

Beginn der Vorlesung

- 740.750 Einführung in die Pharmazeutische Analytik + 725.042, Analytische Grundvorlesung I
 - wöchentlich; Di, Mi, Do; 11:30 - 13:00, L.EG.220
- Teil 1: Einführung und Gravimetrie
- Website (Skripten und Infos)
<http://www.uibk.ac.at/acrc/mitarbeiter/bonn/lehre.html>
- Fragen bezgl. Vorlesung
 - Email -> christoph.woelger@uibk.ac.at

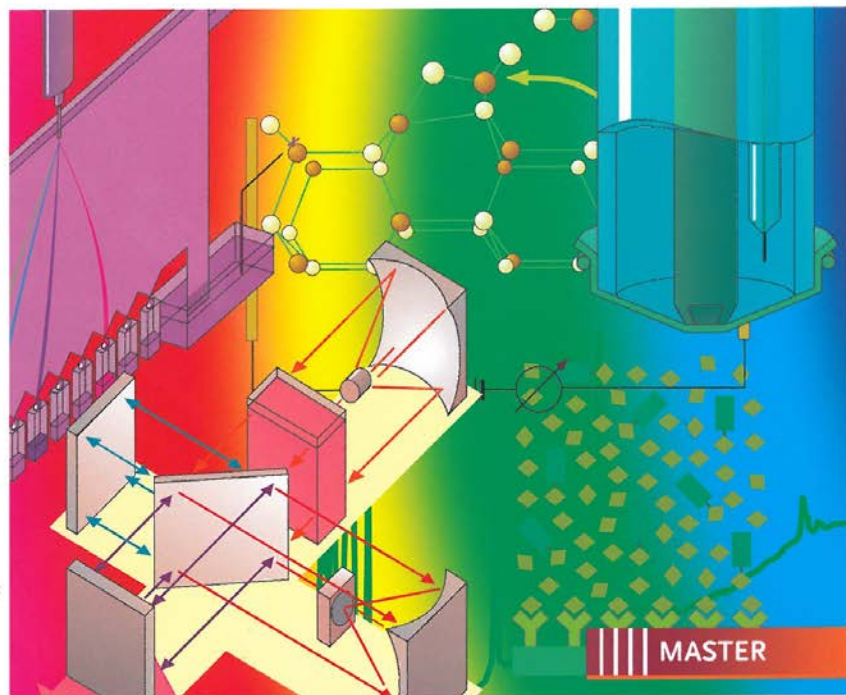
Georg Schwedt

 WILEY-VCH

Analytische Chemie

Grundlagen, Methoden und Praxis

Zweite, vollständig überarbeitete Auflage



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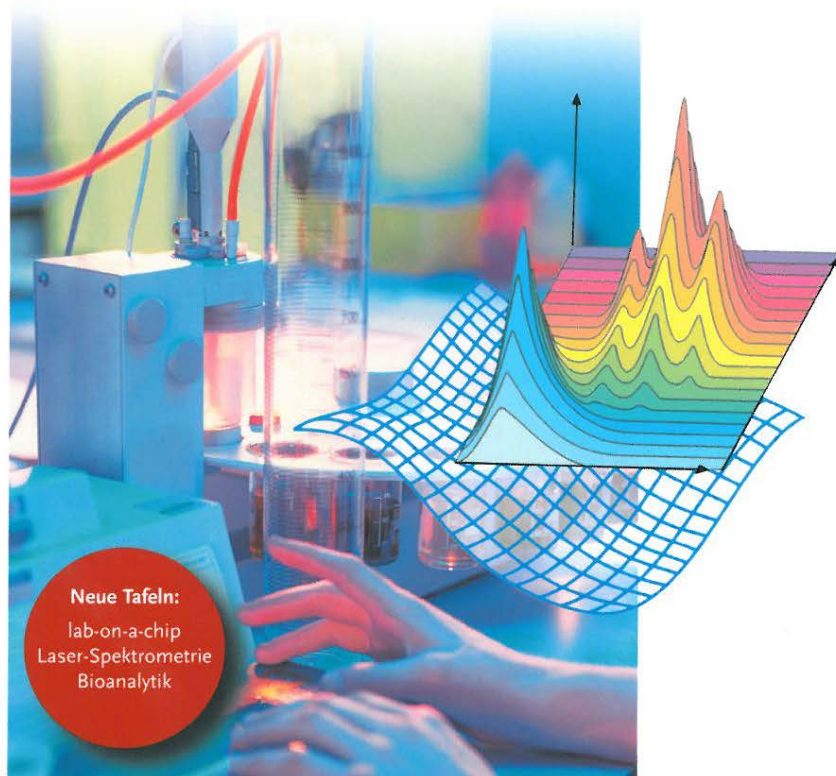
Bindung Litges & Dopf GmbH, Heppenheim

ISBN: 978-3-527-31206-1

Georg Schwedt

Taschenatlas der Analytik

117 Farbtafeln von Joachim Schreiber
3., überarbeitete und erweiterte Auflage



Neue Tafeln:

lab-on-a-chip
Laser-Spektrometrie
Bioanalytik

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Analytische Grundvorlesung I

Quantitative Analyseverfahren

- ❖ Gravimetrie

Fällung eines Niederschlages – Wägung und Berechnung

- ❖ Maßanalyse

Säure-Base Titrationsen, Komplexbildungstitrationen, Fällungstitrationen,
Redox-titrationsen

❖ Optische Analyseverfahren - Spektroskopie

Lambert-Beersches Gesetz, Photometrie, Fluoreszenzspektroskopie,
UV, IR-Spektroskopie

❖ Trennoperationen

Fällung, Destillation, Verteilung, Ionenaustausch etc.

❖ Chromatographie

Theorie, Papierchromatographie, Dünnschichtchromatographie, Flüssigkeitschromatographie,
Gaschromatographie, Apparaturen, Detektionssysteme

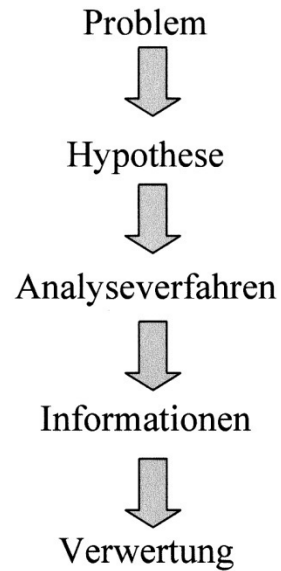
❖ Elektrophorese

Grundlagen, Gelelektrophorese, Kapillarelektrophorese

Was ist Analytische Chemie?

Chemische Analytik ist die Wissenschaft von der Gewinnung und verwertungsbezogenen Interpretation von Informationen über stoffliche Systeme mit Hilfe naturwissenschaftlicher Methoden. (IUPAC 1974)

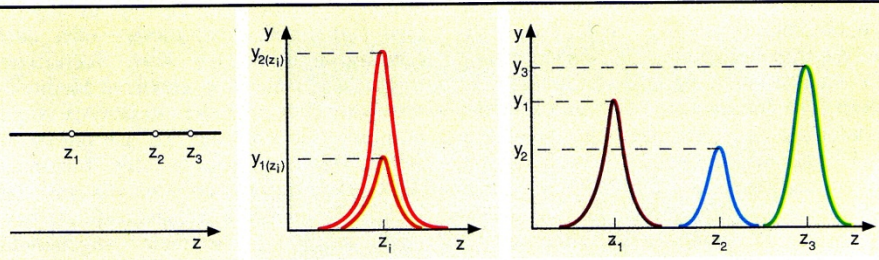
Der analytische Prozess (1)



Grundoperationen („unit operations“)

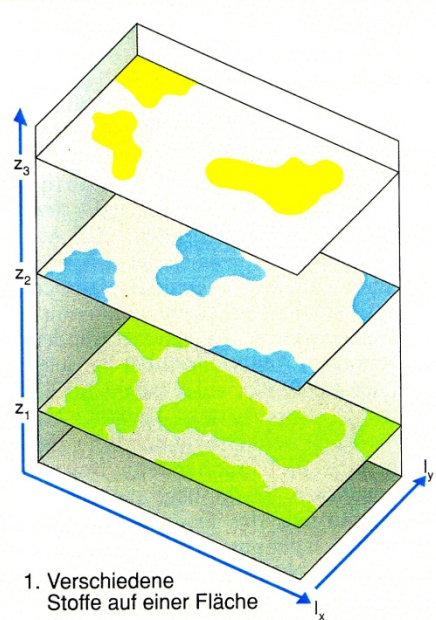
Jede Analyse besteht aus folgenden Teilschritten (Grundoperationen):

- o Probenaufnahme (sampling)
- o Probenvorbereitung (Trocknen, Lösen, Trennen)
- o Messung (Messwert)
- o Berechnung (Analysenwert) und Bewertung

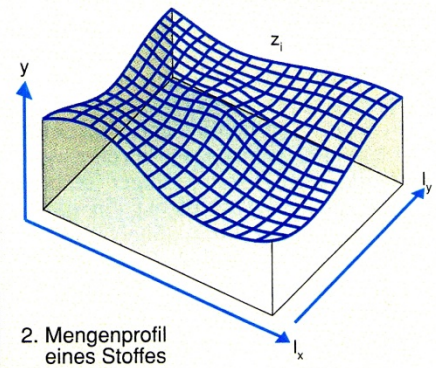


1. Qualitativ 2. Quantitativ 3. Qualitativ und quantitativ

A. Gehaltsanalyse

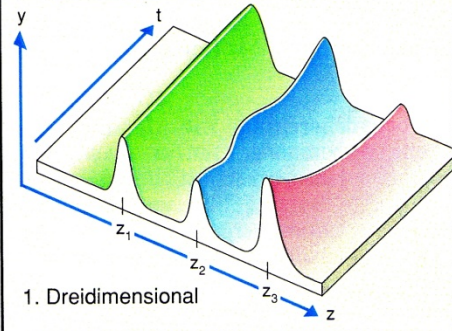


1. Verschiedene Stoffe auf einer Fläche

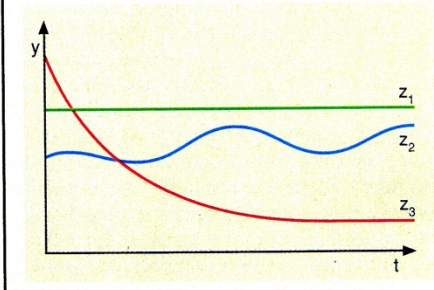


2. Mengenprofil eines Stoffes

B. Verteilungsanalyse

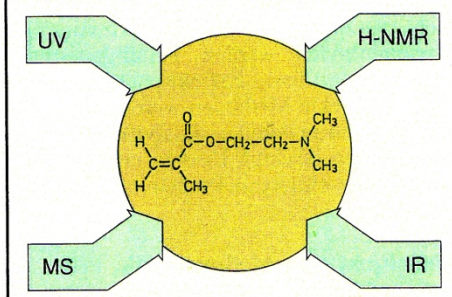


1. Dreidimensional

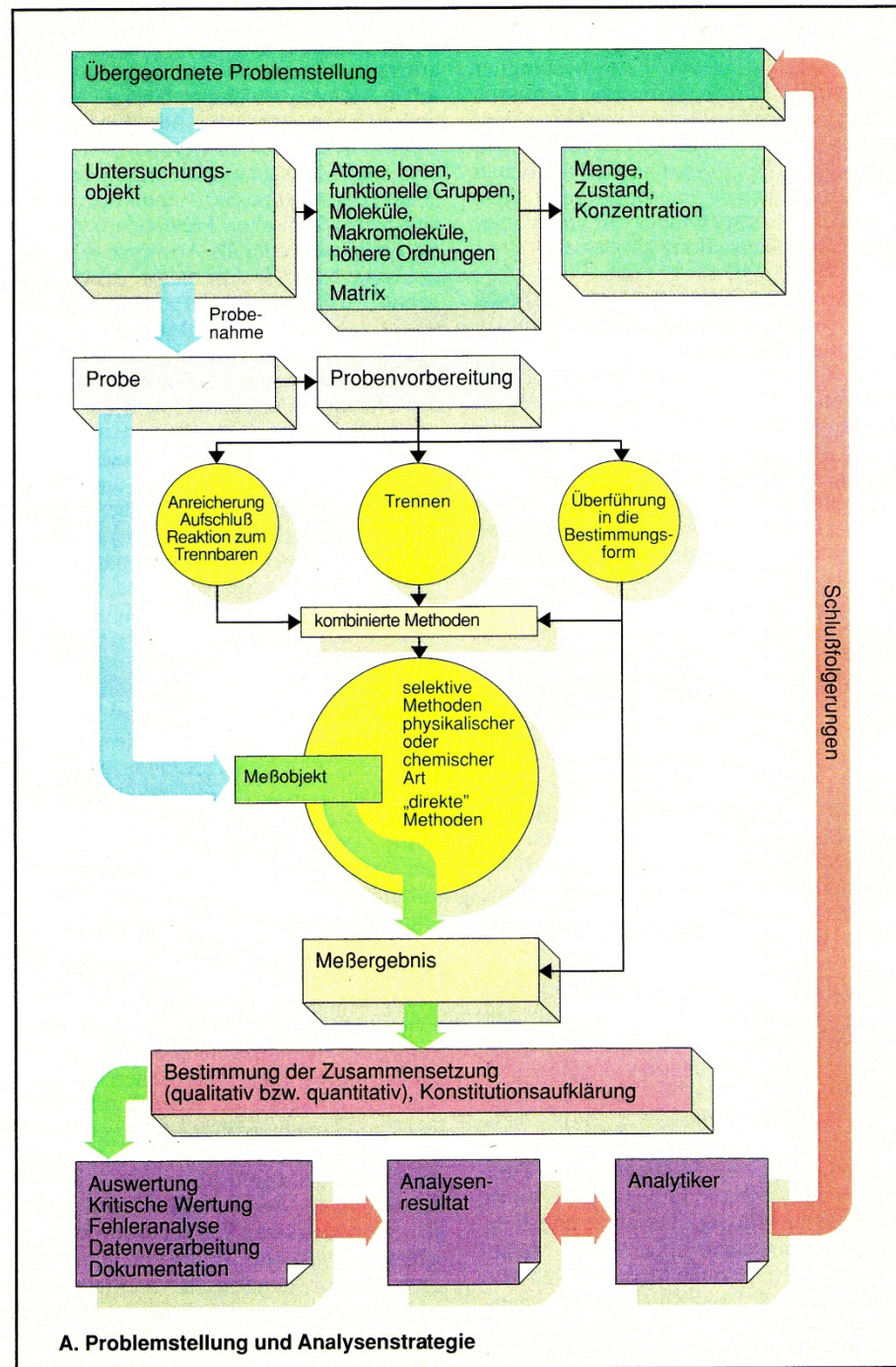


2. Zweidimensional

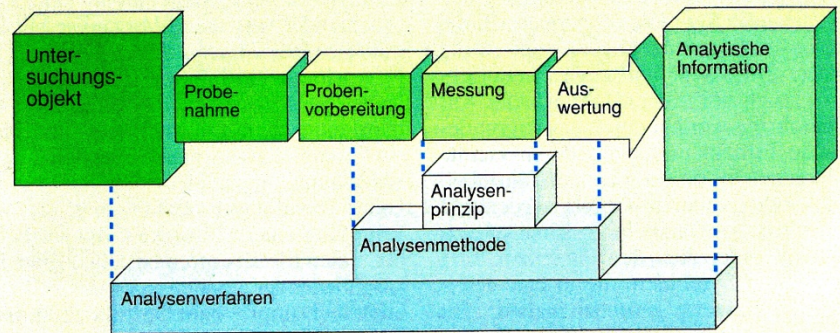
C. Prozessanalyse



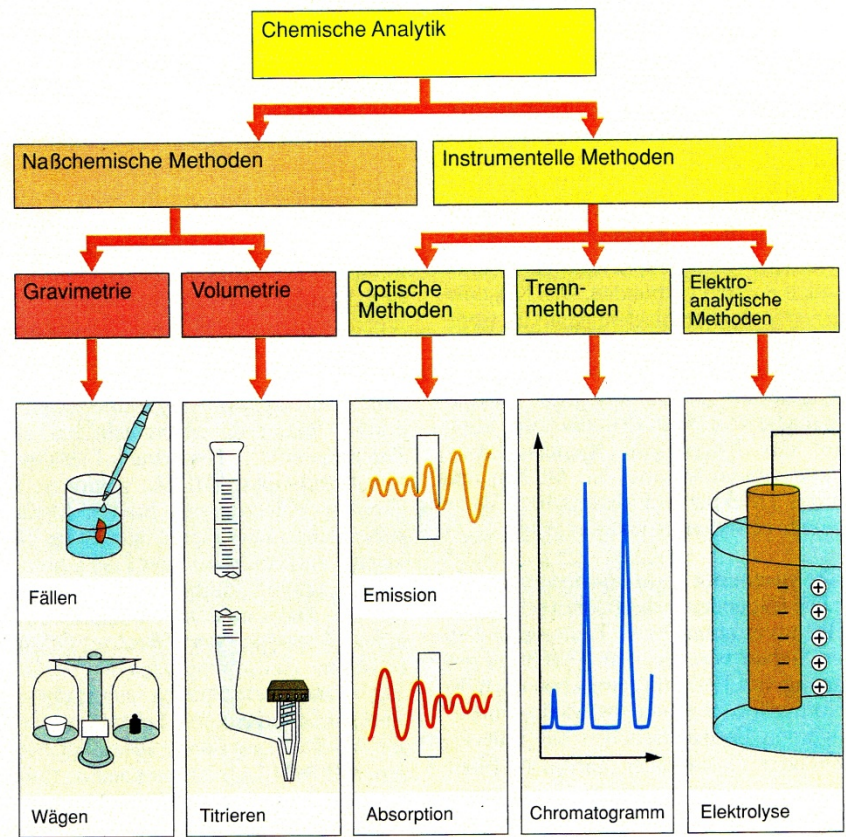
D. Strukturanalyse



A. Problemstellung und Analysenstrategie



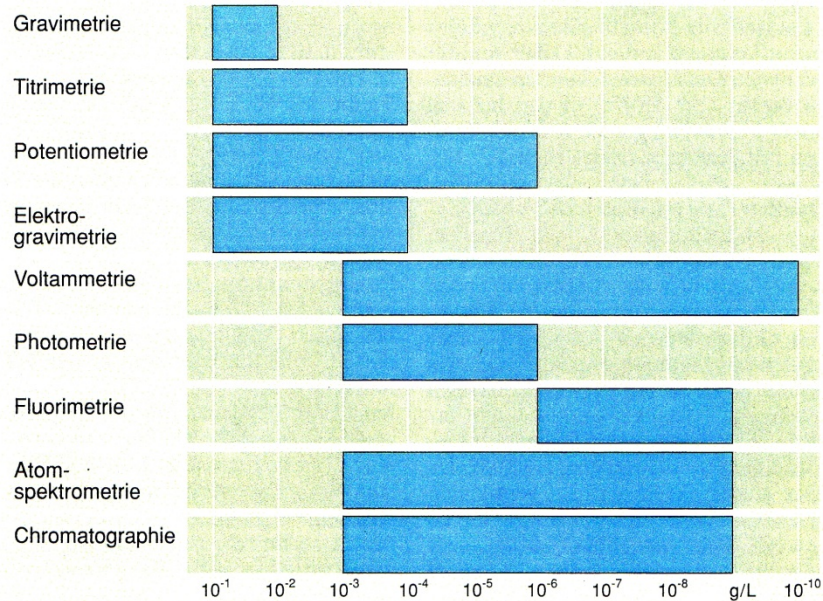
A. Analysenprinzip, Analysenmethode und Analysenverfahren



B. Systematik der Analysenmethoden

p	Probenmassenbereich S Absolutmassenbereich Q	Gehaltsbereich C	p
+2	Grammbereich (g)	Makroprobe	+2
+1	Dezigrammbereich (dg)		+1
0	Zentigrammbereich (cg)	Mesoprobe	0
-1	Milligrammbereich (mg)		-1
-2	Mikrogrammbereich (μg)	Mikroprobe	-2
-3		Submikroprobe	-3
-4		Ultramikroprobe	-4
-5		Spuren	-5
-6	Nanogrammbereich (ng)		-6
-7		Bereich der Spurenbestandteile	-7
-8			-8
-9		Mikrospuren	-9
-10	Picogrammbereich (pg)		-10
-11		Nanospuren	-11
-12	Femtogrammbereich (fg)		-12
-13		Picospuren	-13
-14	Attogrammbereich (ag)		-14
-15		ppm	-15
-16			-16
-17		ppb	-17
-18			-18

A. Arbeitsbereiche



B. Vergleich von Analysenmethoden

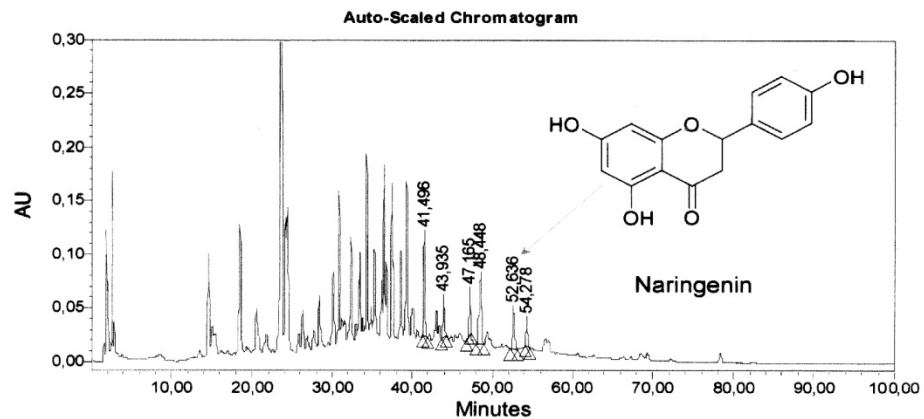
Arzneimittelanalyse

- Reinheit, Qualitätskontrolle
- Von Reinstoffen
- Von Gemischen in der Pharmazie
Zunehmend von Bedeutung : Phytochemische Präparate

Sample Information

SampleName Sinupret 00113011
Vial 2
Injection 1
Injection Volume 20,00 ul
Channel 996
Run Time 160,0 Minutes
Channel Description PDA 190.0 to 400.0 nm at 1.2 nm

Sample Type Unknown
Date Acquired 21.02.2001 19:34:42
Acq Method Set dwell 150 min
Processing Method Naringenin
Date Processed 28.02.2001 15:21:10



Sinupret®

Medizinische Analysen – Diagnostik

- Pharmakologische Wirkung
 - Wirkstoffanalytik
 - Abbauprodukte, Blut
 - Kinetik der Wirksubstanz
- Blut, Harn
- Genomics, Proteomics, Metabolomics

Umweltanalysen

- Luft, Wasser, Boden
 - Schadstoffanalytik, z.B. Dioxine

Lebensmittelanalytik

- Inhaltsstoffe in Lebensmitteln
 - z.B. Wein – Resveratrol

Kriterien für die Wahl des Analysenverfahrens

- Geforderte Information
- Genauigkeit, Präzision
- Probemenge
- Voraussichtlicher Gehalt der Probe
- Empfindlichkeit
- Selektivität
- Qualitative Zusammensetzung
- Analytische Operationen
- Ausrüstung
- Zeit
- Kosten

Probemenge – Gehalt - Arbeitsbereich

❖ Probemenge, Probenbereich (sample size) **P** :

Menge (meist der Masse) der Analysenprobe

Makrobereich: $m > 100 \text{ mg}$

Halbmakrobereich: $100 \text{ mg} > m > 10 \text{ mg}$

Mirkobereich: $m < 10 \text{ mg}$

❖ Gehalt, Gehaltsbereich (content) **G** :

Gehalt der Probe an der bestimmten Komponente

Hauptbestandteil : $w_i > 10\%$

Nebenbestandteil: $10\% > w_i > 1\%$

Spurenbestandteil: $w_i < 1\%$

❖ **Arbeitsbereich A:**

Menge (Masse) der Komponente i, welche mit der Methode bestimmt werden kann

$$A = G \times P$$

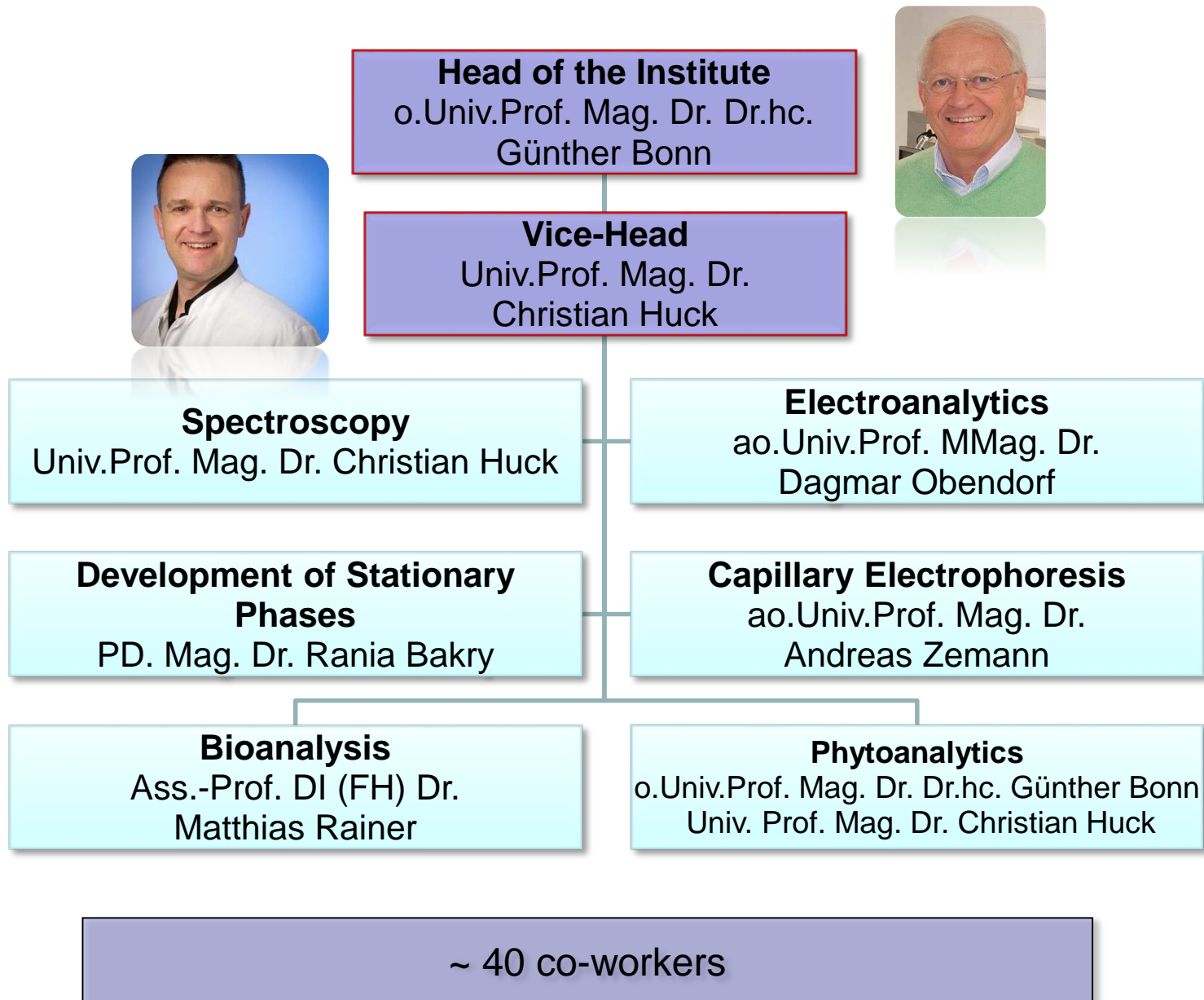
❖ **Nachweisgrenze, Bestimmungsgrenze:**

Untergrenze des Arbeitsbereichs (3s – Regel!)

Größen und Einheiten – heutige und frühere Bezeichnungen

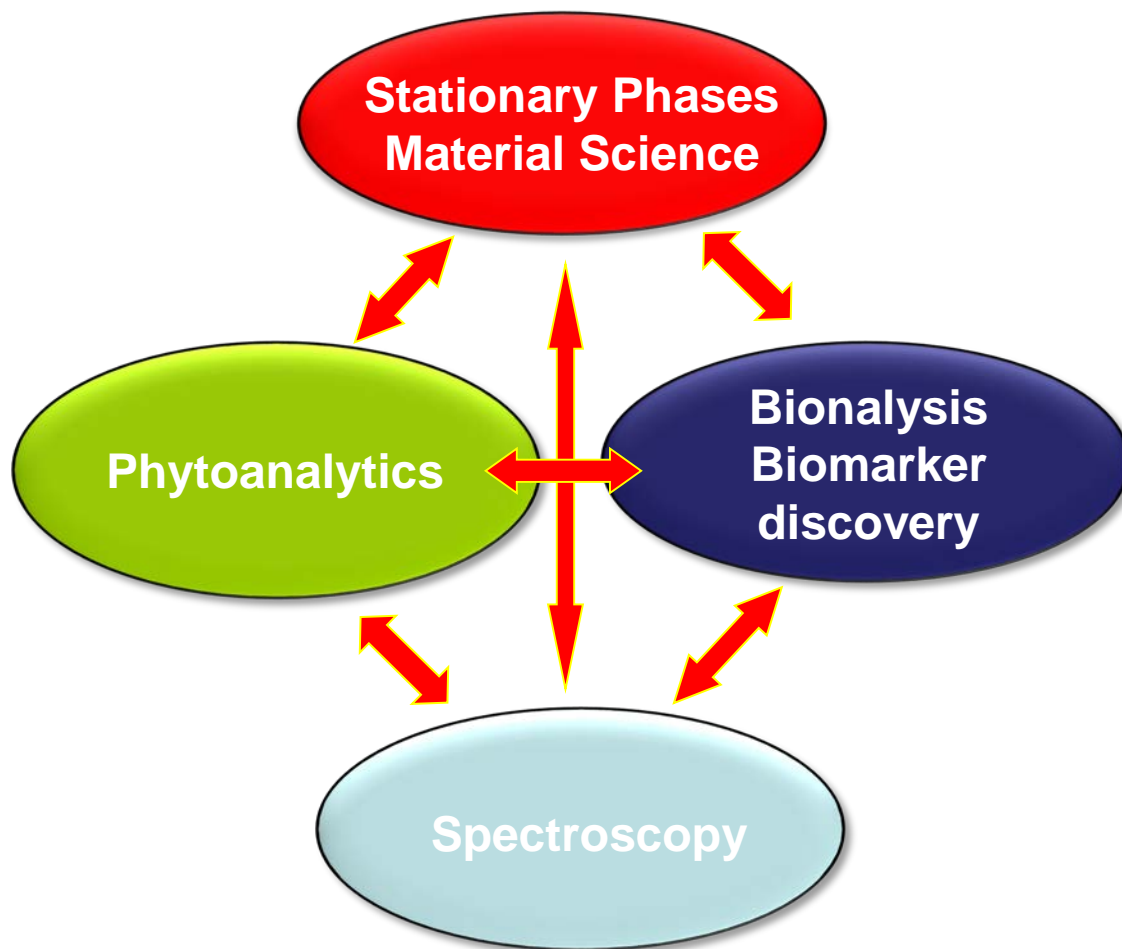
heutige Bezeichnungen	frühere Bezeichnungen
Masse m in kg oder g	Masse, Gewicht, Menge, Gewichtsmenge, Grammenge in kg oder g
Stoffmenge $n(X)$, $n(\text{eq})$ in mol	Menge, Molmenge, Molzahl, Anzahl Mole als Grammatom, Grammion, Grammolekül (Mol), Grammäquivalent (Val)
Molare Masse $M(X)$, $M(\text{eq})$ in g/mol	Atomgewicht, Atommasse, Molekulargewicht, Molgewicht, Molmasse, Äquivalentgewicht, Äquivalentmasse, Formelgewicht, Formelmasse in g, g/Mol, g/Val, als relative Größen ohne Einheit
Massenkonzentration β in g/l	Konzentration, Gehalt in g/l
Stoffmengenkonzentration $c(X)$ in mol/l	Konzentration, Gehalt; molare Konzentration oder Molarität in Mol/l, Zeichen: M
Äquivalentkonzentration $c(\text{eq})$ in mol/l	normale Konzentration oder Normalität in Val/l, Zeichen: N
Massenanteil w in g/g	Gewichtsprozent (Gew.-%), Massenprozent, Massenbruch
Stoffmengenanteil x in mol/mol	Molprozent (Mol-%), Molenbruch, Molgehalt, Atomprozent

Institute of Analytical Chemistry

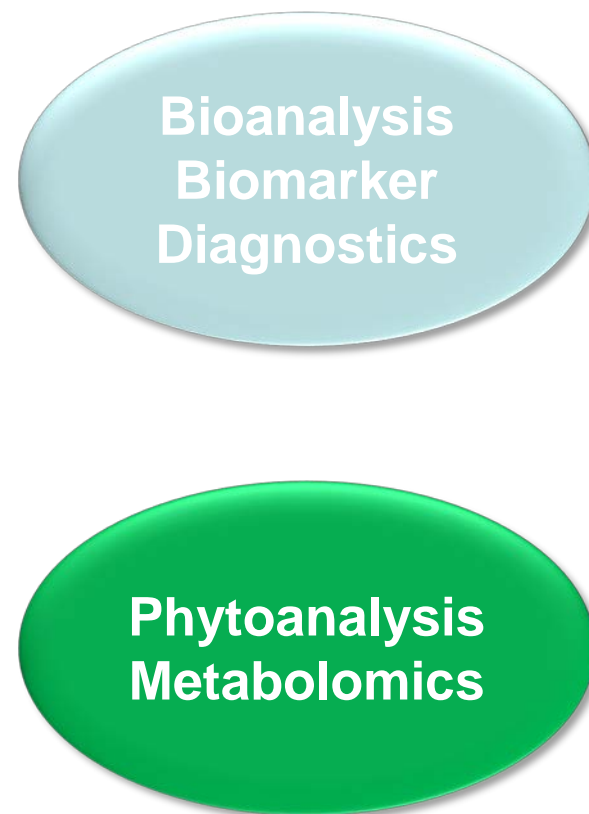


ACRC TOPICS / Synergies

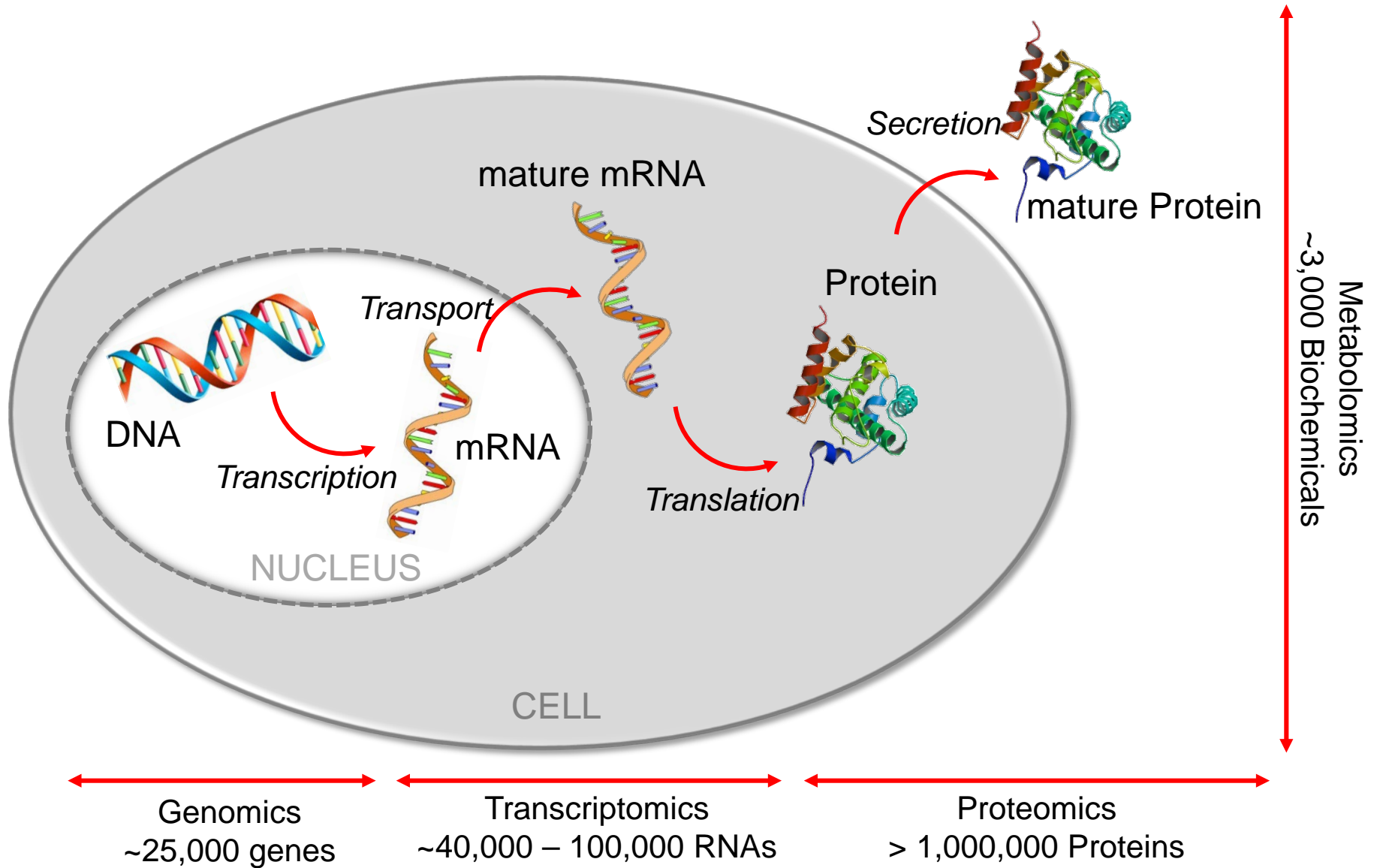
Innovations in Analytical Chemistry



Application Fields



Omics - Overview



Omics - Overview

New Analytical Approaches
Bionalysis

Genomics

Stationary Phases
Material Science
Separation

Metabolomics

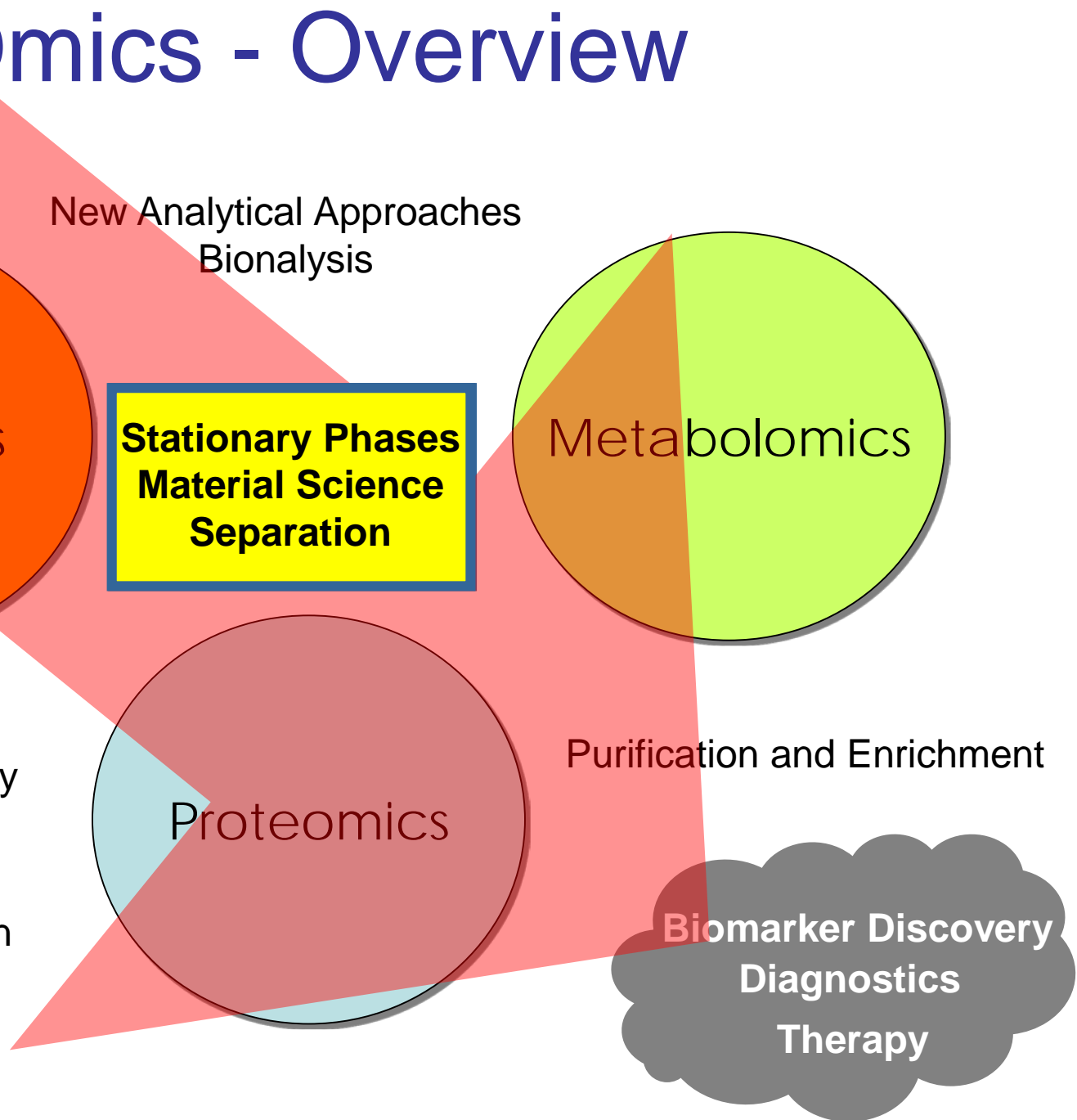
Instrumentation
MS, NMR, Spectroscopy

Purification and Enrichment

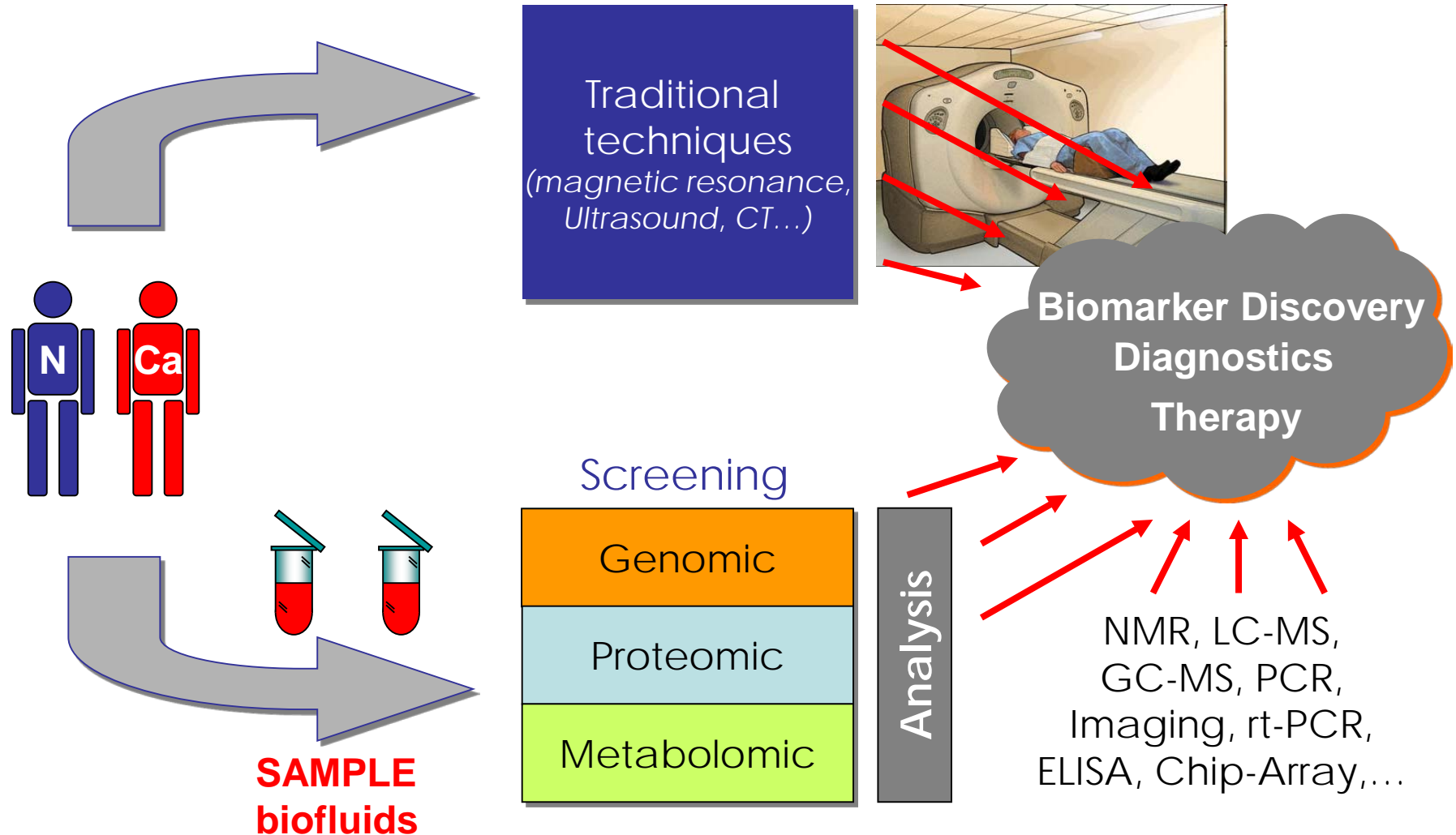
Proteomics

Data Interpretation
Data Analysis

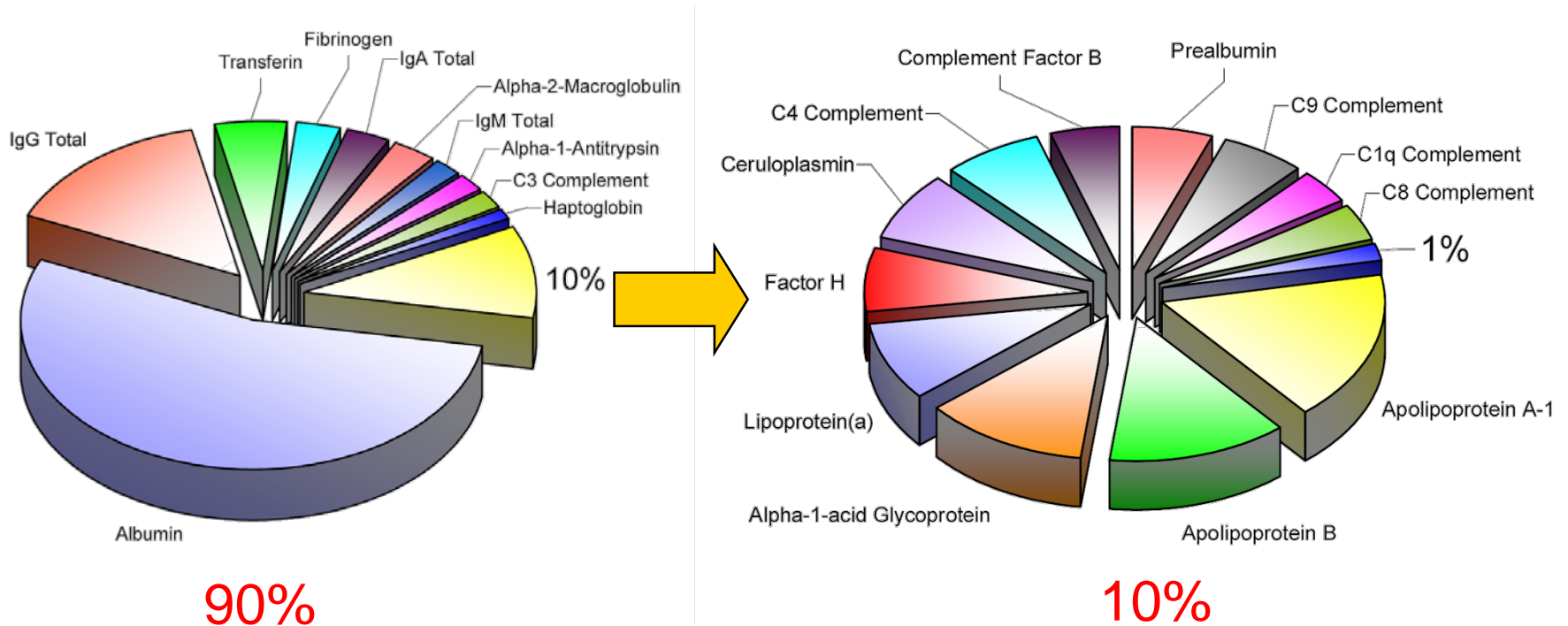
Biomarker Discovery
Diagnostics
Therapy



Omic - Overview



Complexity of Human Serum



22 proteins are approx. 99% of the whole serum proteome

→ Reducing the complexity by pre-analytical approaches!

