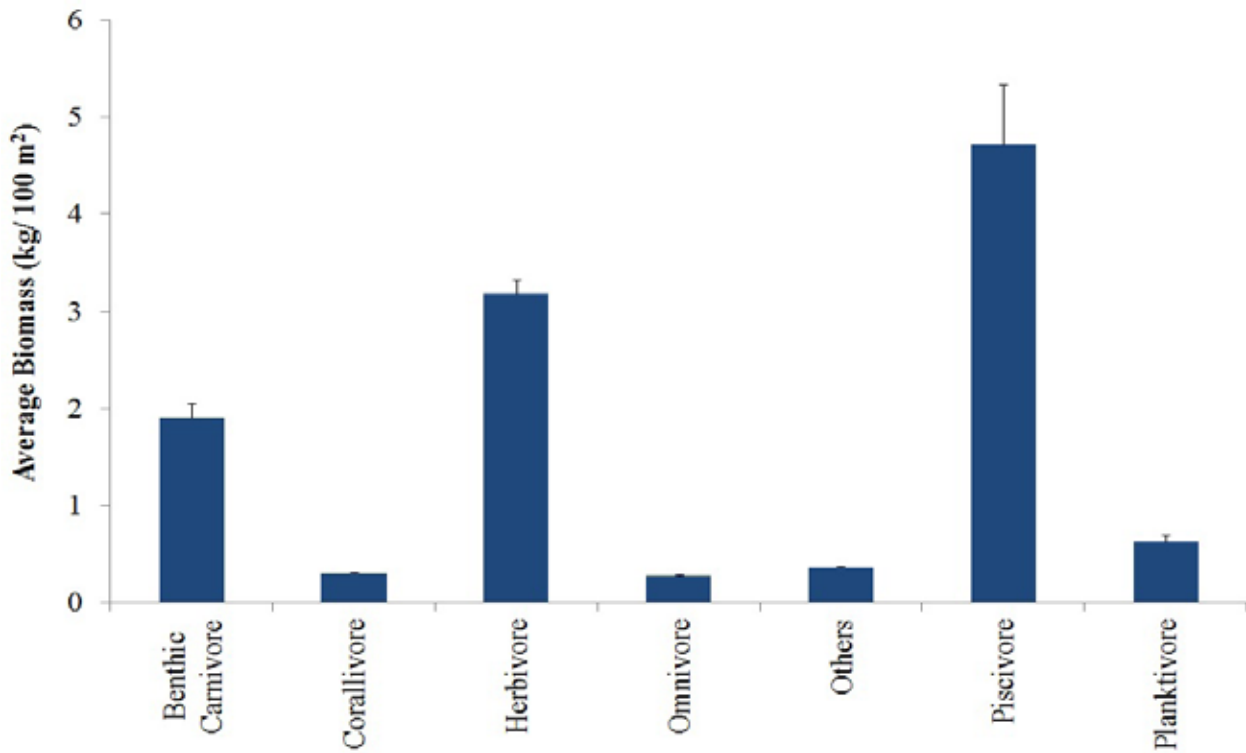


Fig. 112. The average total biomass (kg/100 m² ±SE) recorded by feeding guild.

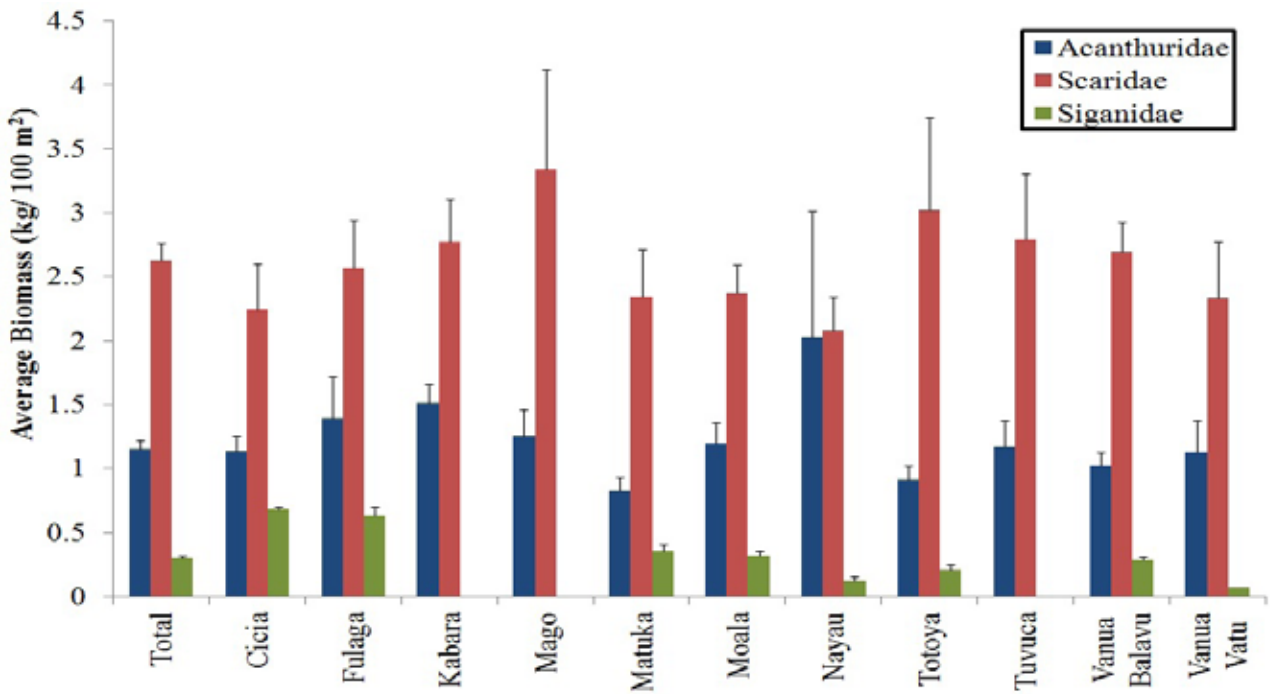


Fig. 113. The total average biomass (kg/100 m² ±SE) recorded for Acanthuridae, Scaridae and Siganidae families in each island.

Two of the feeding guilds recorded an average biomass that exceeded 1 kg/100 m²; piscivores (4.7 kg/100 m²), herbivores (3.2 kg/100 m²), benthic carnivores (1.9 kg/100 m²; Fig. 111). Omnivores and corallivores contained the lowest average biomass of all feeding guilds, with just 0.3 kg/100 m² in each guild.

Herbivore biomass was dominated by Scarids (total average; 2.6 kg/100 m²) and Acanthurid and Siganid biomass were lower, with an average of just 1.15 kg/100 m² and 0.3 kg/100 m² respectively (Figure 113). The largest biomass of Scarids was recorded at Mago (3.34 kg/100 m²) and Totoya (3.02 kg/100 m²). Scarid biomass at the remaining islands was very consistent, ranging between 2.79 and 2.1 kg/100 m². Acanthurid biomass was also consistent throughout the islands, with a maximum recorded in Nayau (2.1 kg/100 m²) and the lowest biomass at Matuka (0.8 kg/100 m²). Siganid biomass was extremely low throughout the islands and never exceeded 1 kg/100 m².

Lutjanids dominated the biomass of predatory families at most islands, with an overall average of 1.6 kg/100 m² (Fig. 113). Epinephelinae and Carangidae had an average of approximately 0.8 kg/100 m². Mago recorded a higher biomass of Lutjanids (2.27 kg/100 m²), which is most likely due to the presence of larger schools and individuals of *Lutjanus gibbus*, *Lutjanus bohar*, *Lutjanus monostigma* and *Macolor niger*. The lowest Lutjanid biomass of 1 kg/100 m² was recorded at Nayau. Grouper biomass was greatest at Kabara (1.26 kg/100 m²), followed by Vanua Balavu (1.16 kg/100 m²). The lowest biomass was observed at Vanua Vatu (0.47 kg/100 m²). Due to large *Carangoides orthogrammus* and *Caranx melampygus* individuals, Carangid biomass was surprisingly high at Kabara (1.93 kg/100 m²), compared with the other islands (range: 0.9 to 0.06 kg/100 m²).

