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The Staphylinidae of Armenia and Nagorno-Karabakh (Coleoptera)

With 65 figures, 8 maps and 4 tables

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Abstract

The previously largely neglected and poorly known staphylinid faunas of Armenia and Nagorno-Karabakh are addressed. Based on a study of more than 31,000 Staphylinidae recently collected in various habitats and using different methods, and on a critical evaluation of previous literature records, a checklist of the faunas of Armenia and Nagorno-Karabakh is compiled. The fauna of Armenia currently includes 675, that of Nagorno-Karabakh 198 named species. Nevertheless, it is concluded that the species inventory of both regions, especially that of Nagorno-Karabakh, is still far from complete. As many as 262 and 183 species are reported from Armenia and Nagorno-Karabakh, respectively, for the first time. A list of 99 species erroneously or doubtfully recorded from the study region is provided. A comparison with the species number and systematic composition of the faunas of other Caucasian countries and regions revealed that (a) their known diversities are significantly lower than should be expected and (b) a remarkably high proportion (nearly 40 %) of Aleocharinae in the faunas of Armenia and Nagorno-Karabakh, with the genus *Atheta* Thomson, 1858 alone accounting for approximately 10 % of the total diversity in Armenia. The faunas of Armenia and Nagorno-Karabakh are primarily composed of widespread species, many of them Caspian (Caucasian) and Iranian elements, and remarkably few endemics. Only 23 species, thirteen of the Aleocharinae (most of them belonging to the genus *Geostiba* Thomson, 1858), four of the Pselaphinae, five of Scydmaeninae, and one of Staphylininae are classified as regional endemics, and one species of Aleocharinae from the peak region of Mount Khustup is classified as a local endemic. The distribution of one addititional species is confined to South Armenia and adjacent parts of North Iran. A number of species is currently known only from Armenia, but of doubtful taxonomic status or unlikely to represent endemics. Records of some species in Armenia and/or Nagorno-Karabakh revealed some remarkably discontinuous distributions with gaps of up to approximately 2,800 km; three of these distributions are mapped. Fourteen species are newly described: *Omalium kociani* Zanetti spec. nov. (Armenia: Jermuk) of the Omaliinae, *Proteinus baculatus* Assing spec. nov. (Armenia; Northeast Turkey) of the Proteininae, *Bryaxis armeniacus* Brachat spec. nov. (Armenia: NW Hrazdan) and *B. meghruicus* Brachat spec. nov. (South Armenia: Meghru range) of the Pselaphinae, *Atheta* (*Paralpinia*) *meghruica* Assing spec. nov. (South Armenia: Meghru range), *Bellatheta khustupica* Assing spec. nov. (South Armenia: Mount Khustup), *Calodera alticola* Assing spec. nov. (Armenia: Mount Karkar), and *Tachyusa unguis* Assing spec. nov. (South Armenia) of the Aleocharinae, *Anotylus hamatoides* Schülke spec. nov. (Armenia) of the Oxytelinae, *Euconnus* (*Tetramelus*) *longilaminatus* Meybohm spec. nov. (North Armenia), *E.* (*T.*) *tavushus* Meybohm spec. nov. (North Armenia), *E.* (*T.*) *karabakhus* Meybohm spec. nov. (Nagorno-Karabakh), *Neuraphes* (*Paraphes*) *gomarantsus* Meybohm spec. nov. (South Armenia: Meghru range), and *N.* (*P.*) *syunikus* Meybohm spec. nov. (South Armenia) of the Scydmaeninae. Eight synonymies and one revalidation are established: *Dialycera minuta* Luze, 1906 = *Phyllodrepa armena* Iablokoff-Khnzorian, 1959, syn. nov.; *Mycetoporus silvaticus* Iablokoff-Khnzorian, 1962 = *M. dispersus* Schülke & Kocian, 2000, syn. nov.;

Aleochara subtumida (Hochhuth, 1859) = *Aleochara khnzoriani* Amiryan, 1999, syn. nov.; *Platystethus cephalotes* Eppelsheim, 1878, revalidated (previously synonym of *P. laevis* Märkel & Kiesenwetter, 1848) = *P. oblongopunctatus* Roubal, 1911, syn. nov.; *Euconnus lalvarensis* Iablokoff-Khnzorian, 1964 = *Euconnus pseudorobustus* Franz, 1986, syn. nov.; *Astenus rufopacus* Reitter, 1909 = *A. baali* Coiffait, 1960, syn. nov.; *Heterothops dissimilis* (Gravenhorst, 1802) = *H. armeniacus* Coiffait, 1977, syn. nov.; *Heterothops praevius* Erichson, 1839 = *Heterothops montanus* Iablokoff-Khnzorian, 1966, syn. nov.