Analytical Approaches

Innovations

Enrichment

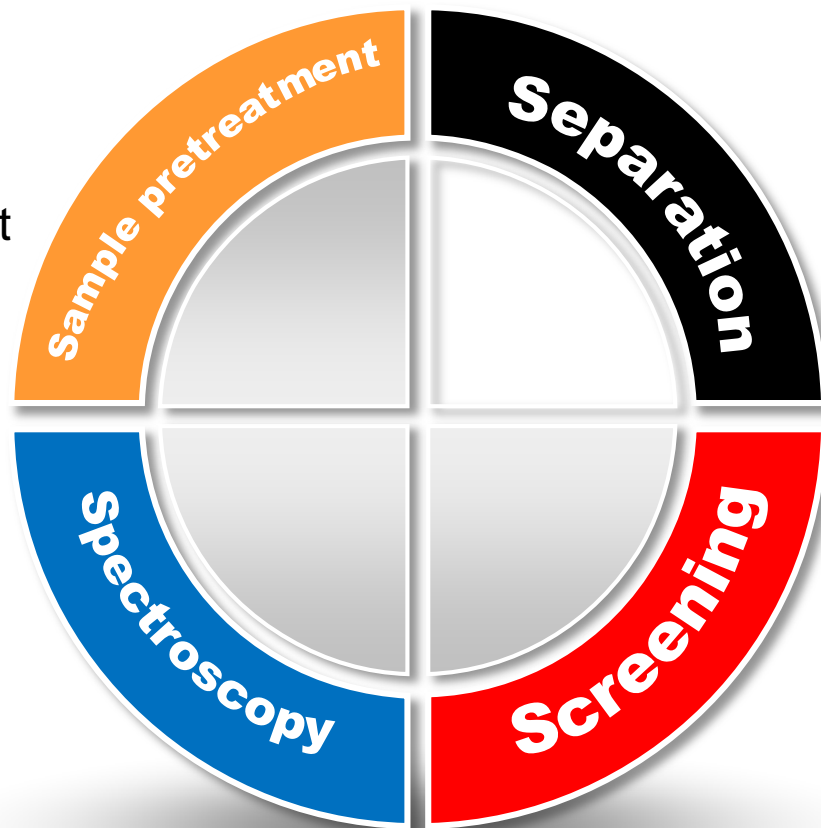
Desalting

High-sample throughput

Near-infrared

Mid-Infrared

Imaging/mapping



LC, LC-MS/MS

μ -LC, μ -LC-MS/MS

CE, CE-MS

CEC

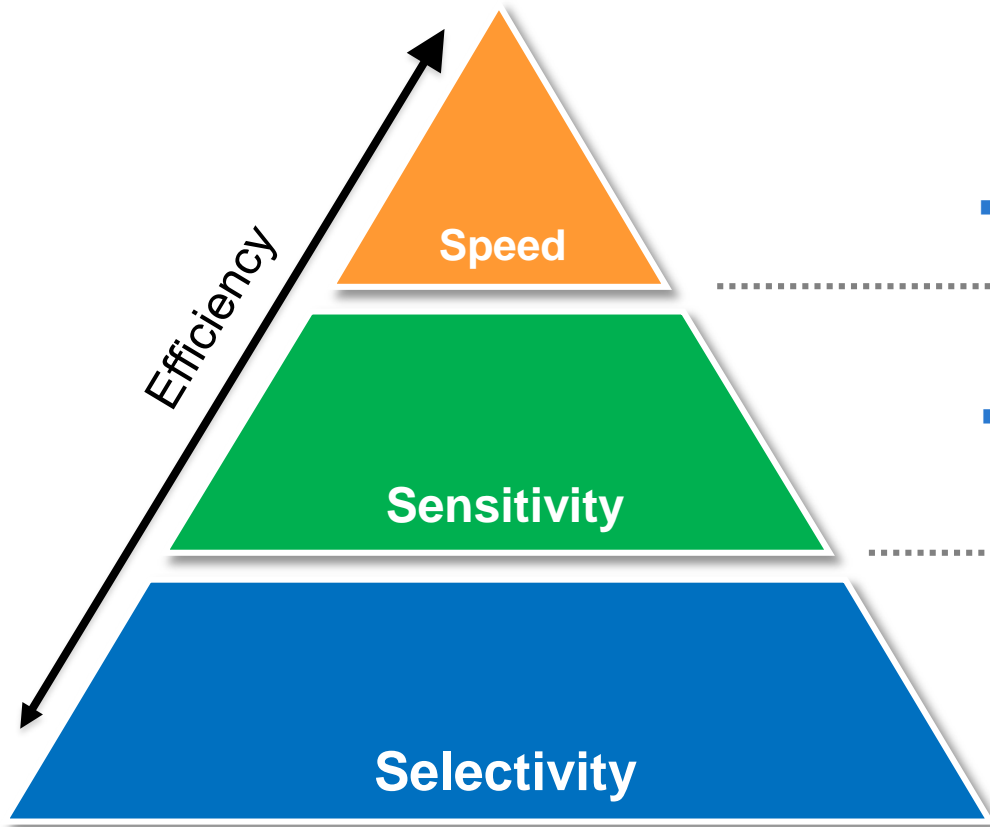
MALDI-TOF-MS/MS

Matrixfree-MALDI

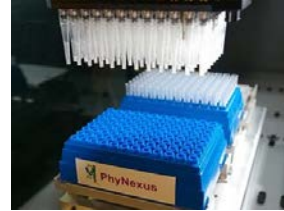
MELDI-TOF

MALDI-imaging/mapping

Why Analytical Innovations?



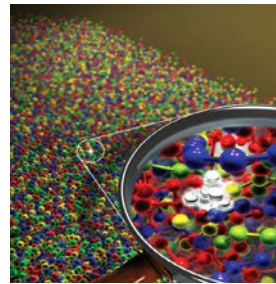
- High sample throughput



- Improved detection limit



- Needle in haystack



„To see what one could not see before“

Highly Efficient Enrichment and Separation of Biomolecules

Enrichment

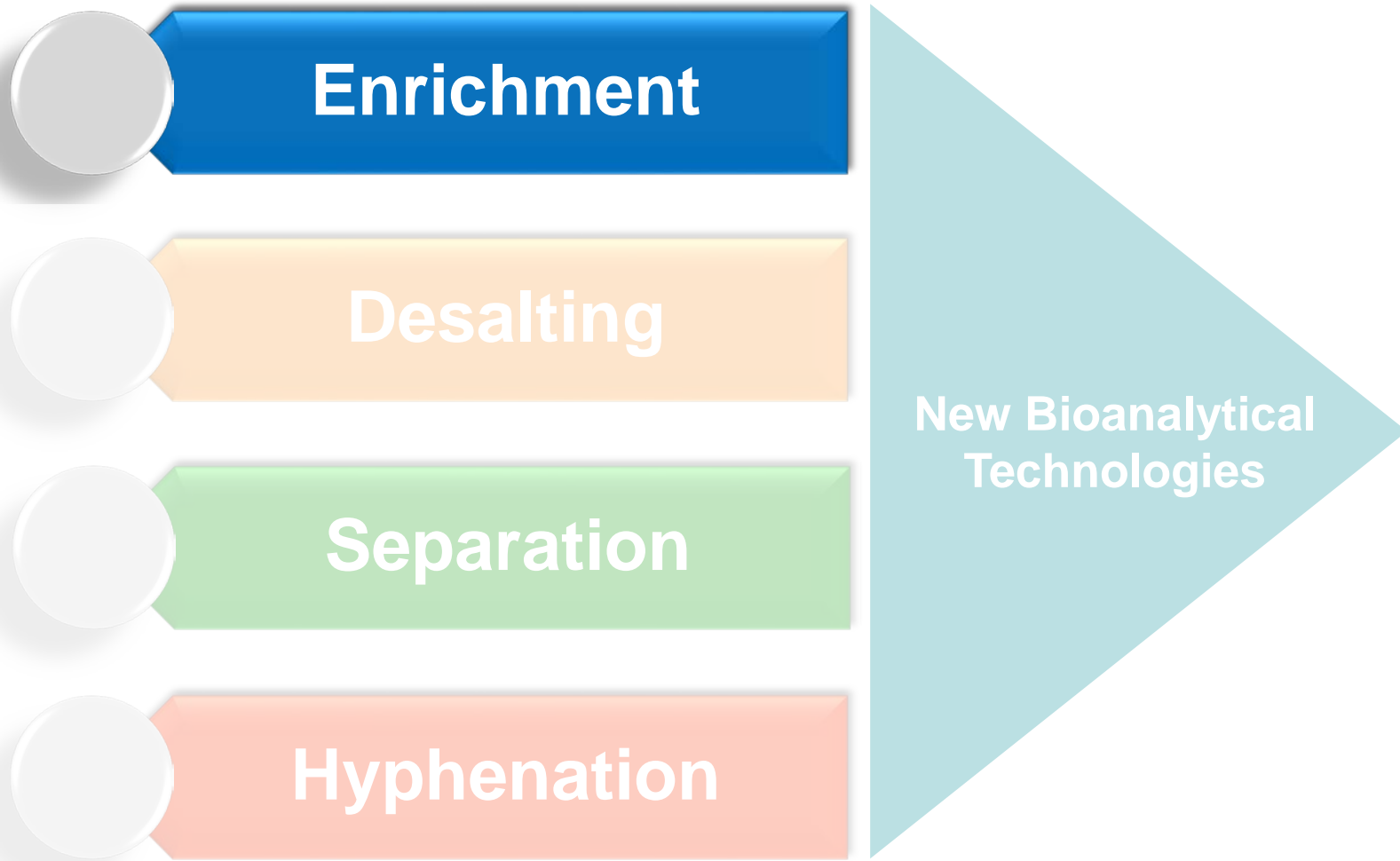
Desalting

Separation

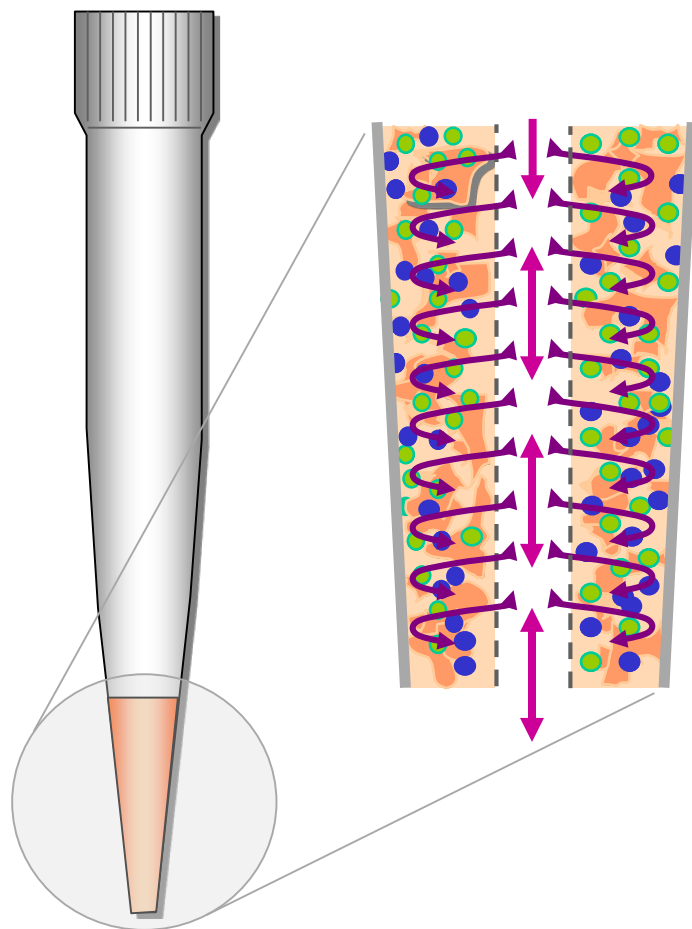
Hyphenation

**New Bioanalytical
Technologies**

Highly Efficient Enrichment and Separation of Biomolecules



Preparation of - Hollow Monolith™



Hollow Monolith™



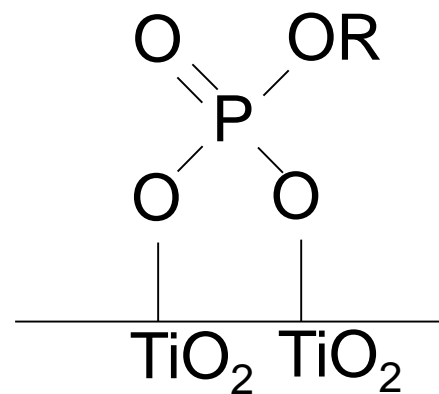
$\text{ZrO}_2 < 100 \text{ nm}$



$\text{TiO}_2 < 100 \text{ nm}$

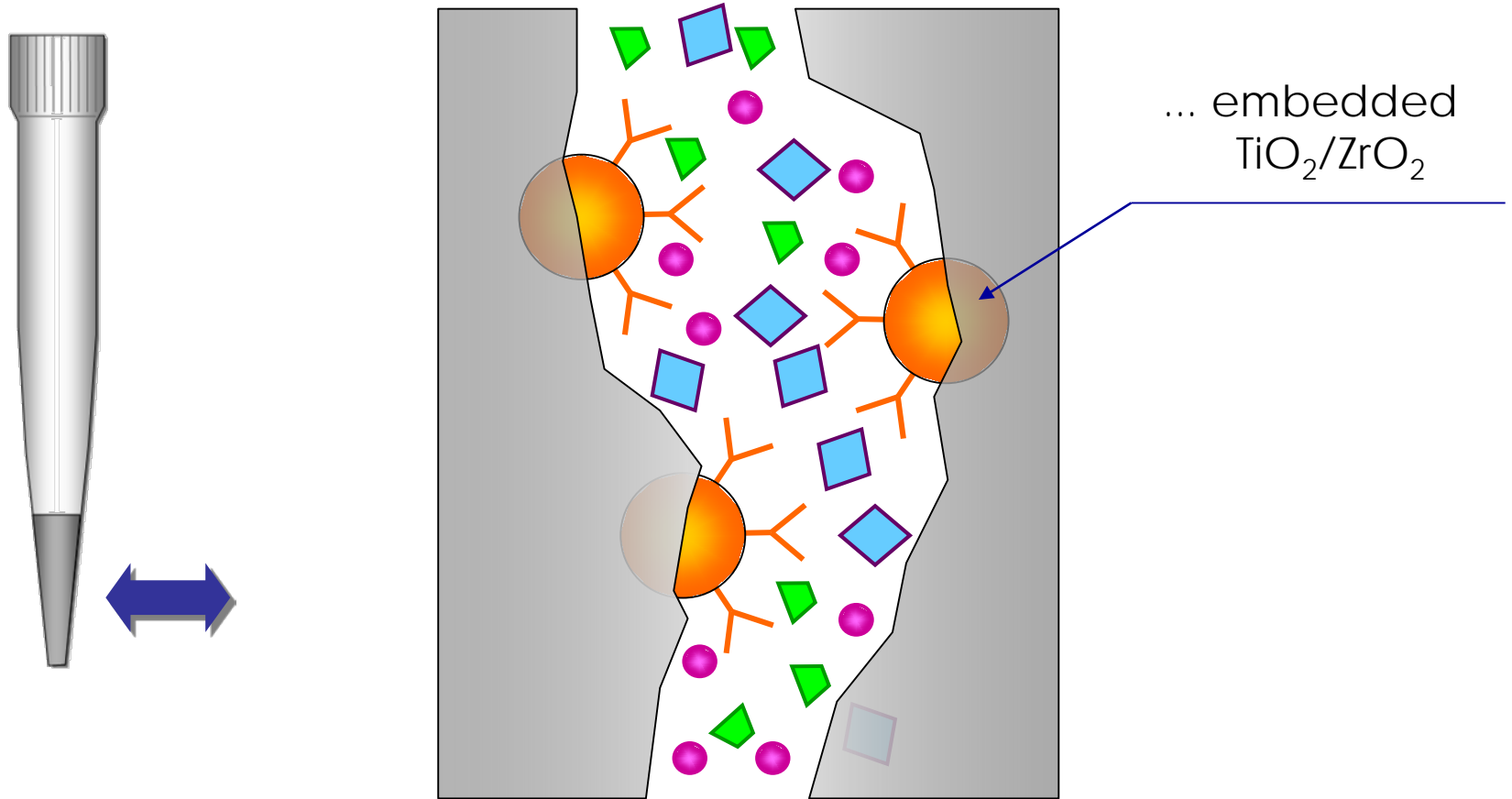


poly(divinylbenzene)



Mechanism: Bridging Bidentate

Enrichment of Phosphopeptides



... embedded
 $\text{TiO}_2/\text{ZrO}_2$

◆ ... Phosphorylated Peptides

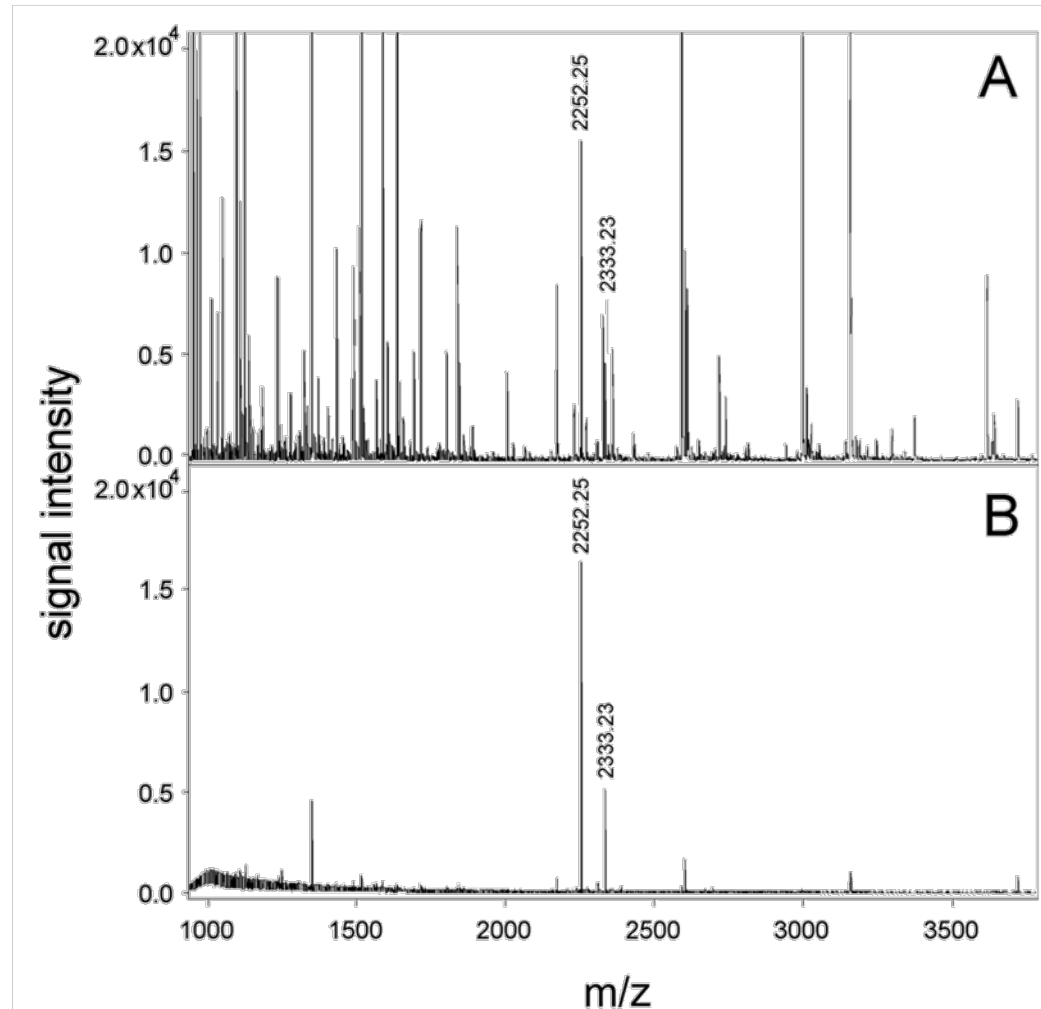
Enrichment of *in vitro* phosphorylated ERK1 digest

MALDI MS spectra:

1.) before enrichment (A)

2.) after enrichment with poly(DVB)-TiO₂/ZrO₂ tips (B)

Signals at m/z 2252.25 and m/z 2332.23 correspond to phosphorylated peptides

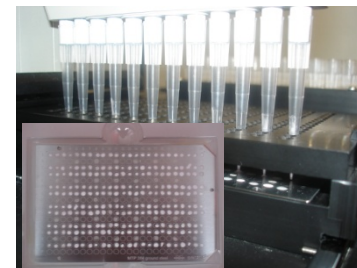


Collaboration with Prof. Lukas Huber, Biocenter - Innsbruck

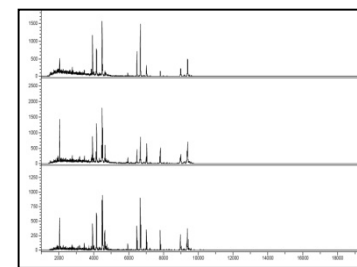
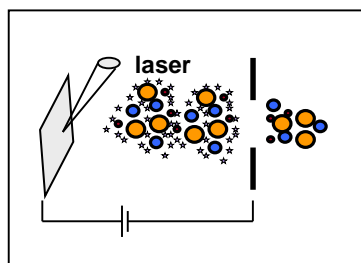
Automation of Sample Preparation



1. sample loading 2. sample spotting



3. sample analysis 4. data processing



- Specific enrichment
- Purification
- Desalting

A collaboration with PhyNexus Inc., San Jose, CA, USA



Proteomics Cover

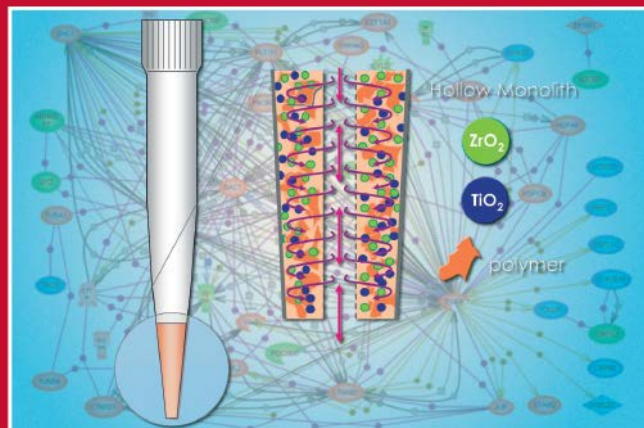
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21'08

www.proteomics-journal.com



**Signal Transduction
Proteomics**

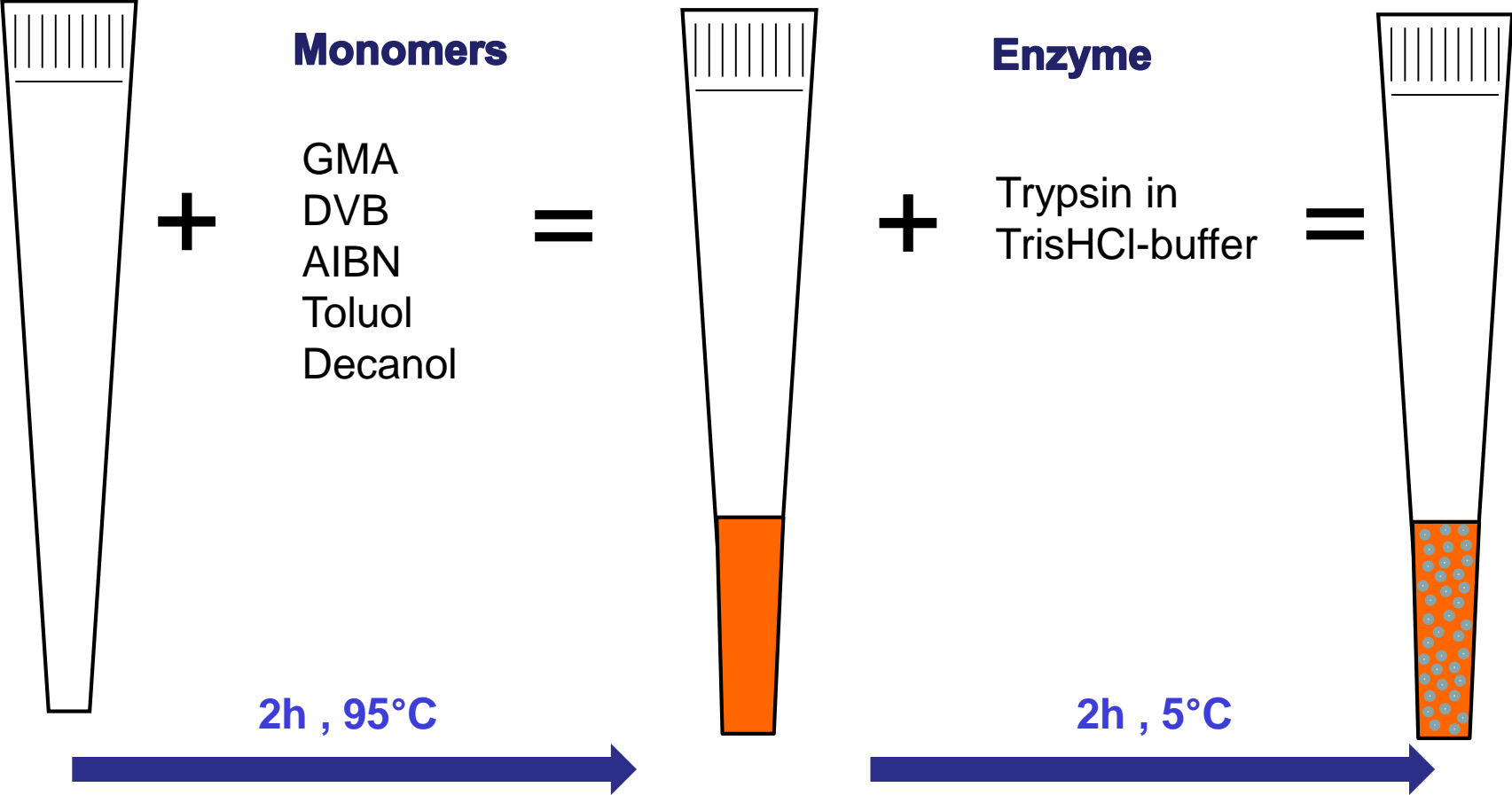
**Editors:
Lukas A. Huber**

Endorsed as an
Official Journal of
HU
Human Proteome Organisation

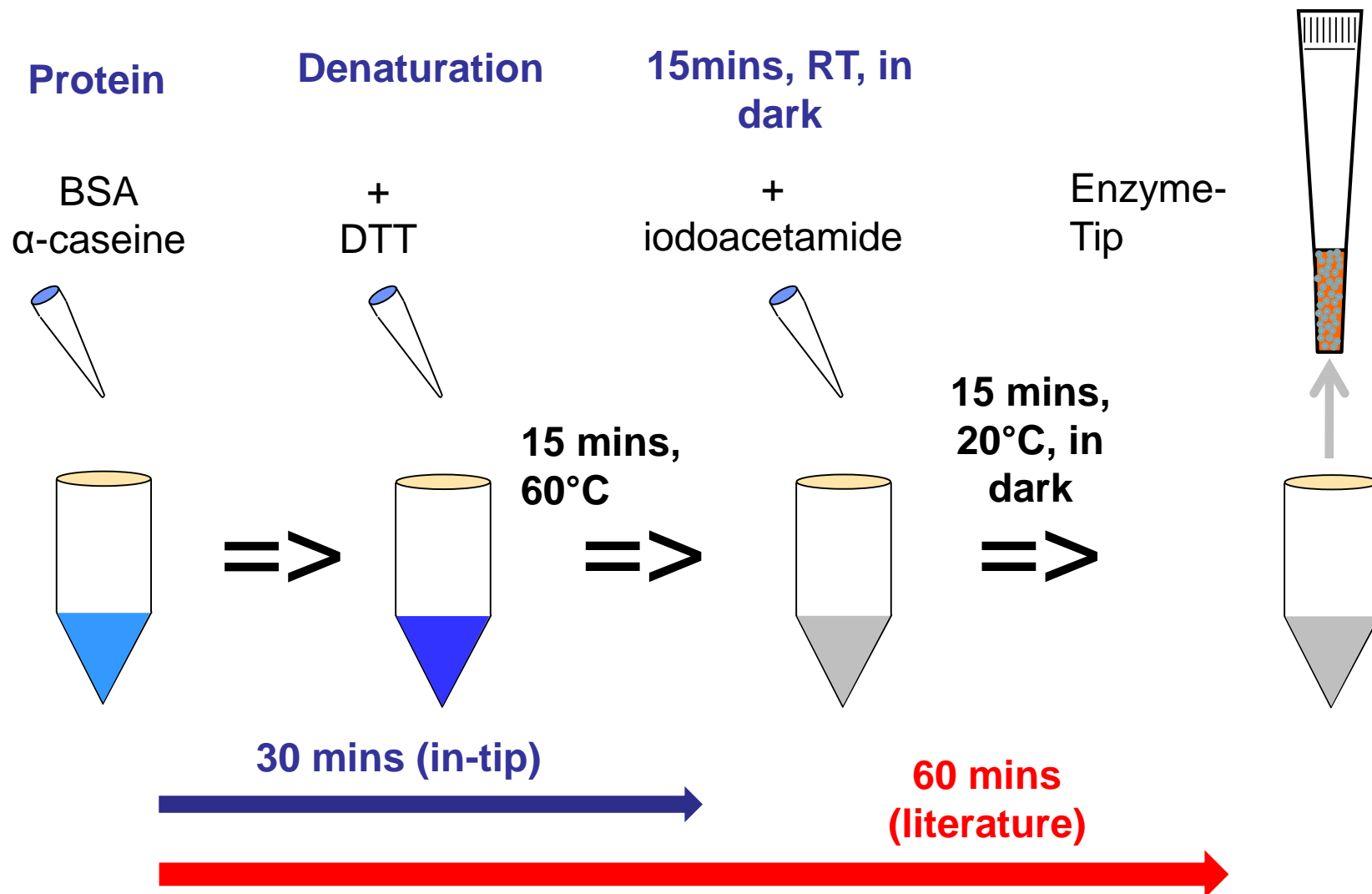
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Monolithic Extraction Tips for Enzymatic Digestion



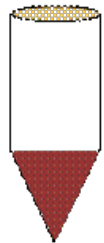
Monolithic Extraction Tips for Enzymatic Digestion



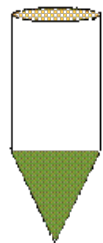
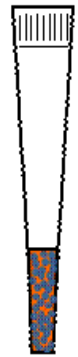
Microwave-Assisted In-Tip Digestion

Denaturation

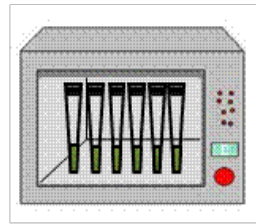
- Bovine Serum Albumin
- Myoglobin
- α -Casein
- Milk
-



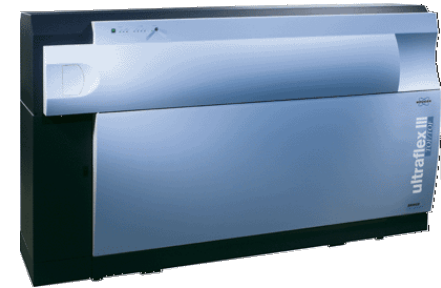
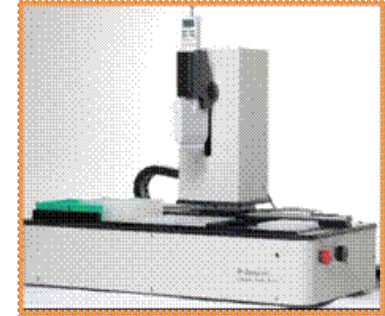
Enzyme-Tip



Microwave In-Tip-Digestion



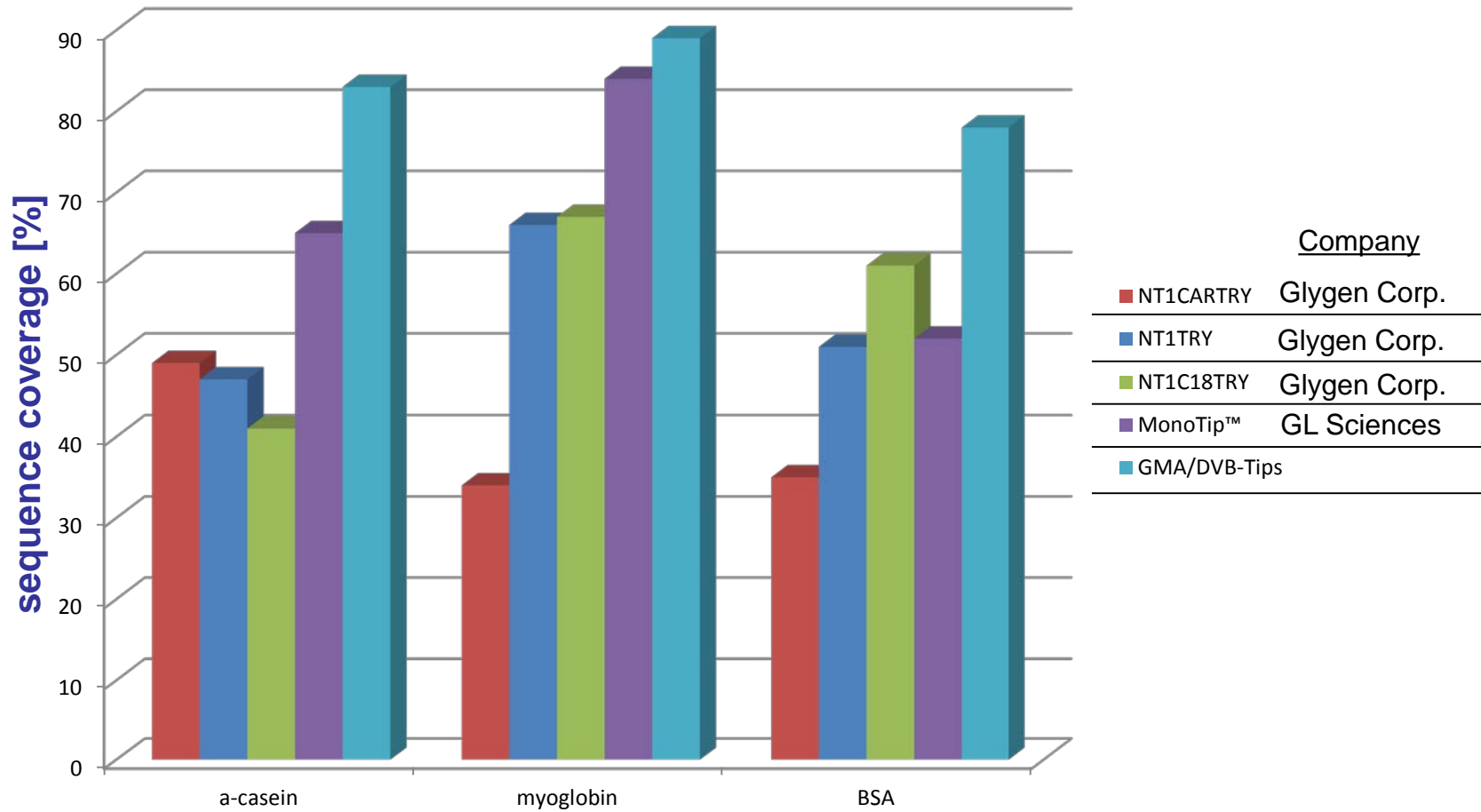
**Eluting and
MALDI TOF
MS**



Total Time 45 min!