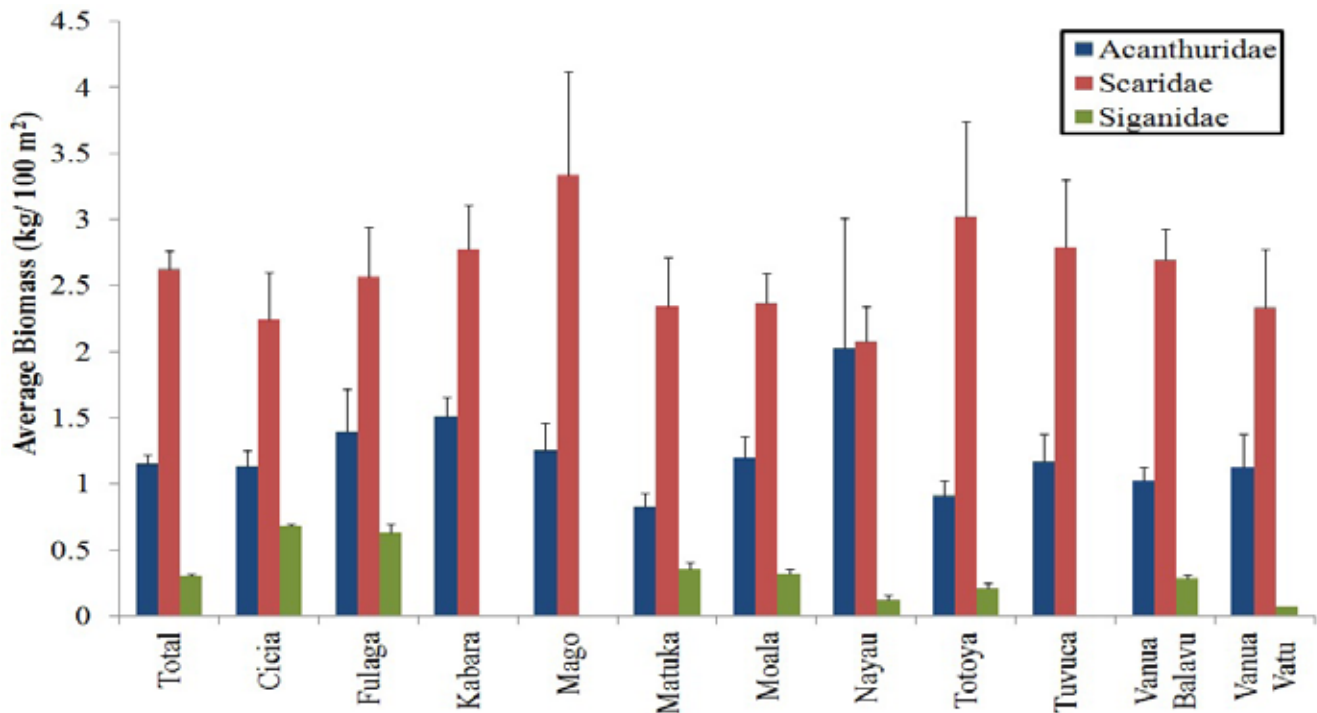


Fig. 114. The total average biomass (kg/100 m² ±SE) recorded for Carangidae, Lutjanidae and Epinephelinae families in each island.

Sea Cucumber Fishery Surveys

A survey of sea cucumber resources and the status of the fishery in Lau Province, Fiji was conducted by Stacy Jupiter of the Wildlife Conservation Society of Fiji, Ron Vave of the University of the South Pacific, and William Saladrau of the Fiji Department of Fisheries, as part of the Khaled bin Sultan Living Oceans Foundation's Global Reef Expedition. Direct in-water resource assessments were carried out using standardized protocols developed by the Secretariat of the Pacific Community (SPC) Regional Fisheries Observatory program. The data were combined with fishermen perception surveys, measurements of dried sea cucumbers processed in villages, and general observations from community discussions to assess the overall status of the fishery.



Fig. 115. Many types of sea cucumbers dry on a rack in Fulaga. The harvest may be too high to be sustainable.

The density of commercially important sea cucumber species varied across the islands of Lau Province, ranging from 0 – 132.81 sea cucumbers per hectare. Densities were below SPC regional indicator values for all species except one, *Pearsonothuria graeffei*. The highest density of sea cucumbers were found on Totoya and Vanuabalavu, but even there the density of sea cucumbers generally fell below the threshold required to avoid reproductive failure (10-50 sea cucumbers of an individual species per hectare). Furthermore, with the exception of some well-managed and long-established community marine protected areas (tabu), sea cucumbers tended to be widely dispersed, which also prevents successful fertilization.

The one optimistic finding was that community-based management does seem to be having a positive effect in preserving some remaining individuals, as total sea cucumber abundance was significantly higher in tabu areas than in areas open to fishing surveyed using belt transects.

The mean size of sea cucumbers measured underwater were generally above minimum recommended wet sizes. There were almost no very small sea cucumbers on the reef, which is of concern and may be indicative of recruitment failure. Village measurements of dried samples indicate that many undersized individuals are being harvested. As prices have increased due to reduction in supply, fishers have not yet been proactive about management measures given that they are still able to meet their daily needs with income derived from sea cucumbers as they are using techniques (e.g. underwater breathing apparatus, free diving with 'bombs') to extract individuals from deep refuges.



Fig. 116. William Saladrau records sea cucumber size.

These surveys demonstrate that the density of sea cucumbers in Lau Province are below what is necessary for successful recruitment. This indicates that sea cucumber populations, and the associated fishery, is likely to continue to decline in the future unless measures are taken to sustainably manage these species. Sea cucumbers are not only an important economic resource for the community, they are critical to maintaining the health of the reefs. The presence of cyanobacterial mats and relative absence of juvenile sea cucumbers suggests that declining sea cucumber populations are already negatively affecting coral reef health in Fiji.

Education Programs

During the Global Reef Expedition’s mission to Fiji, the Foundation provided coral reef education seminars at schools and in communities throughout Lau Province. Overall, the Foundation conducted 14 school and 8 community seminars reaching a total of 1,464 people on ten different islands (Table 8). In addition, 3 stakeholder seminars were held in Suva to discuss the state of coral reefs in Fiji, these seminars reached 80 additional people (Table 9). This was the first time in Foundation’s history that this much education had been provided in a single mission.

These educational seminars were conducted in partnership with representatives from the Lau Provincial Council’s Office, the Pacific Blue Foundation, and the University of the South Pacific. The following representatives donated their time to support this important education mission:

Lau Provincial Council’s Office

Roko Latia Raloa, Protocol Officer

Pacific Blue Foundation

Roko Sau (Roko Josefa Cinavilakeba), High Chief and Representative

University of the South Pacific

Ron Vave, Scientific Diver

Coral Reef Education Seminars

Students participating in the coral reef education seminars learned about the Khaled bin Sultan Living Oceans Foundation, the Global Reef Expedition, and the coral reefs of Fiji. The program was delivered in both English and Fijian. Living Ocean Foundation’s Education Director Amy Heemsoth presented the content, while Roko Sau Josefa Cinavilakebe or Roko Latia Raloa, the education team’s cultural liaisons, translated what she said into Fijian.

As part of the seminars, students were shown where stony corals live throughout the world and discussed why corals can survive in Fiji. Through informal results, when asked if a coral was a plant, animal, or mineral, approximately 95% of the students thought that corals were minerals. Many students said they thought coral was not a living thing since it is hard like a rock, but the education team explained that corals are animals and they identified the relationships between the plant, animal, and mineral components of coral reefs.

Roko Latia Raloa told the education team that “In these remote islands, students are not taught about the environment around them. By providing this knowledge, we hope that they will better understand their resources and how to manage them.”



Fig. 117. Students, teachers, and Amy Heemsoth at a primary school in Mabula, Fiji.