Taxonomic acts

Omalium kociani Zanetti spec. nov. – urn:lsid:zoobank.org:act:5F24413E-6AB4-4D1C-B567-AB93C38380C9
Proteinus baculatus Assing spec. nov. – urn:lsid:zoobank.org:act:3DDB2D69-A000-4FFA-AAC7-48CE4BAB7020
Bryaxis armeniacus Brachat spec. nov. – urn:lsid:zoobank.org:act:01F595CB-9F3B-4DEE-8A4D-C97F39DE26E1
Bryaxis meghruicus Brachat spec. nov. – urn:lsid:zoobank.org:act:AA6E7E2A-4450-48E8-8048-8061C3C6DF5C
Atheta (*Paralpinia*) *meghruica* Assing spec. nov. – urn:lsid:zoobank.org:act:4673F1AA-F8C1-4524-AFA3-99E4E7F502FF
Bellatheta khustupica Assing spec. nov. – urn:lsid:zoobank.org:act:C342B3DE-9EE5-4950-85BD-817DBEE6FE3E
Calodera alticola Assing spec. nov. – urn:lsid:zoobank.org:act:3BF46F7B-FBDB-4F1B-B0D0-89D669B021C2
Tachyusa unguis Assing spec. nov. – urn:lsid:zoobank.org:act:A4B4E5C2-EEB2-4146-B6F2-27D5873D5AEB
Anotylus hamatoides Schülke, spec. nov. – urn:lsid:zoobank.org:act:0167E052-EBAB-421A-BB72-39E33C1EDC97
Euconnus (*Tetramelus*) *longilaminatus* Meybohm spec. nov. – urn:lsid:zoobank.org:act:96692EB3-A82F-4ADB -A278-EC1B64040C82
Euconnus (*Tetramelus*) *tavushus* Meybohm spec. nov. – urn:lsid:zoobank.org:act:7F2BBD70-A7F8-4272-8195-02884B90F2DD
Euconnus (*Tetramelus*) *karabakhus* Meybohm spec. nov. – urn:lsid:zoobank.org:act:A60B66A1-20EB-4341-9950-B6485B1A3853
Neuraphes (*Paraphes*) *gomarantsus* Meybohm spec. nov. – urn:lsid:zoobank.org:act:102F7EEC-B2A5-458F-B271-E3582970195F
Neuraphes (*Paraphes*) *syunikus* Meybohm spec. nov. – urn:lsid:zoobank.org:act:2972F28B-0968-4810-B9F5-471E93F97F43

Key words

Coleoptera, Staphylinidae, Palaearctic region, Caucasus region, Armenia, Nagorno-Karabakh, taxonomy, diversity, zoogeography, endemism, discontinuous distributions, ecology, new species, new synonyms, revalidation, new records, checklist, distribution maps.