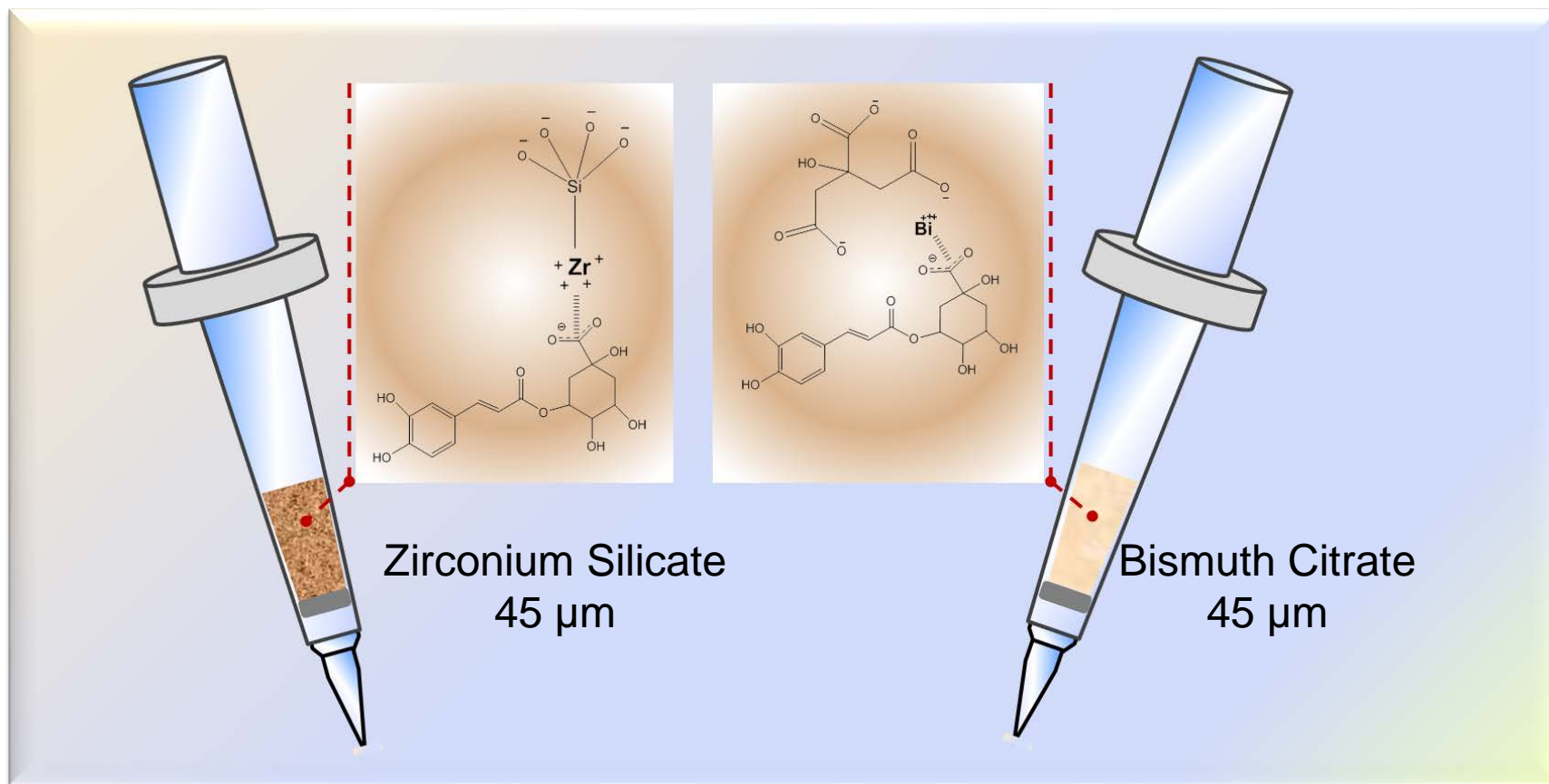
conventional digestion time: 6-10 h

Results - Comparison Study



α-casein digested, eluted from enzyme-Tips with Tip-Technology

Schematic view of spin columns containing sorbents

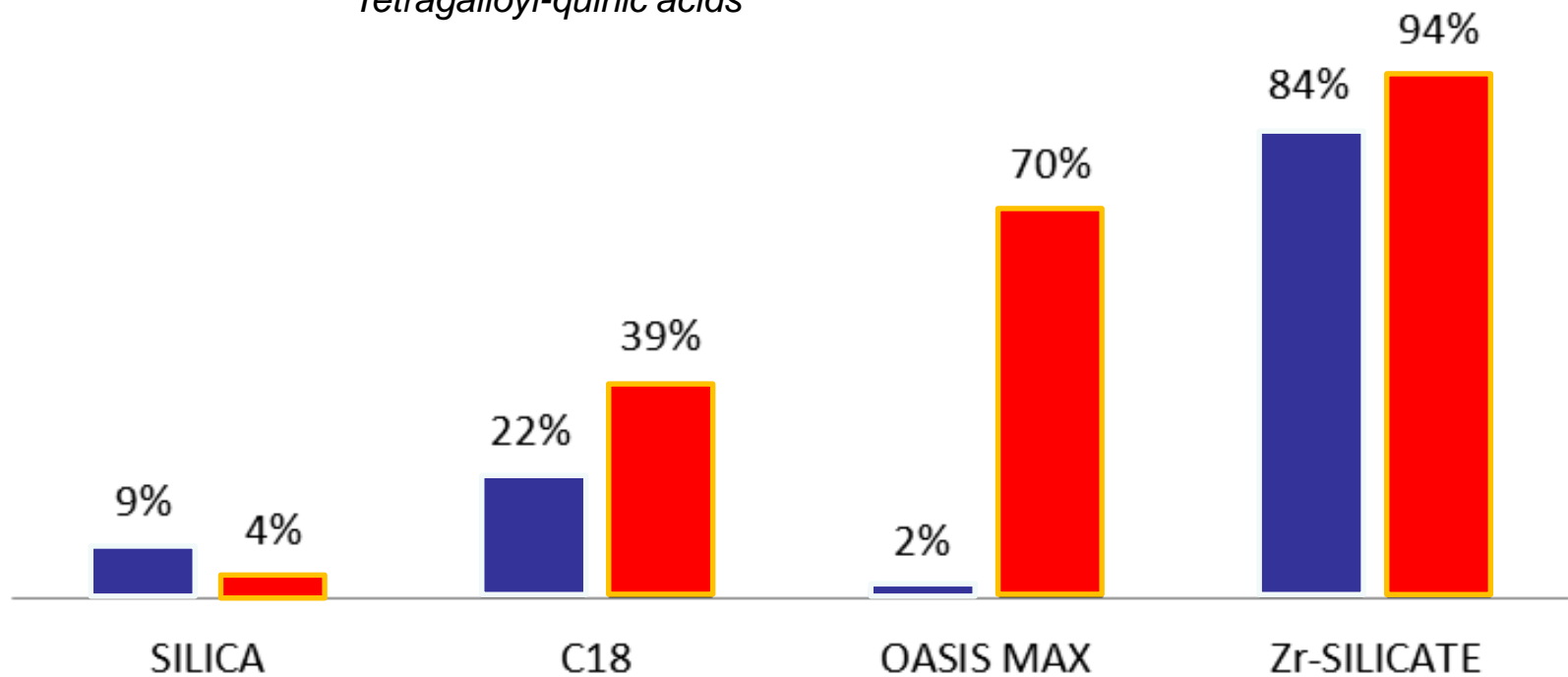


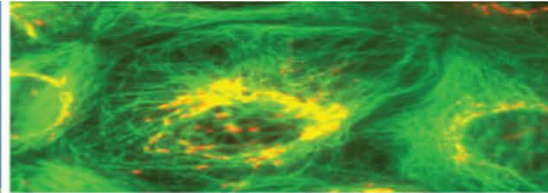
Recovery study of standards

Comparison based upon nature of stationary phase/Type of interaction

■ TGQA % recoveries
Tetragalloyl-quinic acids

■ CYNARIN % recoveries

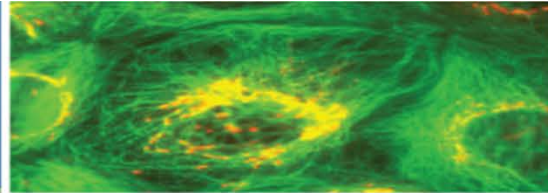




Pyrrolizidine alkaloids

Background

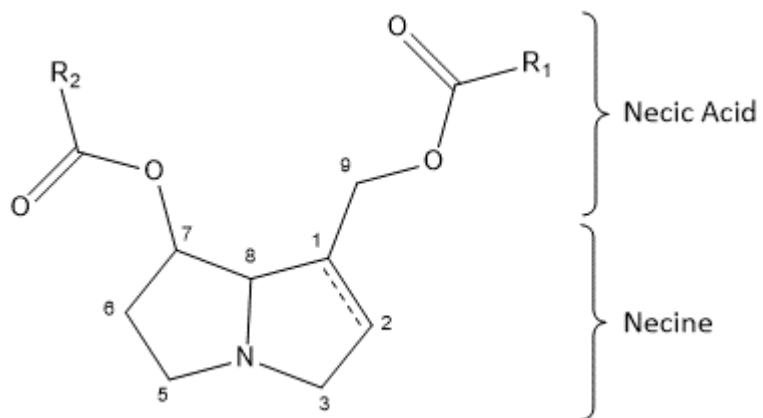
- Pyrrolizidine alkaloids (PA) are **secondary plant metabolites** (for plant protection)
- **400 different PAs** in approximately 6000 plant species are known
 - **Contamination** of plant products during **harvest** or through animals (for example bees)
 - Examples for contaminated food: herbal teas, honey, salads etc.
- Problem: **Hepatotoxic** for animals as well as for humans
 - Safety values for the maximum dose for drugs in Germany:
 - Application up to 6 weeks: 1 µg/day oral, 100 µg/day cortically
 - Application for more than 6 weeks: 0.1 µg/day oral, 10 µg/day cortically



Pyrrolizidine alkaloids

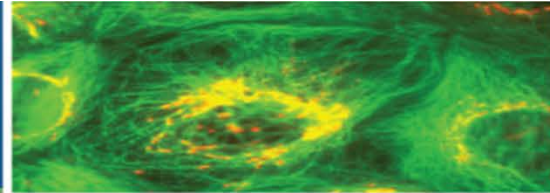
Toxicity

- General structure:



Nearly all types are in
coexistence with their N-oxides

- Requirement(s) for **toxicity**:
 - 1,2-unsaturated necine
 - Esterification of at least one OH-group with a branched acid

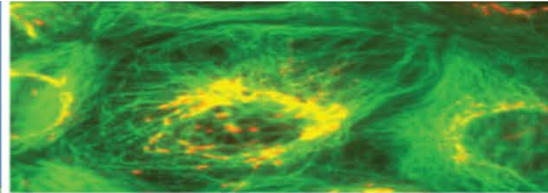


Pyrolizidine alkaloids

Methods for detection

- HPLC-MS (LOD ~ 1 ppb)
- GC-MS (LOD = 3 ppb)
 - Otonecines and N-oxides (without derivatisation) not detectable
- Double antibody ELISA (LOD = 0.1 – 1.5 ppb)
 - Antibodies only against some specific PAs
- HPLC Evaporative Light Scattering Detection (ELSD) (LOD = 40 ppm)
- Nonaqueous capillary electrophoresis (NACE) MS (LOD < 7.5 ppm)
- Photometric detection with Ehrlich reagent (LOD = 10 ppm)

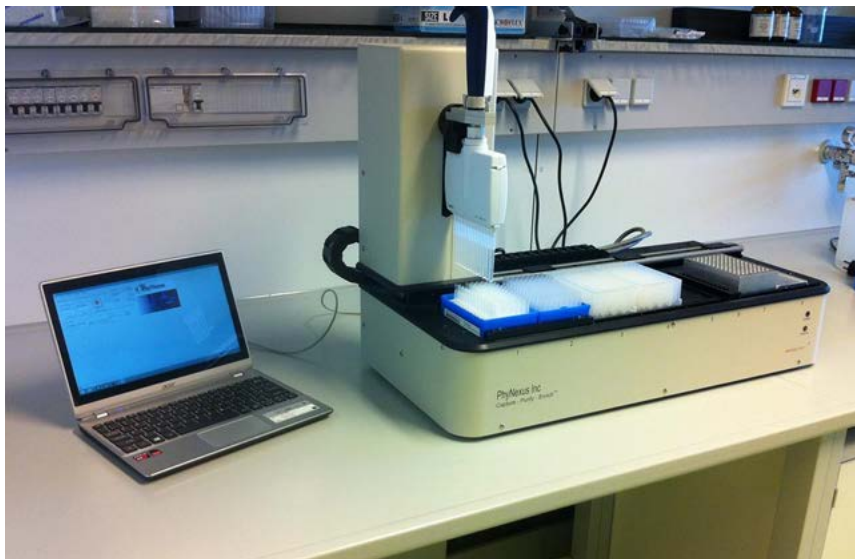
- General problem for PA analysis: **Only about 25 standards are available**
 - Validation is limited to a small spectrum of PAs
- ➡ **Isolation of PAs** out of plant material with countercurrent chromatography (CCC) to get more standards (Cooperation with Medical University of Lublin)



Pyrolizidine alkaloids

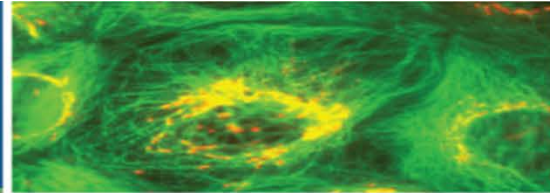
Cation exchange

- Detection only possible in the low ppb-range (for GC- and HPLC-MS)
 - for lower concentrations and for separation from other plant substances enrichment is necessary
- Due to the chemical structure ion exchange is the enrichment method of choice
- Automation with PhyNexus MEA 2 possible



PhyTip with Toyopearl SP-650 resin



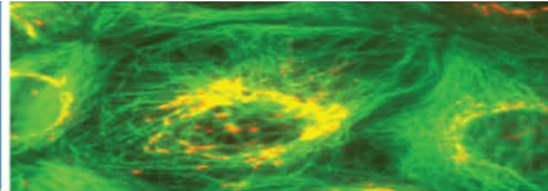


Pyrrrolizidine alkaloids Procedure

Extraction of the plant material

Enrichment of PAs with ion
exchange

Detection with UPLC-
MS



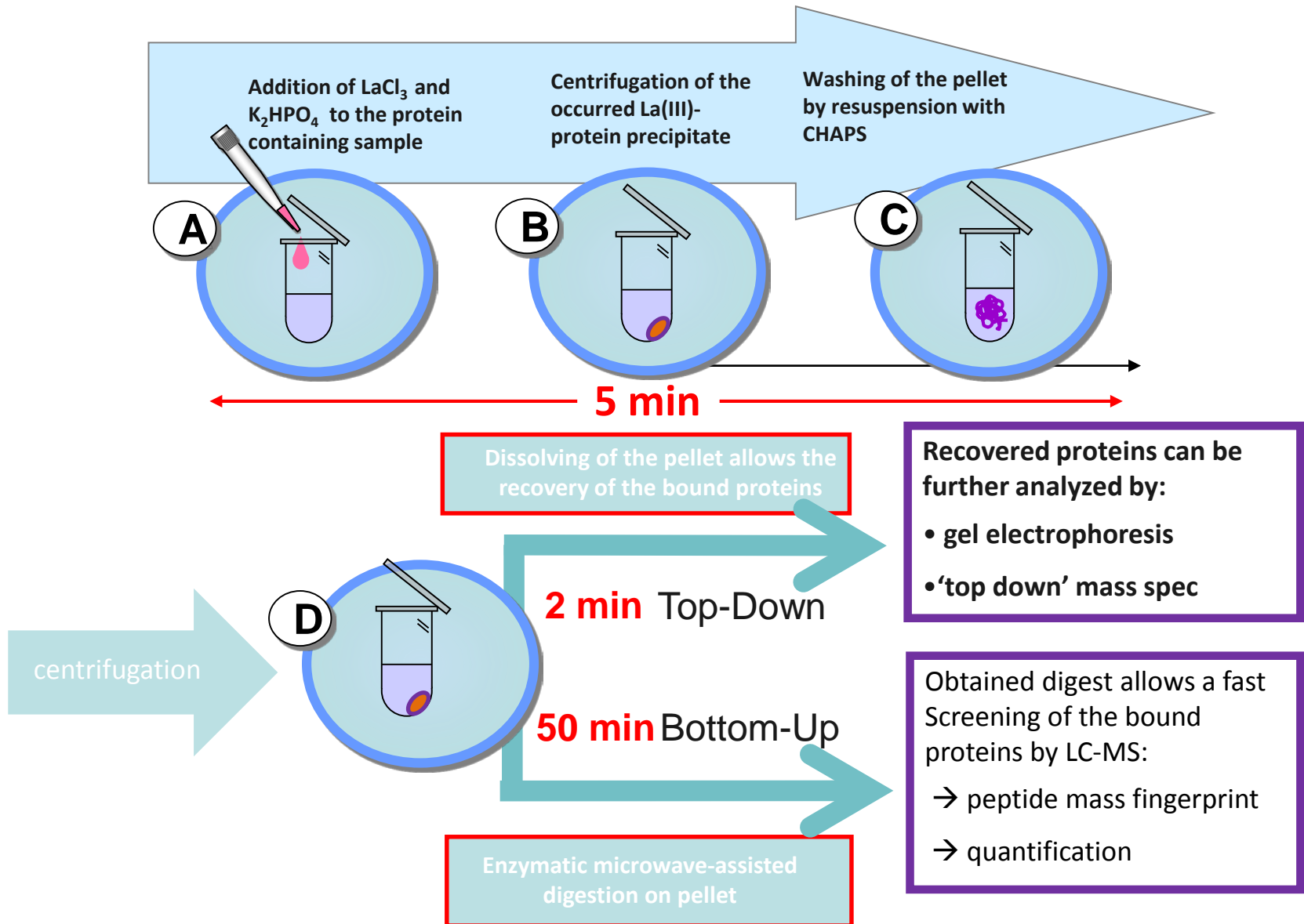
Pyrrolizidine alkaloids

Cation exchange – Comparison of commercial products

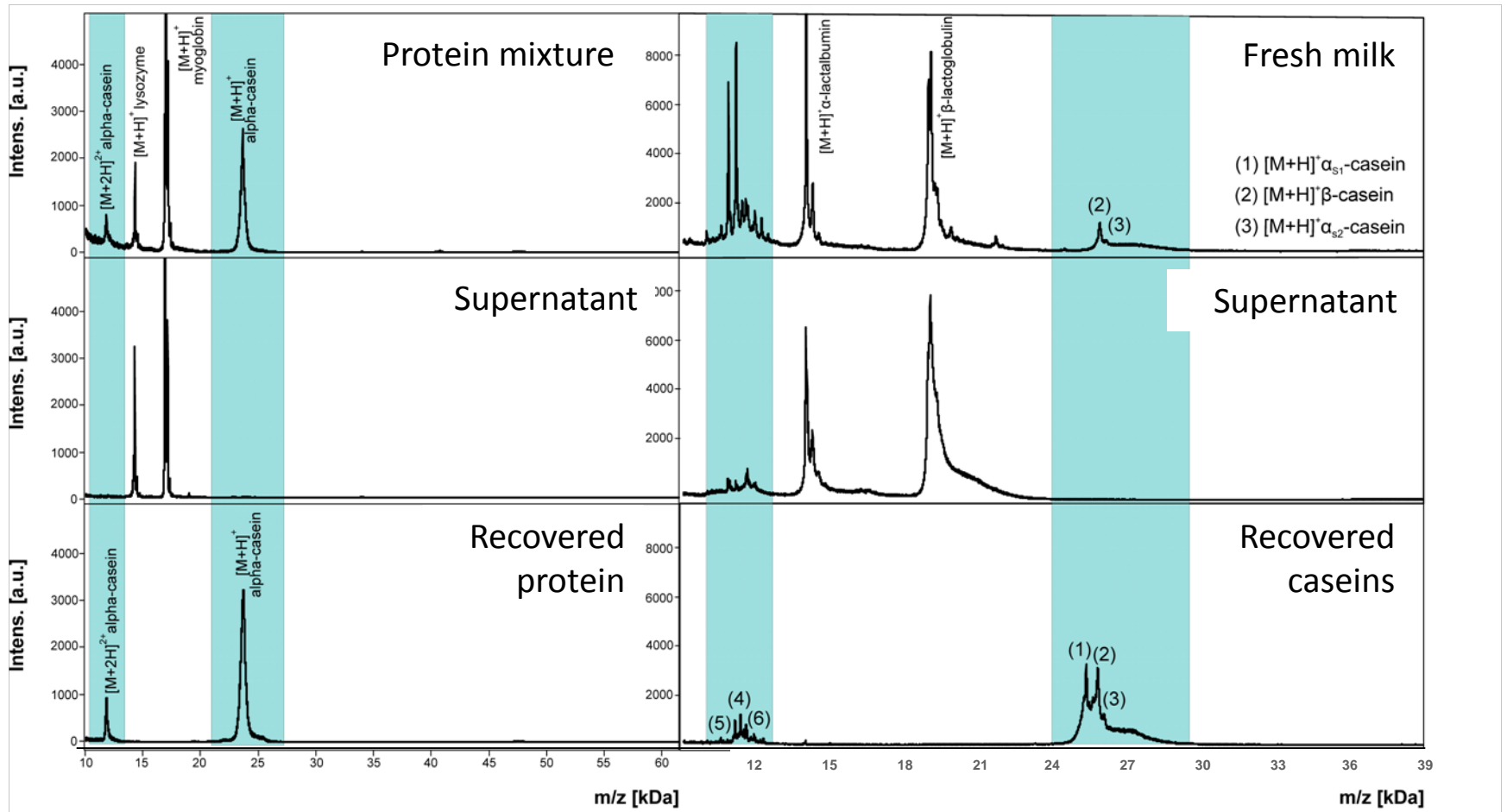
- In terms of enrichment of pyrrolizidine alkaloids different commercial materials were tested
- Best recoveries (**94-99%**) were achieved with a polystyrene-divinylbenzene resin functionalized with sulfonate groups
- Further approach
 - Polystyrene DVB resin in PhyNexus tips and automation of the cation exchange with PhyNexus MEA 2
 - Continuing comparison with other resins (in cartridges and tips)
 - Test of anion exchange resins and Immobilized Metal Ion Affinity Chromatography (IMAC)

Enrichment of Phosphoproteins by Precipitation using Trivalent Lanthanides

Highly Selective Precipitation of Phosphoproteins by Lanthanum



Highly Selective Precipitation of Phosphoproteins by Lanthanum

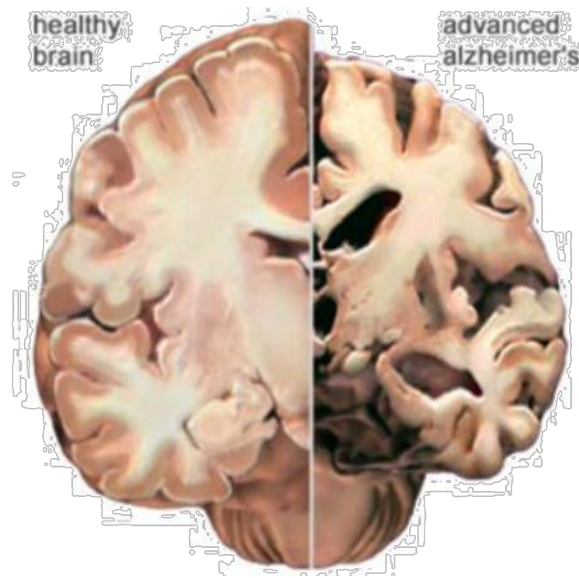


Recovery of intact proteins allows „top down“ protein analysis by LC-MS or gel electrophoresis

Highly Selective Precipitation of Phosphoproteins by Lanthanum

Analysis of Liquor for the Diagnosis of Alzheimer

- Alzheimer's disease is the most common form of dementia.
- Alzheimer's is a brain disorder destroying brain cells, causing problems with memory, thinking and behavior severe enough to affect work, lifelong hobbies or social life.
- An estimated 26.6 million people worldwide had Alzheimer's in 2006; this number may quadruple by 2050.



Analysis of Liquor for the Diagnosis of Alzheimer

Accession number	Protein	Hit	Phosphorylation	Glycosylation
P05067	Amyloid beta A4 protein	(Ray, 2000), (Grimmer, 2009)	+	+
P06396	Gelsolin	(Ray, 2000)	+	-
P01011	Alpha-1-antichymotrypsin	(Eriksson, 1995), (Gollin, 1992)	-	+
P02647	Apolipoprotein A-I	(Yin, 2009)	-	+
P02679	Fibrinogen	(Yin, 2009), (Ojien van, 2005)	-	+
P00738	Haptoglobin	(Yin, 2009)	+	+
P02766	Transthyretin	(Biroccio, 2006)	-	+
P10451	Osteopontin	(Wung, 2007)	-	+
Q14515	SPARC-like protein 1	(Yin, 2009)	+	+
P02649	Apolipoprotein E	(Corder, 1993),	+	+
P10909	Clusterin	(Yin, 2009)	-	+
Q9UBP4	Dickkopf-related protein 3 Pigment	(Zenzmaier, 2004)	+	+
P36955	epithelium-derived factor	(Yin, 2009)	+	+
P02774	Vitamin D-binding protein	(Yin, 2009)	-	+
P01009	Alpha-1-antitrypsin	(Gollin, 1992)	-	+
P02763	Alpha-1-acid glycoprotein 1	(Merritt, 1988)	-	+
P04004	Vitronectin	(Rogers, 2000)	+	+
P01042	Kininogen-1	(Puchades, 2003)	+	+
P01034	Cystatin-C	(Mi, 2009)	+	-
P10645	Chromogranin A	(Lechner, 2004)	+	+

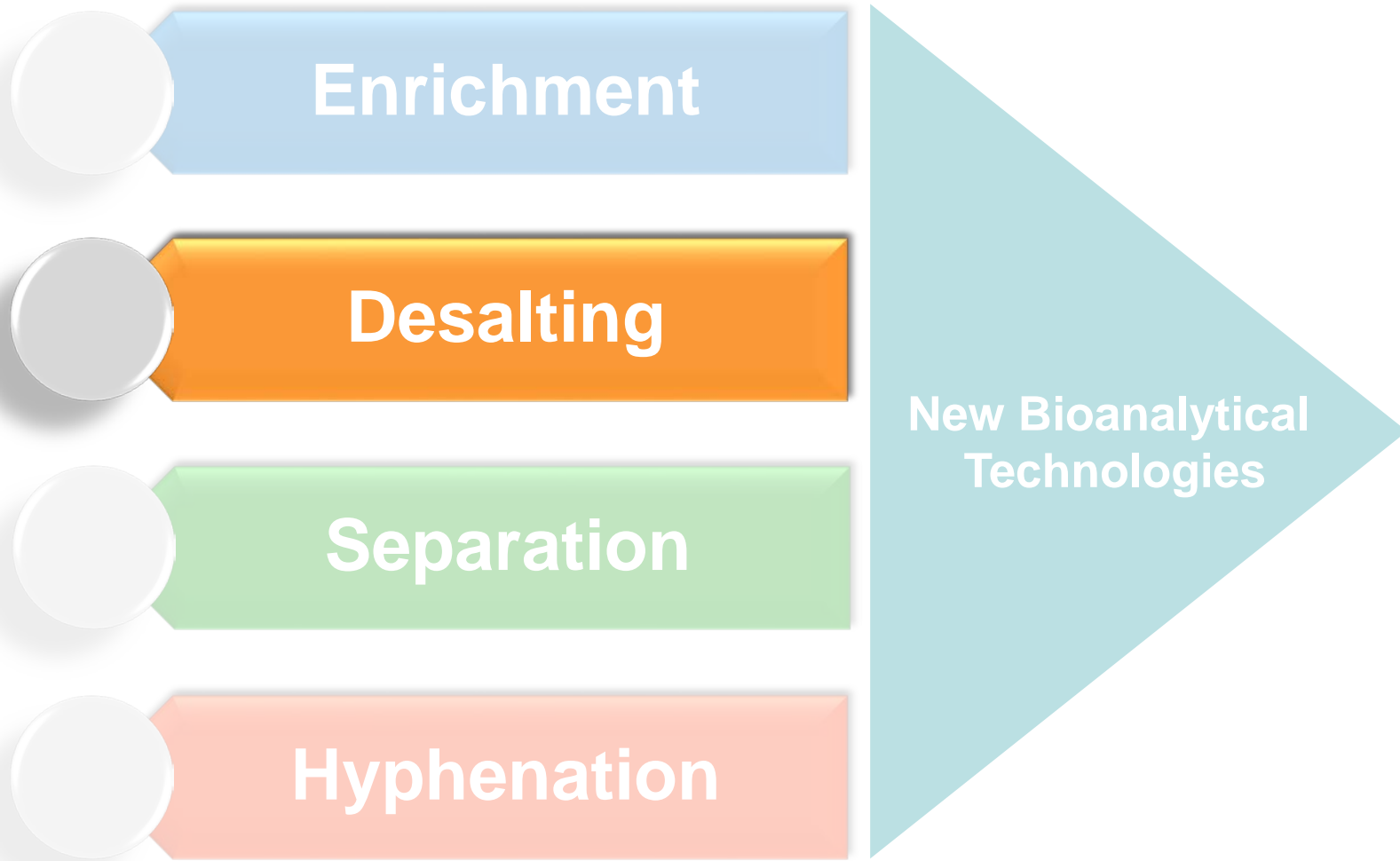
- CSF was analyzed by lanthanum (III) precipitation
- **53 phosphorylated and/or glycosylated proteins were identified** through peptide mass fingerprint
- **20 out of them are potential biomarkers for Alzheimer's disease**
- Highly selective analysis
- 20 biomarkers detected in the same analytical run

After the loading of 7 µl onto the trapping column with a flow rate of 20 µl/min for 4 min, the separation was performed under reversed phase conditions with solvent A 4 % acetonitrile (ACN) and 0.1 % (v/v) formic acid (FA) in water and solvent B 0.1 % (v/v) FA in 60 % ACN, at a flow rate of 300 nl/min and 60 °C. A linear gradient (300 min from 0 % B to 60 % B) was used.

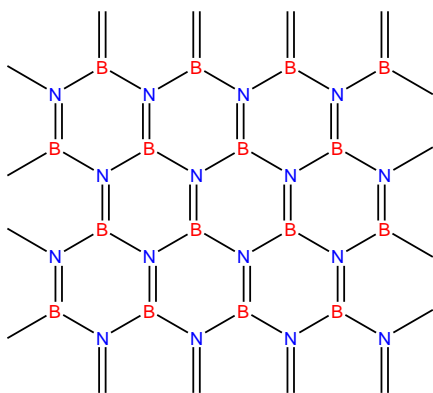
Recovery of Phosphopeptides

[M+H] ⁺ Da	Phosphopeptide Sequences ^a	Phospho- groups	ErCl ₃	HoCl ₃	CeCl ₃	LaCl ₃	EuCl ₃	TmCl ₃	TbCl ₃	TiO ₂
1254.52	EVVGSpAEAGVDAA (Ov-(340–352))	Mono	-	-	-	-	-	-	-	-
1331.53	EQLSpTSpEENSK (α-S2-(141–151))	Mono	-	-	-	-	-	-	-	-
1411.50	EQLSpTSpEENSK (α-S2-(141–151))	Di	-	-	-	-	-	-	-	-
1466.61	TVDMEspTEVFTK (α-S2-(153–164))	Mono	+	+	+	+	+	+	+	+
1594.70	TVDMEspTEVFTKK (α-S2-(153–165))	Mono	+	+	+	+	+	+	+	+
1660.79	VPQLEIVPNSpAEER α(-S1-(121–134))	Mono	+	+	+	+	+	+	+	+
1832.83	YLGEYLIVPNSpAEER (α-S1)	Mono	+	+	+	+	+	+	+	+
1847.69	DIGSESpTEDQAMEDIK (α-S1-(58–73))	Mono	-	+	+	-	+	+	-	-
1927.69	DIGSESpTEDQAMEDIK (α-S1-(58–73))	Di	+	+	+	+	+	+	+	+
1951.95	YKVPQLEIVPNSpAEER (α-S1-(119–134))	Mono	+	+	+	+	+	+	+	+
2061.83	FQSpEEQQQTEDELQDK (β-C-(33–48))	Mono	+	+	+	+	+	+	+	+
2088.89	EVVGSpAEAGVDAASVSEEFR (Ov-(340–359))	Mono	+	+	+	+	+	+	+	+
2432.05	IEKFQSpEEQQQTEDELQDK (β-C-(33–48))	Mono	+	+	+	+	+	+	-	-
2511.13	LPGFGDspIEAQCGTSVNVHSSLR (Ov-(62–84))	Mono	-	-	-	-	-	-	-	-
2556.10	FQSpEEQQQTEDELQDKIHPF (β-C-(48-67))	Mono	+	+	+	+	+	+	-	+
2619.04	NTMEHVSpSpSpEESpIISQETYK (α-S2-(17–36))	Tetra	+	+	+	+	+	-	-	-
2678.01	VNELSpKDIGSpESpTEDQAMEDIK (α-S1-(52–73))	Tri	+	-	-	-	-	-	-	+
2703.50	LRLKKYKVPQLEIVPNSpAEERL(α-S1-(114–135))	Mono	+	+	+	+	+	-	+	+
2720.91	QMEAESpISpSpSpEEIVPNSVEAQK (α-S1-(74–94))	Penta	+	-	-	+	+	-	+	+
2747.10	NTMEHVSpSpSpEESpIISQETYKQ (α-S2-(17–37))	Tetra	+	-	-	-	-	-	-	-
2856.50	EKVNELSpKDIGSpESTEDQAMEDIK (α-S1-(50–73))	Di	+	+	+	-	+	-	-	+
2901.32	FDKLPFGDspIEAQCGTSVNVHSSLR (Ov-(59–84))	Mono	+	-	+	+	-	+	-	-
2935.15	EKVNELSpKDIGSpESpTEDQAMEDIK (α-S1-(50–73))	Tri	+	+	-	+	+	-	-	+
2966.16	ELEELNVPGEIVESpLSpSpSpEESITR (β-C-(17–40))	Tetra	-	-	+	-	-	-	-	-
3008.01	NANEEYSIGSpSpSpEESpAEVATEEVK (α-S2-(61–85))	Tetra	+	+	+	+	+	-	+	+
3042.27	RELEELNVPGEIVESLSpSpSpEESITR (β-C-(16–40))	Tetra	+	+	+	+	+	-	+	-
3087.99	NANEEYSIGSpSpSpEESpAEVATEEVK (α-S2-(61–85))	Penta	+	+	-	+	+	-	-	+
3122.27	RELEELNVPGEIVESpLSpSpSpEESITR (β-C-(16–40))	Tetra	+	+	+	+	+	-	+	+
3132.20	KNTMEHVSpSpSpEESpIISQETYKQEK (α-S2-(16–39))	Tetra	+	+	+	+	+	-	+	+
			23	20	19	20	21	12	14	18

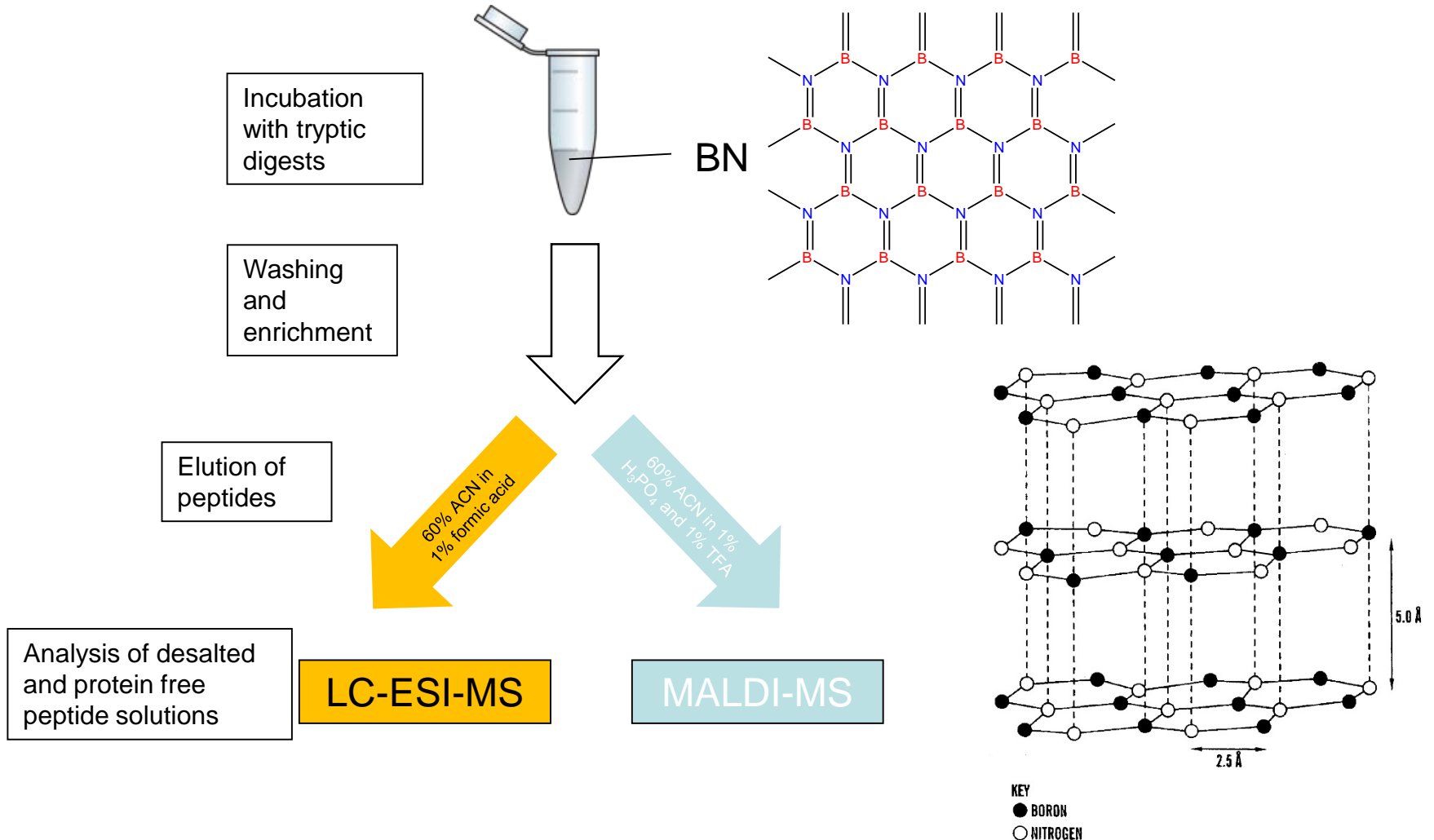
Highly Efficient Enrichment and Separation of Biomolecules



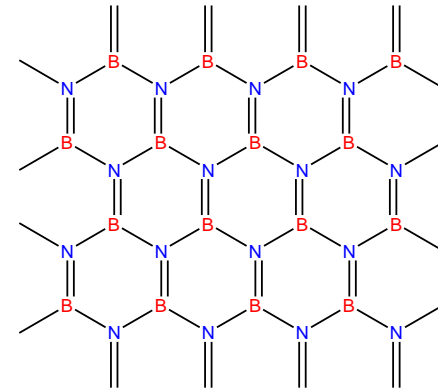
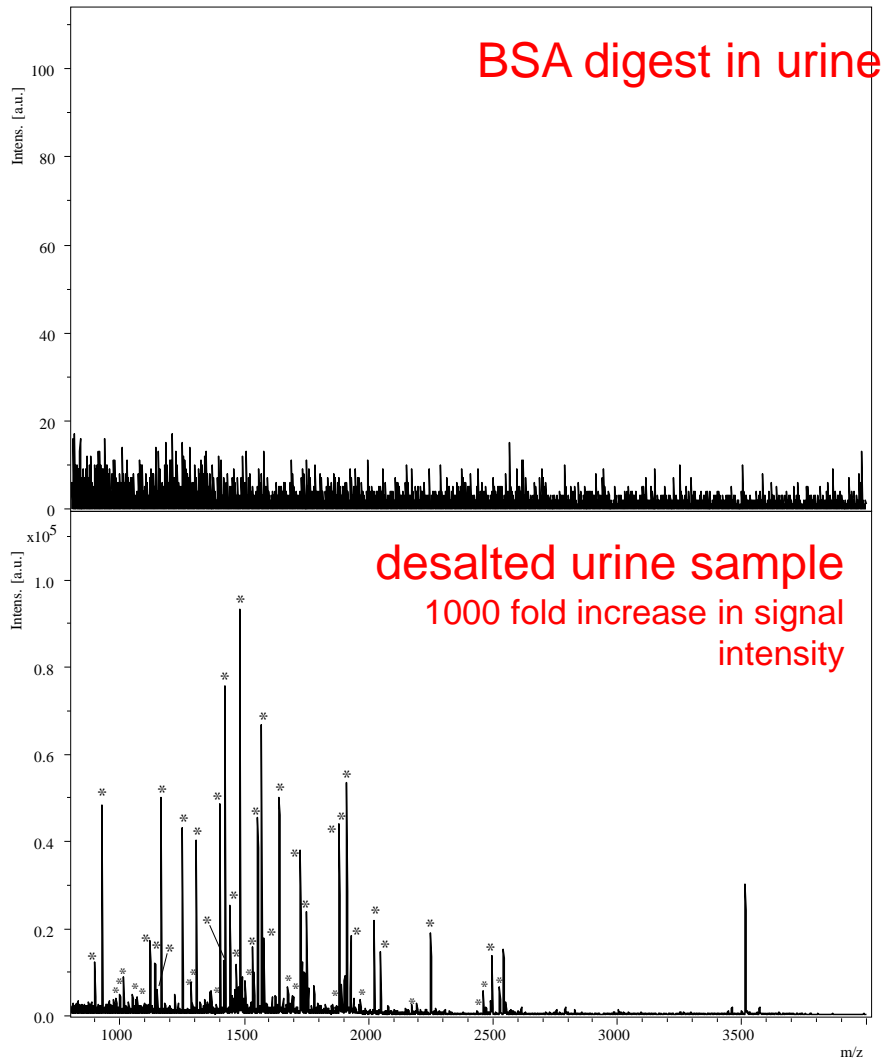
Boron Nitride, a Novel Material for the Enrichment and Desalting of Protein Digests and the Protein Depletion



Boron Nitride, a Novel Material for the Enrichment and Desalting of Protein Digests and the Protein Depletion



Boron Nitride, a Novel Material for the Enrichment and Desalting



MALDI Spectra:

The identified peptides are labeled with asterisk. Sequence Coverage of desalted BSA > 68 %

Bisphenol derivatives

	MW	pKa	log K _{ow}	R ²	LOD [ng/ml]	LOQ [ng/ml]
BADGE·2H₂O	376.44	14.7 ^b	2.05 ^b	0.999	27.0	82.0
BPF	200.23	7.5 ^a	2.91 ^a	0.999	27.0	82.0
BPA	228.29	9.6 ^a	3.32 ^a	0.999	33.0	99.0
BPZ	268.35	9.7 ^b	4.53 ^b	0.999	28.0	85.0
BADGE	340.41	-	4.02 ^a	0.999	30.0	90.0

BPF

BPA

BPZ

BADGE·2H₂O

BADGE

Recovery for the enrichment of 5 Phenols

	Phenol [12.5 µg/ml]	4-Nitrophenol [12.5 µg/ml]	2-Chlorphenol [12.5 µg/ml]	2-Nitrophenol [12.5 µg/ml]	Dimethylphenol [12.5 µg/ml]
	87.51	94.74	108.08	91.29	100.71
	93.97	102.95	114.70	95.20	111.82
	91.36	101.91	113.24	96.88	105.84
Average	90.95	99.87	112.01	94.46	106.13
RSD	2.66	3.65	2.84	2.34	4.54

5 mg of BN was incubated with a phenol mixture and afterward eluted with 80% acetonitrile in water.