Coral Reef Education Seminars in Fiji

Island	Village	Activity Type	# of Participants			
			Adults	Teachers	Principals	Students
Totoya	Tovu	Primary school seminar (Grades K-8)	4	2	1	38
Matuku	Lavukai	Village/school seminar (GRADES K-5)	28		1	28
Moala	Naroi	Village seminar	99			
		Primary school seminar (Grades K-8)		4	1	96
		Secondary school seminar (Grades 9-12)		12	1	103
Fulaga	Muanaicake	Primary school seminar (Grades K-8)	6	3	1	61
		Village seminar	36			
Kabara	Naikeleya	Primary school seminar (Grades K-5)	14	2	1	24
		Village seminar	35			
Kabara	Tokalau	Primary school seminar (Grades K-8)	15	3	1	50
Vanuavatu	Taira	Primary school seminar (Grades K-8)	3	2		29
Nayau	Salia	Village seminar	70			13
Tuvuca	Tuvuca	Primary school seminar (Grades K-8)		4	1	36
		Village seminar	56			
Cicia	Taruka	Primary school seminar (Grades K-8)		4	1	67
Cicia	Mabula	Primary school seminar (Grades K-8)		3	1	101
		Secondary school seminar (Grades 9-12)	6	12	1	145
Vanua Balavu	Adi Maopa	Primary school seminar (Grades K-8)		4	1	67
Vanua Balavu	Mualevu	Secondary school seminar (Grades 9-12)		12	1	100
Vanua Balavu	Mavana	Village seminar	53			
Vanua Balavu	Malaka	Village seminar	37			3
10 Islands	15 Villages	14 Schools	48	67	13	906
		11 Primary Schools		31	10	558
		3 Secondary Schools		36	3	348
		8 Village Talks	414			16
TOTAL			1,464 PEOPLE			

Table 8. Summary of participation in coral reef education seminars given by Amy Heemsoth in Fiji.

The coral reef education seminars also discussed the benefits of coral reefs with students, including food, income, tourism, jobs, coastal protection, medicine, and as habitat for other organisms. In the Lau Province, villages depend on the reefs for seafood, which is their main source of protein, and for some, income. The students quickly realized the importance of coral reefs, not only for their family, but also for the health of the overall ecosystem.

The education team also covered coral reef health, and explained that not all coral reefs around the world are healthy due to various threats that they face. They discussed the main threats to Fiji’s coral reefs, including marine and land pollution, overfishing and destructive fishing practices, and outbreaks of crown of thorns starfish (COTS). Using photos, graphics, and videos the team described the various threats. The Fijian team members explained to the students that they were fortunate to have such healthy coral reefs, and if they didn’t take care of them, they too could become unhealthy.

After a school talk in Vanua Vatu, with no instruction, a student began collecting batteries that were polluting the beach. Ron Vave, who aided in teaching the seminar that day commented, “The boy was young and it threw me off guard when I saw him collecting rubbish. No one told him to collect the litter. We raised the awareness and it shows that the talk had an impact on this kid. It makes it all worthwhile.”

Each coral reef education seminar concluded on a note of hope, hope for Fiji’s reefs, and hope for coral reefs around the world. As we learn more about coral reefs, the better we will know how to manage them and sustain these resources.

Every school visited on the Global Reef Expedition in Fiji received a DVD of the coral reef education seminar PowerPoint presentation so that they could continue to educate their students about the health and importance of their coral reefs for many years to come. Many of the teachers and principals that participated in the coral reef education seminars thanked the Khaled bin Sultan Living Oceans Foundation education team for bringing this program to their students and teachers.



Fig. 118. Students at a primary school in Tuvuca, Fiji after participating in our coral reef education program.

Stakeholder Seminars

In Suva, the capital of Fiji, the Khaled bin Sultan Living Oceans Foundation Executive Director, Captain Philip G. Renaud held discussions with stakeholders about the state of coral reefs in Fiji. Stakeholder seminars were held with three different groups: the University of the South Pacific, Ministry of Fisheries, and the International Union for the Conservation of Nature (IUCN).

These stakeholder seminars covered an overview of the Global Reef Expedition and the scientific objectives of the mission to Fiji. Captain Renaud also covered the importance of coral reef ecosystems, global and local threats to reefs, and factors that affect their resilience, before opening up the seminar for questions and a discussion with stakeholder groups about the health and resilience of coral reefs in Fiji.

Organization	# of Participants
University of the South Pacific	50 students and faculty
Briefing to the Ministry of Fisheries	5 staff members of the Ministry of Fisheries
IUCN	25 Coral reef stakeholders from a variety of groups
TOTAL	80 People

Table 9. Summary of participation in coral reef seminars given by Captain Philip Renaud in Suva, Fiji.

Conclusions

Coral communities throughout the Lau Province have shown progressive recovery from damage from coral bleaching, outbreaks of corallivores and storms sustained in the early part of the new millennium. In particular, acroporid corals have rebounded and are once again the dominant coral in terms of cover. Most corals were in fairly good health, although some reefs had a low to medium prevalence of coral diseases (especially white syndrome). Bleaching was not observed during the mission. Signs of predation were predominantly limited to fish bites, with localized impacts from *Drupella* gastropods and a single outbreak of crown of thorns starfish documented on the fore reef at Cicia. It is interesting to note that these reefs also had the highest live coral cover and the most *Acropora* spp. recorded in the Lau Province.

In general, benthic communities could support continued recruitment and growth of corals, as the cover of macroalgae was relatively low. The only exceptions were some deeper reef communities, lagoonal reefs and vertical walls. The main pest species that had a negative impact benthic substrate was cyanobacteria, which formed dense mats on the sediment, rubble and often adjacent to corals in many locations. One reason for the notable presence of this algae may be the localized depletion of sea cucumbers, which are key detritivores that play a critical role in maintaining clean, aerated sediments. *Xenia* soft corals were also notably abundant on some deeper reefs in areas where stony coral had been damaged. Most reefs had high cover of crustose coralline algae, but a high prevalence of lethal orange disease was observed on many reefs. Considerable damage to fore reef and lagoonal reef communities was also attributed to *P. lacrymatus* damselfish algal lawns.

Traditional management and a lack of commercial fisheries within Lau Province have resulted in reef environments that sustain healthy reef fish assemblages. A high number of reef fish species (482 species) was recorded throughout this mission. Species richness was slightly higher than previously documented in another Fijian District (approx. 340

species, Kubulau District; Goetze, 2009; Jupiter et al., 2010).

Total mean biomass was moderately high (1126 kg/ha, range 1941-801 kg/ha) and surpassed other locations both in the Pacific Ocean (e.g., PNG 378-301 kg/ha, Cinner et al., 2006) and in Fiji (e.g., Vatu-i-Ra, \bar{x} 730 kg/ha, Marnane et al., 2003). Our result was similar to those found in areas closed to fishing around Kubulau District, Bua Province (see Jupiter and Egli, 2010).

The size distribution of reef fish was skewed towards those under 20 cm total length, with few exceeding 50 cm. Fish assemblage was dominated by smaller bodied genera, primarily pomacentrids. This result was surprising as human population and fishing pressure have both declined throughout this province (Turner et al., 2007). However the notably high algal cover may explain part of this trend, by supporting the large aggregations and abundance of herbivorous farming damselfish throughout the islands.

Results from other studies in Lau Province identified a temporal and spatial decline in the abundance of small fish species, mostly in coral-associated species (Wilson et al., 2008, Wilson et al., 2010). The reduced abundance was attributed to declines in live coral cover (primarily Acroporids), which decreased habitat complexity and food availability, and reduced fishing pressure which altered predator-prey dynamics. Our surveys suggest that recovery of branching corals may have occurred between these previous studies. This may have resulted in an increase in abundance of small-bodied species as habitat rugosity improved (Pratchett et al., 2012; Graham and Nash, 2013). Although fishing in Lau is for subsistence, net fishing does occur and our results suggest that this method favors the larger bodied and slow growing species. Hence, very few large surgeonfish, parrotfish, groupers or snapper were recorded which is concerning as smaller individuals generally produce fewer and weaker offspring.

Wilson et al. (2008) identified a 50-70% decline in coral cover at both Matuka and Totoya, with a maximum of 24% coral cover in 2006. In 2013, coral cover was still lower at these islands compared with other locations in our surveys. This corresponded with the two lowest total mean densities of reef fish, particularly corallivores. However, total mean biomass was greatest in Totoya which indicates that fish assemblage here is dominated by large, roving species and not smaller species that depend on coral for refuge or food. Sharks, large parrotfish, jobfish and unicornfish were commonly observed around this area.