Zusammenfassung

Über die Staphylinidenfauna von Armenien and Bergkarabach war bisher wenig bekannt. Auf Basis der Bearbeitung von mehr als 31.000 in den letzten Jahren in verschiedenen Habitaten und mit unterschiedlichen Methoden gesammelten Staphyliniden sowie einer kritischen Auswertung von Literaturnachweisen wird eine Checkliste der Fauna von Armenien und Bergkarabach zusammengestellt. Aus Armenien sind derzeit 675, aus Bergkarabach 198 Arten nachgewiesen. Trotzdem ist das Arteninventar bei weitem noch nicht vollständig erfasst. Insgesamt werden 262 Arten aus Armenien und 183 Arten aus Bergkarabach erstmals gemeldet. Frühere Nachweise von insgesamt 99 Arten beruhen wahrscheinlich auf Fehldeterminationen oder wurden irrtümlich Armenien zugeordnet. Ein Vergleich der Artenzahlen und systematischen Zusammensetzung mit den Faunen anderer Länder und Gebieten der Kaukasusregion ergab, dass die derzeit bekannte Diversität deutlich geringer als zu erwarten ist. Darüber hinaus fällt der bemerkenswert hohe Anteil (40 %) der Aleocharinae in den Faunen von Armenien und Bergkarabach auf, wobei allein die Gattung *Atheta* Thomson, 1858 in Armenien 10 % der Gesamtdiversität ausmacht. Der Artenbestand in Armenien und Bergkarabach wird vor allem durch weit verbreitete Arten geprägt, von denen viele als kaspische (kaukasische) oder iranische Faunenelemente einzuordnen sind. Nur 23 Arten, darunter 13 Aleocharinae (die meisten davon aus der Gattung *Geostiba* Thomson, 1858), vier Pselaphinae, fünf Scydmaeninae und eine Art der Staphylininae, werden als Regionalendemiten, eine Art der Aleocharinae vom Mount Khustup als Lokalendemit klassifiziert. Die Verbreitung einer weiteren Art ist auf das südliche Armenien und angrenzende Gebiete des Nordiran beschränkt. Eine beträchtliche Zahl von Arten ist zwar derzeit ausschließlich aus Armenien bekannt, allerdings entweder taxonomisch ungeklärt oder sehr wahrscheinlich auch in angrenzenden Ländern verbreitet. Die Nachweise einiger Arten aus Armenien und/oder Bergkarabach offenbarten eine Reihe von auffälligen Disjunktionen mit Verbreitungslücken von bis zu 2800 km; drei solcher Verbreitungsgebiete werden anhand von Karten veranschaulicht. Vierzehn Arten werden erstmals beschrieben: *Omalium kociani* Zanetti spec. nov. (Armenien: Jermuk) aus der Unterfamilie Omaliinae, *Proteinus baculatus* Assing spec. nov. (Armenien; Nordosttürkei) (Proteininae), *Bryaxis armeniacus*

Brachat spec. nov. (Armenien: NW Hrazdan) und *B. meghruicus* Brachat spec. nov. (Südarmenien: Meghru range) (Pselaphinae), *Atheta* (*Paralpinia*) *meghruica* Assing spec. nov. (Südarmenien: Meghru-Gebirge), *Bellatheta khustupica* Assing spec. nov. (Südarmenien: Mount Khustup), *Calodera alticola* Assing spec. nov. (Armenien: Mount Karkar) und *Tachyusa unguis* Assing spec. nov. (Südarmenien) der Aleocharinae, *Anotylus hamatoides* Schülke spec. nov. (Armenien) der Oxytelinae, *Euconnus* (*Tetramelus*) *longilaminatus* Meybohm spec. nov. (Nordarmenien), *E.* (*T.*) *tavushus* Meybohm spec. nov. (Nordarmenien), *E.* (*T.*) *karabakhus* Meybohm spec. nov. (Bergkarabach), *Neuraphes* (*Paraphes*) *gomarantsus* Meybohm spec. nov. (Südarmenien: Meghru-Gebirge) und *N.* (*P.*) *syunikus* Meybohm spec. nov. (Südarmenien) der Scydmaeninae. Acht Namen werden synonymisiert und einer revalidiert: *Dialycera minuta* Luze, 1906 = *Phyllodrepa armena* Iablokoff-Khnzorian, 1959, syn. nov.; *Mycetoporus silvaticus* Iablokoff-Khnzorian, 1962 = *M. dispersus* Schülke & Kocian, 2000, syn. nov.; *Aleochara subtumida* (Hochhuth, 1859) = *Aleochara khnzoriani* Amiryan, 1999, syn. nov.; *Platystethus cephalotes* Eppelsheim, 1878, revalidiert (vorher Synonym von *P. laevis* Märkel & Kiesenwetter, 1848) = *P. oblongopunctatus* Roubal, 1911, syn. nov.; *Euconnus lalvarensis* Iablokoff-Khnzorian, 1964 = *Euconnus pseudorobustus* Franz, 1986, syn. nov.; *Astenus rufopacus* Reitter, 1909 = *A. baali* Coiffait, 1960, syn. nov.; *Heterothops dissimilis* (Gravenhorst, 1802) = *H. armeniacus* Coiffait, 1977, syn. nov.; *Heterothops praevius* Erichson, 1839 = *Heterothops montanus* Iablokoff-Khnzorian, 1966, syn. nov.