Risk associated with bisphenols

- The French Agency for Food, Environmental and Occupational Health & Safety showed that there are ‘recognized’ effects in animals (**effects on reproduction**, on the mammary gland, on metabolism, the brain and behaviour) and other ‘suspected’ effects in humans (on reproduction, metabolism and **cardiovascular diseases**). These effects could be observed, even at low levels of exposure, during sensitive phases of an individual’s development.
- The French Agency for Food, Environmental and Occupational Health & Safety endorses the conclusions of the Expert Committee on Assessment of the risks related to chemical substances relating to the risks associated with BPA for human health, and on toxicological data and data on the use of bisphenols M, S, B, AP, AF, F and BADGE.

- **Endokrine Disruptoren**

Tolerable Daily Intake (TDI) of **50 µg BPA/kg** body weight (b.w.)/day as set by EFSA in 2006.

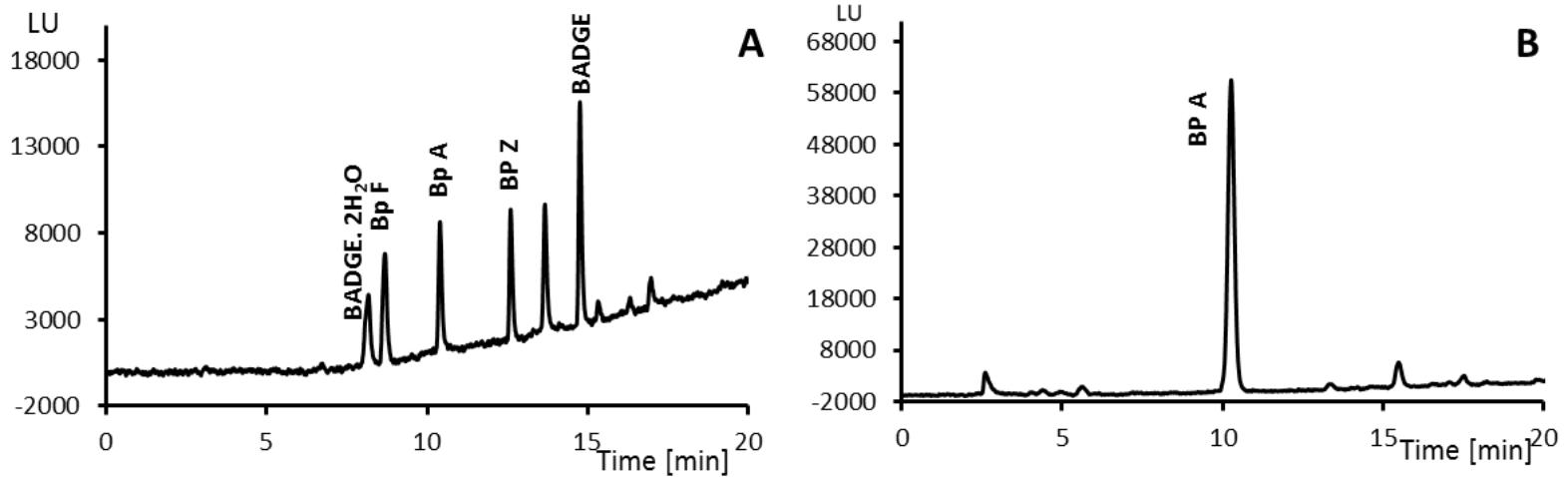
The migration limit of BPA set by European Union of **600 µg/kg** food.

Tolerable Daily Intake (TDI) of **150 µg** BADGE and its hydrolytic products/kg body weight (b.w.)/day.

The migration limit of BADGE and its hydrolytic products **9000 µg/kg** food.

<http://www.efsa.europa.eu/en/topics/topic/bisphenol.htm>

Analysis of 5 bisphenol derivatives after enrichment with BN by LC and fluorescence detector



HPLC-FLD chromatograms of (A) standard solution containing 5 bisphenol derivatives [100 ng/mL] and (B) bisphenol leached from baby bottle after enrichment with h-BN-60.

Chromatographic conditions: Shimadzu LC-10 Acvp, Phenomenex Luna 5u C18 250*4,6 mm, isocratic 3 min 30% B, gradient 30-70% B in 15 min, flow rate 1mL/min, 50°C, inj.: 30 µL, FLD: 230/303 nm

Determination of bisphenol derivatives

	Found \pm SD [ng/ml]				
	Recovery rates \pm SD [%]				
	BADGE·2H ₂ O	BPF	BPA	BPZ	BADGE
river water^a	nd (92.13 \pm 2.69)	nd (102.62 \pm 3.73)	nd (107.14 \pm 6.50)	nd (104.95 \pm 1.92)	nd (98.74 \pm 2.57)
drinking water^a	nd (90.16 \pm 2.40)	nd (98.56 \pm 3.51)	nd (99.16 \pm 1.60)	nd (101.14 \pm 2.58)	nd (96.68 \pm 3.35)
cola^b	4.63 \pm 0.14 (103.41 \pm 4.39)	2.23 \pm 0.21 (96.98 \pm 2.39)	10.34 \pm 0.41 (96.93 \pm 5.72)	nd (92.46 \pm 6.58)	1.49 \pm 0.04 (96.57 \pm 3.84)
apple juice^b	1.93 \pm 0.15 (96.43 \pm 3.63)	3.11 \pm 0.30 (90.77 \pm 3.63)	nd (84.22 \pm 2.20)	nd (98.74 \pm 1.23)	1.36 \pm 0.22 (98.22 \pm 2.71)
lemon soda^b	8.14 \pm 0.09 (82.46 \pm 3.01)	1.40 \pm 0.07 (84.45 \pm 5.44)	nd (97.60 \pm 1.81)	nd (87.61 \pm 2.94)	5.70 \pm 0.04 (94.00 \pm 2.61)
citrus soft drink^b	5.32 \pm 0.71 (103.90 \pm 2.58)	nd (97.82 \pm 5.82)	1.04 \pm 0.31 (98.31 \pm 5.67)	nd (98.05 \pm 4.04)	1.56 \pm 0.042 (99.63 \pm 4.42)
herbal soda^b	nd (100.45 \pm 5.15)	nd (99.07 \pm 5.09)	nd (104.43 \pm 6.20)	nd (105.07 \pm 7.21)	nd (108.42 \pm 4.83)
energy drink^b	4.46 \pm 0.07 (98.54 \pm 9.21)	1.87 (91.55 \pm 4.21)	5.57 \pm 0.81 (93.55 \pm 5.84)	nd (100.76 \pm 2.04)	nd (103.60 \pm 1.91)
canned mushroom liquid^b	20.42 \pm 1.19 (99.06 \pm 6.88)	nd (91.35 \pm 0.79)	8.60 \pm 0.37 (98.47 \pm 8.37)	nd (103.71 \pm 6.17)	1.81 \pm 0.12 (99.63 \pm 0.62)
pickled cucumber liquid^b	1.81 \pm 0.15 (97.53 \pm 1.12)	2.23 \pm 0.21 (96.02 \pm 2.45)	125.00 \pm 2.18 (109.12 \pm 0.77)	nd (103.36 \pm 3.80)	1.44 \pm 0.06 (92.67 \pm 0.71)
pickled onion liquid^b	2.98 \pm 0.08 (105.80 \pm 3.90)	nd (96.53 \pm 8.20)	13.45 \pm 0.63 (92.03 \pm 4.70)	nd (97.91 \pm 3.12)	1.74 \pm 0.00 (95.20 \pm 0.63)
urine^b	nd (100.45 \pm 3.13)	nd (105.67 \pm 0.68)	nd (92.80 \pm 2.52)	nd (105.26 \pm 4.00)	nd (103.70 \pm 3.45)

Leaching of BPA from polycarbonate products

	leached BPA [ng/ml]	± SD (%)
Babybottle 1. treatment	20.79	1.05
Babybottle 2. treatment	19.82	0.72
muffin form	2.26	0.07
Syringe	0.85	0.03

Treatment conditions: boiling water for 1 h



Peking University



University of Innsbruck

Health Science Center



Institute of Analytical Chemistry and
Radiochemistry
Horvath Lab of Bioseparation



Chinese - Austrian Center for Biomarker Discovery at Peking University Health Science Center

TEACHING COURSES

- **Fundamentals** - separation sciences e.g. electrophoresis, HPLC, capillary electrophoresis
 - mass spectrometry, LC-MS, MALDI
 - biomarker discovery
 - including practical exercises

Lecturers from Austria, PUHSC and guests; **Duration** per year: 6 – 14 days

SCIENTIFIC PROJECTS

- **Development of new biomarker methods** as well as new materials for separation and MELDI
- **Development of disease related biomarker discovery methods including bioinformatics** e.g. liver cancer and prostate cancer
 - **Application studies**
 - **Natural Product Proteomics** (Phyto-Proteomics)

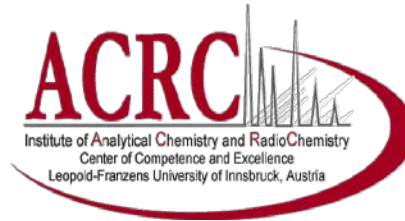
FELLOWSHIP PROGRAMS - Exchange on different levels

Academics, Students and Non-academic positions – technicians

Participation at the International Proteomics Conference, Seefeld Austria

Duration: 2 weeks, 3 months, up to 3 years (PH.D.) fellowships

Collaborations



Prof. Günther Bonn



International Agency for Research on Cancer
Centre International de Recherche sur le Cancer

WHO – Lyon, France



北京大学医学部
Peking University
Health Science Center

Prof. Ke Yang



Beijing Institute of Genomics,
Chinese Academy of Sciences

Prof. Siqi Liu



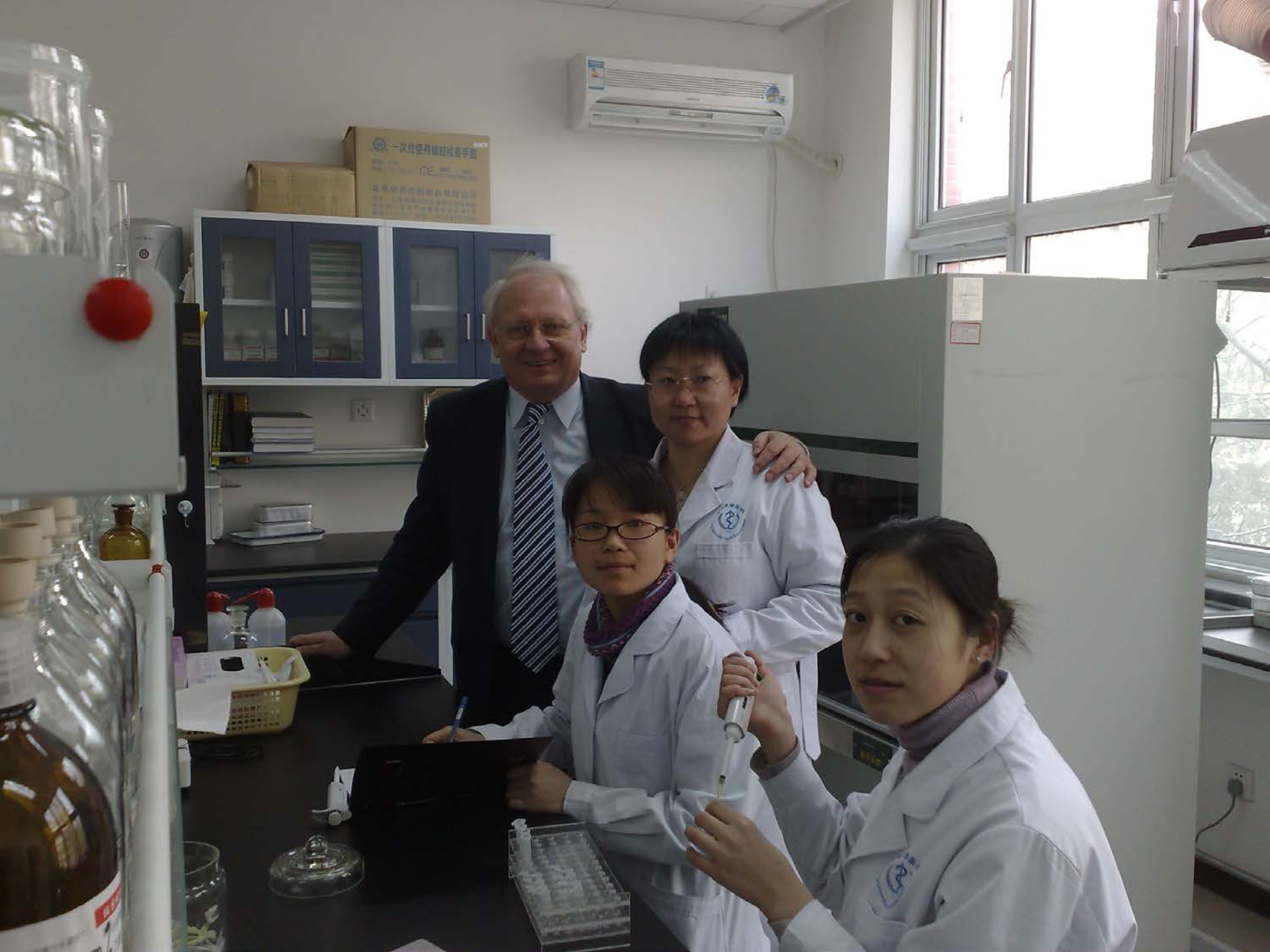
北京大学医学部

国际合作交流中心

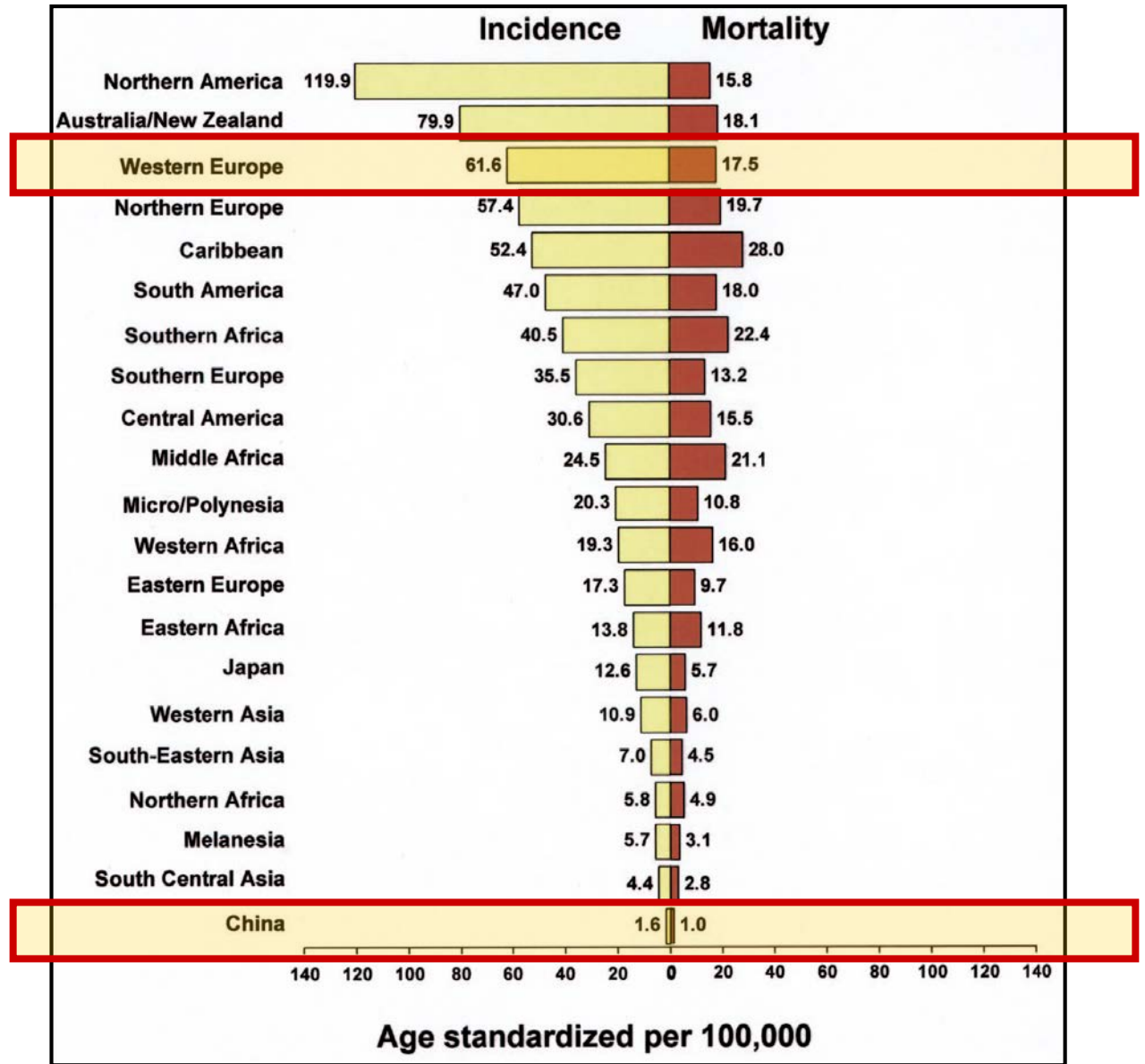


中英生物标志物研究中心
Sino-Austrian Bio-Marker Center





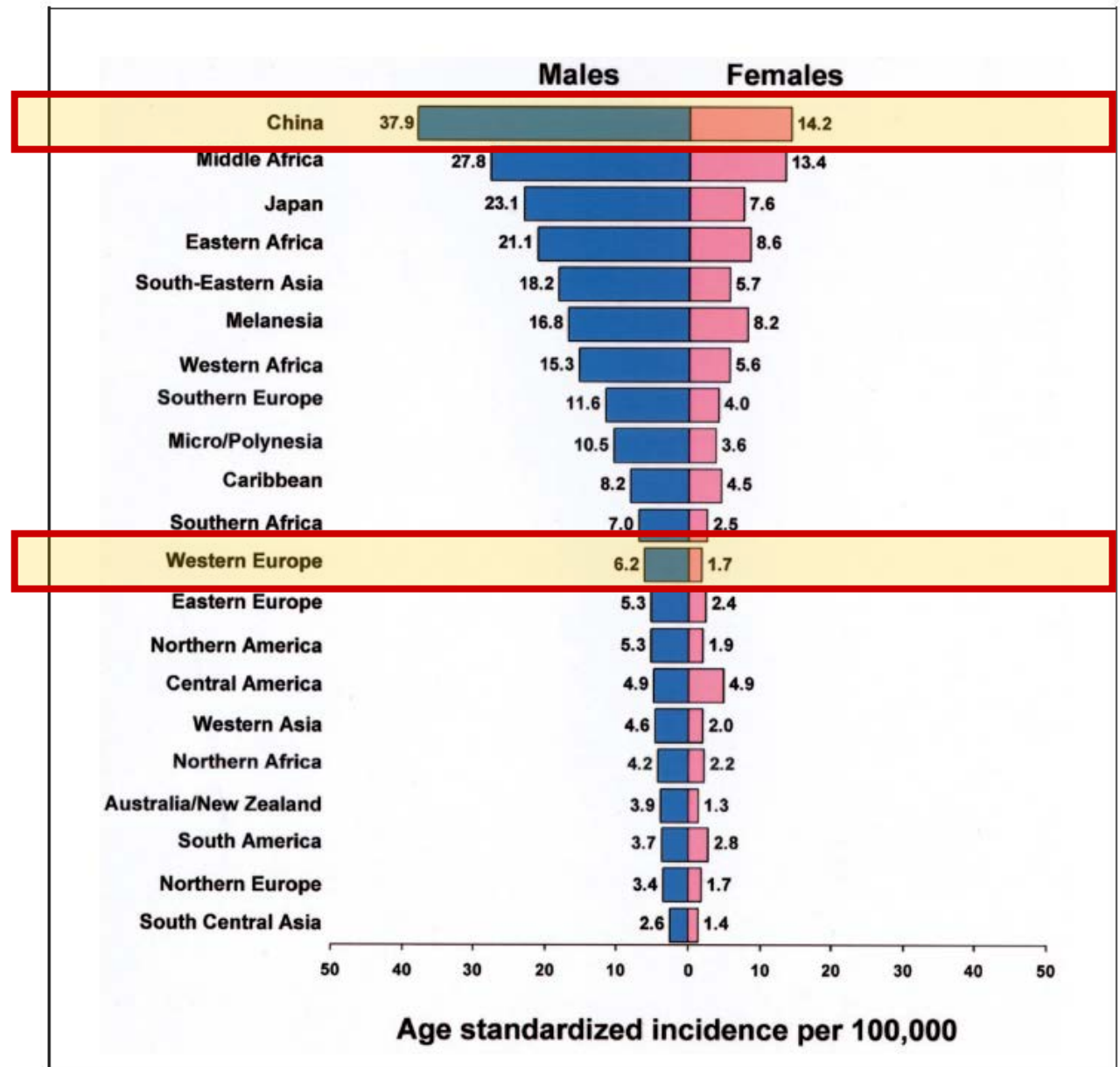
Prostate Cancer



D. Max Parkin, Freddie Bray, J. Ferlay and Paola Pisani *CA Cancer J Clin* 2005;55;74-108

Age-standardized Incidence and Mortality Rates for Prostate Cancer. Data shown per 100,000.

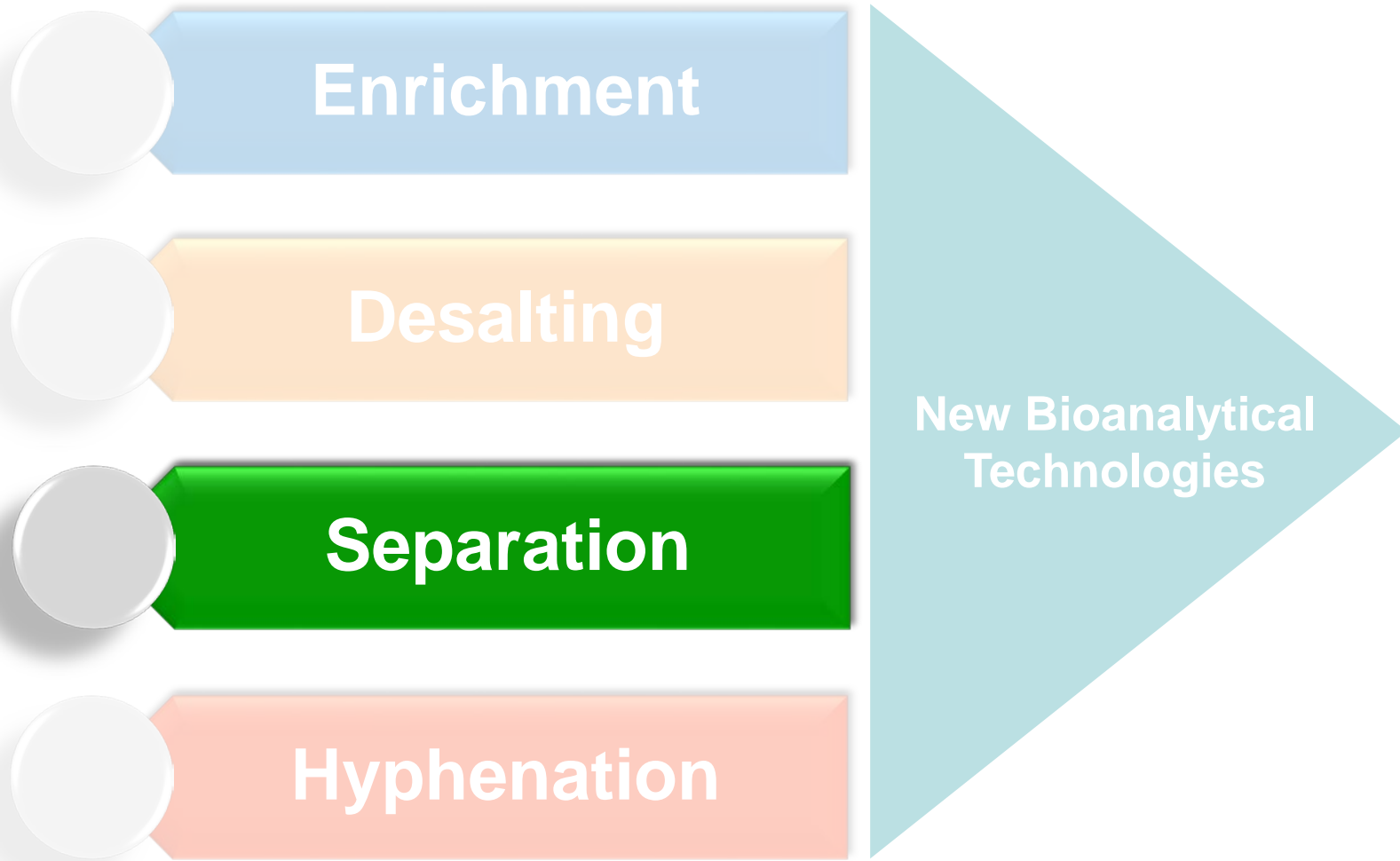
Liver Cancer



D. Max Parkin, Freddie Bray, J. Ferlay and Paola Pisani *CA Cancer J Clin* 2005;55:74-108

Age-standardized Incidence and Mortality Rates for Liver Cancer. Data shown per 100,000.

Highly Efficient Enrichment and Separation of Biomolecules

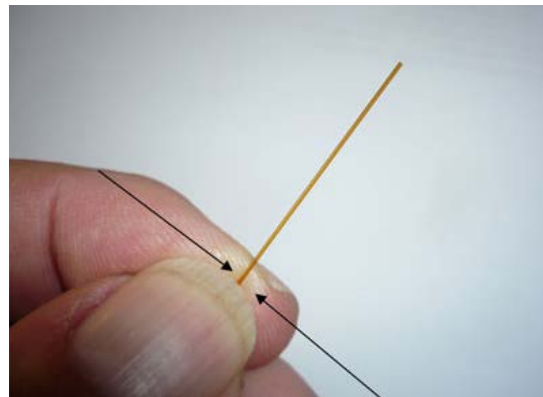


Stationary Phases – Formats – Strategies

HPLC - column



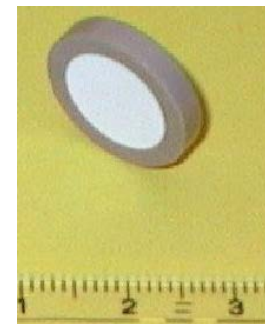
μ -LC - capillary



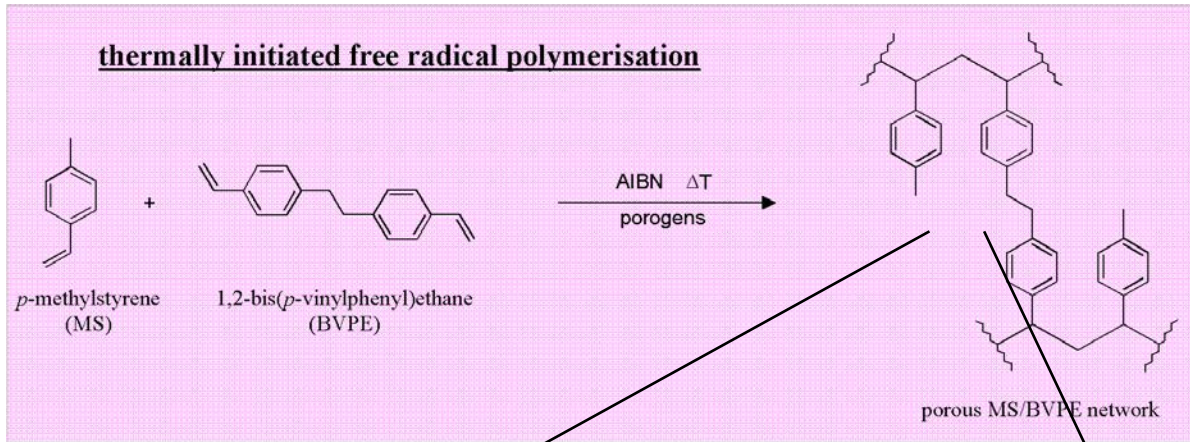
**Chip -
LC**



Disc - technology

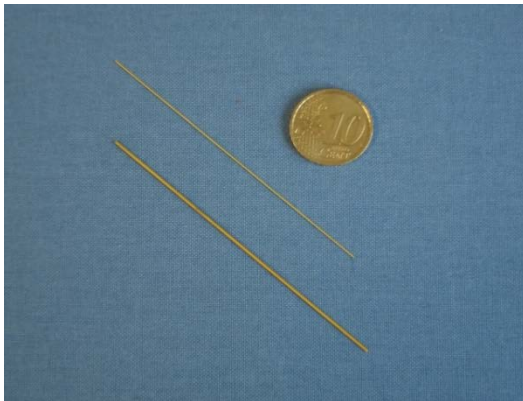


MONOLITHIC MS/BVPE



- highly crosslinked
- high crosslinking homogeneity due to non-conjugated crosslinker
- minimised swelling
- high pressure stability

→ **Applicable to HPLC**



capillary columns
different dimension

80 x 0.2 mm
80 x 0.533 mm



conventional HPLC columns

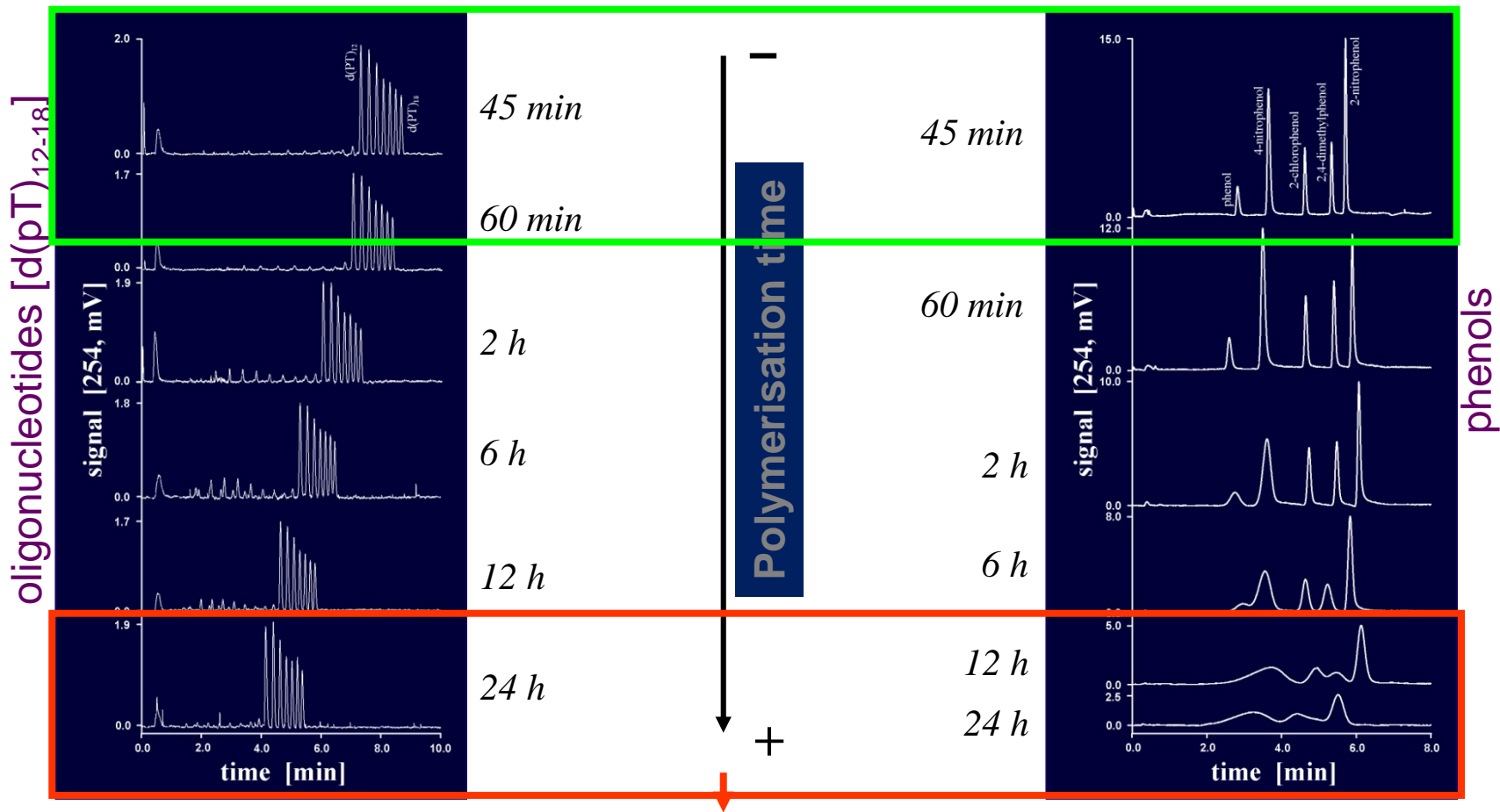
90 x 3.0 mm

Monolithic Separation

Impact on the Separation Efficiency of Monolithic MS/BVPE

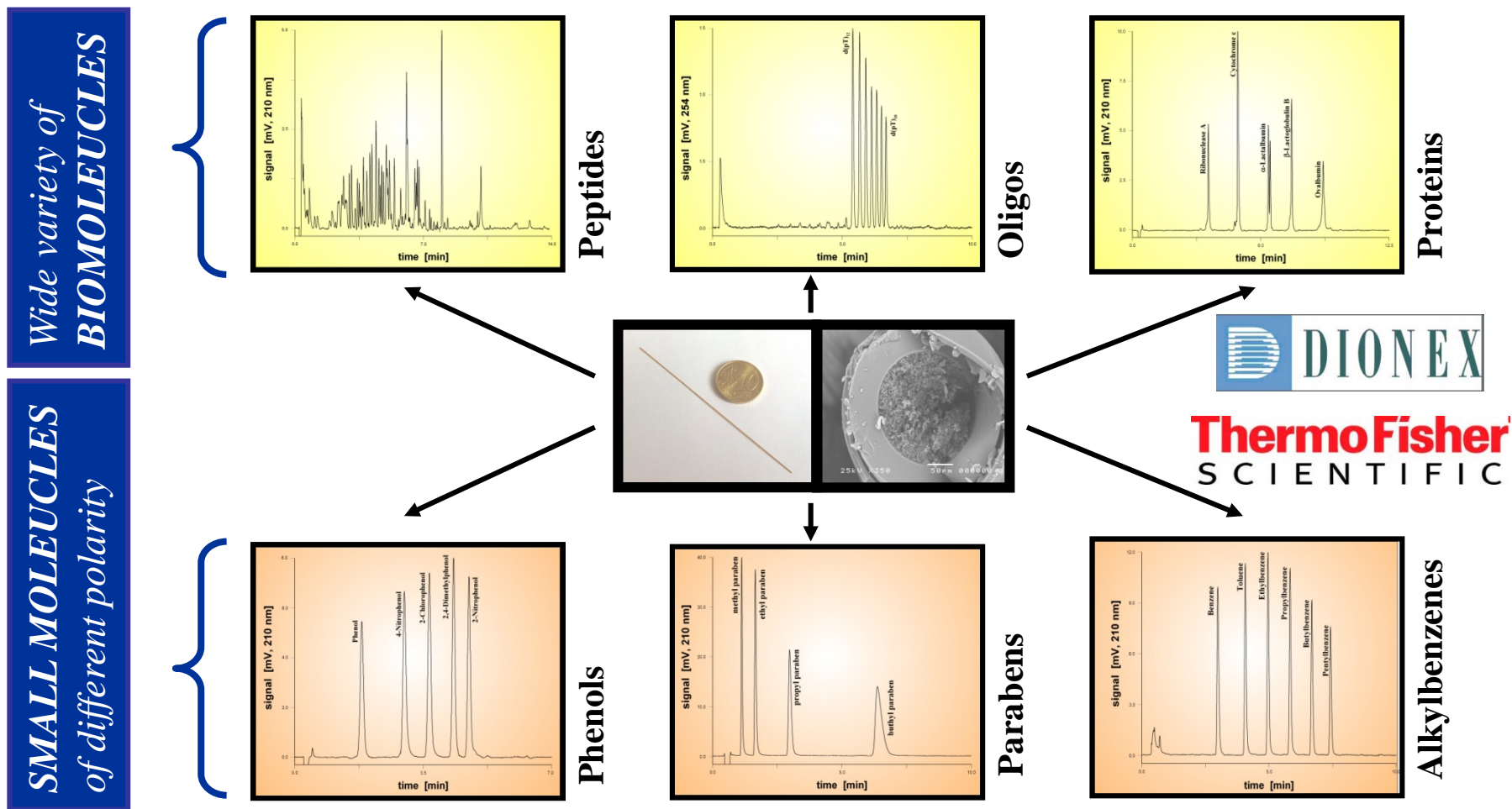
Biomolecules

Low-molecular-weight compounds



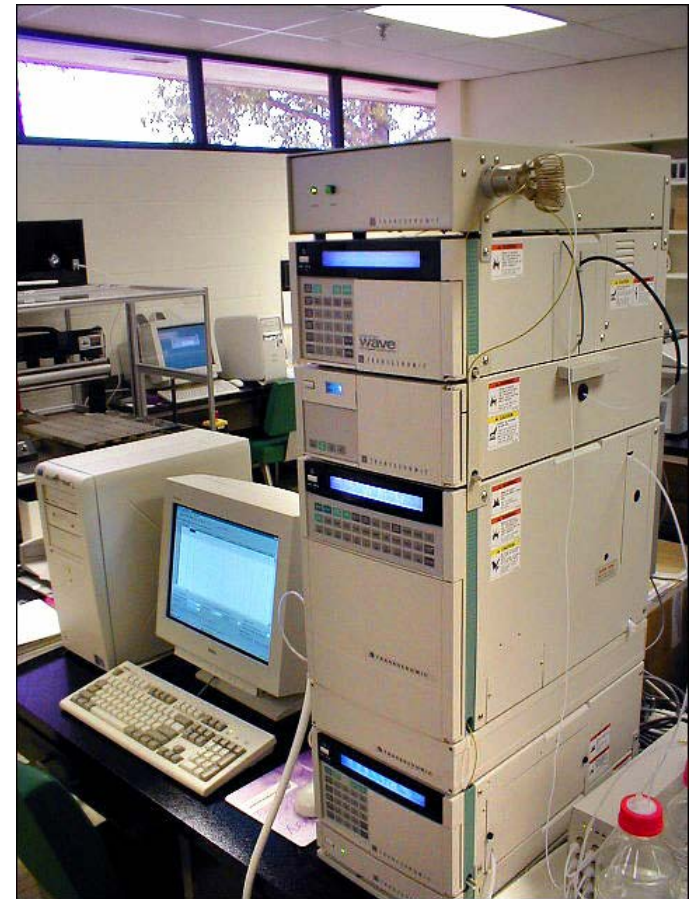
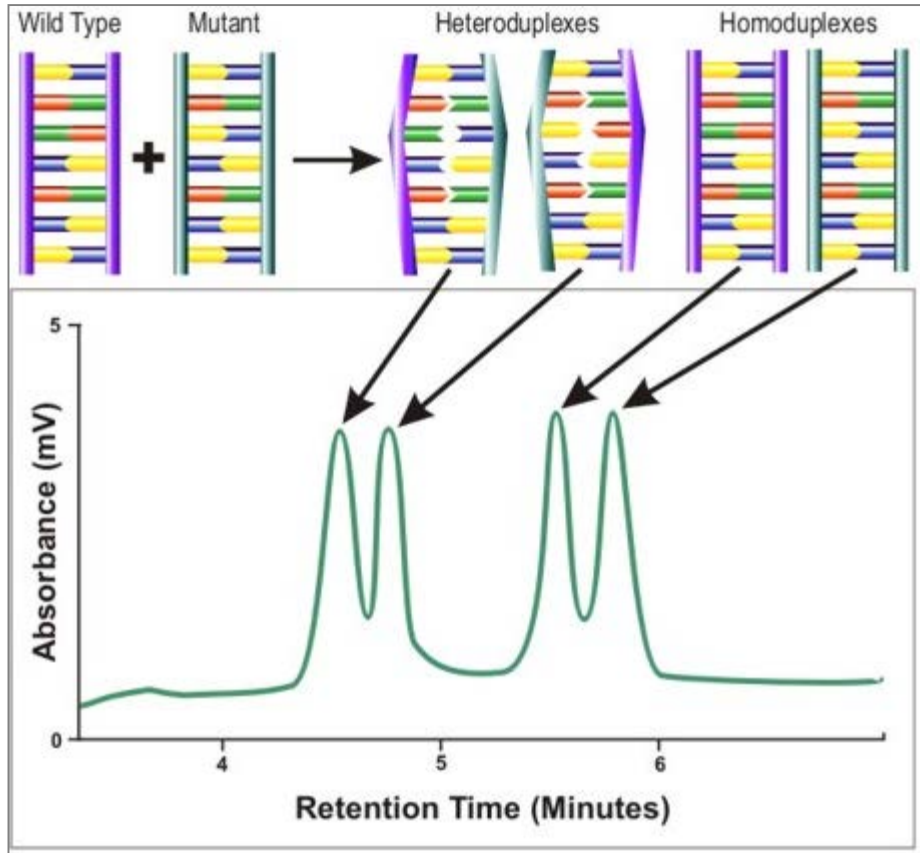
Achievable also with commercial monolithic organic capillary columns

Extended Applicability of the shortly Polymerised Monoliths



→ Separation of biopolymers as well as small molecules of various polarity can be separated on the same monolithic capillary column

Transgenomic Wave-System - Mutation Analysis

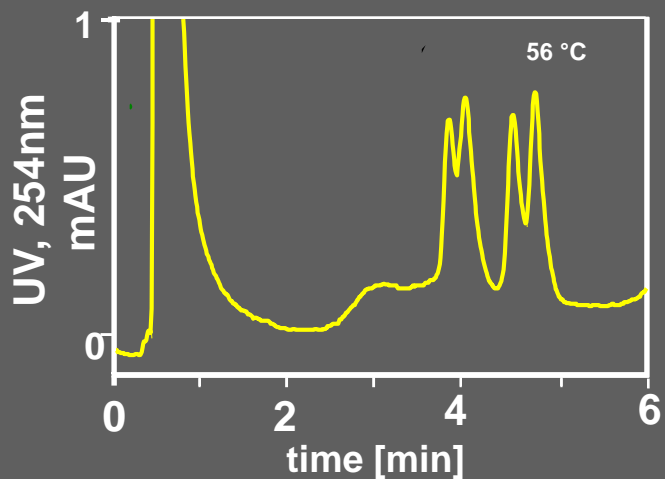
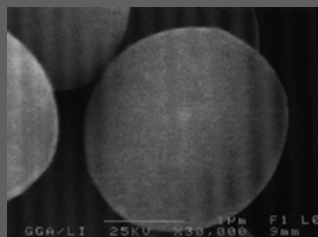


Transgenomic Wave HPLC-System

Bonn, Gunther; Huber, Christian; Oefner, Peter. **Separation of nucleic acid fragments with alkylated nonporous polymer beads.** *PCT Int. Appl.* (1994), 30 pp.