Many of these reefs are traditionally managed, but would benefit from additional fisheries management, particularly to increase the size structure of assemblages. Despite the remarkably low fish biomass seen elsewhere in Lau Province, reefs in Totoya had remarkably high biomass and large individuals from key predatory and herbivorous families. Hence these reefs would benefit from additional protection to continue sustaining high biomass. Interestingly, the greatest abundance of fish over 30 cm in length occurred around Moala, which also had the greatest human population. The protection of larger-bodied fish appears to be working in Moala and should be continued in order to maintain current assemblages. Of all the islands surveyed, Matuka supported the lowest overall biomass and density of reef fish. Although, several large (>50 cm) sharks, snappers,

unicornfish and parrotfish were recorded. Despite the fact that human population is not exceptionally high for the area, fishing intensity is amongst the greatest of Lau's islands and this may contribute to reduced density and biomass of fish (Dulvy et al., 2004).



Fig. 120. Crown of thorns starfish were seen throughout the Lau Province at low densities, with a single outbreak documented at Cicica.



Fig. 119. Macroalgae overgrowing a Diploastrea colony.



Fig. 121. A damselfish within its territory of algae.

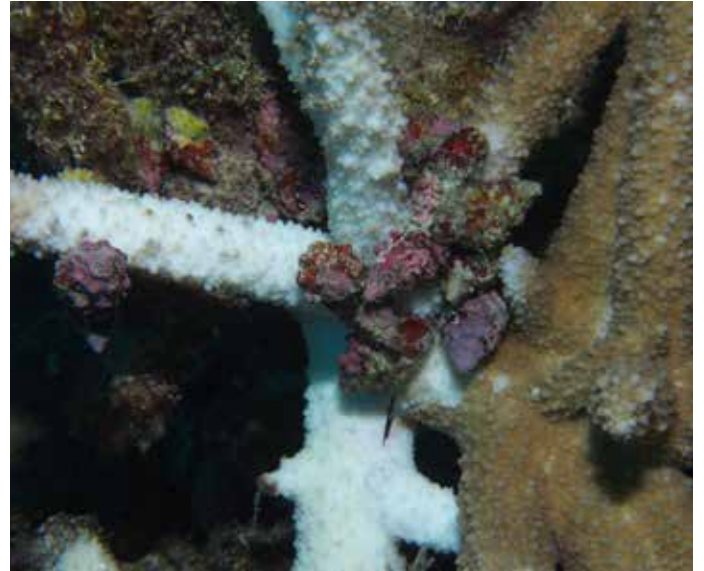
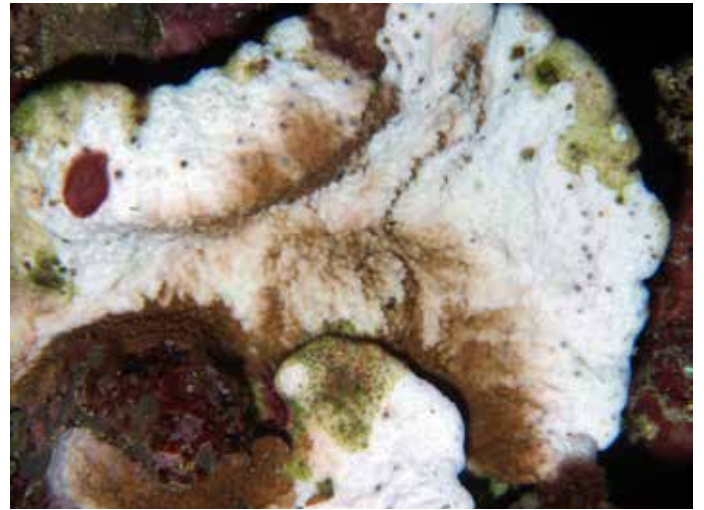


Fig. 122. Signs of stress on coral reefs in the Lau Province. White syndrome on a table acroporid (top left). yellow band disease on *Goniastrea* (top right). Lethal orange disease on CCA (center left). Bleached colony of *Montipora* (center right). Cyanobacteria overgrowing *Porites* (bottom left). *Drupella* snails on *Acropora* (bottom right).

Acknowledgements

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The Foundation worked closely with Wildlife Conservation Society, University of the South Pacific, Ministry of Fisheries, the Pacific Blue Foundation, Lau Provincial Council, and members of the Fiji Locally Managed Marine Area (FLMMA) Network on all aspects of this project. Roko Sau, the paramount chief of Totoya, helped facilitate communications with various chiefs, traditional leaders and community members. He also provided us with critical information on the local community traditional protocols and requirements for the scientific team throughout the islands we examined in the Lau Province. We are grateful for the collaborations with Stacy Jupiter, Laitia Ralao, Ron Vave and William Saladrau in our coral reef research and assistance they provided in our education and outreach activities.

The research missions to Fiji could never have been completed without the leadership, vision, and generosity of His Royal Highness Prince Khaled bin Sultan. We are deeply appreciative of his financial support and for the generous use of his research vessel, the *M/Y Golden Shadow*. His vision of *Science Without Borders*® was materialized in the research mission to Fiji through the partnerships and involvement by scientists from the following countries: Fiji, USA, New Zealand, Australia, Portugal, the Philippines, and Taiwan.

The Living Oceans Foundation appreciates the skill and dedication of the scientific divers that aided the Foundation in data collection, especially our international partners from NOVA Southeastern University, University of the Philippines, University

of the Azores, University of Miami, NOAA, University of Wellington, Florida Aquarium, and Taiwan Museum. I am particularly grateful for the dedicated efforts of each scientist, and thank each of you for your contributions, especially the detailed data you gathered, which has been incorporated into this report.

The Foundation would specifically like to thank William Saladrau from the Fijian Fisheries Department for his contribution to the Expedition. William passed away in 2014. He was very passionate about his work and the future of Fiji's reefs. He will be sorely missed.

The research missions benefited from Captain Steve Breen, Captain Mike Hitch and the officers and crew of the *M/Y Golden Shadow*. They were responsible for getting us safely to our research sites and conducting all logistical operations of the dive and research vessels. They ensured that each researcher has access to the study sites and proper working tools and equipment needed to complete the work, and had highly capable engineers and electricians that repaired and fabricated gear when we ran into complications. Behind the scenes, the crew worked at all hours to support the scientists on the Global Reef Expedition.

As the deliverables from this research project are completed, we look forward to continuing these partnerships to ensure that the information and data from this project is applied towards conservation needs and goals for the Lau Province.



Fig. 123. *M/Y Golden Shadow* in Fiji.

Appendix I: Participants



Name	Institution	Function
Philip Renaud	Khaled bin Sultan Living Oceans Foundation	Executive Director
Andrew Bruckner	Khaled bin Sultan Living Oceans Foundation	Chief Scientist
Amy Heemsoth	Khaled bin Sultan Living Oceans Foundation	Director of Education
Badi Samaniego	University of the Philippines, KSLOF Fellow	Fish Surveyor
Joao Monteiro	University of Azores, KSLOF Fellow	Coral Fluorescence
Jeremy Kerr	Nova Southeastern University, KSLOF Fellow	Groundtruthing / Habitat Mapping
Anderson Mayfield	National Museum of Marine Biology and Aquarium of Taiwan, KSLOF Fellow	Coral Genetics
Gwilym Rowlands	Nova Southeastern University National Coral Reef Institute	Groundtruthing / Habitat Mapping
Steve Saul	Nova Southeastern University National Coral Reef Institute (NCRI)	Groundtruthing / Habitat Mapping
Roko Sau Joesefa Cinavilakeba	Pacific Blue Foundation and Vice Chairman of the Lau Provincial Council	Traditional Leaders and Community Representative
Stacy Jupiter	Wildlife Conservation Society of Fiji	Local Scientist
Laitia Ralao	Lau Provincial Office	Protocol Officer
Ron Vave	University of the South Pacific	Scientific Diver
William Saladrau	Fiji Department of Fisheries	Scientific Diver
Ken Marks	Atlantic and Gulf Rapid Reef Assessment Program (AGRRA)	Photo Transects
Alexandra Dempsey	Nova Southeastern University National Coral Reef Institute (NCRI)	Benthic Surveyor
Dawn Bailey	Dive-In OCEAN Foundation	Coral Surveyor
Janet Eyre	Reef Environmental Education Foundation (REEF)	Fish Surveyor
Derek Manzello	NOAA	Ocean Acidification
Katie Hillyer	Victoria University, Wellington New Zealand	Benthic Surveyor
Nick Cautin	Dive Safety Officer	Diving Operations

Table 10 Participants and their roles on the Global Reef Expedition: Fiji.