Schlüsselwörter

Coleoptera, Staphylinidae, Paläarktis, Kaukasusregion, Armenien, Bergkarabach, Taxonomie, Diversität, Zoogeographie, Endemismus, Ökologie, neue Arten, neue Synonymien, Erstnachweise, Checkliste, Verbreitungskarten

1 Introduction

Its area covering a mere 29,800 km^2, Armenia is a small country in the northeast of the Armenian Highlands and to the south of the Lesser Caucasus. It borders on Georgia in the north, Turkey in the west, the Azerbaijanian exclave Nakhchivan in the southwest, Iran in the south, and Azerbaijan and the Republic of Artsakh (Nagorno-Karabakh) in the east. Most of the area is dominated by mountains and high plateaus, with a mean altitude of approximately 1800 m, the highest peak (Mount Aragats) at 4090 m, numerous summits at more than 3000 m, and the lowest elevation at 380 m in the Aras river valley in the south.

Nagorno-Karabakh, officially named Republic of Artsakh since 2017, is a de facto independent state. It formed part of Azerbaijan prior to the Nagorno-Karabakh War and is still recognized as such by the UN today. Its small territory of barely 11,500 km^2 shares borders with Azerbaijan in the north and east, with Armenia in the west, and Iran in the south. Like Armenia, Nagorno-Karabakh is mostly covered by mountains and highlands, with an average altitude of approximately 1100 m and with two major mountain ranges, the Mrav range in the north (highest peaks Gomshasar and Mrav at 3720 and 3340 m, respectively) and the Karabakh range in the west with several peaks at above 2500 m.

The mountainous topography and geology of the entire Armenian Highland, which includes the territories of Armenia and Nagorno-Karabakh, was primarily shaped by the collision of the Arabian with the Eurasian plate. Since the Arabian plate is moving northwards at 2.5 cm per year even today (Neukirchen 2011), the Armenian Plateau is still tectonically and seismically active, the last major earthquake dating back only to 1988. The geological history also explains why the region is characterized by a mix of various mesozoic and palaeogenic sediments, neogenic continental sediments, and of volcanic rocks of various ages (Plöchinger 1979).

According to Fauyvush & Aleksanyan (2016), four major geomorphological regions can be distinguished in Armenia. They are the mountains and valleys in the northeast, which have been subject to massive erosion, areas covered by lava of Pliocene origin not significantly affected by erosion, ridged mountains in the south with signs of heavy erosion, and the Ararat valley with a cover of alluvial and proluvial sediments.

The climate is essentially dry continental, with dry and hot summers and with cold winters. However, pronounced gradients in temperature and precipitation can be observed along different altitudinal regimes, the mean annual temperature varying between 2.7 and 14.1 °C and the mean annual precipitation between 250 (Ararat plain) and 1000 mm (Mount Aragats).

As a result of the diversity of topography, geology, and climate in the region, Armenia (and Nagorno-Karabakh) are shaped by a wide range of landscapes such as desert (Transcaucasian sand desert), semi-deserts, and salt marshes (at altitudes of 480–1200 m), steppes and forests at altitudes of 1200–2200 m, as well as subalpine and alpine meadows, carpets, and wetlands at 2200–4000 m. Approximately 3800 species of vascular plants, 428 species of algae, 399 species of mosses, 4207 species of fungi, 464 species of lichens, and 549 species of vertebrates have been recorded from Armenia, with the flora including 142 endemic plant species (Fauyvush &

Aleksanyan 2016). According to the website of the Office of the Nagorno Karabakh Republic (www.nkrusa.org), the flora of Nagorno-Karabakh is composed of 2000 – probably vascular – plant species.

The Armenian forest habitats are composed of 110 tree and 152 shrub species. The dominant tree species are *Quercus* spp., *Fagus orientalis*, and *Carpinus betulus*, which alone account for approximately 81 % of the forest cover. The remainder is mostly composed of pine (mostly secondary), juniper, hornbeam coppice, lime, ash, and maple. The forest cover has significantly decreased, primarily due to deforestation, from 35 % to 7–8 % during the past 6000 years. However, large areas especially in Northwest, West, and Central Armenia have not supported forests since the Tertiary period. Today, 62 % of the remaining forested areas are found in the northeast, 36 % in the southeast, and only 2 % in the central parts of Armenia (Moreno-Sanchez & Sayadyan 2005). The western and northern parts of Nagorno-Karabakh, by contrast, still feature extensive primary forests.