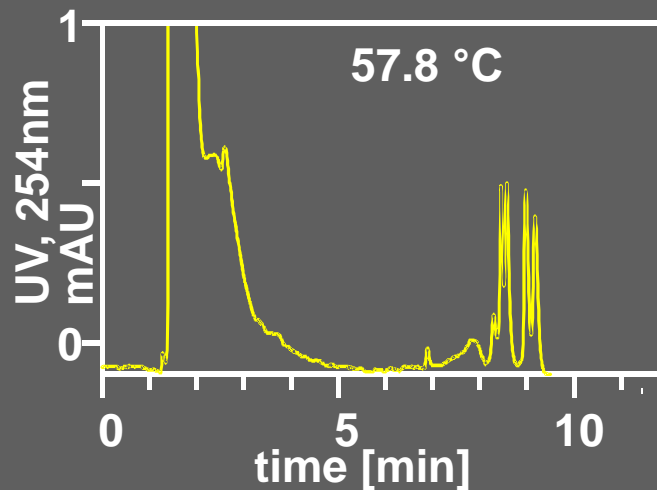
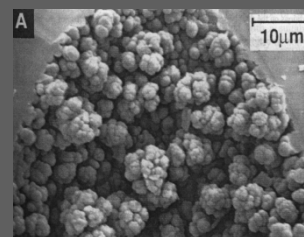
Mutation Detection by Denaturing HPLC using Packed Columns and Monolithic Capillaries

Packed Column



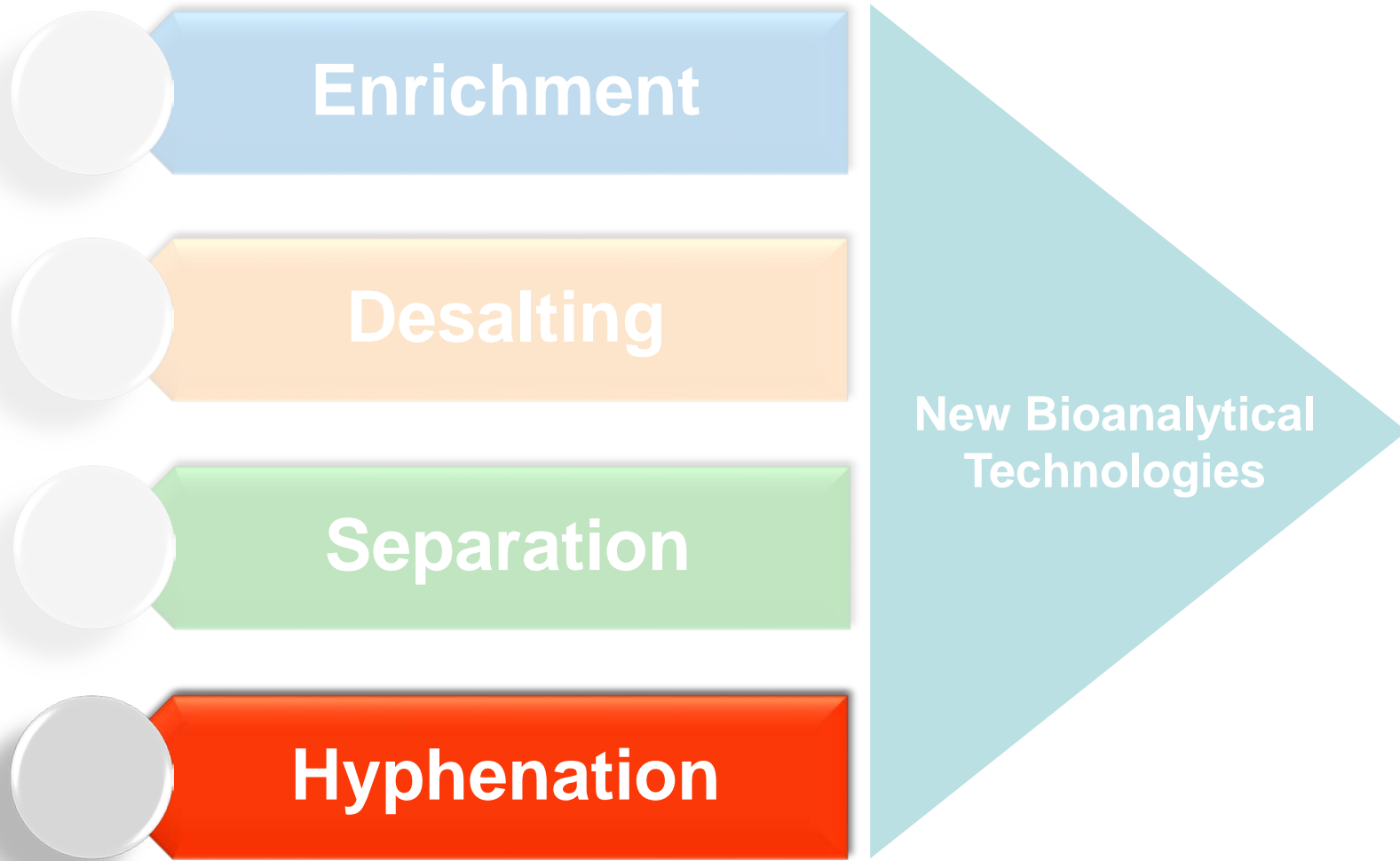
column: 50 x 4.6 mm I.D.
flow rate: 1 ml/min
injection vol.: 5 μ l

Monolithic Capillary

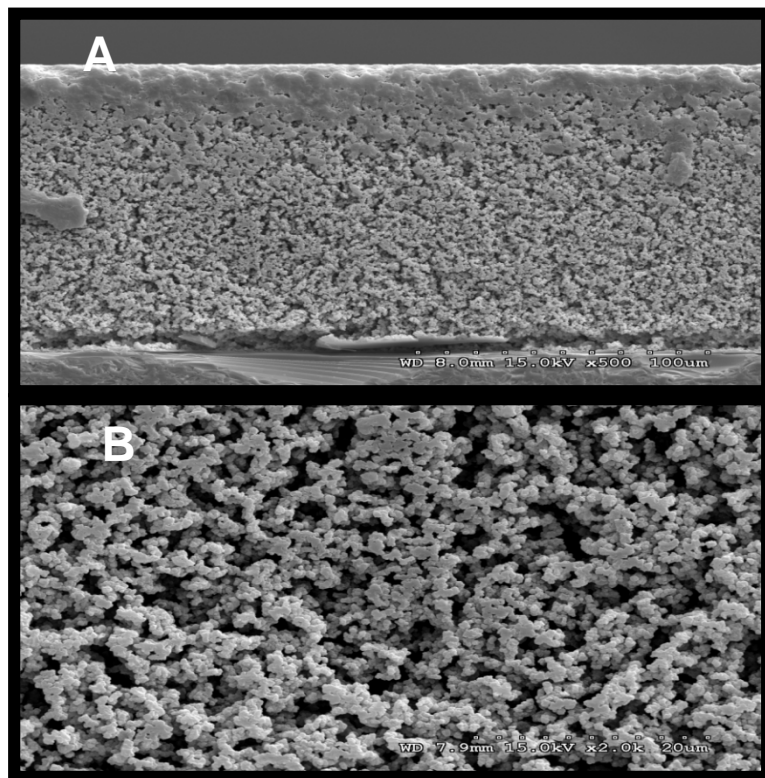


column: 50 x 0.2 mm I.D.
flow rate: 3.0 μ l/min
injection vol.: 500 nl

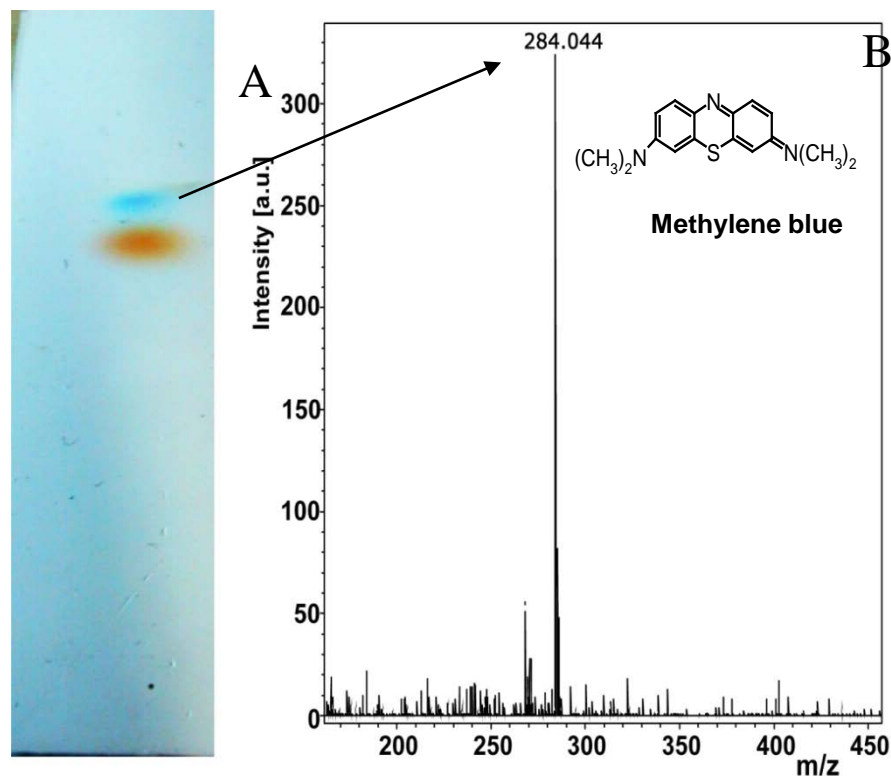
Highly Efficient Enrichment and Separation of Biomolecules



Monolithic Porous Polymer Layer For Thin-Layer Chromatography Coupled with MALDI- TOF-MS



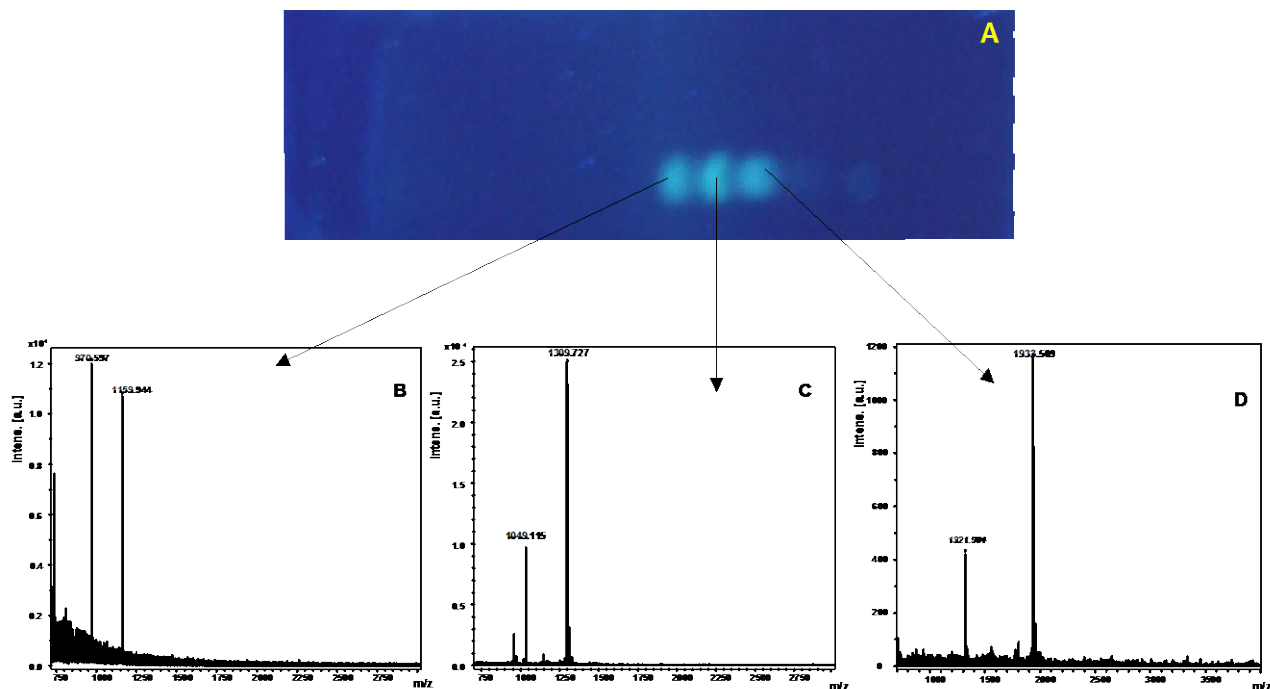
SEM micrographs of porous structure of 150 μm thick poly(butyl acrylate-co-ethylene dimethacrylate) monolith attached to a glass plate



TLC separation of methylene blue and methyl red on monolithic layer attached to a glass plate using ethyl acetate-ethanol-water mixture (6:4:3) as the mobile phase (A) and MALDI spectrum from the spot of methylene blue obtained “from-plate” without using any matrix (B).

Collaboration with Prof. F. Svec, Lawrence Berkeley National Lab

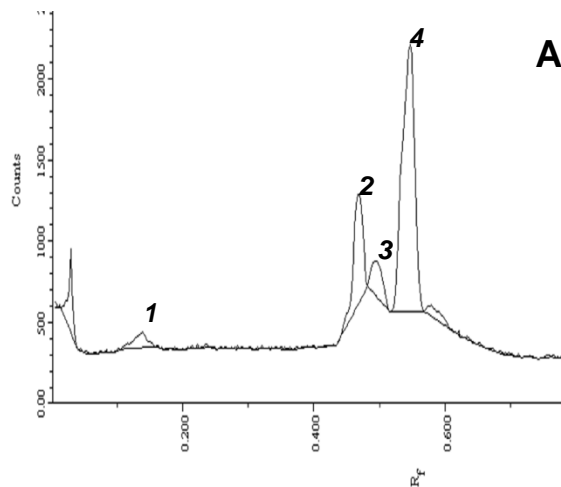
Monolithic Porous Polymer Layer For Thin-Layer Chromatography Coupled with MALDI-TOF-MS



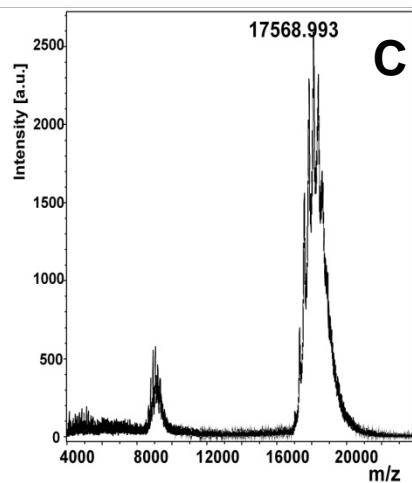
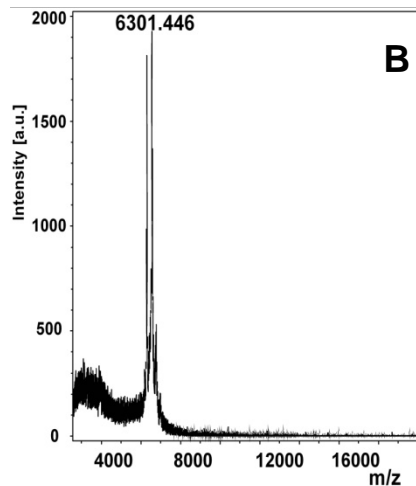
TLC separation of mixture of peptides labelled with fluorescamine on 150 μm thick poly(butyl acrylate-*co*-ethylene dimethacrylate) monolithic layer attached to a glass plate using 0.1% TFA in 45% aqueous acetonitrile as the mobile phase (A). Sample volume 0.5 μL MALDI-TOF MS spectra of fluorescently labelled [Sar1,Ile8]-angiotensin II (B), angiotensin II (C), and neurotensin (D) obtained “from-plate” using HCCA as matrix.

Collaboration with Prof. F. Svec, Lawrence Berkeley National Lab

Monolithic Porous Polymer Layer For Thin-Layer Chromatography Coupled with MALDI- TOF-MS

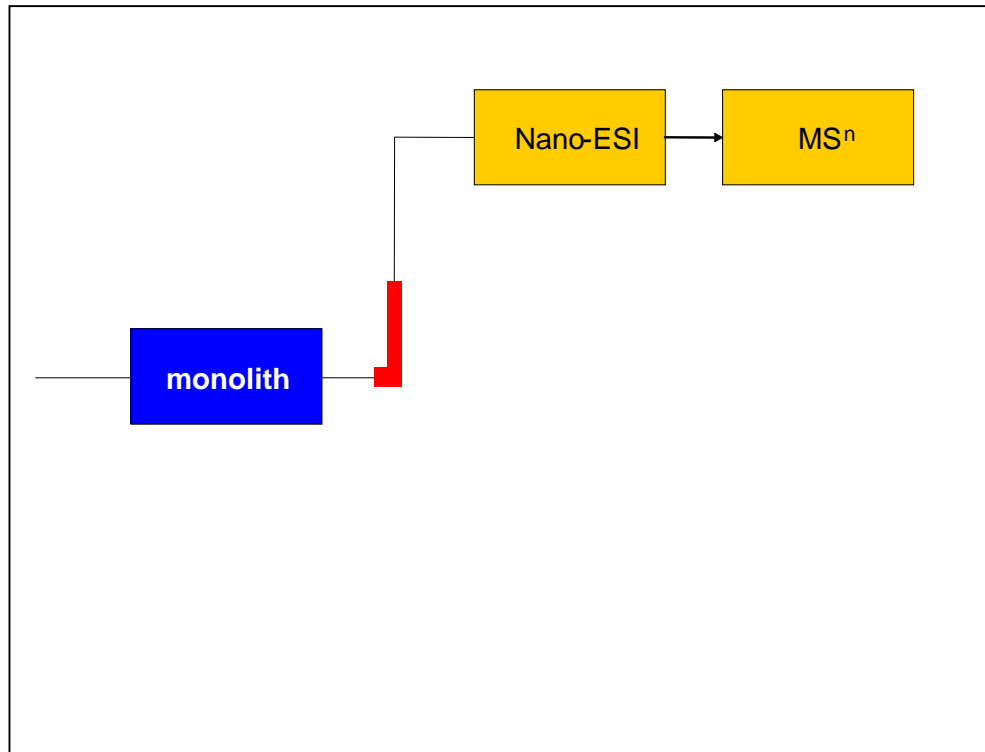


TLC separation of mixture of proteins labelled with fluorescamine (A). Sample volume 0.5 μ L. Peaks: Insulin (1), cytochrom c (2), lysozyme (3), and myoglobin (4). MALDI-TOF MS spectra of fluorescently labelled insulin (B), and myoglobin (C) obtained “from-plate” using sinapinic acid as matrix.



Structural Elucidation with Hyphenated techniques for Phyto-Metabolomics

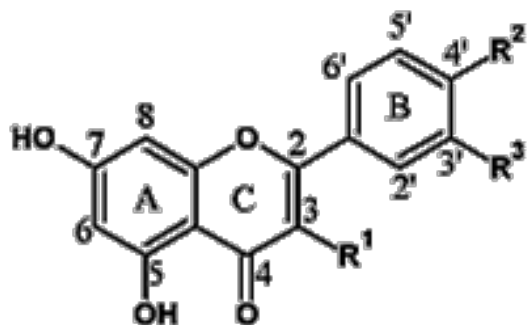
μ -HPLC-ESI-MS



Electrospray Ionisation (ESI-MS) – online

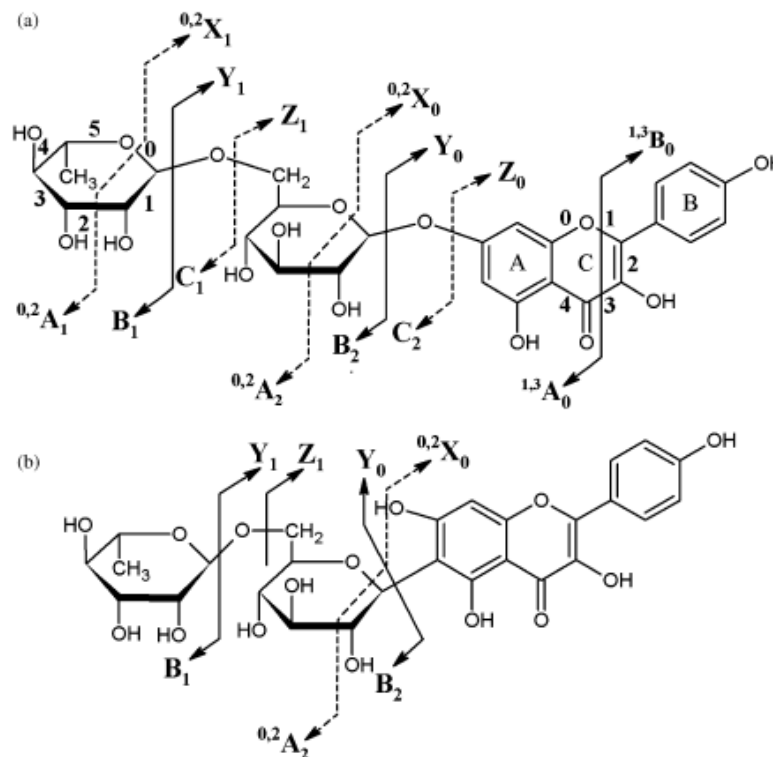
→ Increase speed of separation in case of MS instruments with high scan rates

Qualitative Analysis of Flavonoids of Heartsease (*Viola tricolor* L.)



	MW	R ¹	R ²	R ³
apigenin	270	H	OH	H
luteolin	286	H	OH	OH
kaempferol	286	OH	OH	H
chrysoeriol	300	H	OH	OCH ₃
diosmetin	300	H	OCH ₃	OH
quercetin	302	OH	OH	OH
isorhamnetin	316	OH	OH	OCH ₃

Core structure of flavonoid aglycones

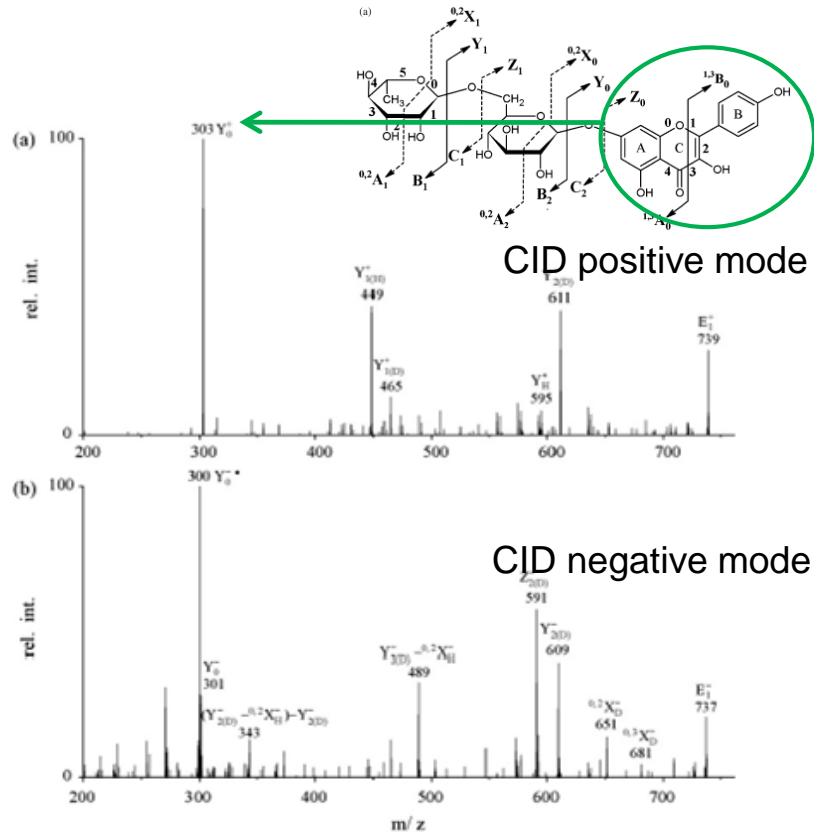


Fragment nomenclature applied for

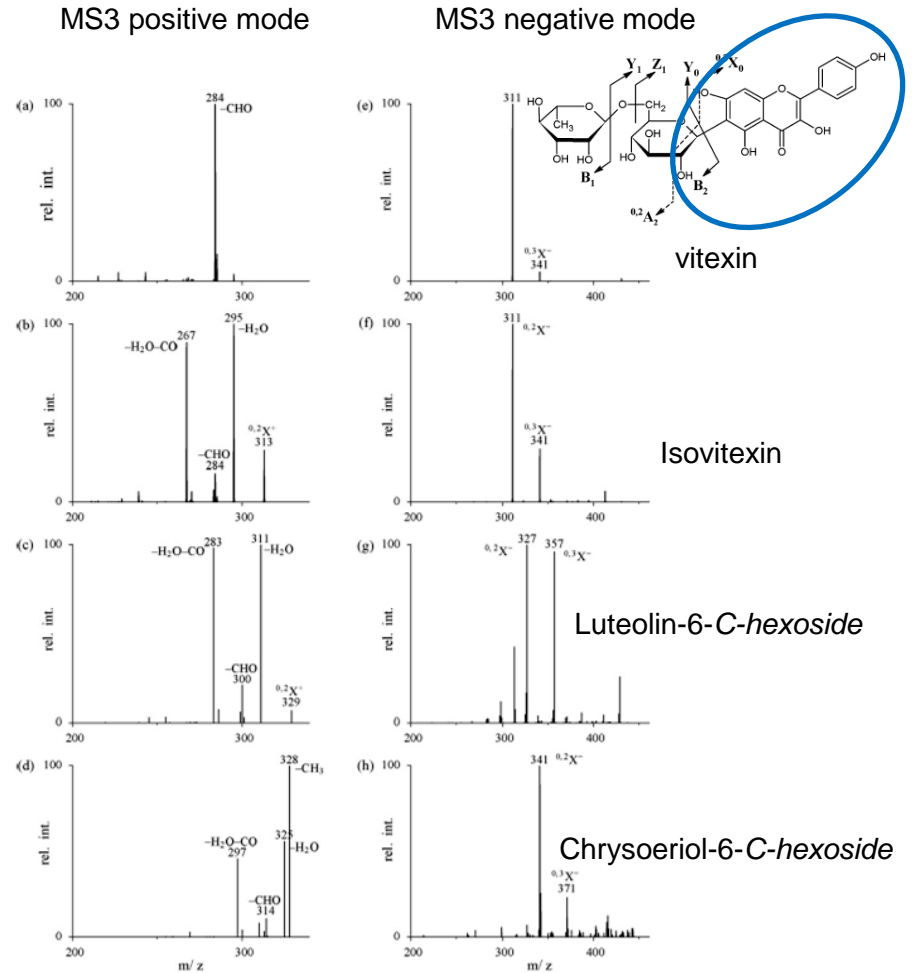
(a) O-glycosides

(b) C,O-glycosides and C-glycosides

Qualitative Analysis of Flavonoids of Heartsease (*Viola tricolor* L.)



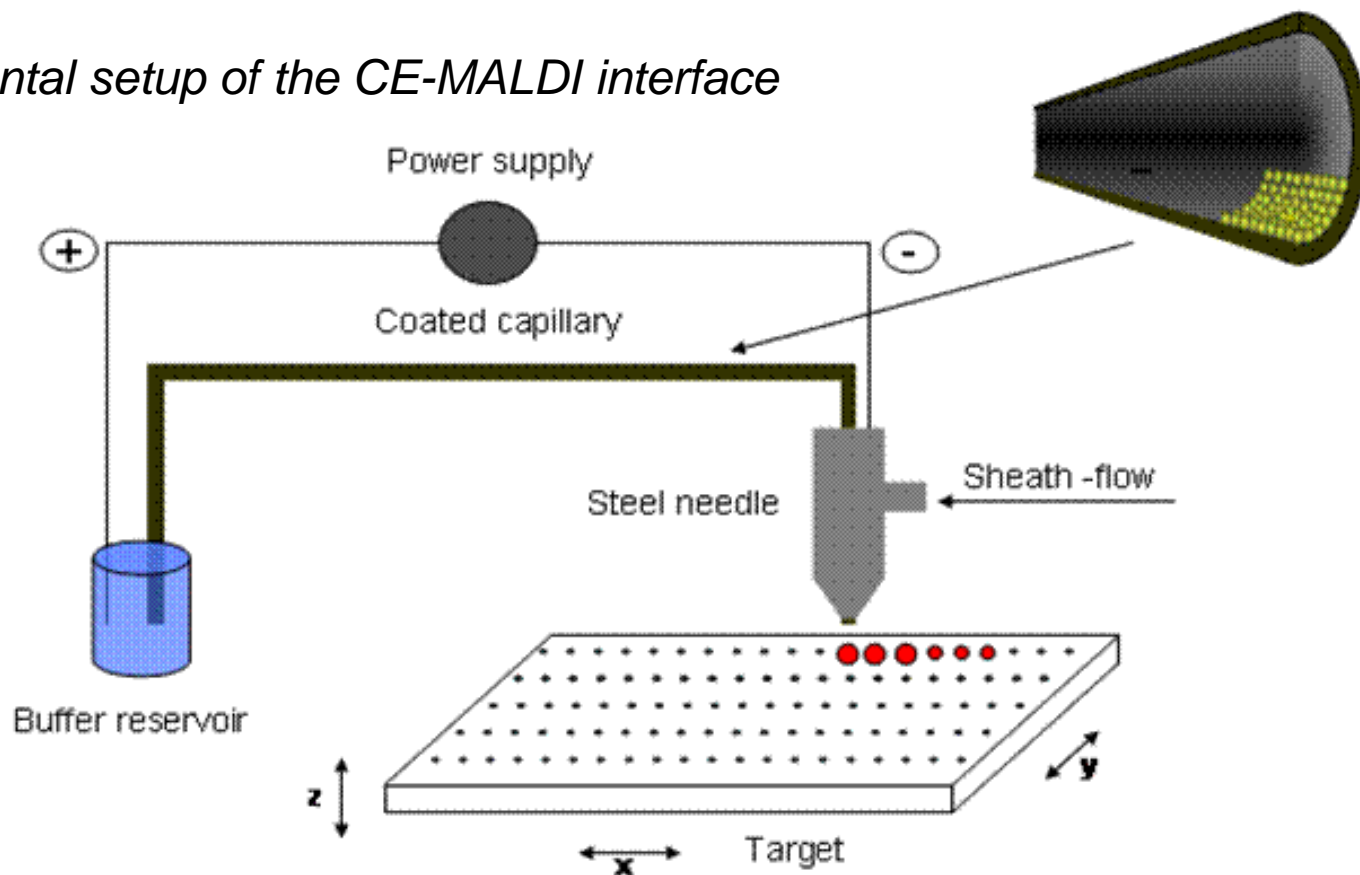
Fragment spectra of an O-glycoside
(Quercetin-3-O-deoxyhexosylhexoside-7-O-deoxyhexoside)



Fragment spectra of C-glycosides
Positive (a-d) and negative (e-h) ion MS3 spectra
of $[M+H]^+ \rightarrow 0, 2X^+$

Hyphenation of CE with MALDI

Experimental setup of the CE-MALDI interface

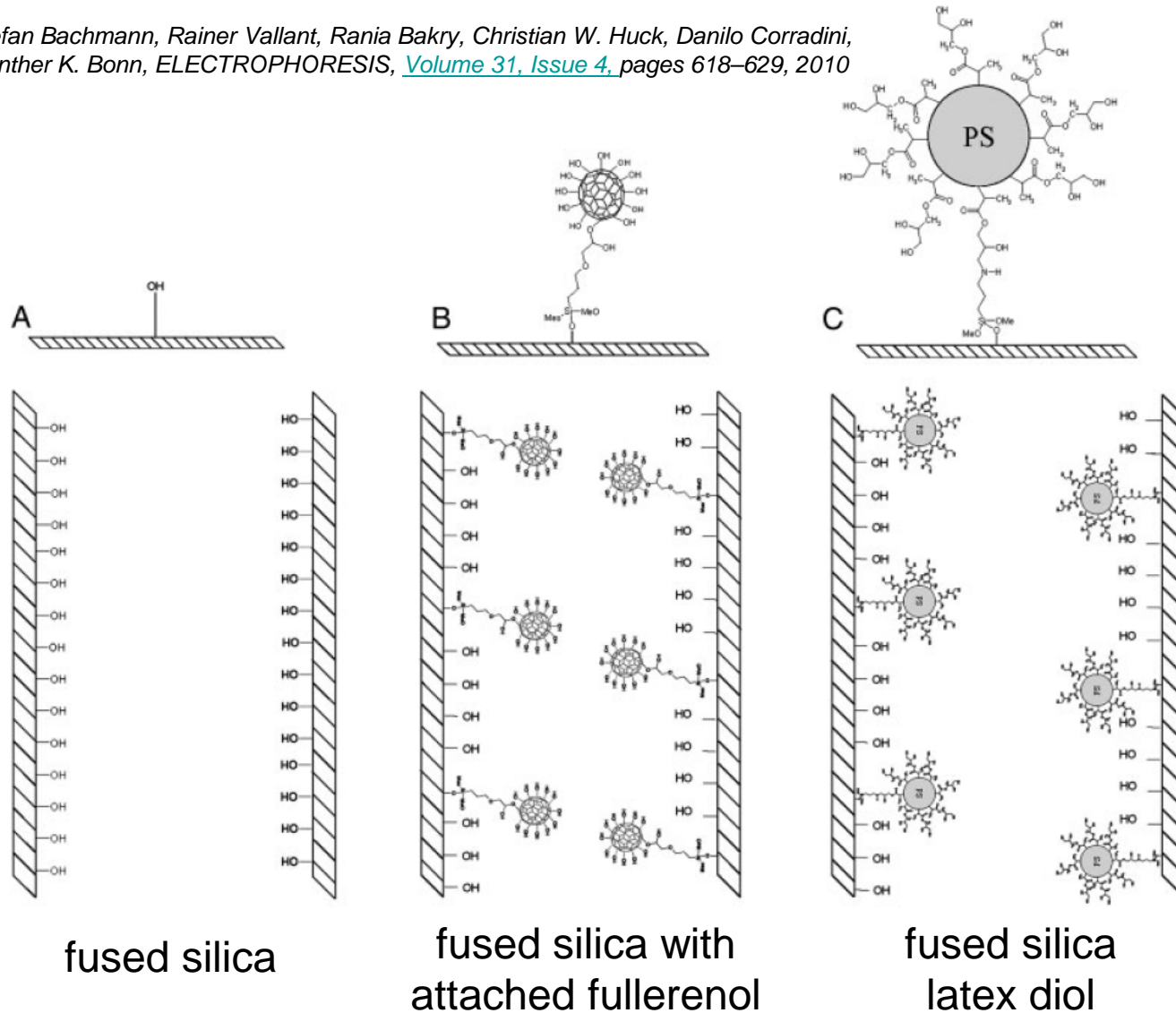


 *Institute of Chemical Methodologies*

Collaboration with Prof. Danilo Corradini

CE coupled to MALDI with novel covalently coated capillaries

Stefan Bachmann, Rainer Vallant, Rania Bakry, Christian W. Huck, Danilo Corradini, Günther K. Bonn, *ELECTROPHORESIS*, [Volume 31, Issue 4](#), pages 618–629, 2010

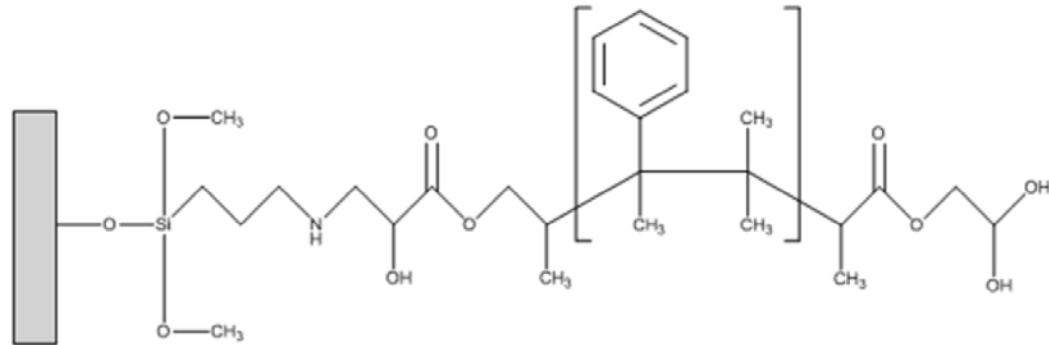


Characterization of coated capillaries for CE by infrared spectroscopy

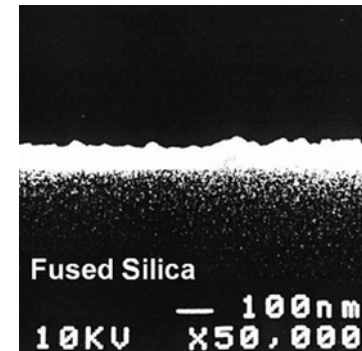
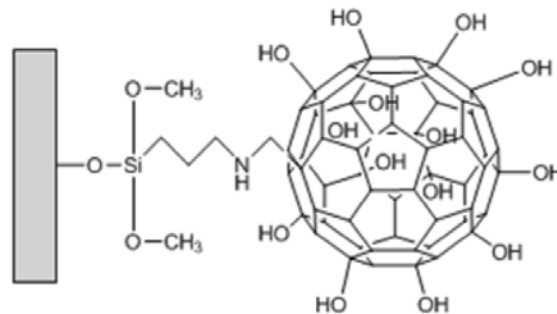
fused silica Polymicro capillary
(uncoated)



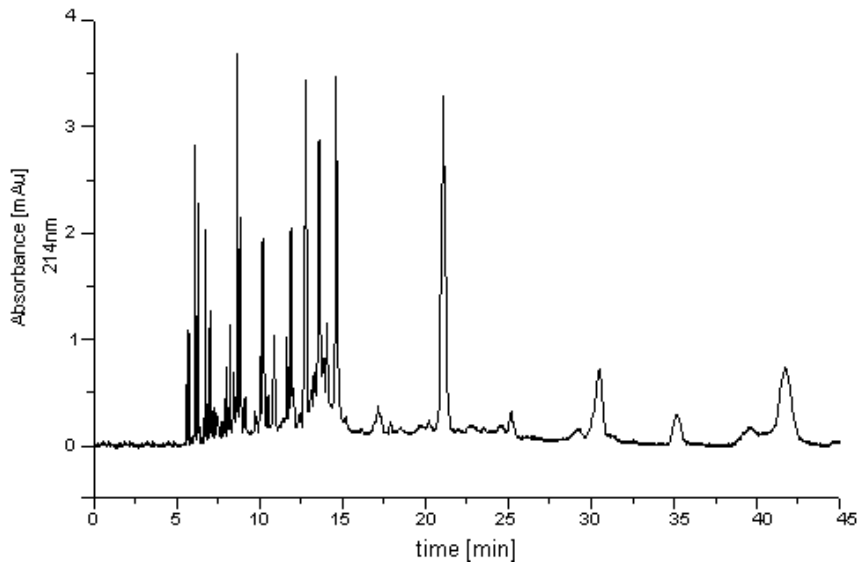
fused silica Polymicro capillary
(latex diol coated)



fused silica Polymicro capillary
(fullerenol coated)

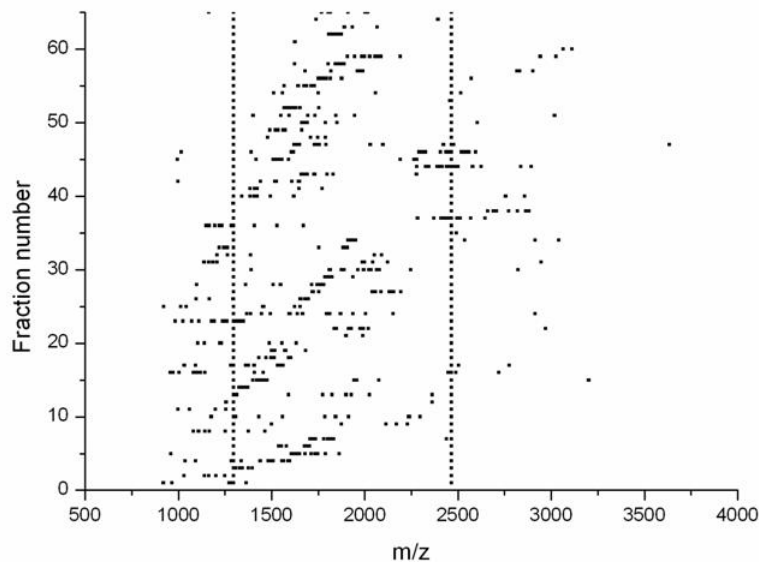


Hyphenation of MALDI with CE



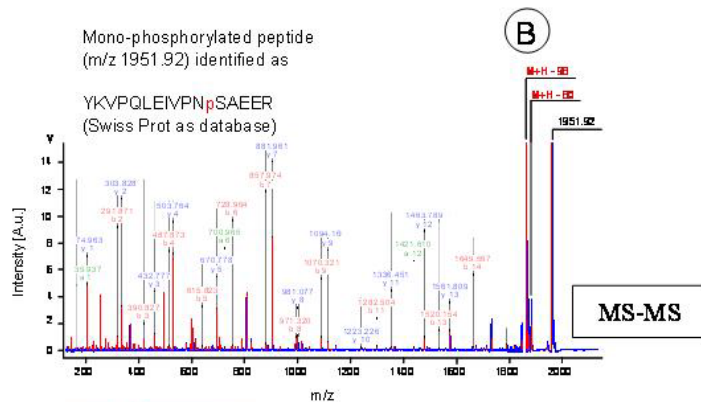
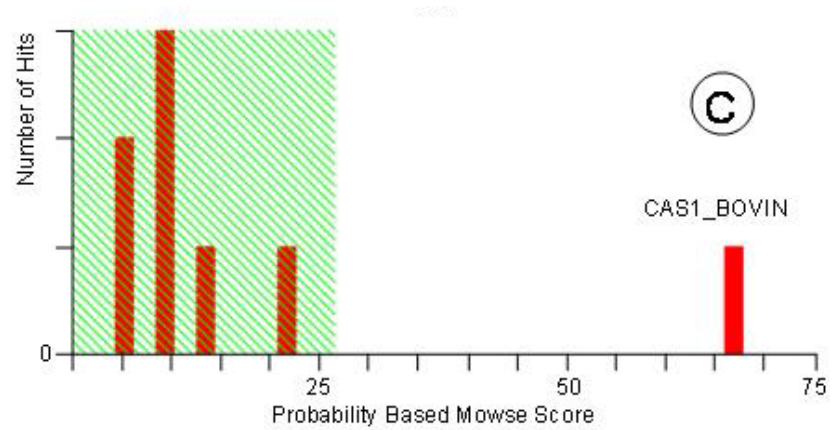
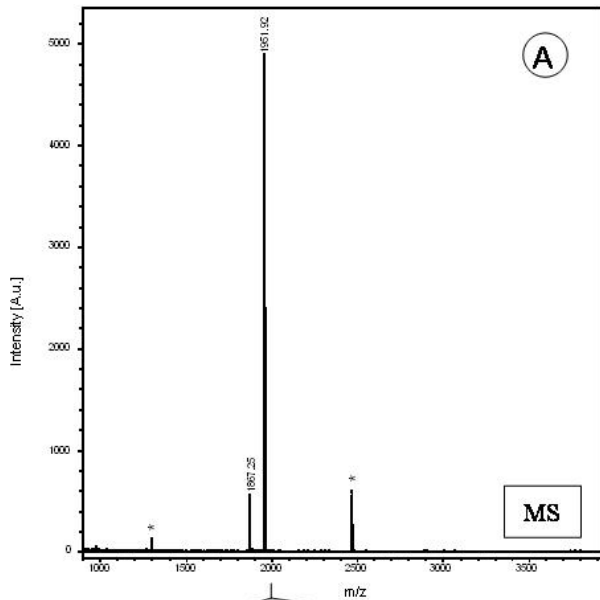
Electropherogram for the separation of α -casein tryptic digest obtained with latex diol.