Appendix II. Fish Species Lists

Fish Species List: Planktivores

Scientific Name	# of Survey Sites	Scientific Name	# of Survey Sites
<i>Acanthurus albipectoralis</i>	11	<i>Malacanthus latovittatus</i>	4
<i>Acanthurus nubilus</i>	1	<i>Genicanthus melanospilos</i>	12
<i>Acanthurus sp.</i>	1	<i>Genicanthus watanabei</i>	4
<i>Acanthurus thompsoni</i>	30	<i>Abudefduf sexfasciatus</i>	23
<i>Naso annulatus</i>	3	<i>Abudefduf vaigiensis</i>	9
<i>Naso brachycentron</i>	1	<i>Amblyglyphidodon aureus</i>	18
<i>Naso brevirostris</i>	26	<i>Amblyglyphidodon curacao</i>	6
<i>Naso caesioides</i>	2	<i>Amblyglyphidodon leucogaster</i>	1
<i>Naso hexacanthus</i>	15	<i>Amblyglyphidodon orbicularis</i>	28
<i>Naso lopezi</i>	3	<i>Chromis acares</i>	3
<i>Naso minor</i>	1	<i>Chromis agilis</i>	6
<i>Naso thynnoides</i>	1	<i>Chromis alpha</i>	30
<i>Naso tonganus</i>	6	<i>Chromis amboinensis</i>	50
<i>Naso vlamingii</i>	34	<i>Chromis atripectoralis</i>	16
<i>Caesio caerulea</i>	25	<i>Chromis atripes</i>	60
<i>Caesio lunaris</i>	6	<i>Chromis cf. leucura</i>	1
<i>Caesio teres</i>	15	<i>Chromis chrysur</i>	19
<i>Pterocaesio digramma</i>	4	<i>Chromis iomelas</i>	62
<i>Pterocaesio lativittata</i>	1	<i>Chromis lepidolepis</i>	36
<i>Pterocaesio pisang</i>	14	<i>Chromis margaritifer</i>	52
<i>Pterocaesio tile</i>	42	<i>Chromis retrofasciata</i>	48
<i>Pterocaesio trilineata</i>	38	<i>Chromis ternatensis</i>	52
<i>Decapterus macarellus</i>	2	<i>Chromis vanderbilti</i>	16
<i>Decapterus sp.</i>	1	<i>Chromis viridis</i>	32
<i>Hemitaenichthys polylepis</i>	27	<i>Chromis weberi</i>	22
<i>Heniochus acuminatus</i>	19	<i>Chromis xanthura</i>	60
<i>Platax pinnatus</i>	1	<i>Neopomacentrus metallicus</i>	1
<i>Myripristis adusta</i>	2	<i>Pomacentrus callainus</i>	63
<i>Myripristis berndti</i>	31	<i>Pomacentrus cf. adelus</i>	1
<i>Myripristis kuntee</i>	41	<i>Pomacentrus opisthostigma</i>	1
<i>Myripristis murdjan</i>	2	<i>Pomacentrus pavo</i>	6
<i>Myripristis sp.</i>	1	<i>Pomachromis richardsoni</i>	5
<i>Myripristis violacea</i>	26	<i>Nemateleotris magnifica</i>	26
<i>Myripristis vittata</i>	9	<i>Ptereleotris evides</i>	41
<i>Cirrhilabrus exquisitus</i>	13	<i>Ptereleotris heteroptera</i>	3
<i>Cirrhilabrus punctatus</i>	48	<i>Ptereleotris microlepis</i>	3
<i>Cirrhilabrus scottorum</i>	15	<i>Ptereleotris zebra</i>	3
<i>Cirrhilabrus sp.</i>	3	<i>Grammatorhynchus bilineatus</i>	1
<i>Pseudocoris yamashiroi</i>	8	<i>Rastrelliger kanagurta</i>	1
<i>Thalassoma amblycephalum</i>	22	<i>Pseudanthias pascualis</i>	40
<i>Hoplolatilus starcki</i>	7	<i>Pseudanthias squamipinnis</i>	8
<i>Malacanthus brevirostris</i>	6	<i>Serranocirrhites latus</i>	35

Table 11. These species of planktivorous fish were present on the following number of the 70 SCUBA fish abundance and diversity surveys we conducted in Lau Province, Fiji.

Fish Species List: Benthic Carnivores

Scientific Name	# of Survey Sites	Scientific Name	# of Survey Sites
<i>Apogon angustatus</i>	2	<i>Eviota dorsogilva</i>	2
<i>Apogon kallopterus</i>	2	<i>Eviota guttata</i>	14
<i>Apogon nigrofasciatus</i>	3	<i>Eviota parasites</i>	3
<i>Archamia biguttata</i>	1	<i>Eviota sebreei</i>	14
<i>Archamia fucata</i>	1	<i>Eviota sigillata</i>	5
<i>Cheilodipterus quinquelineatus</i>	12	<i>Eviota smaragdus</i>	1
<i>Nectamia savayensis</i>	1	<i>Exyrias bellissimus</i>	1
<i>Aulostomus chinensis</i>	29	<i>Fusigobius duospilus</i>	1
<i>Balistoides conspicillum</i>	10	<i>Fusigobius melacron</i>	1
<i>Balistoides viridescens</i>	16	<i>Fusigobius signipinnis</i>	3
<i>Odonus niger</i>	1	<i>Gnatholepis cauerensis</i>	3
<i>Pseudobalistes flavimarginatus</i>	7	<i>Gobiodon sp.</i>	8
<i>Rhinecanthus aculeatus</i>	1	<i>Istigobius rigilius</i>	1
<i>Rhinecanthus lunula</i>	3	<i>Koumansetta rainfordi</i>	9
<i>Rhinecanthus rectangulus</i>	1	<i>Trimma annosum</i>	1
<i>Sufflamen bursa</i>	55	<i>Trimma benjamini</i>	1
<i>Sufflamen chrysopterus</i>	23	<i>Trimma caesiura</i>	1
<i>Sufflamen fraenatum</i>	1	<i>Trimma cf. tevegae</i>	5
<i>Xanthichthys auromarginatus</i>	1	<i>Trimma sp.</i>	3
<i>Ecsenius bicolor</i>	3	<i>Trimma taylori</i>	1
<i>Ecsenius fijiensis</i>	8	<i>Valenciennea helsdingenii</i>	18
<i>Ecsenius sp. (checkered)</i>	1	<i>Valenciennea parva</i>	1
<i>Meiacanthus bundoon</i>	30	<i>Valenciennea puellaris</i>	1
<i>Meiacanthus oualanensis</i>	21	<i>Valenciennea strigata</i>	10
<i>Diplogrammus goramensis</i>	1	<i>Plectorhinchus chaetodonoides</i>	6
<i>Synchiropus morrisoni</i>	1	<i>Plectorhinchus gibbosus</i>	1
<i>Caracanthus maculatus</i>	1	<i>Plectorhinchus picus</i>	3
<i>Selaroides leptolepis</i>	1	<i>Plectorhinchus vittatus</i>	1
<i>Chaetodon flavirostris</i>	3	<i>Neoniphon argenteus</i>	1
<i>Chaetodon lunula</i>	14	<i>Neoniphon opercularis</i>	5
<i>Heniochus monoceros</i>	41	<i>Neoniphon sammara</i>	31
<i>Neocirrhitis armatus</i>	3	<i>Sargocentron caudimaculatum</i>	33
<i>Gorgasia maculata</i>	3	<i>Sargocentron cornutum</i>	1
<i>Heteroconger sp.</i>	1	<i>Sargocentron diadema</i>	3
<i>Dasyatis kuhlii</i>	2	<i>Sargocentron ittodai</i>	1
<i>Amblyeleotris guttata</i>	19	<i>Sargocentron rubrum</i>	3
<i>Amblyeleotris steinitzi</i>	9	<i>Sargocentron spiniferum</i>	22
<i>Amblyeleotris wheeleri</i>	3	<i>Sargocentron tiere</i>	4
<i>Amblygobius phalaena</i>	5	<i>Anampses caeruleopunctatus</i>	13
<i>Amblygobius sp.</i>	2	<i>Anampses melanurus</i>	1
<i>Bryaninops natans</i>	4	<i>Anampses neoguinaicus</i>	34
<i>Bryaninops yongei</i>	1	<i>Anampses twistii</i>	59
<i>Ctenogobiops crocineus</i>	1	<i>Bodianus anthioides</i>	17

Table 12. These species of benthic carnivorous fish were present on the following number of the 70 SCUBA fish abundance and diversity surveys we conducted in Lau Province, Fiji.