According to Fauyvush & Aleksanyan (2016), the Armenian invertebrate fauna is composed of 17200 species. However, this figure is probably based on rough estimates rather than long-term modern studies and most likely requires revision. At least this can be said for the Staphylinidae (rove beetles), the most speciose of all beetle families and in fact the most speciose of all families of organisms. The two editions of the Palaearctic Catalogue (Smetana 2004, Schülke & Smetana 2015) list 340 and 395 species, respectively, for Armenia. No such data are available for Nagorno-Karabakh, since it was treated as a part of Azerbaijan in these works. However, even the recent figures for Armenia are not only incomplete, but also misleading and largely erroneous, primarily because of the changing interpretations, definitions, and boundaries of the geographic term "Armenia" over the centuries. Most of what was regarded as Armenia and East Armenia in the 19th century belongs to Turkey, Georgia, or Azerbaijan today. The same is true of the geographic terms "Hochland von Armenien" (Armenian Highland) and "Armenisches Gebirge" (Armenian Mountains), the latter of which includes the Lesser Caucasus and adjacent mountain ranges such as the Pontic Mountains. Moreover, numerous species reported from the historical locality "Araxestal" (or equivalent) (e.g., Eppelsheim 1890a, b) have erroneously been referred to Armenia. This German term was used for what is Ordubad in the Azerbaijanian exclave Nakhchivan today and at the same time denotes "Aras river valley". The Aras river forms the border between Turkey and Armenia, as well as that between Iran and Armenia and Azerbaijan, respectively.

By around 1830, the regions of the Caucasus and Transcaucasica were under Russian administration. In the following period, this area was visited by several coleopterists such as Ménétriés (1827–31), Scovicz (1827–1830), Motschulsky (1834–35, 1837–38), Kolenati (1843–45), Chaudoir (1845), and Gotsch (1845). At least Kolenati (see Kolenati 1858: 32 ff.), Motschulsky (see Motschulsky 1850: 18 ff.), Gotsch (see Hochhuth 1847: 449), and probably also Scovicz (see Motschulsky 1850: 19, Faldermann 1837: 42 f.) collected material in the region that is Armenia or Nagorno-Karabakh today.

The first descriptions of Armenian Staphylinidae were probably published by Faldermann (1835), who studied the material collected by Scovicz, in his "Coleoptera Persico-Armeniaca". However, since he did not specify localities in this work, the exact origin of the type material is unknown. Kolenati (1846) reported eleven species of Staphylinidae, six of them newly described, from "Karabagh" and eight species, five of them new, from Armenia, but only the localities of one of these species are in Armenia today. Hochhuth (1849) described numerous species of Staphylinidae, mostly without specifying localities, based on material from Chaudoir, Gotsch, and Motschulsky. In some cases he indicated Armenia as the origin, but it is rather unlikely that the respective localities are within the boundaries of modern Armenia (see above). In the second half of the 19th century, the German entomologist Hans Leder significantly contributed to the knowledge of the beetle fauna of the Caucasus region by conducting long-term collecting expeditions in the years 1875–1881. From 1882 to 1888 he lived in Helenendorf (today Göygöl in Azerbaijan). His earlier expeditions (1875–1879) did not, however, include the territory of modern Armenia (Schneider & Leder 1878: 23 ff., Leder 1880: 451 ff., 1881: 501 ff.), and his later collecting activities focused on the environs of Ordubad (Hetschko 1922). In 1875, Schneider (see Schneider & Leder 1878: 6 ff.) collected beetles in North and Central Armenia (Gyumri, Yerevan, Vagharshapat). His Staphylinidae were studied by Eppelsheim (1878), who reported eleven species from Armenia, one of them new. In the early 20th century (1915–1916), the Polish entomologist W. Eichler collected beetles in the environs of Vagharshapat. The list of species published by Eichler (1930) includes 87 species of Staphylinidae.