Conditions: capillaries, (65/57.5 cm eff., 75 μ m ID, 360 μ m OD); BGE, pH 2.3: 40 mM TEA titrated with phosphoric acid mixed with 20% (v/v) ACN.



2D Plot CE-MALDI-MS data obtained from the analysis of a tryptic digest of five proteins (myoglobin, cytochrom c, BSA, α -casein, ovalbumin).

Hyphenation of MALDI with CE



(A) MALDI-TOF mass spectrum of **CE fraction no. 34**. **(B)** The fragment ion spectrum of peptide 1951.92 Da. **(C)** The identification probability plot from the swiss prot database.

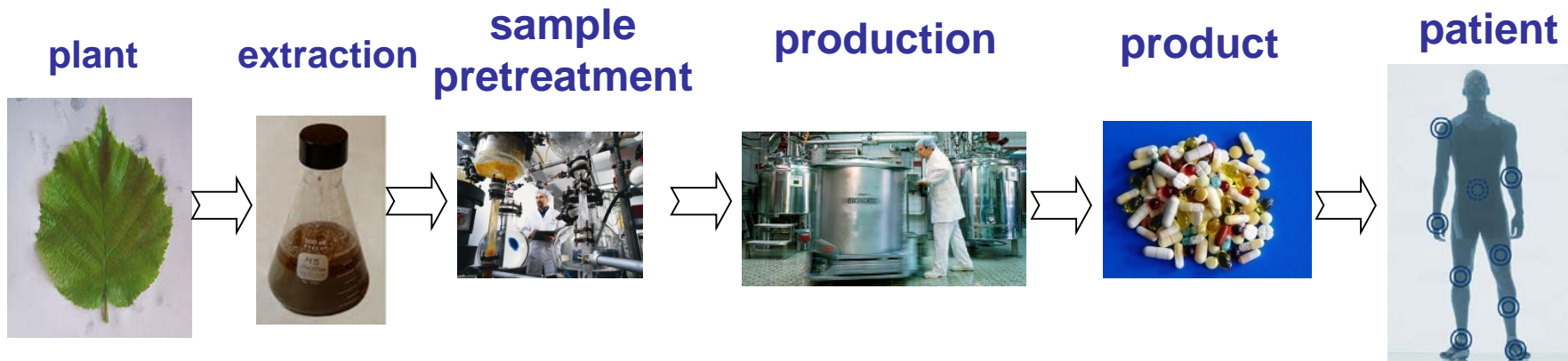
The internal standards are indicated with asterisks

A photograph of a lush green forest. In the foreground, a large, dark, weathered log lies on the ground, partially covered with fallen leaves. The forest floor is covered in a layer of brown and orange leaves. The trees are tall and thin, with dense green foliage. The lighting is soft, suggesting a dappled sunlight effect. A semi-transparent green banner is overlaid across the middle of the image, containing the text.

New Analytical Approaches in “Phytoanalytics”

Analytical Chemistry in Phytopharmacy

Steps for Quality Control



Selective materials for sample pretreatment:

particles (spherical, irregular)
monoliths
SPE, columns, disks

Stationary phases for HPLC and μ -HPLC:

particles (different mechanisms)
monoliths

Screening: MALDI targets/MELDI materials:

matrix free
with matrix

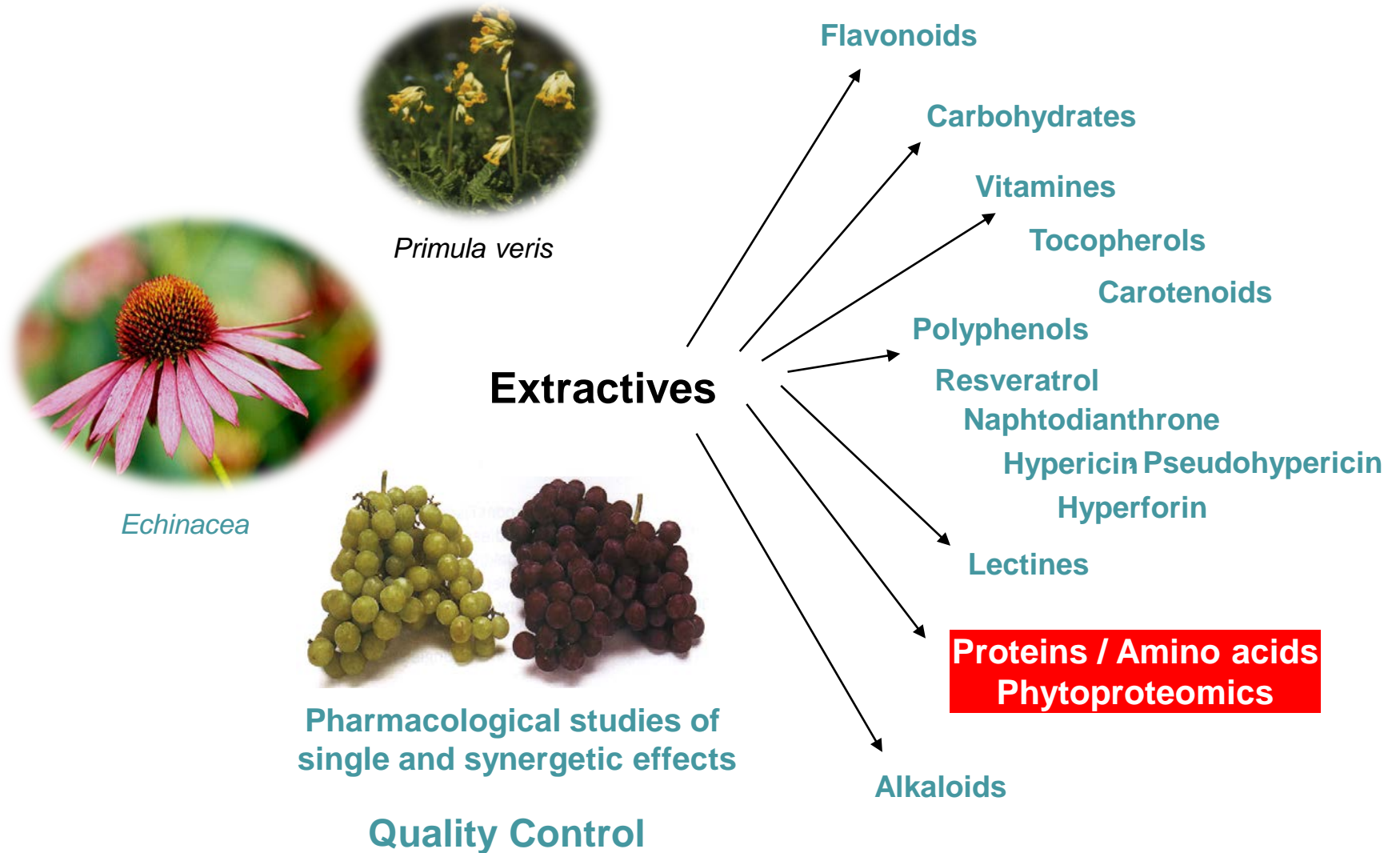
Hyphenated techniques:

μ -HPLC-ESI-MS
 μ -HPLC-MALDI-TOF MS

Infrared Spectroscopy:

non-invasive quality control

Natural Products - Extraction and Purification - Strategies



Extraction Scheme

Plant

Plant material
e.g. Scutellaria,
St. John's Wort etc.

Extraction

Extraction principle

- Hydrothermolysis (flow through, p, T)
- Acidic hydrolysis
- Microwave
- Accelerated Solvent Extraction (ASE, Dionex)
- Mazeration

Analytical Tests

Microbiological and pharmacological tests
z.B. antimicrobial, anti-inflammatory (COX, LOX) properties

Extracts

Analysis

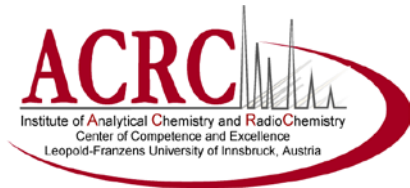
- Solid-phase extraction (SPE)
- Separation (HPLC, μ -LC, LC-MS, CE, CE-MS, CEC)
- Vibrational spectroscopy (MIR, NIR, ATR, Raman)
- Antioxidative potential (DPPH, FRAP)

Process analysis/screening
(e.g. IR, LC, etc.)

AIM

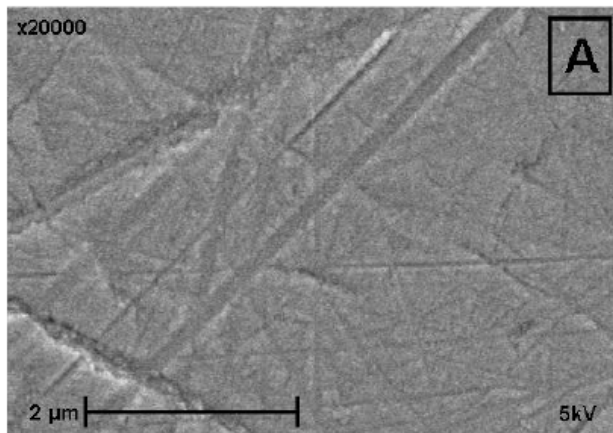
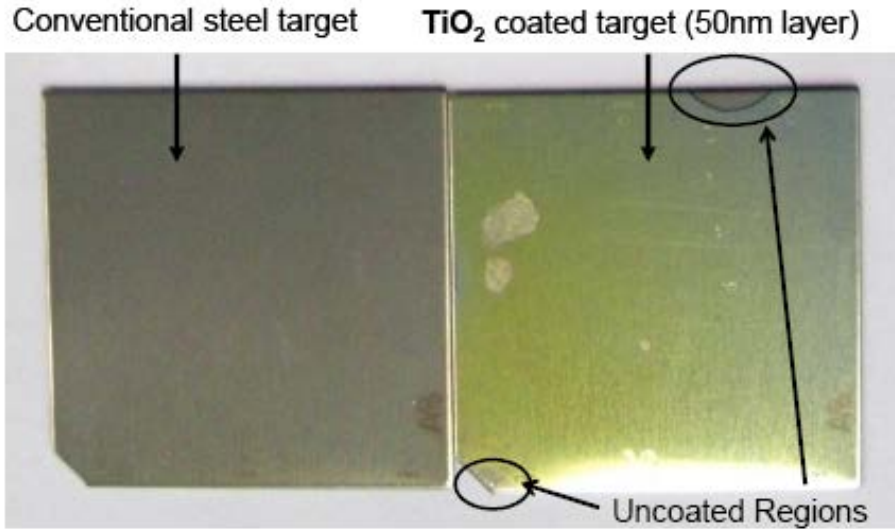
- Increased extraction yield
- Increased selectivity and specificity

Cooperation with Bionorica®



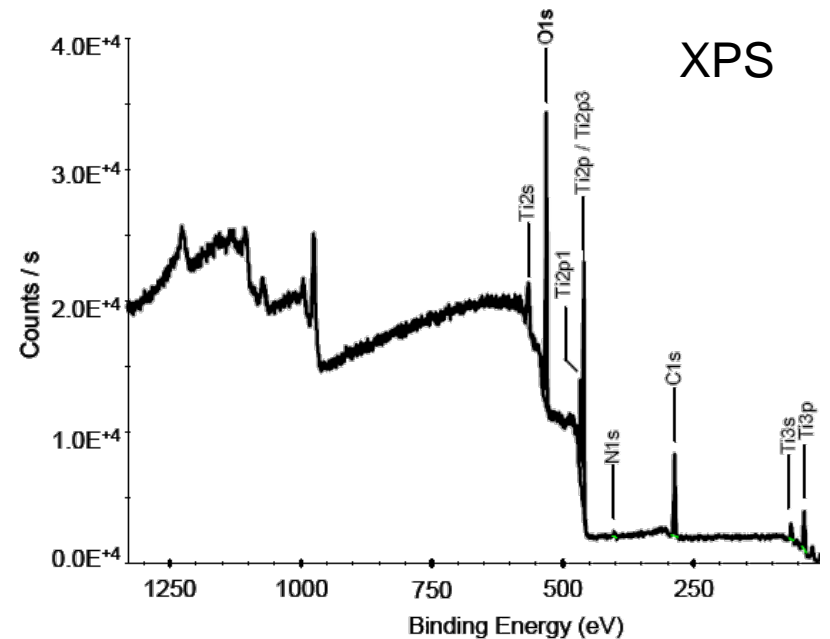
In the field of phytoanalytics
and antimicrobial activities of plants

TiO₂ coated Targets for matrix-free LDI MS



SEM picture

50 nm TiO₂ coating



TiO₂ coated Targets for matrix-free LDI MS

Analysis of Artichoke Samples

Cynara scolymus

Apigenin (A),

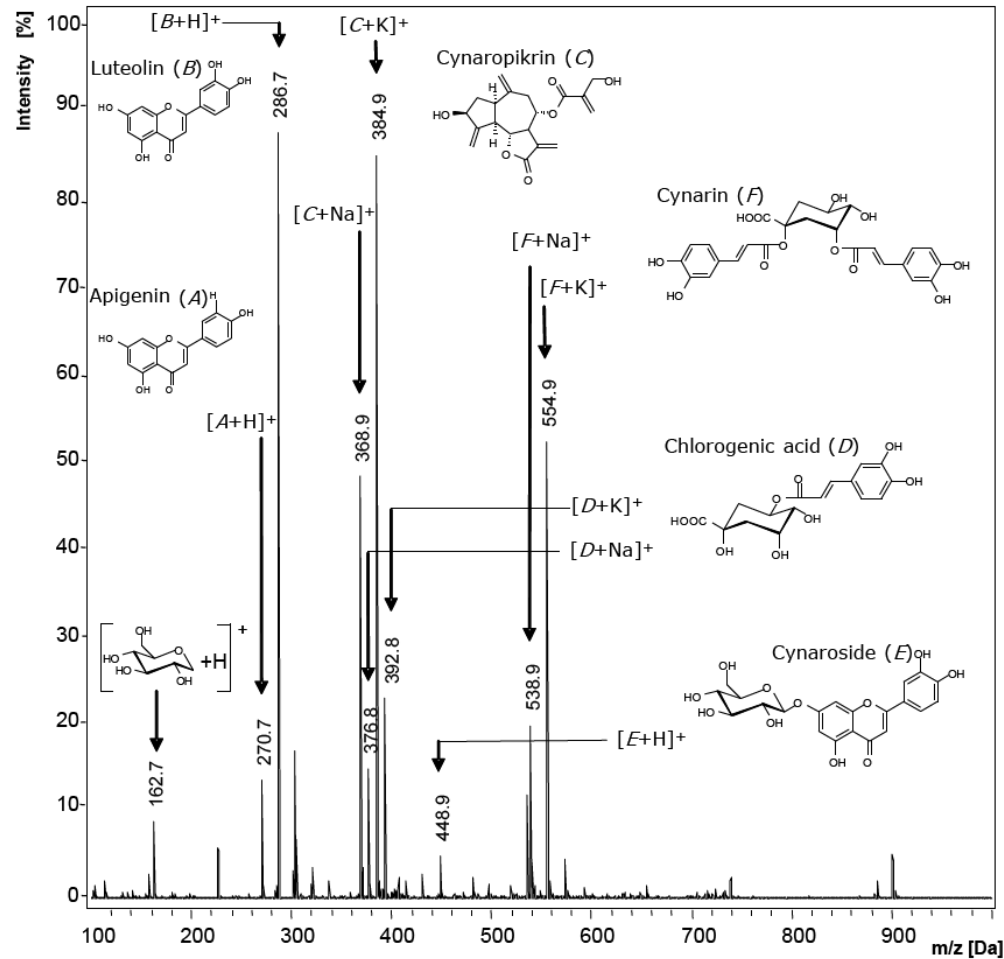
Luteolin (B),

Cynaropikrin (C),

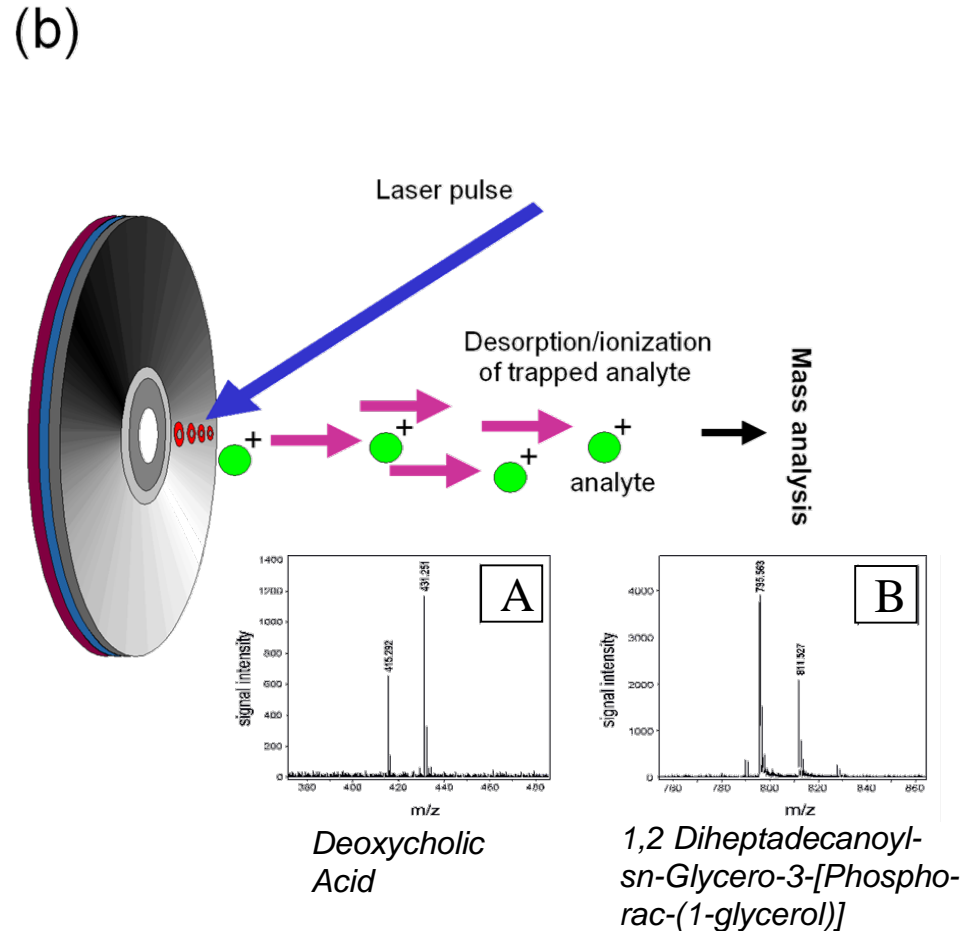
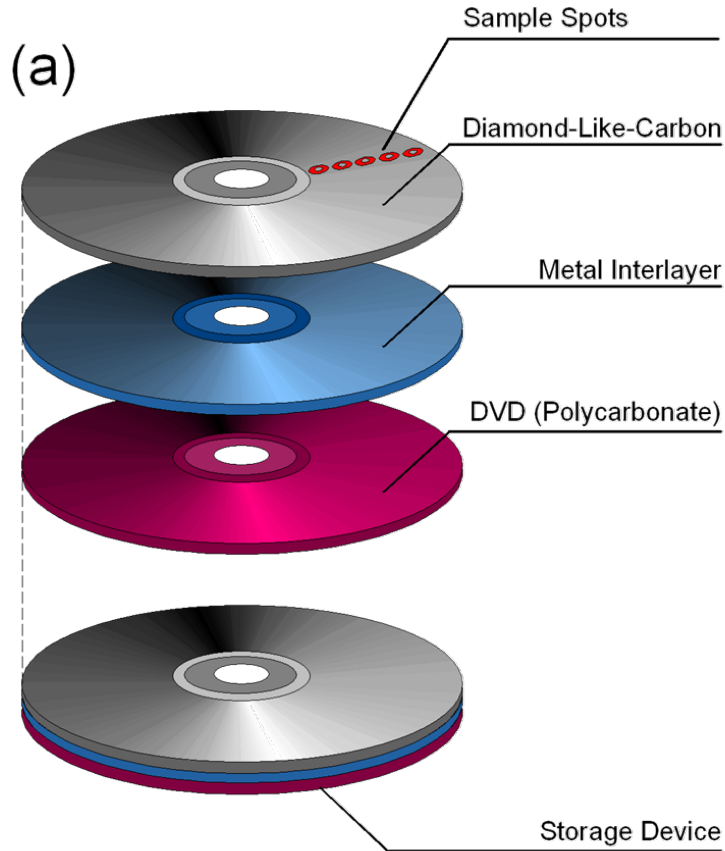
Chlorogenic acid (D),

Cynaroside (E) and

Cynarin (F),



Matrix-free LDI MS for the Detection of Low Molecular Weight Compounds



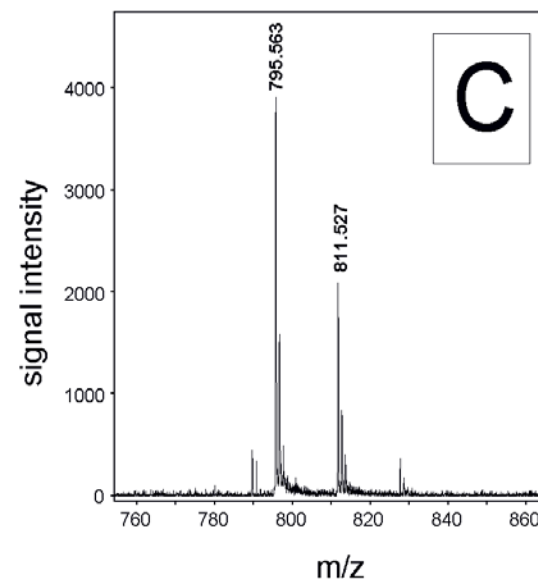
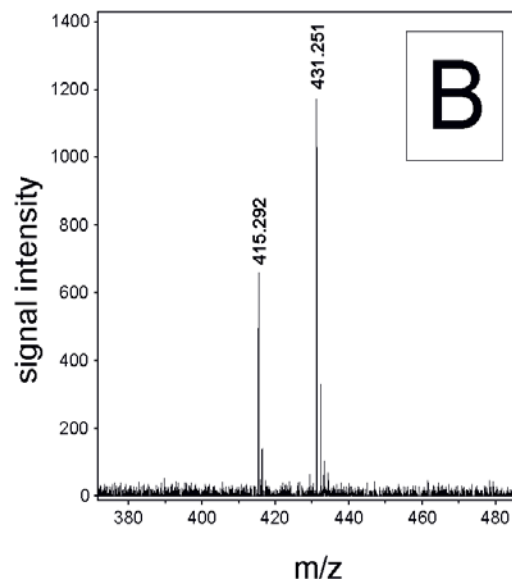
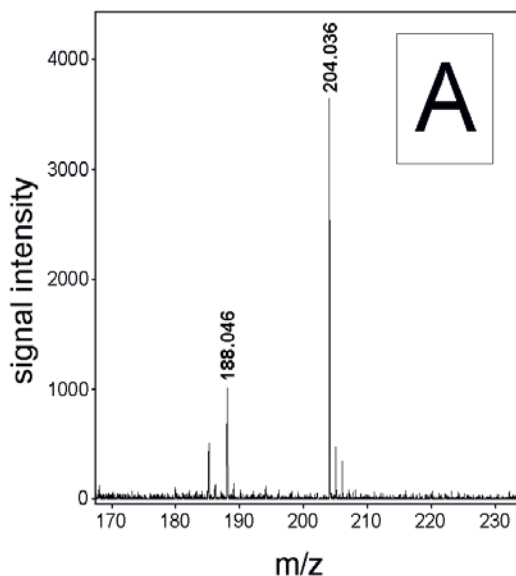
Collaboration with SONY

Laser Desorption/Ionization Mass Spectrometry on Diamond for the Application in Metabolomics

A.) Amino Acids
e.g. *L*-Phenylalanine

B.) Blood Metabolites
e.g. *Deoxycholic Acid*

C.) Lipids
e.g. *1,2 Diheptadecanoyl-sn-Glycero-3-[Phospho-rac-(1-glycerol)]*



Na⁺ and K⁺ adducts.

Invited Interview in Analytical Chemistry 2008

10.1021/AC801668W © 2008 AMERICAN CHEMICAL SOCIETY
Published on Web 08/29/2008

OCTOBER 1, 2008 / ANALYTICAL CHEMISTRY 7183

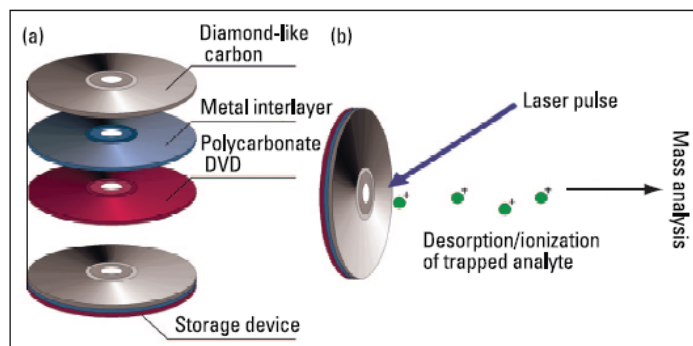
RESEARCH PROFILE

Femtomolar sensitivity with matrix-free LDI MS

Scientists who want to analyze small molecules with MALDI MS have long been frustrated by unwelcome signals from the matrix. And until now, matrix-free LDI has not reached the femtomolar sensitivity required to, for example, perform clinical blood tests for metabolites. But Günther Bonn and colleagues at Leopold-Franzens University (Austria) report in a new *AC* paper (DOI 10.1021/ac801190e) that they have boosted sensitivity into the femtomolar range by replacing their MALDI target with a DVD coated with diamond-like carbon (DLC). “That it worked was really a surprise,” says Bonn, who hopes to use the technology to create metabolomic tests for prostate cancer.

The porous silicon most commonly used as a target for matrix-free LDI is easily contaminated and can't be stored for

layer, says Matthias Rainer, another group member. “Their electrical, thermal, and optical properties improve the efficiency of laser transfer, leading to high reproducibility.”



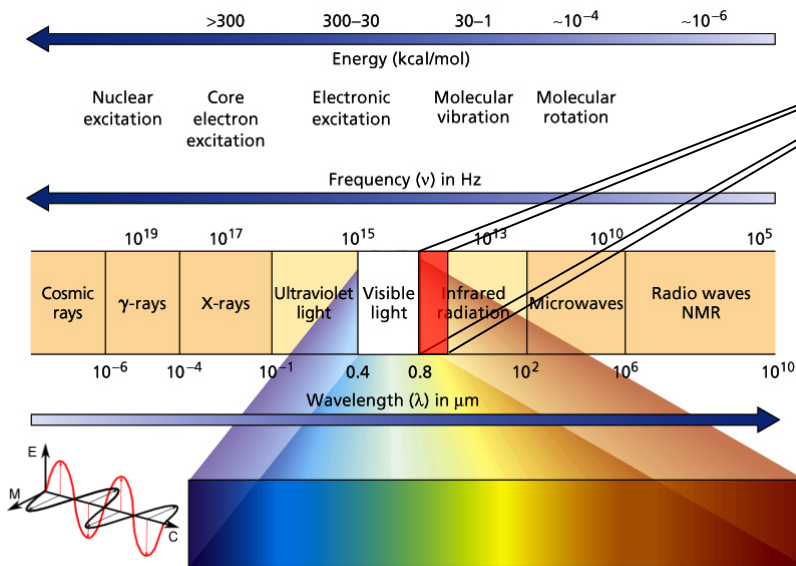
(a) The matrix-free target is a polycarbonate DVD coated with an interlayer of molybdenum and topped with a layer of DLC. Even after the DVD is coated, it can store data. (b) A laser pulse desorbs and ionizes the sample from the carbon layer for analysis by MS.

Bonn wanted to use DVDs so he could burn analytical data onto their undersides. This technique is particularly useful, he points out, when hundreds of

The researchers also detected peptides that fall into the same mass range as vaporized matrix would and successfully measured the m/z of peptides in a BSA digest, an amino acid, a phospholipid, and an emulsifier found in the small intestine. The masses of these test compounds ranged from 100 to 1500 Da. “The different analytes reported here show the broader application of DLC,” Rainer says. “This is important, as many matrix-free systems are reported to be viable for a fixed kind of analyte.”

The group is trying to increase sensitivity even further by decreasing the thickness of the DLC layer and chemically modifying its surface. For example, they are applying aromatic compounds, such as sinapinic acid, to boost UV sensitivity and thereby expand the mass range. Before the technology can be commercialized, they must also

Basics of Near-infrared Spectroscopy (NIRS)



- **NIR wavelength range: 780 – 2500 nm (4000 – 12800 cm^{-1})**

- interaction between sample and electromagnetic radiation

- Pre – condition: change of dipole moment

- overtone and combination bands appear

- use of chemometrical and statistical tools

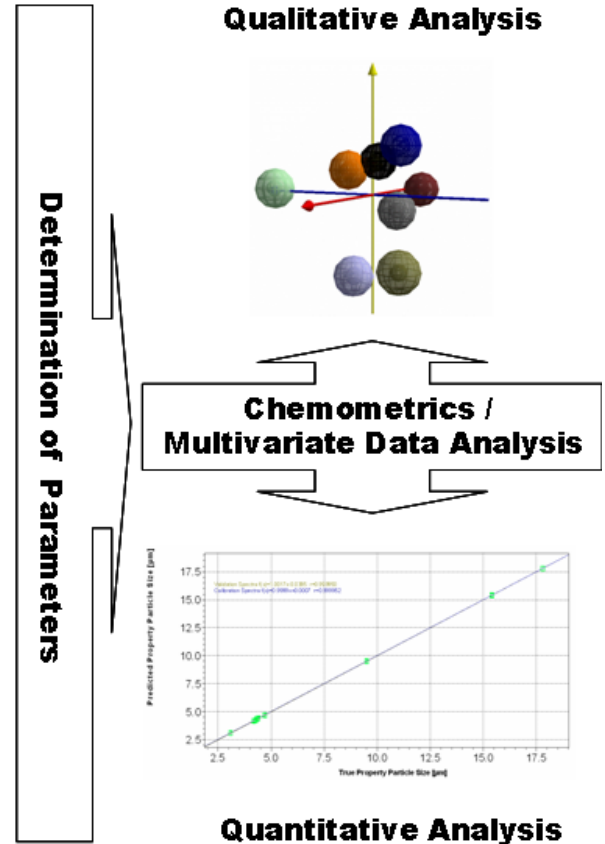
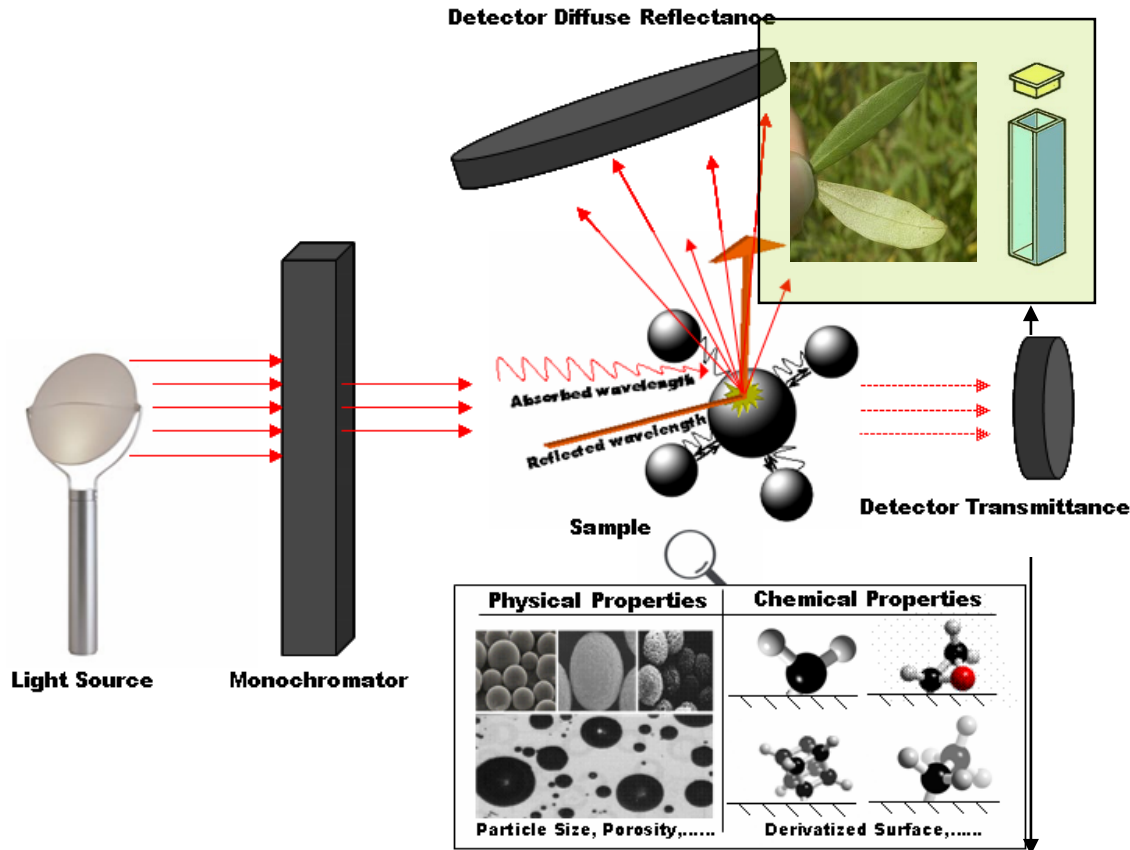
➔ Absorption of NIR radiation by organic molecules → mainly O-H, C-H, N-H and C=O groups whose fundamental molecular stretching and bending absorb in the mid – IR region (400 – 4000 cm^{-1})

➔ These overtone and combination bands are anharmonic → very complex and not direct interpretable as in other spectral regions → need for chemometric evaluation

➔ Chemical (e.g. functional groups) as well as physical parameters (particle size, pore size, specific surface area) can be detected

Basics of Near-infrared Spectroscopy (NIRS)

- NIRS Setup -



Lambert Beer's law
$$A = \epsilon * c * d = -\log \frac{I}{I_0}$$

MIR + NIR

Phytomics

Chemical
paramters

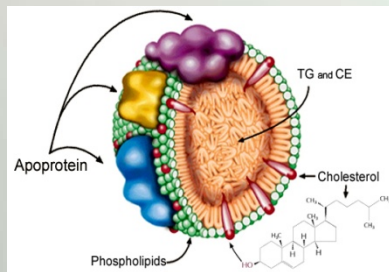
Physical
parameters



Bioanalysis

Tissue
analysis

Blood
ingredients



Pharmacy

Chemical
parameters

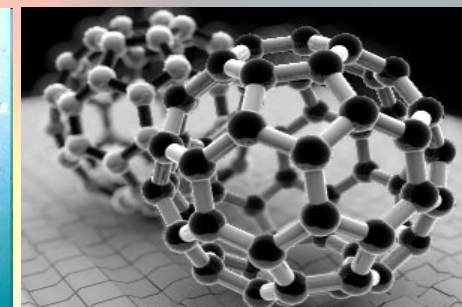
Physical
parameters



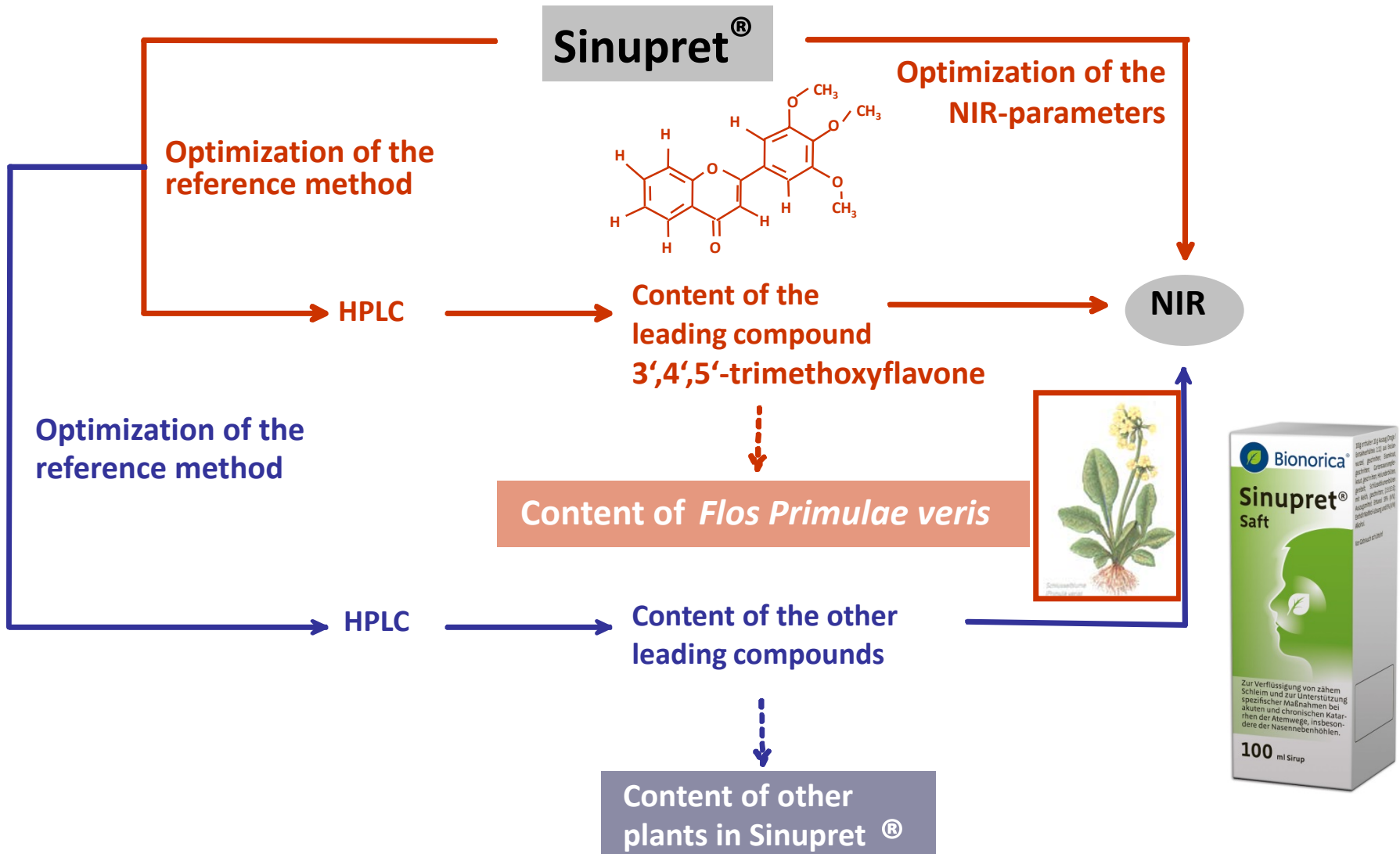
**Material
Science**

Morphological
properties

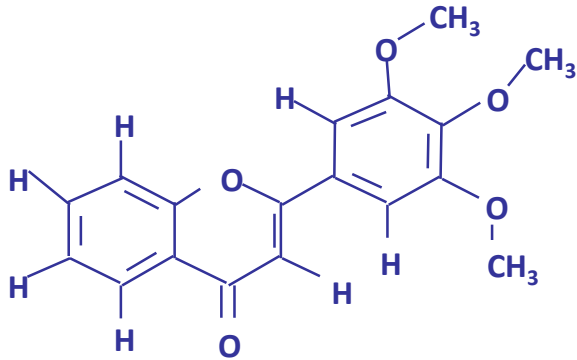
Physical
properties



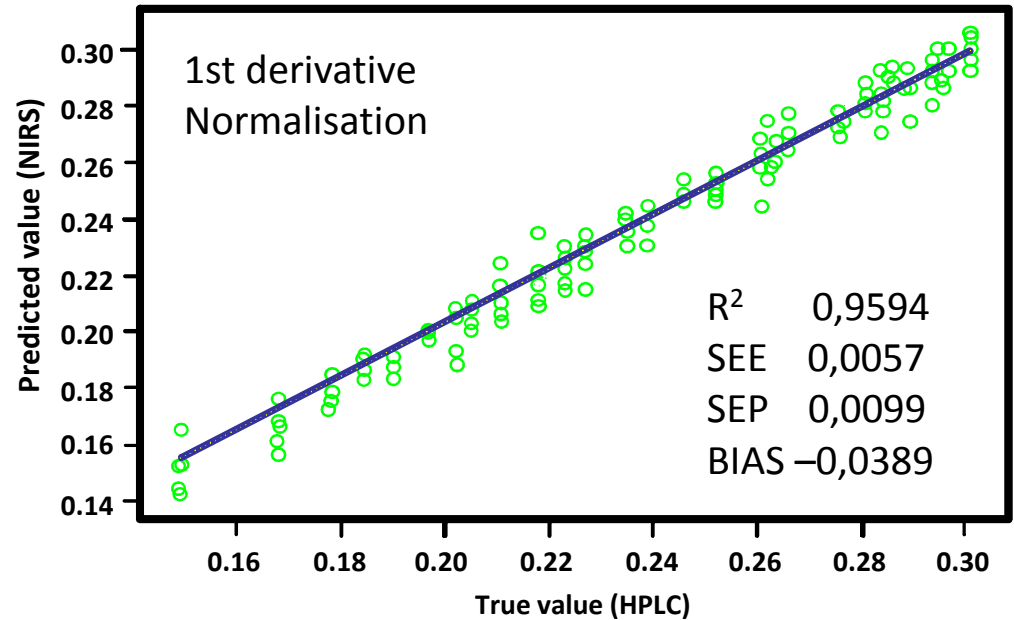
NIR Strategy of Analysis



NIR Strategy of Analysis

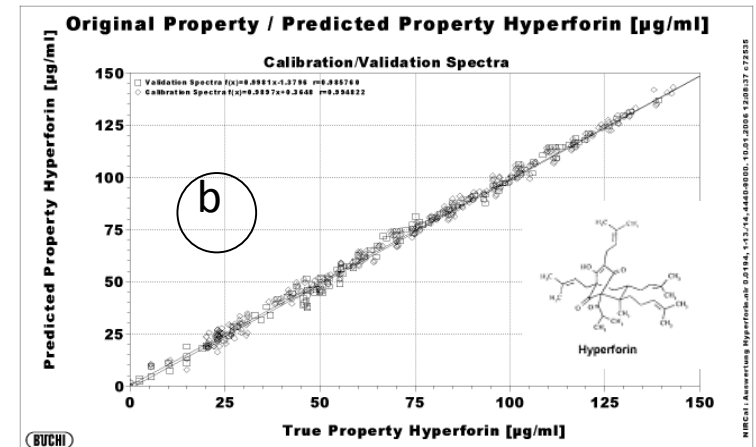
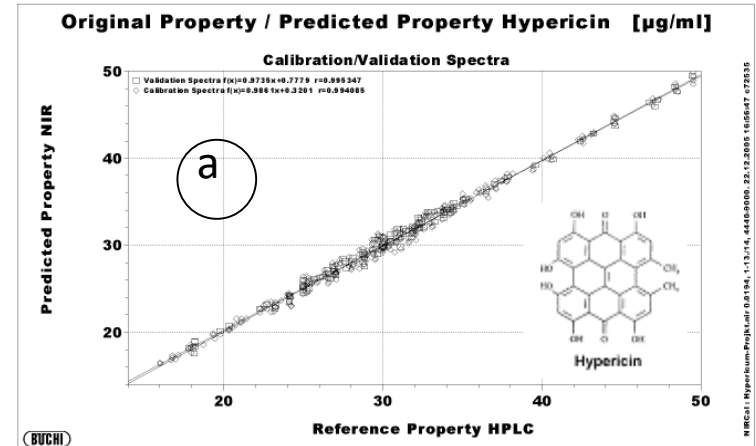
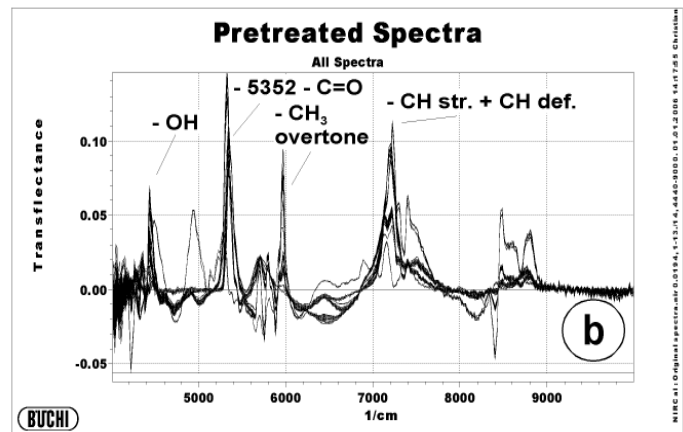
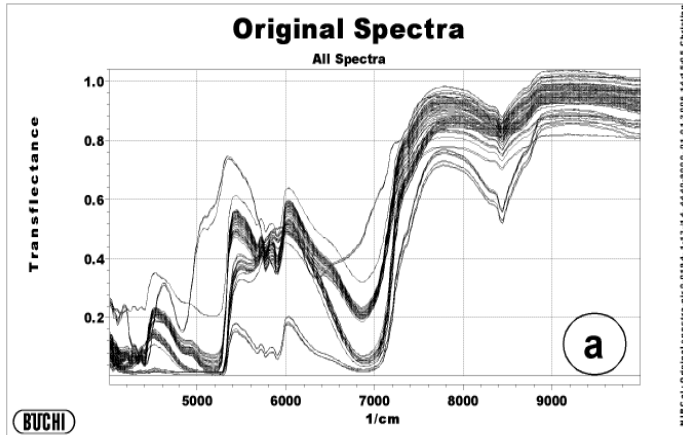