Fish Species List: Benthic Carnivores (cont.)

Scientific Name	# of Survey Sites	Scientific Name	# of Survey Sites
<i>Bodianus axillaris</i>	57	<i>Thalassoma purpureum</i>	1
<i>Bodianus dictynna</i>	4	<i>Thalassoma quinquevittatum</i>	26
<i>Bodianus loxozonus</i>	43	<i>Thalassoma trilobatum</i>	1
<i>Bodianus mesothorax</i>	29	<i>Gnathodentex aureolineatus</i>	24
<i>Cheilinus chlorourus</i>	15	<i>Lethrinus obsoletus</i>	10
<i>Cheilinus fasciatus</i>	25	<i>Monotaxis grandoculis</i>	29
<i>Cheilinus oxycephalus</i>	52	<i>Monotaxis heterodon</i>	66
<i>Cheilinus trilobatus</i>	39	<i>Cantherhines longicaudus</i>	1
<i>Cheilinus undulatus</i>	14	<i>Cantherhines pardalis</i>	21
<i>Choerodon jordani</i>	5	<i>Mulloidichthys flavolineatus</i>	5
<i>Coris batuensis</i>	6	<i>Mulloidichthys vanicolensis</i>	9
<i>Coris dorsomacula</i>	10	<i>Parupeneus barberinoides</i>	3
<i>Coris gaimard</i>	31	<i>Parupeneus barberinus</i>	33
<i>Coris sp. (3 stripes) (C. notialis)</i>	1	<i>Parupeneus ciliatus</i>	3
<i>Zanclus cornutus</i>	63	<i>Parupeneus crassilabris</i>	48
<i>Gomphosus varius</i>	66	<i>Parupeneus cyclostomus</i>	61
<i>Halichoeres biocellatus</i>	59	<i>Parupeneus multifasciatus</i>	67
<i>Halichoeres claudia</i>	22	<i>Parupeneus pleurostigma</i>	33
<i>Halichoeres hortulanus</i>	62	<i>Gymnothorax javanicus</i>	1
<i>Halichoeres margaritaceus</i>	4	<i>Aetobatus narinari</i>	1
<i>Halichoeres marginatus</i>	19	<i>Manta birostris</i>	1
<i>Halichoeres prosopion</i>	15	<i>Scolopsis bilineatus</i>	50
<i>Halichoeres richmondi</i>	4	<i>Scolopsis trilineatus</i>	1
<i>Halichoeres trimaculatus</i>	6	<i>Opistognathus sp.</i>	1
<i>Hemigymnus fasciatus</i>	53	<i>Ostracion meleagris</i>	21
<i>Hemigymnus melapterus</i>	14	<i>Ostracion solorensis</i>	23
<i>Hologymnosus annulatus</i>	11	<i>Heteropriacanthus cruentatus</i>	1
<i>Hologymnosus doliatus</i>	6	<i>Pempheris oualensis</i>	11
<i>Macropharyngodon meleagris</i>	26	<i>Parapercis clathrata</i>	15
<i>Macropharyngodon negrosensis</i>	3	<i>Parapercis hexophthalma</i>	26
<i>Novaculichthys taeniourus</i>	9	<i>Parapercis sp. (Allen)</i>	2
<i>Oxycheilinus rhodochrous</i>	3	<i>Pempheris schwenkii</i>	1
<i>Pseudocheilinus evanidus</i>	51	<i>Apolemichthys trimaculatus</i>	2
<i>Pseudocheilinus hexataenia</i>	65	<i>Centropyge bicolor</i>	36
<i>Pseudocheilinus octotaenia</i>	35	<i>Pygoplites diacanthus</i>	58
<i>Pseudocheilinus tetrataenia</i>	1	<i>Cypho purpurascens</i>	8
<i>Stethojulis bandanensis</i>	40	<i>Pictichromis porphyreus</i>	3
<i>Stethojulis notialis</i>	2	<i>Solenostomus paegnius</i>	1
<i>Stethojulis trilineata</i>	4	<i>Arothron cf. caeruleopunctatus</i>	1
<i>Thalassoma hardwicke</i>	44	<i>Arothron hispidus</i>	3
<i>Thalassoma janseni</i>	10	<i>Arothron meleagris</i>	1
<i>Thalassoma lunare</i>	33	<i>Arothron nigropunctatus</i>	23
<i>Thalassoma lutescens</i>	54	<i>Arothron stellatus</i>	2

Table 12 (continued). These species of benthic carnivorous fish were present on the following number of the 70 SCUBA fish abundance and diversity surveys we conducted in Lau Province, Fiji.

Fish Species List: Corallivores, Omnivores & Other Species

Corallivores

Scientific Name	# of Survey Sites
<i>Exallias brevis</i>	3
<i>Chaetodon baronessa</i>	44
<i>Chaetodon bennetti</i>	24
<i>Chaetodon citrinellus</i>	42
<i>Chaetodon ephippium</i>	52
<i>Chaetodon kleinii</i>	14
<i>Chaetodon lineolatus</i>	8
<i>Chaetodon lunulatus</i>	66
<i>Chaetodon melannotus</i>	10
<i>Chaetodon mertensii</i>	49
<i>Chaetodon ornatissimus</i>	10
<i>Chaetodon pelewensis</i>	65
<i>Chaetodon plebeius</i>	29
<i>Chaetodon rafflesii</i>	47
<i>Chaetodon reticulatus</i>	63
<i>Chaetodon trifascialis</i>	41
<i>Chaetodon ulietensis</i>	45
<i>Chaetodon unimaculatus</i>	24
<i>Chaetodon vagabundus</i>	55
<i>Forcipiger flavissimus</i>	61
<i>Forcipiger longirostris</i>	37
<i>Heniochus chrysostomus</i>	58
<i>Heniochus singularis</i>	6
<i>Heniochus varius</i>	32
<i>Coris aygula</i>	14
<i>Labrichthys unilineatus</i>	30
<i>Labropsis australis</i>	30
<i>Labropsis xanthonota</i>	49
<i>Cantherhines dumerilii</i>	18
<i>Oxymonacanthus longirostris</i>	30
<i>Plectroglyphidodon dickii</i>	30
<i>Plectroglyphidodon johnstonianus</i>	44

Omnivores

Scientific Name	# of Survey Sites
<i>Balistapus undulatus</i>	64
<i>Melichthys vidua</i>	38
<i>Chaetodon auriga</i>	31
<i>Chaetodon semeion</i>	2
<i>Kyphosus cinerascens</i>	5
<i>Aluterus scriptus</i>	1
<i>Pervagor janthinosoma</i>	6
<i>Pervagor melanocephalus</i>	1
<i>Ostracion cubicus</i>	4
<i>Pomacanthus imperator</i>	9
<i>Amphiprion barberi</i>	11
<i>Amphiprion chrysopterus</i>	47
<i>Amphiprion clarkii</i>	4
<i>Amphiprion fijiensis</i>	8
<i>Amphiprion perideraion</i>	19
<i>Dascyllus aruanus</i>	14
<i>Dascyllus reticulatus</i>	54
<i>Neoglyphidodon carlsoni</i>	17
<i>Pomacentrus brachialis</i>	1
<i>Pomacentrus imitator</i>	67
<i>Pomacentrus vaiuli</i>	67
<i>Priacanthus hamrur</i>	7

Other Species

Scientific Name	# of Survey Sites
<i>Canthigaster solandri</i>	11
<i>Canthigaster valentini</i>	20
<i>Ctenochaetus binotatus</i>	38
<i>Ctenochaetus cyanocheilus</i>	43
<i>Ctenochaetus striatus</i>	67
<i>Aspidontus taeniatus</i>	1
<i>Plagiotremus flavus</i>	34
<i>Plagiotremus rhinorhynchus</i>	4
<i>Plagiotremus tapeinosoma</i>	9
<i>Labroides bicolor</i>	41
<i>Labroides dimidiatus</i>	67
<i>Labroides rubrolabiatus</i>	2

Table 13, 14 & 15. These species of fish were present on the following number of the 70 SCUBA fish abundance and diversity surveys we conducted in Lau Province, Fiji.

Fish Species List: Herbivores

Scientific Name	# of Survey Sites	Scientific Name	# of Survey Sites
<i>Acanthurus blochii</i>	8	<i>Pomacentrus nigromanus</i>	2
<i>Acanthurus grammoptilus</i>	1	<i>Pomacentrus nigromarginatus</i>	11
<i>Acanthurus guttatus</i>	3	<i>Pomacentrus spilotoceps</i>	19
<i>Acanthurus lineatus</i>	45	<i>Stegastes fasciolatus</i>	38
<i>Acanthurus mata</i>	7	<i>Stegastes nigricans</i>	21
<i>Acanthurus nigricans</i>	22	<i>Calotomus carolinus</i>	14
<i>Acanthurus nigricauda</i>	56	<i>Cetoscarus bicolor</i>	42
<i>Acanthurus nigrofuscus</i>	57	<i>Chlorurus bleekeri</i>	11
<i>Acanthurus olivaceus</i>	8	<i>Chlorurus frontalis</i>	5
<i>Acanthurus pyroferus</i>	62	<i>Chlorurus microrhinos</i>	51
<i>Acanthurus triostegus</i>	12	<i>Chlorurus sordidus</i>	66
<i>Naso lituratus</i>	59	<i>Hipposcarus longiceps</i>	30
<i>Naso unicornis</i>	19	<i>Scarus altipinnis</i>	40
<i>Zebrasoma scopas</i>	65	<i>Scarus chameleon</i>	43
<i>Zebrasoma veliferum</i>	50	<i>Scarus dimidiatus</i>	10
<i>Cirripectes castaneus</i>	9	<i>Scarus forsteni</i>	39
<i>Cirripectes stigmaticus</i>	38	<i>Scarus frenatus</i>	33
<i>Cirripectes variolosus</i>	1	<i>Scarus ghobban</i>	7
<i>Kyphosus vaigiensis</i>	6	<i>Scarus globiceps</i>	36
<i>Pseudodax moluccanus</i>	10	<i>Scarus longipinnis</i>	6
<i>Pseudojuloides cerasinus</i>	7	<i>Scarus niger</i>	62
<i>Amanses scopas</i>	6	<i>Scarus oviceps</i>	10
<i>Paraluteres prionurus</i>	4	<i>Scarus psittacus</i>	52
<i>Centropyge bispinosus</i>	65	<i>Scarus rivulatus</i>	2
<i>Centropyge flavissimus</i>	62	<i>Scarus rubroviolaceus</i>	16
<i>Centropyge heraldi</i>	10	<i>Scarus schlegeli</i>	64
<i>Paracentropyge multifasciata</i>	6	<i>Scarus sp.</i>	1
<i>Chrysiptera rollandi</i>	2	<i>Scarus spinus</i>	33
<i>Chrysiptera sp. (brown)</i>	1	<i>Siganus argenteus</i>	16
<i>Chrysiptera talboti</i>	51	<i>Siganus cf. javus</i>	1
<i>Chrysiptera taupou</i>	47	<i>Siganus doliatus</i>	6
<i>Dascyllus trimaculatus</i>	54	<i>Siganus punctatissimus</i>	18
<i>Dischistodus sp.</i>	1	<i>Siganus punctatus</i>	10
<i>Plectroglyphidodon lacrymatus</i>	63	<i>Siganus spinus</i>	1
<i>Pomacentrus bankanensis</i>	5	<i>Siganus stellatus</i>	1
<i>Pomacentrus coelestis</i>	6	<i>Siganus uspi</i>	22
<i>Pomacentrus maafu</i>	27	<i>Canthigaster amboinensis</i>	
<i>Pomacentrus microspilus</i>	5		

Table 16. These species of herbivorous fish were present on the following number of the 70 SCUBA fish abundance and diversity surveys we conducted in Lau Province, Fiji.

Fish Species List: Piscivores

Scientific Name	# of Survey Sites	Scientific Name	# of Survey Sites
<i>Cheilodipterus artus</i>	3	<i>Lutjanus kasmira</i>	23
<i>Cheilodipterus isostigmus</i>	1	<i>Lutjanus monostigma</i>	27
<i>Cheilodipterus macrodon</i>	14	<i>Lutjanus semicinctus</i>	10
<i>Tylosurus crocodilus</i>	1	<i>Macolor macularis</i>	28
<i>Carangoides ferdau</i>	2	<i>Macolor niger</i>	30
<i>Carangoides orthogrammus</i>	20	<i>Gymnothorax flavimarginatus</i>	2
<i>Carangoides plagiotaenia</i>	1	<i>Gymnosarda unicolor</i>	14
<i>Caranx ignobilis</i>	1	<i>Sarda orientalis</i>	2
<i>Caranx melampygus</i>	7	<i>Sarda sp.</i>	1
<i>Elagatis bipinnulata</i>	3	<i>Scomberomorus commerson</i>	5
<i>Scomberoides lysan</i>	4	<i>Pterois antennata</i>	1
<i>Carcharhinus amblyrhynchos</i>	6	<i>Anyperodon leucogrammicus</i>	9
<i>Carcharhinus melanopterus</i>	1	<i>Belonoperca chabanaudi</i>	11
<i>Negaprion acutidens</i>	1	<i>Cephalopholis argus</i>	52
<i>Triaenodon obesus</i>	23	<i>Cephalopholis leopardus</i>	17
<i>Chanos chanos</i>	1	<i>Cephalopholis miniata</i>	1
<i>Cirrhitichthys falco</i>	17	<i>Cephalopholis sexmaculata</i>	2
<i>Paracirrhites arcatus</i>	55	<i>Cephalopholis spiloparaea</i>	15
<i>Paracirrhites forsteri</i>	47	<i>Cephalopholis urodeta</i>	55
<i>Paracirrhites hemistictus</i>	1	<i>Epinephelus caeruleopunctatus</i>	1
<i>Echeneis naucrates</i>	11	<i>Epinephelus cyanopodus</i>	1
<i>Fistularia commersonii</i>	4	<i>Epinephelus fasciatus</i>	4
<i>Epibulus brevis</i>	9	<i>Epinephelus howlandi</i>	1
<i>Epibulus insidiator</i>	59	<i>Epinephelus maculatus</i>	1
<i>Oxycheilinus bimaculatus</i>	3	<i>Epinephelus merra</i>	22
<i>Oxycheilinus digrammus</i>	67	<i>Epinephelus microspilos</i>	1
<i>Oxycheilinus unifasciatus</i>	11	<i>Epinephelus ongus</i>	1
<i>Gymnocranius microdon</i>	4	<i>Epinephelus polyphekadion</i>	14
<i>Lethrinus atkinsoni</i>	6	<i>Epinephelus tauvina</i>	2
<i>Lethrinus erythracanthus</i>	6	<i>Gracila albomarginata</i>	16
<i>Lethrinus erythropterus</i>	3	<i>Plectropomus laevis</i>	28
<i>Lethrinus harak</i>	1	<i>Plectropomus leopardus</i>	13
<i>Lethrinus olivaceus</i>	3	<i>Variola albimarginata</i>	1
<i>Lethrinus rivulatus</i>	1	<i>Variola louti</i>	16
<i>Lethrinus sp.</i>	1	<i>Sphyraena barracuda</i>	3
<i>Lethrinus xanthochilus</i>	4	<i>Sphyraena flavicauda</i>	1
<i>Aphareus furca</i>	55	<i>Sphyraena helleri</i>	24
<i>Aprion virescens</i>	10	<i>Sphyraena jello</i>	1
<i>Lutjanus bohar</i>	41	<i>Saurida gracilis</i>	2
<i>Lutjanus fulvus</i>	17	<i>Synodus dermatogenys</i>	2
<i>Lutjanus gibbus</i>	40	<i>Synodus variegatus</i>	21

Table 17. These species of picivorous fish were spotted on the following number of the 70 SCUBA fish abundance and diversity surveys we conducted in Lau Province, Fiji.