**3',4',5'- Trimethoxyflavone
Leading Compound**



Charge	Area ($\mu\text{V}\cdot\text{sec}$)	Primula (g/100g)	Conc. LC (ng/ μl)	NIR-MW (ng/ μl)	Water (%)	EtOH (%)
91102064 28.10.91	70490	0,493	0,219	0,219	80,38	15,7
91112182 05.11.91	71740	0,500	0,222	0,210	80,83	15,2
91112191 08.11.91	61346	0,442	0,197	0,230	80,25	15,7
91112302 22.11.91	66799	0,472	0,210	0,186	81,33	14,5
91112311 28.11.91	81551	0,555	0,245	0,193	79,69	15,7

NIRS of Naphthodianthrones and Phloroglucines in St. John's Wort



NIR spectra

(a) Original spectra, (b) pretreated spectra

Predicted (NIRS) vs. True values (LC)

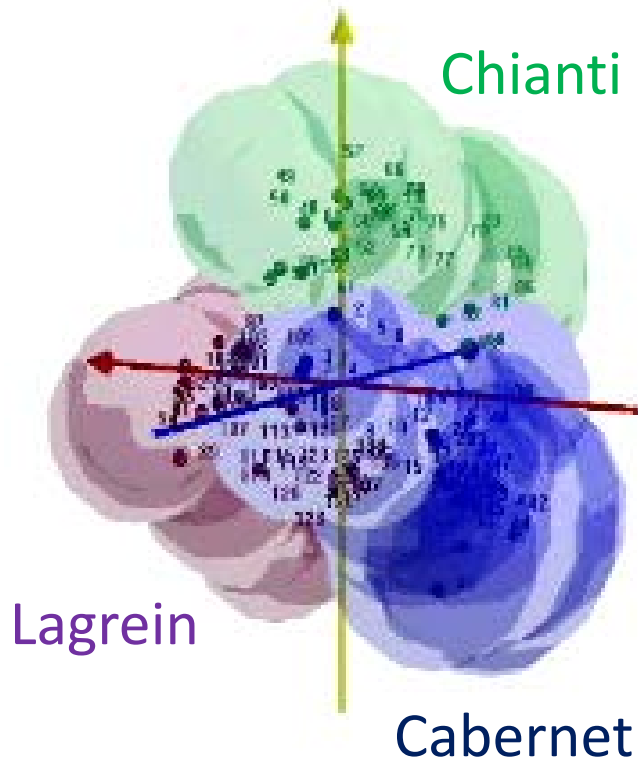
(a) Hypericin, (b) Hyperforin

(n=80). (a) $R^2=0.99$; $SEP=0.68$; (b) $R^2=0.99$; $SEP=0.72$

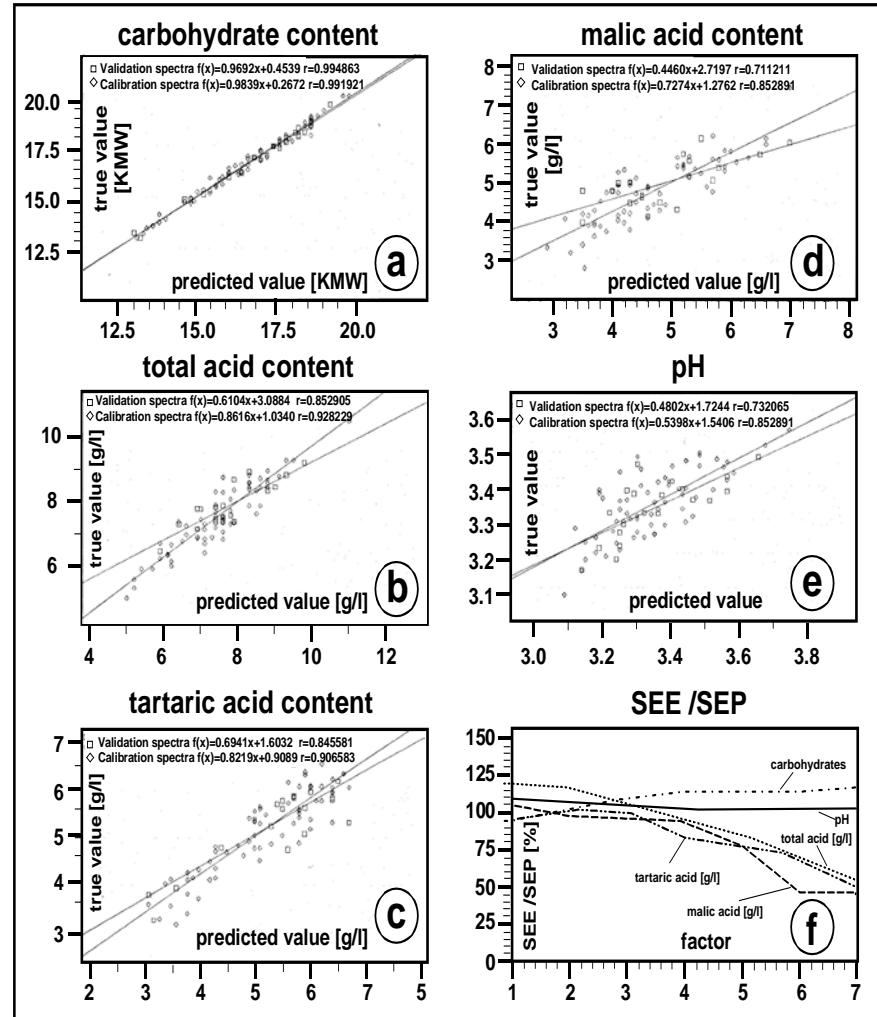
Wine Analysis using NIRS



Wine Analysis using NIRS



Factor plot of 141 spectra of different wines (Lagrein, Chianti, Cabernet Sauvignon). Conditions: Normalisation, 1st derivative; wavenumber range, 4500 - 10000 cm^{-1} ; thickness, 3 mm; scans, 10; temperature, 23°C.

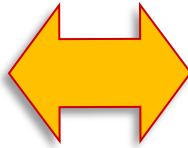
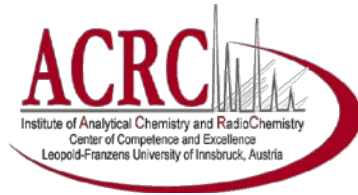


Coffee Analysis using NIRS

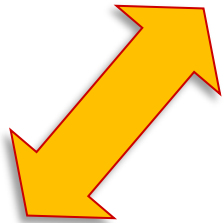
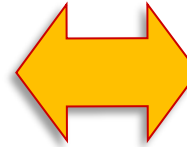


International Collaborations

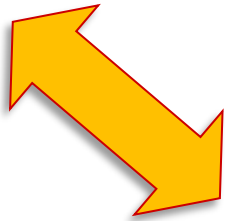
Prof. Günther Bonn



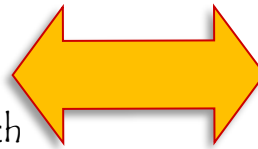
Prof. Lukas Huber



WHO – Lyon, France



Sino-Austrian
Center for Biomarker Research



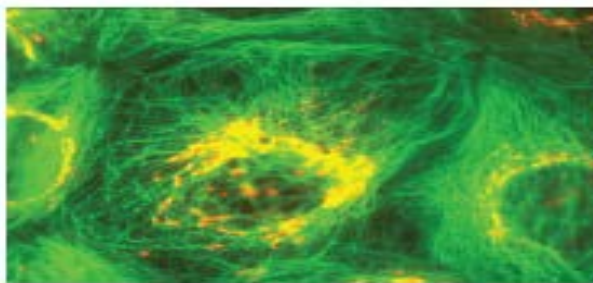
Prof. Ke Yang



מכון ויצמן למדע
WEIZMANN INSTITUTE OF SCIENCE

Prof. David Mirelmann





Austrian Drug Screening Institute (ADSI)

Prof. Lukas A. Huber

Scientific Director – Biological Division

Cell Biology, Biocenter

Innsbruck Medical University

Prof. Günther K. Bonn

Scientific Director – Analytical Division

Institute of Analytical Chemistry and Radiochemistry

University of Innsbruck

BM.W.F^a



Tyrol as a region of cancer and phyto research: stepwise development

Projects of Bionorica SE and Bionorica Research

at the Universities and Medical Universities
in Innsbruck, Graz and Vienna



Austrian TCM Project with
The Chinese Academy of Traditional
Chinese Medicine



2008/9

2006/7

2005

2004

2003

Special Research
Program SFB021:
Cell Proliferation and
Cell Death in Tumors

Doctoral Program
in Oncoscience

Austrian Proteomics
Platform (APP)

Formation of
Biocenter Innsbruck

Oncology in the
scientific focus at the I-
MED

FP6-Project
Growthstop

Proposal for COMET
Center ONCOTYROL

First „Clinical Trial Center“
in Austria established at
the I-MED


GEN-AU II

Launch of the
„ONCOTYROL –
Center for
Personalized Cancer
Medicine“

4x START grants at
the I-MED Biocenter

Drug Screening
Platform Concept

GEN-AU III

SFB021
4x START grants
Gen-Au:






basic research | target identification

target validation | hit generation | lead development

lead optimization | pre-clinical development | clinical development

validated targets

validated hits

lead candidates

Projects Bionorica
Austrian TCM Projects



Bionorica SE  Bionorica®
Bionorica Research, Austria

academic research

industry-academia partnership

industrial development

Academic Drug Screening Centres - USA

UNIVERSITY	CHEMICAL LIBRARY SIZE	DIRECTOR	WEBSITE
Broad Institute of MIT and Harvard	500,000	Michael Foley	www.broad.mit.edu/node/139
Columbia University	118,000	James Rothman	www.columbia.edu
Emory University	200,000	Ray Dingleline	www.pharm.emory.edu
Harvard Medical School	250,000	Caroline Shamu	iccb.med.harvard.edu
Johns Hopkins University	180,000	Min Li	www.hopkinschemcore.org
Rockefeller University	46,000	J. Fraser Glickman	www.rockefeller.edu
Stanford University	130,000	David Solow-Cordero	htbc.stanford.edu
University of California	55,000	Scott Lokey	chemistry.ucsc.edu
University of Cincinnati	250,000	William L. Seibel	www.gri.uc.edu
University of Illinois	200,000	Carson Putt	www.scs.uiuc.edu/htsf
University of Kansas	110,000	Rathnam Chagaturu	www.hts.ku.edu
University of Michigan	56,000	Martha Larsen	www.umich.edu
University of Minnesota	5,000	Marc von Keitz	www.btl.umn.edu/htba
University of New Mexico	231,000	Larry Sklar	nmmlsc.health.unm.edu
University of Pennsylvania	218,000	Scott Diamond	www.seas.upenn.edu/~pcmd/hts
University of Pittsburgh	280,000	John Lazo	www.upddi.pitt.edu
University of Rochester	23,000	Alan V. Smrcka	www.urmc.rochester.edu/hts
University of Texas Southwestern Medical Center	200,000	Michael Roth	www.utsouthwestern.edu
University of Wisconsin Madison	105,000	F. Michael Hoffmann	www.hts.wisc.edu
Vanderbilt University	260,000	Charles David Weaver	www.vanderbilt.edu
Washington University	140,000	Jayne Marasa	mic.wustl.edu/Cores/HighThroughputCore
Yale University	30,000	Paul Fletcher	cgp.yale.edu/chemical/chem_info

Academic Drug Screening Centers - Europe

	Library size
Medical Research Council Technology (Mill Hill, UK)	51-100.000
Imperial College Drug Discovery Centre (London, UK)	11- 50.000
Scottish Hit Discovery Facility (Dundee, UK)	51-100.000
Max Planck Institutes Chemical Genomics Centre (Dortmund, Germany)	11- 50.000
EMBL/DKFZ Chemical Biology Core Facility (Heidelberg, Germany)	51-100.000
USEF (Santiago de Compostela, Spain)	<10.000
HTS, Spanish National Cancer Research Centre (Madrid, Spain)	51-100.000
LiMoNe, K.U.Leuven (Leuven, Belgium)	11-50.000
IBITEC-S, CEA Saclay (Paris, France)	11-50.000
PCBIS, Universite Louis pasteur (Strasbourg, France)	11-50.000
CMBA (Grenoble, France)	11-50.000
Biomolecular Screening Facility, EPFL (Lausanne, Switzerland)	11-50.000
European Screening Port (Hamburg, Germany)	151 - 300.000

High-Content Screening prevails at European Academic Screening Centres

Austrian Drug Screening Institute, Tirol

Comparison of early drug discovery approaches at industry and academia

	high throughput screening (industry)	high content screening (academia)
throughput	high to ultra-high	low to medium
content	low	high
format	1536- and 3456-well plates	96- and 384-well plates
assay	simple biochemical assays (cell-free)	complex biological assays (cell-cell interactions)
detection	absorbance, fluorescence, luminiscence	advanced biooptics, mass spectrometry
capacity	> 100.000 compounds/day	< 10.000 compounds/day
screen size	up to 2 millions compounds	< 100.000 compounds
compounds	own synthetic libraries	external libraries (synthetic and natural)
focus	selection of active compounds	understanding of the mode of action
purpose	generation of new drugs for profit	generation of new methods and research tools
IPR strategy	protection of active compounds	protection of methods and research tools
IPR owner	company	project owner
equipment	> 10.000.000 €	< 5.000.000 €

ADSI management

scientific advisory board

Günther Bonn

ANALYTICAL DIVISION

- mass spectrometry
- HPLC
- LC-MS, MS/MS
- IR/NMR spectroscopy
- computational chemistry
- drug pull downs
- new materials for separation technology & enrichment
- advanced electrophoresis
- bioinformatics

Lukas Huber

BIOLOGICAL DIVISION

- biological tests
- immunological tests
- high content biooptics
- molecular screening
- structural biology
- cell –cell interaction assays
- drug signatures
 - Proteomics
 - Genomics

Management

Shareholder University of Innsbruck (100%)

CEO:

Dr. Laco Kacani

Scientific Directors:

Prof.Mag.DDr.h.c. Günther Bonn – Analytical Division

Prof.Dr. Lukas Huber – Biological Division

Scientific Board

Prof.Dr. Peter Boyle

Dr. Danilo Corradini

Montelibretti, Italy

DDr. Friedrich Lottspeich

Prof.Dr. David Mirelman

Mag. Markus Pasterk

Austria

Prof.Dr. Siqi Liu

IPRI , Lyon, France

Institute of Chemical Methodologies, Chromatography
and Capillary Electrophoresis Unit,

Max Planck Institute of Biochemistry,
Martinsried, Germany

Weizmann Institute of Science, Rehovot, Israel

Biobanking and Biomolecular Research Infrastructure, Graz,

Chinese Academy of Sciences, Beijing Institute of
Genomics, Beijing, China

Examples of Screening Projects

Screening in oncology – partnership with ONCOTYROL:

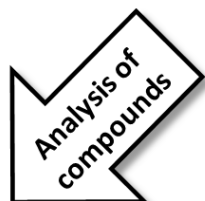
Treatment of prostate cancer, breast cancer, lymphoma & myeloma

Screening of natural compounds – partnership with phytopharmacy:

Treatment of inflammatory, immunological & infectious diseases

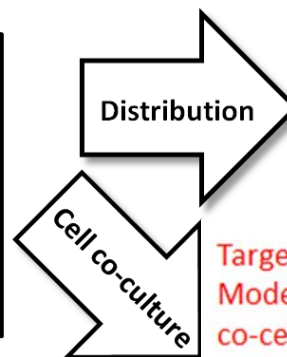
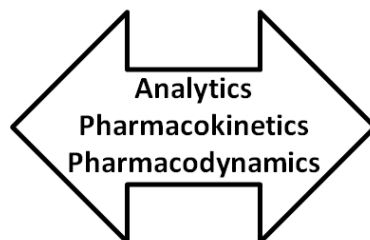
Screening for external partners – partnership with Austrian academy:

External projects – library screening, drug signatures, mode of action



Analytics

Oc1cc(O)c2c(c1)oc(O)c2O[C@@H]3O[C@H](O)[C@@H](O)[C@H]3O
 Isoquercetin



Cell co-culture

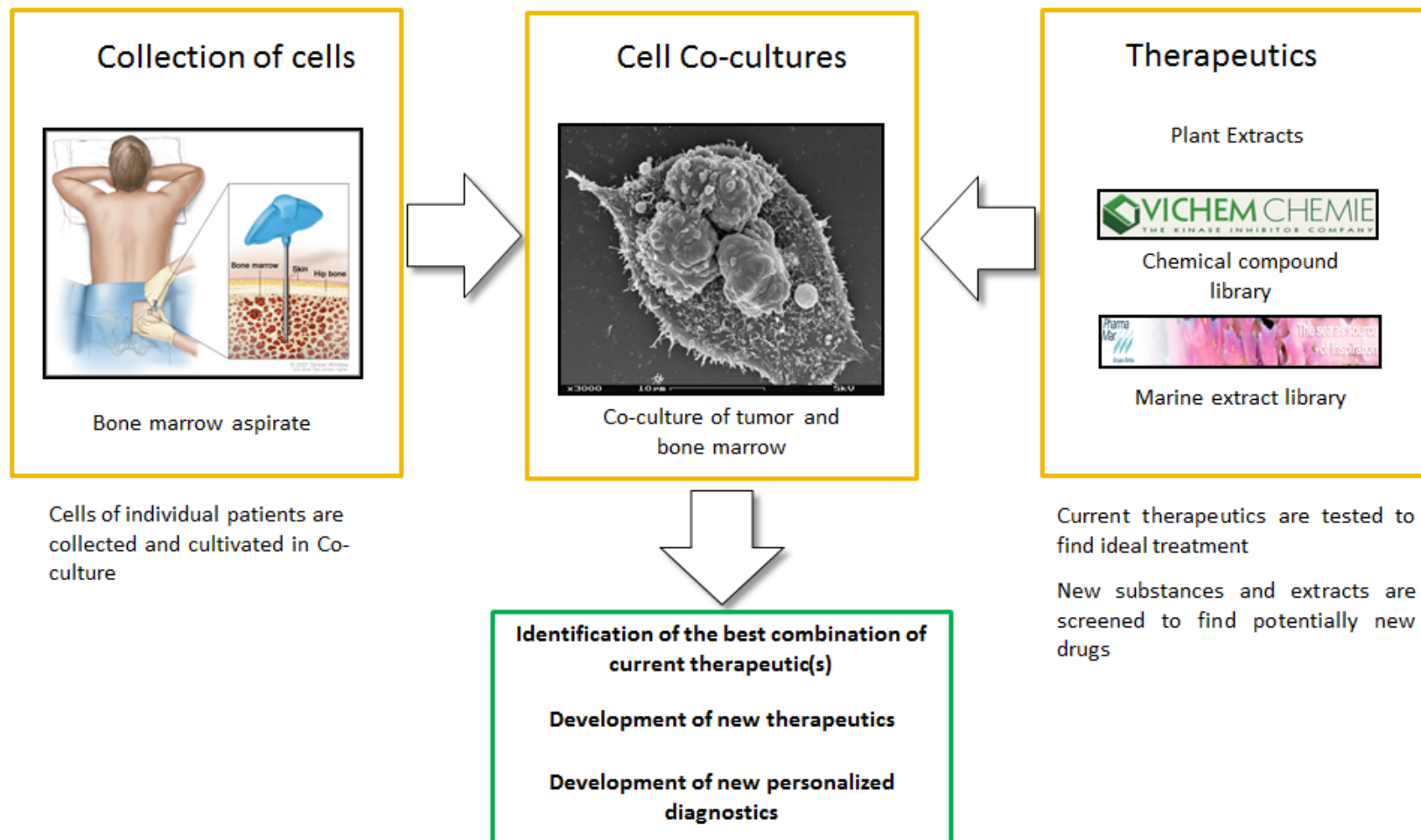
Projects:
 Metabolic Syndrome
 Inflammatory disease
 Liver protection



Target oriented cell-enzyme
 Models and specific cell- and
 co-cell culture models



Project Multiple Myeloma



Equipment

Extraction



ASE 350



Rotavapor



Moisture Analyzer



Aquasolv



CertoClav
Sterilization

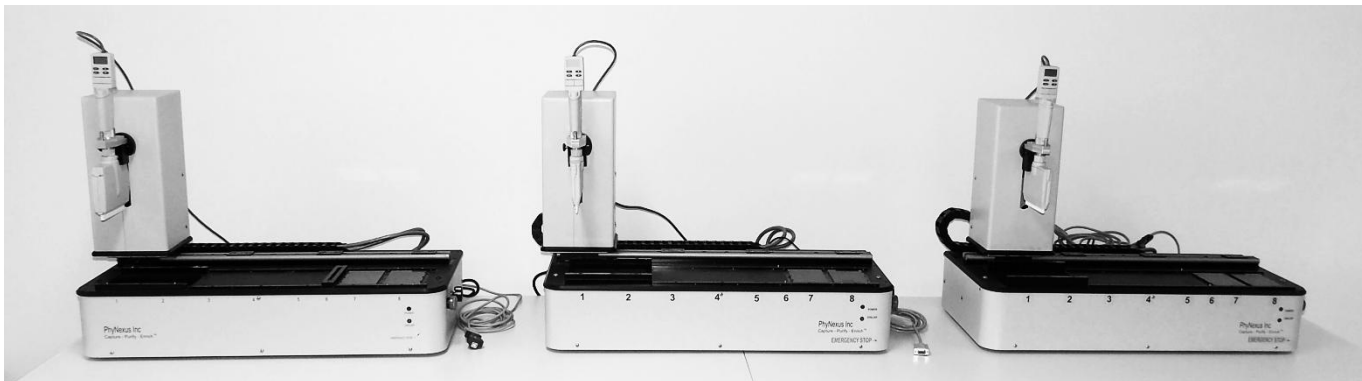
Sample Preparation



Proteiner
Fractionation, LC-MALDI coupling



ImagePrep
MALDI Imaging



PhyNexus MEA System
liquid handling robotic system

Mass Spectrometry



2x Maxis Impact

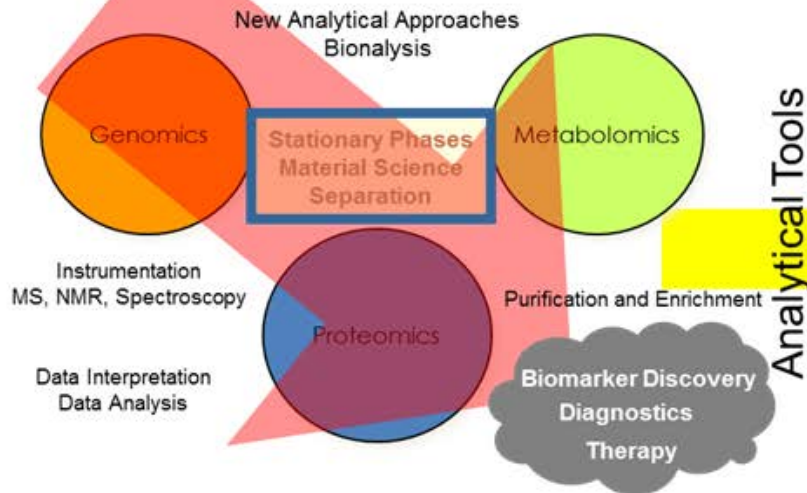


UPLC
2D, analytical and nano

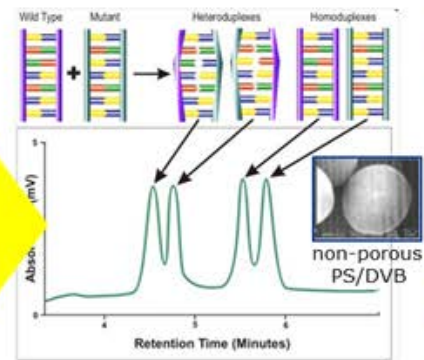


Autoflex Speed

Drug Screening



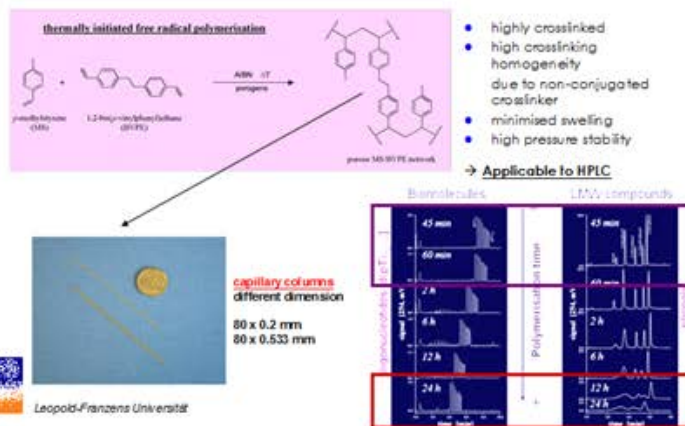
Transgenomic Wave-System - Mutation Analysis



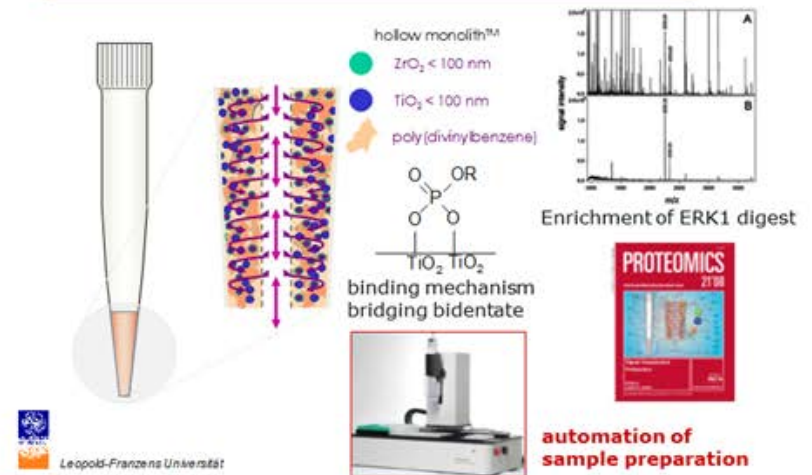
Transgenomic Wave HPLC-System

Bonn, Gunther; Huber, Christian; Oefner, Peter. Separation of nucleic acid fragments with alkylated nonporous polymer beads. US Patent (1994)

Stationary Phases for Separation Science



Preparation of hollow monolith™



Trojer L. et al. Monolithic poly(p-methylstyrene-co-1,2-bis(p-vinylphenyl) ethane) capillary columns as novel styrene stationary phases for biopolymer separation. *Journal of Chromatography, A* (2006), 1117(1), 56-66.

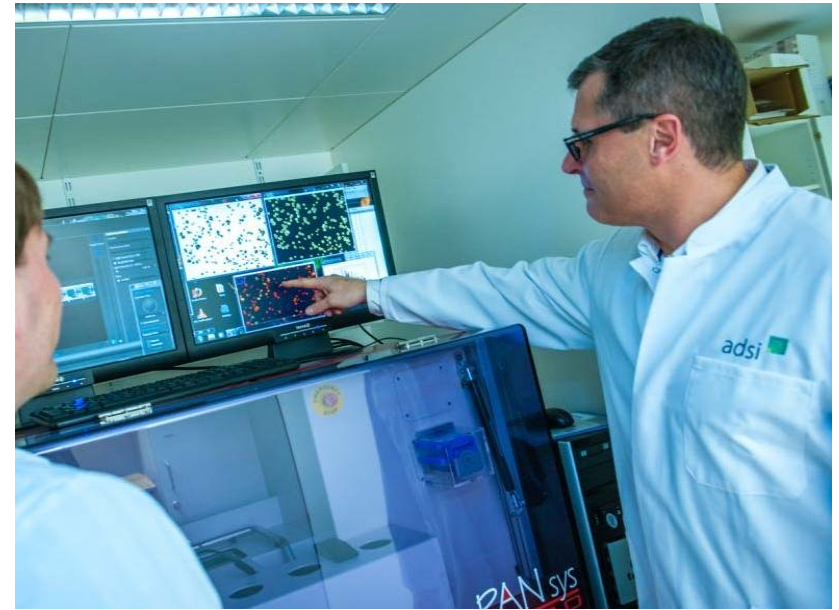
Bonn, Alexander; Bonn, Guenther; Huck, Christian; Maerk, Bernhard; Sonderegger, Harald; Rainer, Mathias; Gjerde, Douglas T. Pipette tip containing particle-filled polymer monolith. U.S. Pat. Appl. Publ. (2010)



Development of **co- culture testsystems**



**Guava easyCyte
8HT Milipore**
Cell sorter



PanSys 4000
Fully automated
cell culture system

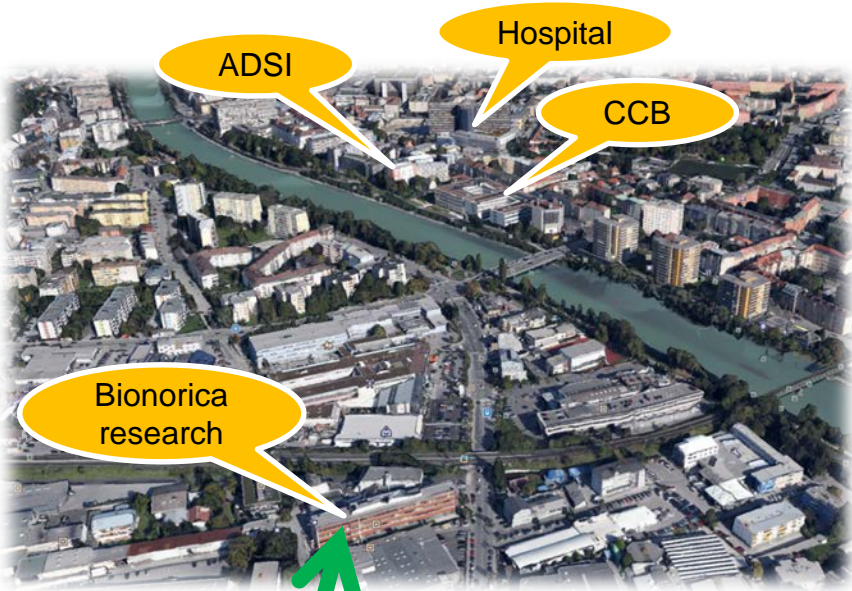
Prof. Dr. Surapote Wongyai

UNIDO Expert in Herbal Medicinal Products Development, Trieste, Italy

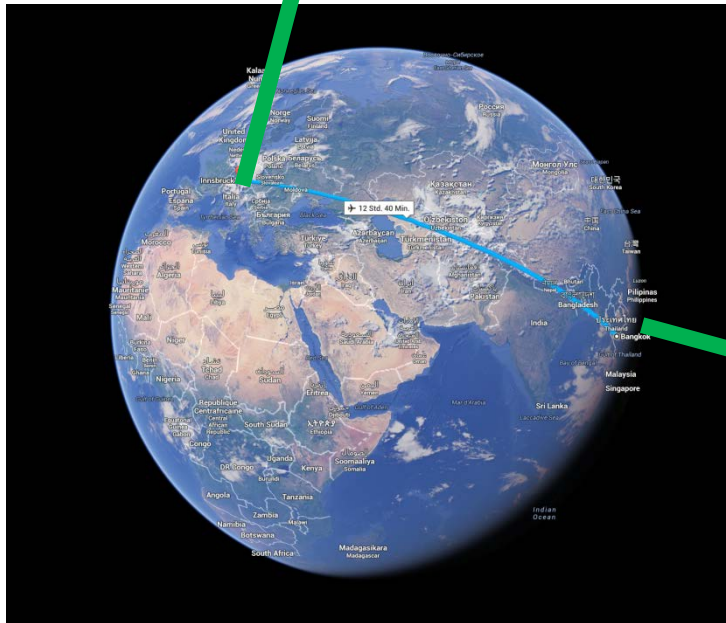
Thai FDA Expert in ASEAN Harmonization of Traditional Medicine and Health Supplement (TMHS)

Founder Dean and Dean, Faculty of Oriental Medicine, Thailand





Association of Southeast Asian Nations (ASEAN), 10 Countries



Rangsit University
 Faculty of Oriental Medicine
 Bangkok, Thailand
**Standardisation/Validation
 Traditional Medicine and
 Health Supplements
 Trainingcenter**
 Prof. Dr. Surapote

Trennung durch Fällung und Gravimetrie

Fällung

Ziel: Bildung einer neuen Phase

Parameter: Löslichkeit (KL, pH, Komplexbildung, Teilchengröße

Mitfällung, Adsorption

Filtration

Ziel: Phasentrennung

Anmerkung: gemeinsam mit Waschen

Waschen

Ziel: Entfernung der „Mutterlauge“

Parameter: Löslichkeit, Komplexbildung,
Peptisation

Trocknen, Glühen

Ziel: stöchiometrische Zusammensetzung

Anmerkung: Analysenfunktion

Messung: Wägen

Ziel: Bestimmung des Messwertes

Anmerkung: Wägefehler

Berechnung und Bewertung

Ziel: Analysenwert,

Vertrauensbereich

Parameter: stöchiometrische

Faktoren

Gravimetrische Grundoperationen

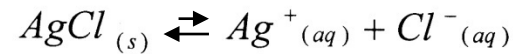
- Lösen der Analysensubstanz
- Fällern eines Niederschlages
- Abtrennen des Niederschlages von der flüssigen Phase durch Filtrieren
- Auswaschen des Niederschlages
- Trocknen und/oder Glühen bis zur Gewichtskonstanz
- Auswiegen der Wägeform des Niederschlages

Das Löslichkeitsprodukt

Kontakt von schwerlöslichen Verbindungen und H₂O

→ Gleichgewicht ($v_{\text{Auflösung}} = v_{\text{Wiederausscheidung}}$)

→ Gleichgewicht d. festen Stoffes und d. gesättigten Lösung des festen Stoffes

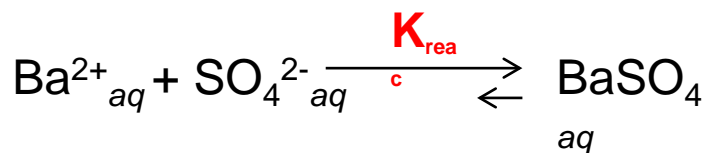


$$K = \frac{[\text{Ag}^{+}] \cdot [\text{Cl}^{-}]}{[\text{AgCl}]}$$

Berechnung von K_L : $K \cdot [\text{AgCl}] = [\text{Ag}^{+}] \cdot [\text{Cl}^{-}]$ [] ... c

$K_L = [\text{Ag}^{+}] \cdot [\text{Cl}^{-}]$ c ... Ionenkonzentration

K_L oder L_p



$\Delta G_f^\ominus \text{BaSO}_4_{aq}$	-722,874842 E_h
$\Delta G_f^\ominus \text{Ba}^{2+}_{aq}$	-25,166661 E_h
$\Delta G_f^\ominus \text{SO}_4^{2-}_{aq}$	-697,670227 E_h
$\Delta G_f^\ominus \text{BaSO}_4_{aq}$ reac	-0,037954 E_h
$\Delta G_f^\ominus \text{BaSO}_4_{aq}$ reac	-23,82 kcal/mol
K_{reac}	$2,90 \cdot 10^{17}$



$\Delta G_f^\ominus \text{AgCl}_{aq}$	-606,030719 E_h
$\Delta G_f^\ominus \text{Ag}^+_{aq}$	-146,329850 E_h
$\Delta G_f^\ominus \text{Cl}^-_{aq}$	-459,659021 E_h
$\Delta G_f^\ominus \text{AgCl}_{aq}$ reac	-0,041848 E_h
$\Delta G_f^\ominus \text{AgCl}_{aq}$ reac	-26,26 kcal/mol
K_{reac}	$1,79 \cdot 10^{19}$

Programm: Gaussian 09 C.01

Methode: Møller-Plesset Strörungstheorie (MP/2)

Basissatz: Karlsruhe double- ζ mit Polarisationsfunktionen
Small Core ECPs für Ag und Ba

Solvatisierung: Polarisierbares Kontinuumsmodell (PCM)

E_h : 627,5095 kcal/mol

R : 8,3145 J/mol K

T : 298,15 K

Fällungsreaktionen

Ionenprodukt:

- Ionenprodukt $< K_L$:
Lösung nicht gesättigt
→ weitere Substanz kann gelöst werden

- Ionenprodukt $= K_L$:
Lösung gesättigt
Lösung im Gleichgewicht mit ungelöster Substanz

- Ionenprodukt $> K_L$:
Lösung übersättigt
→ Fällung bis L erreicht ist

Löslichkeitsprodukte^a

		pK _L			pK _L	
Ag	AgBr	12,4	Cu	CuBr	7,4	
	AgCl	10,0		CuCl	6,0	
	AgCN ^b	11,4		CuI	11,3	
	Ag ₂ CO ₃	11,3		Cu(OH) ₂	19,8	
	Ag ₂ CrO ₄	11,7		Cu ₂ S	46,7	
	AgI	16		CuS	37–44	
	AgOH	7,7		CuSCN	10,8	
	AgSCN	12				
	Ag ₂ S	49				
Al	Al(OH) ₃	32,7	Fe	Fe(OH) ₂	13,5	
				Fe(OH) ₃	37,4	
				FeS	18–21	
Ba	BaCO ₃	8,16	Hg	Hg ₂ Cl ₂	17,5	
	BaC ₂ O ₄	6,77		HgS	52	
	BaCrO ₄	9,70	K	KClO ₄	2,05	
	BaSO ₄	10		K ₂ [PtCl ₆]	5,85	
Ca	CaCO ₃	7,92				
	CaC ₂ O ₄	8,07				
	CaF ₂	10,46				
	CaSO ₄	4,32				

^a aus [1]. Leider fehlt die Temperaturangabe; vermutlich beziehen sich die Werte auf 18–25 °C.

^b $2[\text{AgCN}]_f \rightleftharpoons \text{Ag}^+ + [\text{Ag}(\text{CN})_2]^-$

212 Anhang

Löslichkeitsprodukte (Fortsetzung)

Mg	MgCO ₃	3,7	Pb	Pb(OH) ₂	15,6
	MgC ₂ O ₄	4,1		PbSO ₄	8,0
	MgF ₂	8,2		PbS	28–29
	Mg(OH) ₂	10,9	Sn	Sn(OH) ₂	15,3 ^a
	MgNH ₄ PO ₄	12,6		Sn(OH) ₄	56
Na	NaHCO ₃	2,1	SnS	28	
			Zn	ZnCO ₃	10,2
Pb	PbCl ₂	4,8		Zn(OH) ₂	16,8
	PbCO ₃	13,5		ZnS	23–25
	PbCrO ₄	13,8			
	PbF ₂	7,5			

^a Originalangabe 25,3 [1], wahrscheinlich Druckfehler (vgl. [2]).