Appendix III. Coral Species List

Scientific Name	Scientific Name	Scientific Name
<i>Acanthastrea echinata</i>	<i>Cyphastrea serailia</i>	<i>Hydnophora exesa</i>
<i>Acanthastrea hemprichii</i>	<i>Diploastrea heliopora</i>	<i>Hydnophora grandis</i>
<i>Acanthastrea hillae</i>	<i>Echinophyllia aspera</i>	<i>Hydnophora microconos</i>
<i>Acanthastrea ishigakiensis</i>	<i>Echinophyllia echinata</i>	<i>Hydnophora rigida</i>
<i>Acropora aspera</i>	<i>Echinopora gemmacea</i>	<i>Leptastrea inaequalis</i>
<i>Acropora bifurcata</i>	<i>Echinopora horrida</i>	<i>Leptastrea pruinosa</i>
<i>Acropora brueggemanni</i>	<i>Echinopora lamellosa</i>	<i>Leptastrea purpurea</i>
<i>Acropora cerealis</i>	<i>Echinopora mammiformis</i>	<i>Leptastrea transversa</i>
<i>Acropora cuneata</i>	<i>Echinopora pacificus</i>	<i>Leptoria irregularis</i>
<i>Acropora cytherea</i>	<i>Euphyllia cristata</i>	<i>Leptoria phrygia</i>
<i>Acropora digitifera</i>	<i>Favia fавus</i>	<i>Leptoseris explanata</i>
<i>Acropora divaricata</i>	<i>Favia helianthoides</i>	<i>Leptoseris hawaiiensis</i>
<i>Acropora florida</i>	<i>Favia laxa</i>	<i>Leptoseris mycetoseroides</i>
<i>Acropora formosa</i>	<i>Favia maritima</i>	<i>Leptoseris yabei</i>
<i>Acropora gemmifera</i>	<i>Favia matthaii</i>	<i>Lobophyllia corymbosa</i>
<i>Acropora globiceps</i>	<i>Favia pallida</i>	<i>Lobophyllia hataii</i>
<i>Acropora grandis</i>	<i>Favia rotumana</i>	<i>Lobophyllia hemprichii</i>
<i>Acropora granulosa</i>	<i>Favia rotundata</i>	<i>Lobophyllia pachysepta</i>
<i>Acropora horrida</i>	<i>Favia speciosa</i>	<i>Merulina ampliata</i>
<i>Acropora humilis</i>	<i>Favia stelligera</i>	<i>Merulina scabricula</i>
<i>Acropora hyacinthus</i>	<i>Favites abdita</i>	<i>Montastrea curta</i>
<i>Acropora listeri</i>	<i>Favites chinensis</i>	<i>Montastrea magnistellata</i>
<i>Acropora macrostoma</i>	<i>Favites complanata</i>	<i>Montastrea valenciennesi</i>
<i>Acropora microphthalmalma</i>	<i>Favites flexuosa</i>	<i>Montipora angulata</i>
<i>Acropora palifera</i>	<i>Favites halicora</i>	<i>Montipora australiensis</i>
<i>Acropora retusa</i>	<i>Favites pentagona</i>	<i>Montipora calcarea</i>
<i>Acropora robusta</i>	<i>Favites russelli</i>	<i>Montipora caliculata</i>
<i>Acropora speciosa</i>	<i>Fungi spp.</i>	<i>Montipora capitata</i>
<i>Acropora spicifera</i>	<i>Galaxea astreata</i>	<i>Montipora capricornis</i>
<i>Acropora subulata</i>	<i>Galaxea fascicularis</i>	<i>Montipora cebuensis</i>
<i>Acropora tenella</i>	<i>Galaxea horrescens</i>	<i>Montipora confusa</i>
<i>Acropora tenuis</i>	<i>Gardineroseris planulata</i>	<i>Montipora danae</i>
<i>Astreopora myriophthalma</i>	<i>Goniastrea aspera</i>	<i>Montipora efflorescens</i>
<i>Astreopora randalli</i>	<i>Goniastrea edwardsi</i>	<i>Montipora effusa</i>
<i>Caulastrea furcata</i>	<i>Goniastrea favulus</i>	<i>Montipora floweri</i>
<i>Coscinaraea columna</i>	<i>Goniastrea pectinata</i>	<i>Montipora foliosa</i>
<i>Coscinaraea exesa</i>	<i>Goniastrea retiformis</i>	<i>Montipora foveolata</i>
<i>Coscinaraea wellsi</i>	<i>Halomitra pileus</i>	<i>Montipora grisea</i>
<i>Ctenactis echinata</i>	<i>Herpolitha limax</i>	<i>Montipora hispida</i>
<i>Cyphastrea microphthalmalma</i>	<i>Herpolitha weberi</i>	<i>Montipora spumosa</i>

Table 18. These species of corals were spotted on at least one of the SCUBA surveys we conducted at 70 dive sites in Lau Province, Fiji.

Coral Species List - Continued

Scientific Name	Scientific Name
<i>Montipora venosa</i>	<i>Stylocoeniella armata</i>
<i>Montipora verrucosa</i>	<i>Stylocoeniella guentheri</i>
<i>Mycedium elephantotus</i>	<i>Stylophora pistillata</i>
<i>Pachyseris rugosa</i>	<i>Stylophora subseriata</i>
<i>Pachyseris speciosa</i>	<i>Symphyllia radians</i>
<i>Pavona clavus</i>	<i>Symphyllia recta</i>
<i>Pavona explanulata</i>	<i>Turbinaria frondens</i>
<i>Pavona maldivensis</i>	<i>Turbinaria mesenterina</i>
<i>Pavona minuta</i>	<i>Turbinaria peltata</i>
<i>Pavona varians</i>	<i>Turbinaria reniformis</i>
<i>Pavona venosa</i>	
<i>Physogyra lichtensteini</i>	
<i>Platygyra daedalea</i>	
<i>Platygyra pini</i>	
<i>Plerogyra simplex</i>	
<i>Plerogyra sinuosa</i>	
<i>Pocillopora damicornis</i>	
<i>Pocillopora elegans</i>	
<i>Pocillopora eydouxi</i>	
<i>Pocillopora kelleheri</i>	
<i>Pocillopora ligulata</i>	
<i>Pocillopora meandrina</i>	
<i>Pocillopora verrucosa</i>	
<i>Pocillopora woodjonesi</i>	
<i>Porites cylindrica</i>	
<i>Porites lichen</i>	
<i>Porites lobata</i>	
<i>Porites rus</i>	
<i>Psammocora contigua</i>	
<i>Psammocora digitata</i>	
<i>Psammocora explanulata</i>	
<i>Psammocora haimeana</i>	
<i>Psammocora nierstraszi</i>	
<i>Psammocora obtusangula</i>	
<i>Psammocora profundacella</i>	
<i>Psammocora superficialis</i>	
<i>Sandalolitha robusta</i>	
<i>Scapophyllia cylindrica</i>	
<i>Seriatopora caliendrum</i>	
<i>Seriatopora hystrix</i>	

Table 18 (continued). These species of corals were spotted on at least one of the SCUBA surveys we conducted at 70 dive sites in Lau Province, Fiji.

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