Löslichkeit L

- Löslichkeit nennt man die maximale Menge eines Stoffes, die ein Lösungsmittel bei einer bestimmten Temperatur aufnehmen kann.

- Angabe der Löslichkeit:

Mol/l (Molarität, molare Löslichkeit)

Md/ 1000g Lösemittel (Molalität)

Literatur: Analytische Chemie

Latscha, Klein

Springer Verlag

Tabelle 10. (aus dem Europäischen Arzneibuch EuAB)

Bezeichnung	Ungfähige Anzahl Volumenteile Lösemittel für 1 Massenteil Substanz			
sehr leicht löslich	weniger als	1 Teil	bis	10 Teile
leicht löslich	von	1 Teil	bis	30 Teile
löslich	über	10 Teile	bis	100 Teile
wenig löslich	über	30 Teile	bis	1 000 Teile
schwer löslich	über	100 Teile	bis	10 000 Teile
sehr schwer löslich	über	1 000 Teile	bis	10 000 Teile
praktisch unlöslich	mehr als	10 000 Teile		

Tabelle 11. Löslichkeit einiger Salze in Abhängigkeit von der Temperatur in g/100 g Lösung

Verbindung	0°C	20°C	30°C	40°C	100°C
NaCl	26,28	26,39	26,51	26,68	28,15
Na ₂ SO ₄	4,5	16,1	28,8	32,5	29,9
Na ₂ CO ₃	6,6	17,8	29,0	33,2	31,1
MgSO ₄	20,5	26,2	29,0	31,3	40,6
KNO ₃	11,6	24,1	31,5	46,2	71,1
AgNO ₃	53,5	68,3	73,8	77,0	90,1
AgCl		$1,5 \cdot 10^{-4}$			$2,2 \cdot 10^{-3}$
AgBr		$1,3 \cdot 10^{-4}$			$3,7 \cdot 10^{-4}$
Ca(OH) ₂		$1,2 \cdot 10^{-1}$			$6,0 \cdot 10^{-2}$
Mg(OH) ₂		$8,5 \cdot 10^{-4}$			$4,0 \cdot 10^{-3}$
CuSO ₄		$2,0 \cdot 10^{-1}$			$6,5 \cdot 10^{-2}$
SrSO ₄		$1,2 \cdot 10^{-2}$			$1,8 \cdot 10^{-2}$
BaSO ₄		$2,4 \cdot 10^{-4}$			$3,9 \cdot 10^{-4}$
PbSO ₄		$4,4 \cdot 10^{-3}$			$6,0 \cdot 10^{-3}$

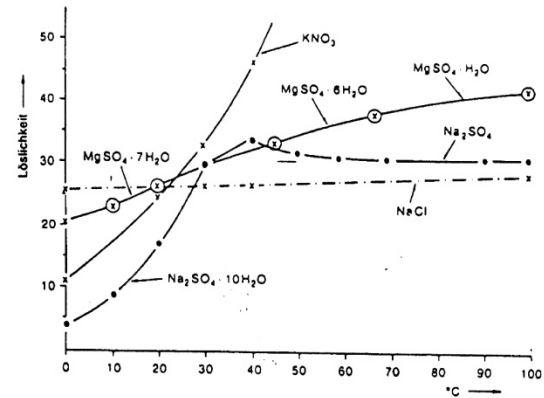


Abb. 21. Temperaturabhängigkeit der Löslichkeit einiger Salze. L = g/100 g Lösung

Anmerkung: Ca-Citrat ist ausnahmsweise in kaltem Wasser leicht löslich, aber in heißem schwer löslich

Tab. 1 Wichtige gravimetrische Verfahren [2, 5]

Ion	Fällungsform	Wägeform	Störungen
K^+	$K [B(C_6H_5)_4]^a$	$K [B(C_6H_5)_4]$	NH_4^+ , Rb^+ , Cs^+
Ag^+	$AgCl$	$AgCl$	Hg_2^{2+}
$M = Mg^{2+}, Zn^{2+}$	$MNH_4PO_4 \cdot 6 H_2O$ $M(oxinat)_2 \cdot 2 H_2O$	$M_2P_2O_7$ $M(oxinat)_2$	alle Metalle außer Na viele Metalle
Ca^{2+}	$CaC_2O_4 \cdot H_2O$	$CaCO_3 (CaO)$	alle Metalle außer Na Mg (Ca, Sr)
$M = Ba^{2+}, Pb^{2+}$	$MSO_4, MCrO_4$	$MSO_4, MCrO_4$	
Ni^{2+}	$Ni(diacetyldioximat)_2$	$Ni(diacetyldioximat)_2$	—
$M = Al^{3+}, Fe^{3+}$	$M(OH)_3 \cdot aq$	M_2O_3	Schwermetalle
Si (IV)	$SiO_2 \cdot aq$	SiO_2	Sn
Sn (IV)	$SnO_2 \cdot aq$	SnO_2	Si, Sb
F^-	$PbCIF$	$PbCIF$	SO_4^{2-}, PO_4^{3-}
Cl^-	$AgCl$	$AgCl$	Br^-, I^-, CN^-, SCN^-
SO_4^{2-}	$BaSO_4$	$BaSO_4$	$NO_3^-, ClO_3^-, PO_4^{3-}$
PO_4^{3-}	$MgNH_4PO_4 \cdot 6 H_2O$ $(NH_4)_3 [P(Mo_{12}O_{40})] \cdot aq^b$	$Mg_2P_2O_7$ $(NH_4)_3 [P(Mo_{12}O_{40})] (300^\circ C)(AsO_4^{3-})$	CO_3^{2-}

^a Die Kaliumbestimmung als $KClO_4$ ($K_L = 10^{-2}$) ist viel zu ungenau und sollte endlich aus den Lehrbüchern verschwinden.

^b Statt NH_3 wird auch Oxin als Base empfohlen [19].

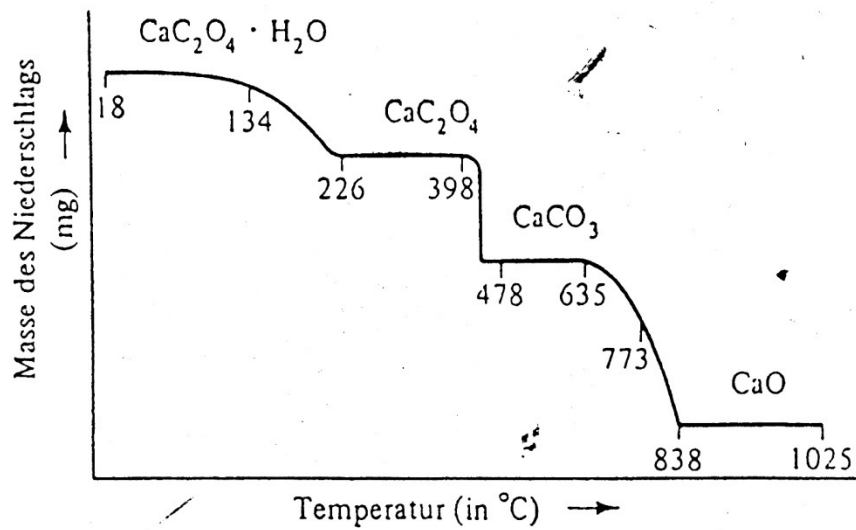
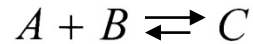


Bild 4-3 Thermogravimetrische Messung: Gewichtsverlust von $\text{CaC}_2\text{O}_4 \cdot \text{H}_2\text{O}$ beim Erhitzen.
 [Nach Cl. Duval, *Inorganic Thermogravimetric Analysis* (Elsevier, Amsterdam 1953) S. 36]

Gleichioniger Zusatz L_c



Anteil Dissoziationsprodukte gering

Löslichkeit richtet sich nach dem Ion mit der geringeren Konzentration

$$[A^+] > [B^-]$$

$$L_c = [B^-] = \frac{K_L}{[A^+]} = \frac{K_L}{C_{A^+}^0 + [A^+]} \sim \frac{K_L}{C_{A^+}^0}$$

$$K_L = [A^+] \cdot [B^-]$$

$$[B^-] = \frac{K_L}{[A^+]}$$

$C_{A^+}^0$... Zugesezte Menge A^+

$[A^+]'$... der durch Dissoziation entstandene Anteil

$$(C_{A^+}^0) \gg [A^+]'$$

$L_c < L$; Die Löslichkeit bei gleichionigem Zusatz nimmt ab

Beispiel: AgCl

$$C_{Ag^+}^0 = 10^{-2} \text{ mol/l}$$

$$L_c = \frac{K_L}{C_{Ag}^0} = \frac{10^{-10}}{10^{-2}} = 10^{-8} \text{ mol/l}$$

Fremdioner Zusatz L_a

Aktivitätskoeffizient = f (Ionenstärke)

⇒ Änderung der Löslichkeit eines Stoffes bei Fremdsalzzusatz

$$K_L = a_{A^+} \cdot a_{B^-} = [A^+] [B^-] \cdot f_{A^+} \cdot f_{B^-}$$

$$a = c \cdot f$$

a ... Aktivität

$$f = 1$$

c ... Konzentration

f ... Aktivitätskoeff.

Löslichkeit bezieht sich auf stöchiometrische Konzentrationen

-

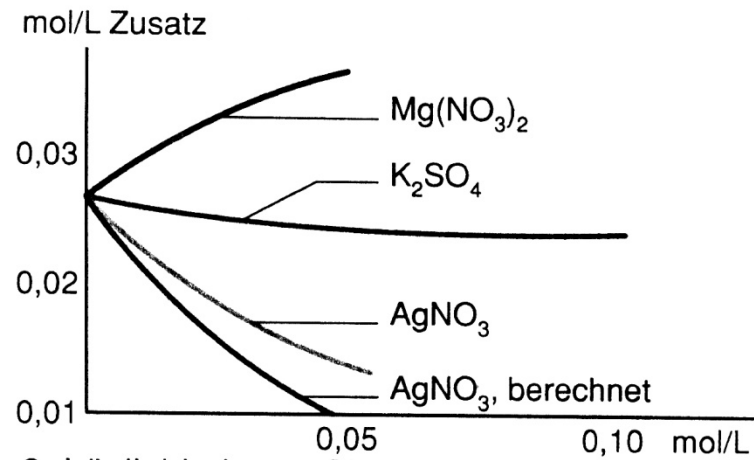
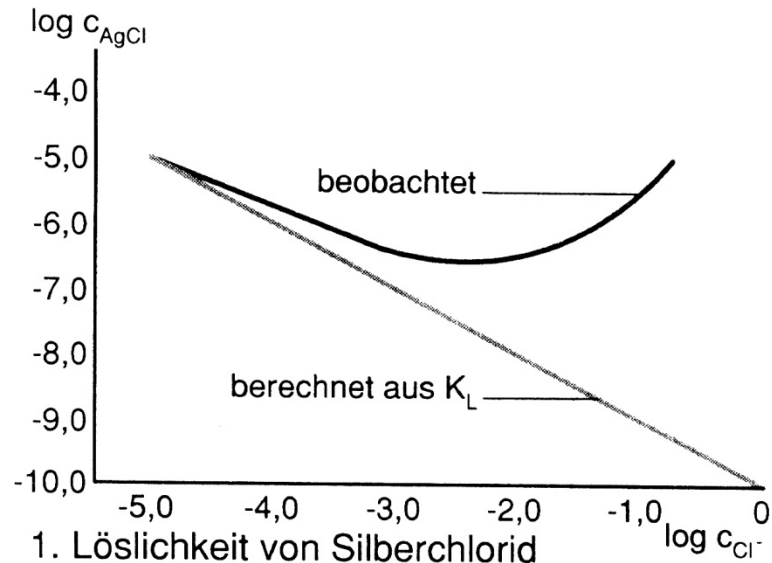
$$\rightarrow K_L = L_a \cdot f_{A^+} \cdot f_{B^-}$$

$$L_a = \sqrt{\frac{K_L}{f_{A^+} \cdot f_{B^-}}}$$

$$f < 1$$

$$L_a > L$$

→ Löslichkeit nimmt bei Anwesenheit von Fremdionen zu → Alkaliaufschluss



C. Beeinflußung der Löslichkeit

NIEDERSCHLAGSBILDUNG -

MECHANISMUS

FÄLLEN

THERMODYNAMISCH = FÄLLUNG

PHASENBILDUNG

FÄLLUNG

INDUKTIONSPERIODE

(KRISTALLKEIME $10^9 - 10^{12}$ KEIME
pro Mol)

KEIMBILDNER

(IMPFKRISTALL, VERUNREINIGUNG,
GLASWAND)

WIRKUNG: - VERRINGERUNG DER GRENZFLÄCHEN-
SPANNUNG
- VERGRÖßERUNG DES TEILCHENRADIUS

KOLLOIDBILDUNG

$As_2O_3 + H_2S \nrightarrow$ kein Niederschlag

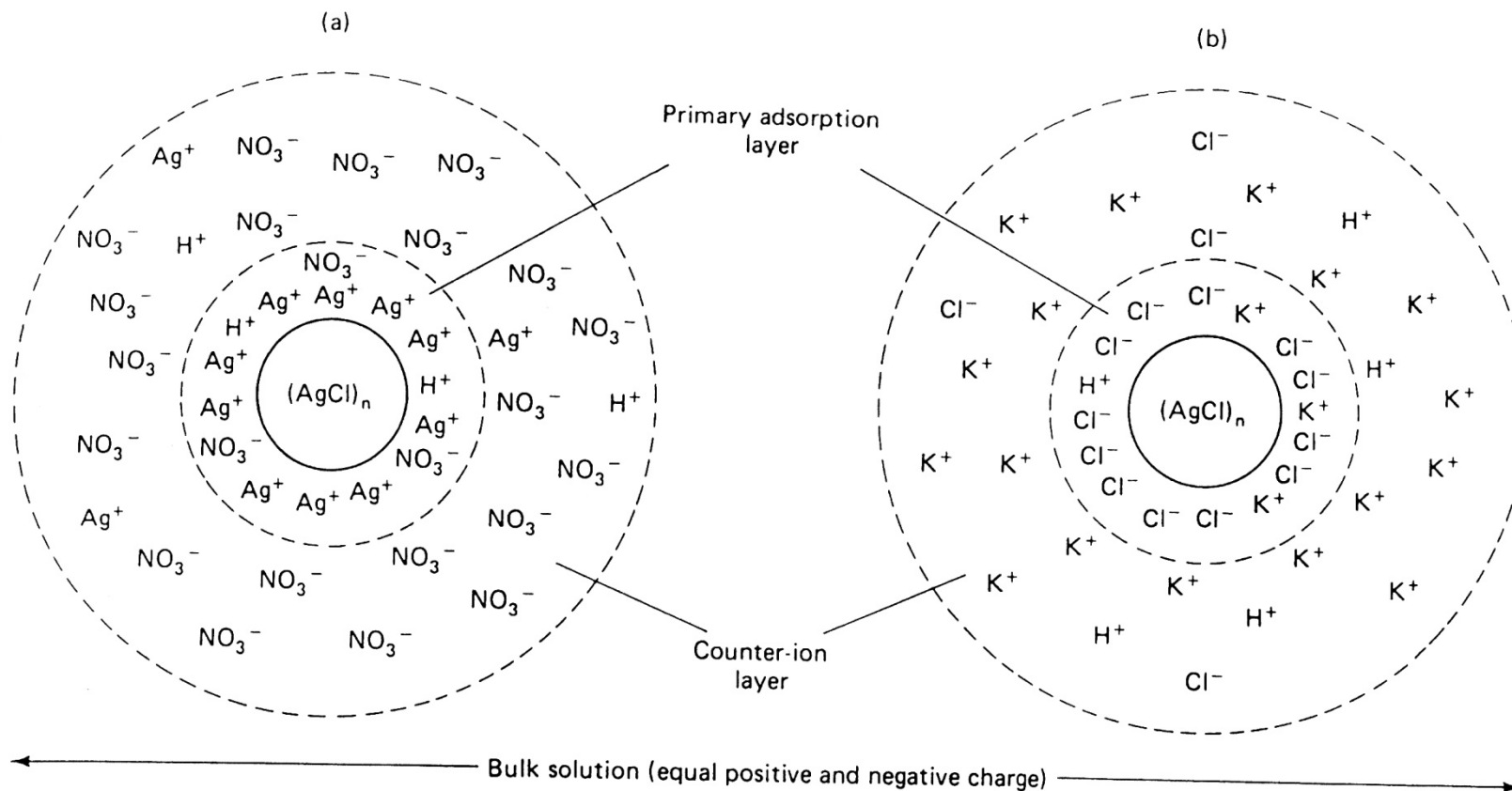
trübe, gelbe Lösung As_2S_3

	TEILCHENGRÖÖÖE [cm]
SUSPENSION	$\gg 10^{-5}$
KOLLOID	$10^{-5} - 10^{-7}$
LÖSUNG	$\leq 10^{-7}$

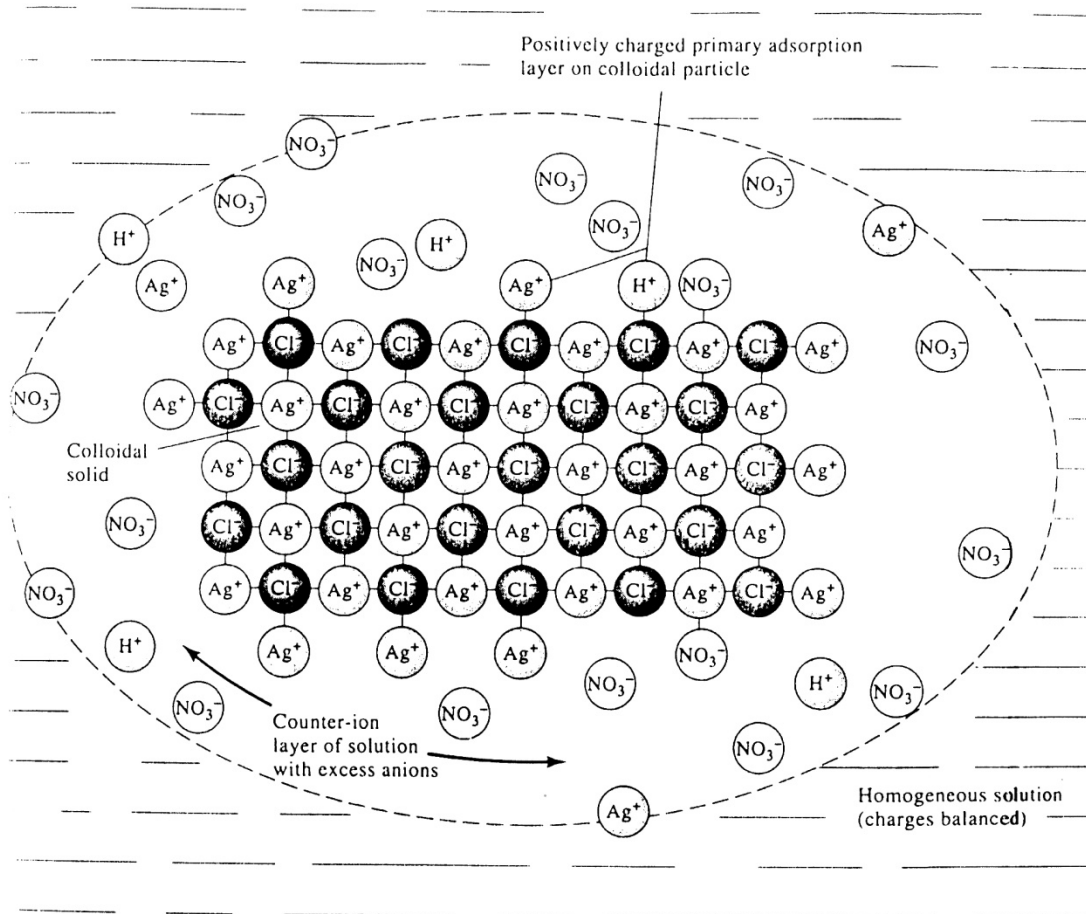
ABTRENNUNG: ZENTRIFUGIEREN
DIALYSE (Diffusion durch
Membranen)
NICHT DURCH FILTER! ⚠

FÄLLUNG ODER KOAGULATION
DURCH SALZZUSATZ ("AUSSALZEN")

KOLLOID	$\xrightarrow{\text{Koagulation}}$	Niederschlag
SOL	$\xleftarrow{\text{Peptisation}}$	gel



5B Properties of Precipitates and Precipitating Age



MITTFÄLLUNG

- EIGENIONEN
- FREMDIONEN

- ADSORPTION

bei Niederschlägen mit aktiver Oberfläche

- OKKLUSION

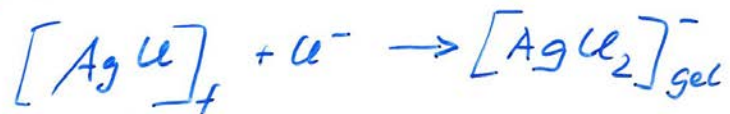
Einschluß von Fremdteilchen im Inneren des Kristalls

- INKLUSION (Mischkristallbildung)

Fremdionen werden direkt in das Kristallgitter eingebaut

KOMPLEXBILDUNG

Niederschläge können durch Komplexbildung ganz oder teilweise wieder in Lösung gehen



ALTERUNG - REIFUNG

physikalisch - chemische Veränderung
nach Fällung als Niederschlag

o REKRISTALLISATION

instabile Kristallbezirke gehen in Lösung

o TEMPERUNG (bei höherer Temp.)

= Ordnungsprozeß durch Diffusion
innerhalb eines Kristalls
⇒ Fehlstellen werden beseitigt

o CHEMISCHE ALTERUNG

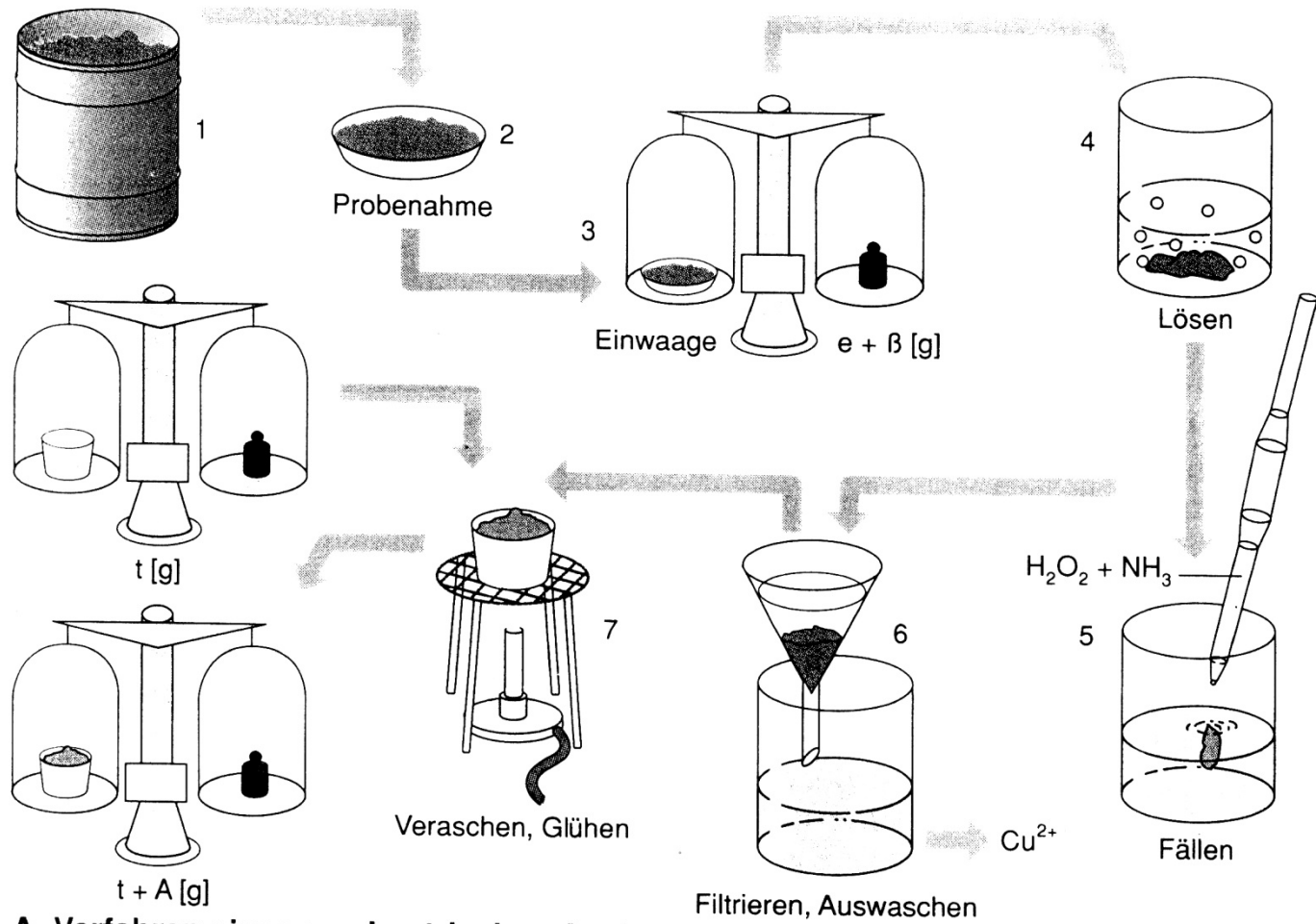
Modifikationsänderung d. Niederschlags



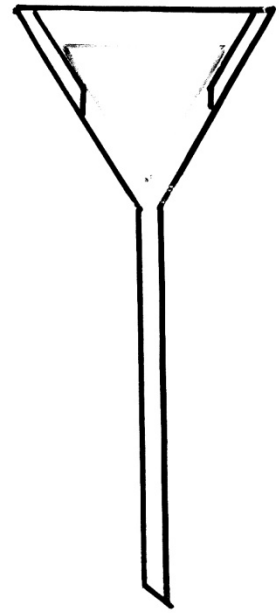
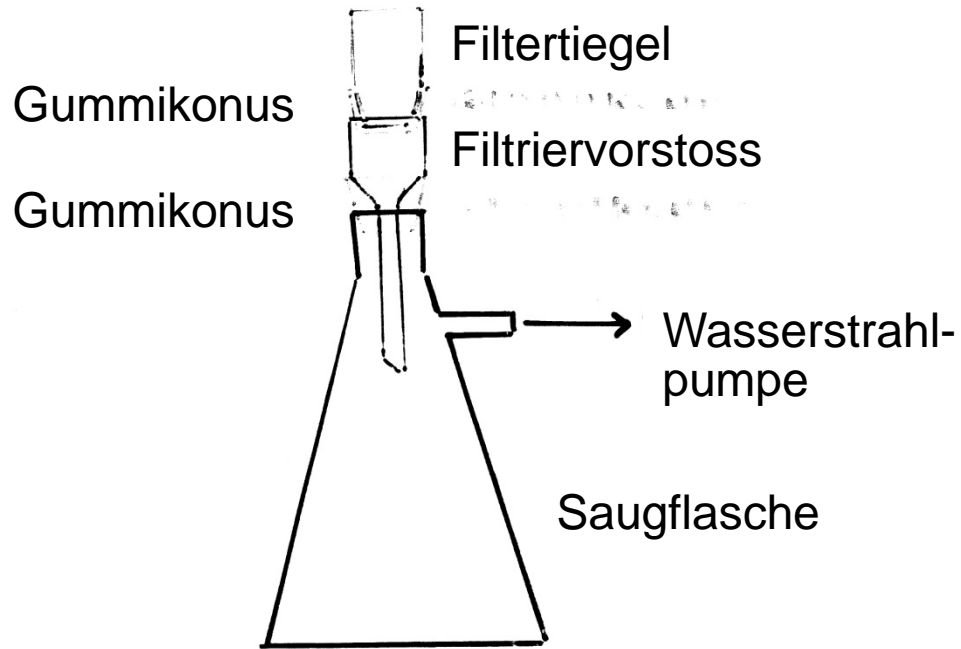
NACHFÄLLUNG

VORGÄNGE, DIE ZUR ÄNDERUNG DER
ZUSAMMENSETZUNG DES NIEDERSCHLAGS
FÜHREN

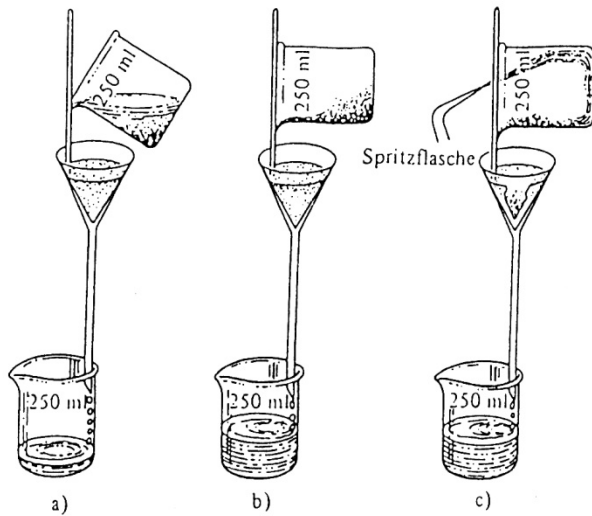




A. Verfahren einer gravimetrischen Analyse



Gravimetrische Verfahren



- Bild 28-7 Dekantieren und Überführen eines Niederschlags in ein Filter.
- Dekantieren, Hauptmenge des Niederschlags verbleibt im Becherglas.
 - Entlang eines Glasrührstabs wird die Hauptmenge des Niederschlags abgegossen.
 - Mit der Spritzflasche wird restlicher Niederschlag in den Trichter gespült (dazu kann auch ein Gummiwischer, oft scherzhaft „Atomwischer“ genannt, erforderlich sein).

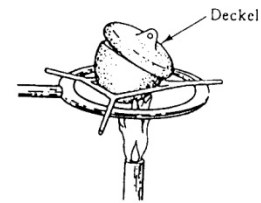


Bild 28-8
Veraschen eines Filters im Tiegel

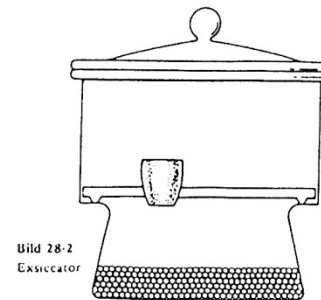
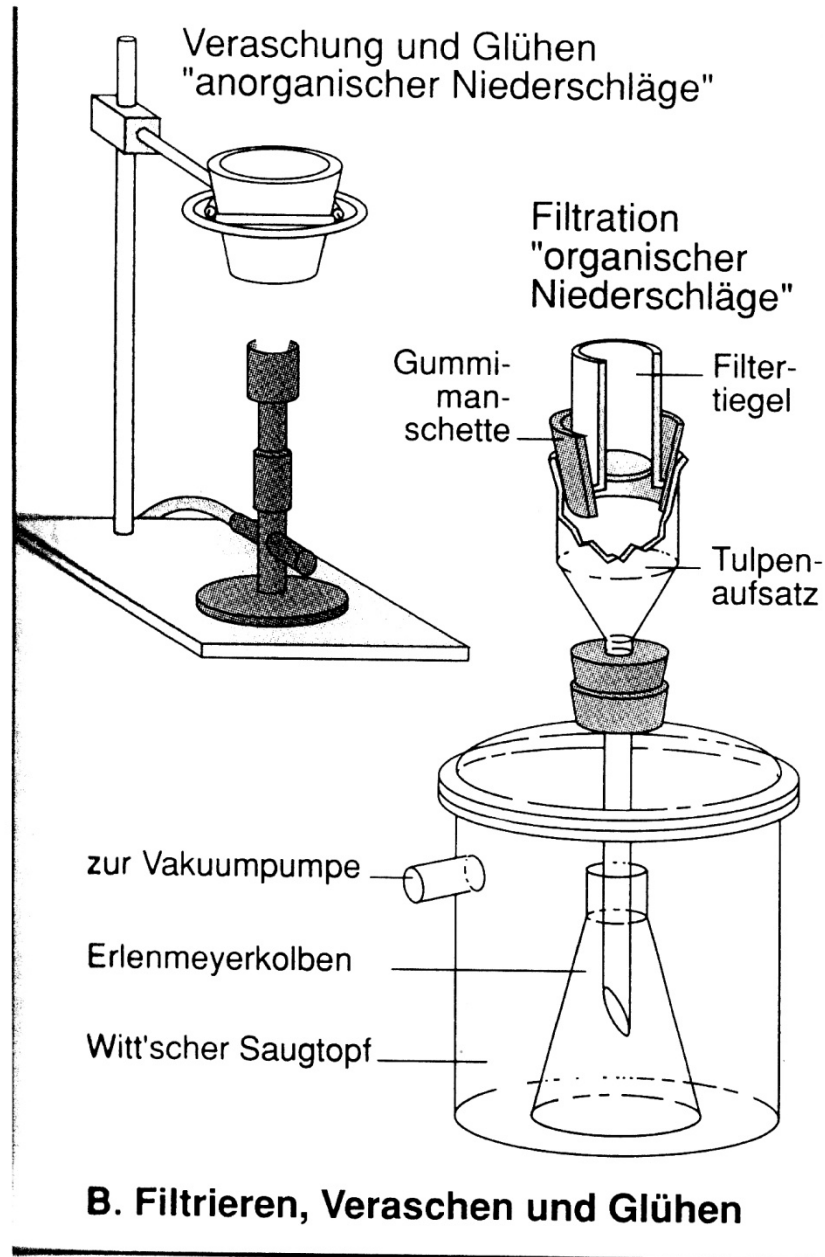


Bild 28-2
EXSICCATOR



Spezifikation von Papierfiltern und Filtertiegeln

Papierfilter Art	Typenbez.	Anwendung, Eigenschaft	Filtertiegel Glas	Porzellan
weich	Schwarzband	großporig	D 1	A 5
		grobkrist. Niederschlag	⋮	⋮
mittel	Weißband	⋮	D 3	A 3
hart	Blauband, Rotband	feinporig feinkrist. Niederschlag	⋮ D 5	⋮ A 1

Gebräuchliche Trockenmittel (nach [18])

Trockenmittel	Partialdruck ^a (mbar)	Anwendbar für	Nicht geeignet für
CaCl ₂ gekörnt	0,20	Neutrale und saure Feststoffe (Gase)	NH ₃ , Br ₂ , HBr, HF; Alkohole, Amine
BaO	1,6 · 10 ⁻⁴	Anorg. und organ.	F ₂ , NO, Säuren
MgO	0,11	Basen (NH ₃ , Amine),	
Al ₂ O ₃ · aq	4,1 · 10 ⁻³	Alkohole	
NaOH geschmolzen	0,21	NH ₃ , Alkohole,	O ₃ , F ₂ , Säuren
KOH geschmolzen	2,8 · 10 ⁻³	organ. Basen	
H ₂ SO ₄ konz.	2,8 · 10 ⁻³	Neutrale und saure Verbindungen; O ₂ , N ₂ , CO, CH ₄ , Halogene, HCl, SO ₂	HBr, HI, HF, NH ₃ , H ₂ S, PH ₃ , NO, NO ₂ , C ₂ H ₂
P ₄ O ₁₀	3,4 · 10 ⁻⁵	O ₂ , CO, C ₂ H ₂ , CS ₂ , CCl ₄ , Stickoxide	Halogene, HHal, NH ₃ , H ₂ S, Ether
Silicagel	3,0 · 10 ⁻²	universell	NH ₃ , HF, Halogene

^aWasserdampf-Partialdruck bei 25 °C (Reines Wasser: 31,3 mbar)

FILTER

ART	KENNUNG	POREN μm	GESCHW.	T	G	VERWENDUNG	BEISPIELE
P A P I E R	BLAU	2-3	langsam	-	+	feinteilige N.	$\text{BaSO}_4, \text{TiO}_2 \cdot \text{aq}$
	WEISS	6-7	mittel	-	+	krist., körnige N.	$\text{CaC}_2\text{O}_4 \cdot \text{aq}$
	SCHWARZ	7-8	rasch	-	+	gelartige N.	$\text{Fe}(\text{OH})_3, \text{Al}(\text{OH})_3$
P O R Z E L L A N	P1, A1	5-6	langsam	+	+	feinstkörnig	BaSO_4
	P2, A2	7-8	mittel	+	+	universell	$\text{AgCl}, \text{Mg}(\text{NH}_4)\text{PO}_4$
	P3, A3	10	rasch	+	+	grobkristallin	PbSO_4
G L A S	(G, D) 0	230		+	-		
	1	110	rasch	+	-	grobkristallin	
	2	50		+	-		
	3	15-40	mittel	+	-	feinkristallin	$\text{Ni}(\text{DAD})_2, \text{Mg}(\text{Ox})_2$
	4	5-15	langsam	+	-	sehr feint.	
	5	3-4					

Waagen

Waagentyp	Maximalgewicht	Auflösung
Analytische Waagen	150-200g	$\pm 0,1\text{mg}$
Semimicrowaagen	75-100g	$\pm 0,01\text{mg}$
Microwaagen	10-30g	$\pm 0,001\text{mg}$

➤Empfindlichkeit

Ausschlag bei einer Auflage von 0,1mg. Die Empfindlichkeit sinkt mit zunehmendem Gewicht. Die Grenze der Auflage ist erreicht, wenn die Waage nur mehr 40 % der Maximalempfindlichkeit hat.

➤Referenz

Urkilogramm in Paris aus Pt/Ir (1887)

➤Bauarten von Waagen

Oberschalige-, Unterschälige-, Elektronische, Digitalwaagen

➤Dämpfung

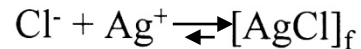
Luft-, magnetische Dämpfung

Fällungsreaktionen in der analytischen Chemie

➤ Sulfatfällung



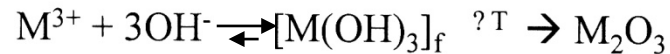
➤ Chloridfällung



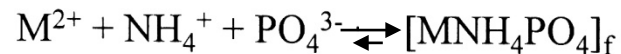
lichtempfindlich (Ag!)

rasch filtrieren, trocknen


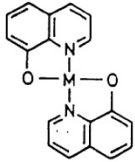
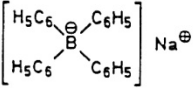
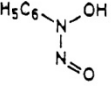
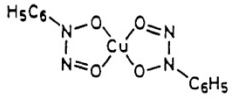
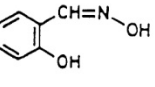
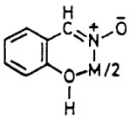
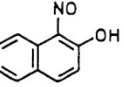
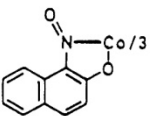
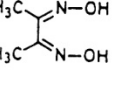
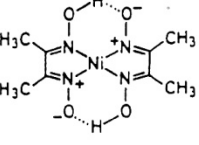
➤ Hydroxidfällung



➤ Phosphatfällung



Organische Fällungsreagenzien

Organische Verbindung	Komplex	Anwendung
 8 - Hydroxyquinolin (Oxin)	 M(II) - oxinat	Mg ²⁺ ; Zn ²⁺ ; Cu ²⁺ ; Cd ²⁺ ; Al ³⁺ ; Fe ³⁺ ; Sb ³⁺ u.a.
 Natriumtetraphenylborat (Kalignost)		K ⁺ (NH ₄ ⁺ , Rb ⁺ , Cs ⁺)
 N-Nitrosophenylhydroxylamin (Kupferron)	 M(II) - oxinat	außer Cu noch für zahlreiche andere Metalle verwendbar
 Salicylaldoxim	 M(II) - Komplex	Cu ²⁺ ; Ni ²⁺ ; Pb ²⁺ ; Bi ³⁺ ; Fe ³⁺ u.a.
 α-Nitroso-β-naphthol	 Co(III) - Komplex	spezifisches Reagenz für Co(III)
 Diacetyldioxim (Dimethylglyoxim)	 Ni(II) - Komplex	spezifisches Reagenz für Ni(II)

Anwendungsbereich - Gravimetrie

mg-Bereich

Vorteile:

Geringer app. Aufwand

Hohe Präzision

Nachteil:

Hoher Zeitaufwand

Keine Serienanalysen

Kaum automatisierbar

$$m(\text{gesuchte Substanz}) = \frac{M(\text{gesuchte Substanz})}{M(\text{Wägeform})} \cdot m(\text{Niederschlag})$$
$$= f_g \cdot m(\text{Niederschlag})$$

$$\text{Prozentgehalt(gesuchte Substanz)} = \frac{m(\text{gesuchte Substanz})}{m(\text{Probe})} \cdot 100$$

$$\text{Prozentgehalt(gesuchte Substanz)} = \frac{100 \cdot f_g \cdot m(\text{Auswaage})}{m(\text{Probe})}$$

m = Masse Gewicht f_g gravimetrischer Faktor

M = molare Masse g/ml

(Atomgewicht alt

Molgewicht)

3.1. Analytische und stöchiometrische

Gesucht	Gegeben	Faktor	lg
K ₂ O	K	1,2046	08 084
	K(C ₆ H ₅) ₃ B	0,1314	11 874
	KCl	0,6317	80 056
	KClO ₄	0,3399	53 142
	K ₂ PtCl ₆ empirisch	0,1931	28 570
	K ₂ SO ₄	0,5406	73 285
	BaSO ₄	0,4036	60 597
	Pt empirisch Pt empirisch ¹⁾	0,4810 0,4809	68 217 68 203
K ₂ O · Al ₂ O ₃ · 6 SiO ₂ }	Al ₂ O ₃	5,460	73 717
	KCl	3,733	57 210
	K ₂ O	5,910	77 153
	K ₂ SO ₄	3,194	50 440
K ₂ SO ₄	K(C ₆ H ₅) ₃ B	0,2432	38 588
	BaSO ₄	0,7466	87 311
	KCl	1,1687	06 770
	K ₂ O	1,850	26 713
La	La ₂ O ₃	0,8527	93 079

¹⁾ Bei Gegenwart von Sulfat nach Finkners Methode

Der Bariumgehalt einer Substanzprobe ergibt sich aus dem Verhältnis des gefundenen Wertes und der Einwaage.

Einwaage: 0,8450 g

$$w_{\text{Ba}} = \frac{0,2034 \text{ g}}{0,8450 \text{ g}} = 0,241 \approx 24,1\%$$

b. Auswaage an Fe₂O₃: 0,2345 g. Gesucht: m_{Fe}

$$m_{\text{Fe}} = \frac{2 M(\text{Fe})}{M(\text{Fe}_2\text{O}_3)} \cdot m(\text{Fe}_2\text{O}_3) = 0,6994 \cdot 0,2345 \text{ g} = 0,1640 \text{ g}$$

In einigen Fällen wird statt des exakten gravimetrischen Faktors ein *empirischer* Faktor verwendet, wenn der Niederschlag signifikant von der stöchiometrischen Zusammensetzung abweicht. Solche empirischen Werte sind allerdings mit gewisser Vorsicht zu betrachten.