In the second half of the 20th century, it was S. M. Iablokoff-Khnzorian who significantly contributed to the knowledge of Armenian beetles, including rove beetles. In total, he described 28 new species of Staphylinidae from Armenia (Iablokoff-Khnzorian 1956, 1957b, 1959, 1960, 1961, 1962, 1964a, 1966, 1989). Besides, he actively engaged in a collaboration with other specialists who described new species and published records based on his Armenian material, e.g., Benick (1974), Coiffait (1965, 1966a, b, 1967a, b, 1968, 1970a, b, d, 1972a, 1977), Likovsky (1971), Pace (1982, 1983), and Ullrich (1975). Around the turn of the millenium, Amiryan (1999a, b) and Amiryan & al. (2000) published three articles on the genus *Tachinus*, a new species of *Aleochara*, and the pholeophilous, nidicolous, and myrmecophilous Staphylinidae of Armenia.

Extensive additional information is scattered in numerous articles, especially comprehensive taxonomic revisions, not specifically dealing with the fauna of Armenia. These include Ryvkin (1990: *Stenus*), Sabella & al. (2004: *Brachygluta*), Assing (2005b: *Leptobium*; 2007b: *Pronomaea*; 2008c: *Sunius*; 2010a: *Luzea*; 2010b: *Achenium*; 2016a: *Anaulacaspis*; 2018e: *Cousya*), Jászay & Hlaváč (2006: *Dropephylla*), and Enushchenko & Semenov (2016: *Gyrophaena*), to name only a few. For additional articles see Tab. 1, 3 and the reference section in this paper.

The present study aims at providing a revised overview and a first critical checklist of the Staphylinidae of Armenia and Nagorno-Karabakh. It is primarily based on the results of three field trips conducted by the authors in June/July in three consecutive years (2016–2018), the first two exclusively to Armenia and the third trip to both Nagorno-Karabakh and Armenia. In addition, material from field trips to Armenia conducted by Matúš Kocian in 2015 and by Jörg Müller and Andrea Jarzabek-Müller in 2016 was examined. The staff of the Zoological Institute in Yerevan, including Mark Kalashian, Tigran Ghrejyan, and Gayane Karagyan, provided substantial material from pitfall trap studies in Armenia. Harald Schillhammer contributed specimens and data of material collected in Armenia in 2001, together with Helena Shaverdo. Finally, a number of additional specimens located in various museum and private collections is included.
Some results of the three field trips conducted by the authors have already been published in separate articles: Assing (2016b–c, 2017b–d, 2018a–c, g–h), Assing & Vogel (2017), and Schülke (2019a–b). Results of revisions of type material in the Iablokoff-Khnzorian collection have been addressed by Assing (2005a, 2017a, 2019a, b).

2 Material and methods

The material treated in this study is deposited in the following public and private collections:

HNHM Hungarian Natural History Museum, Budapest (Gy. Makranczy)
IZAY Armenian National Academy of Sciences, Institute of Zoology, Yerevan (M. Kalashian, T. Ghrejyan, G. Karagyan)
MCSNV Museo Civico di Storia Naturale di Verona (L. Latella)
MNB Museum für Naturkunde Berlin (J. Frisch)
MZH Finnish Museum of Natural History, Zoological Museum, Helsinki (J. Mattila, J. Muona)
NHMW Naturhistorisches Museum Wien (H. Schillhammer)
NMP National Museum of Natural History, Praha (J. Hájek)
cAss private collection Volker Assing, Hannover
cBra private collection Volker Brachat, Geretsried
cFel private collection Benedikt Feldmann, Münster
cKhn Khnzorian collection (currently in private collection Mark Kalashian, Yerevan)
cKoc private collection Matúš Kocian, Prague
cMey private collection Heinrich Meybohm, Großhansdorf
cPut private collection Volker Puthz, Schlitz
cSch collection M. Schülke (MNB)
cWun private collection Paul Wunderle, Mönchengladbach
cZan private collection Adriano Zanetti, Verona

Most of the material collected by the authors is housed in cAss and cSch; the Pselaphinae and Scydmaeninae collected by the first author are deposited in cBra and cMey, respectively, reference material of other groups also in MNB, NHMW, cFel, and cPut. With few exceptions in cAss, cPut, and cSch, specimens found by Harald Schillhammer and Helena Shaverdo are in NHMW. The material from the field trip conducted by M. Kocian is mainly in cKoc, partly also in cAss and cSch. The trap catches made available by J. Müller and T. Ghrejyan is deposited mostly in cAss, partly in IZAY, MNB, and cSch.
Body length was measured from the anterior margin of the mandibles (in resting position) (most subfamilies) or the anterior margin of the clypeus (Tachyporinae, Oxytelinae) to the posterior apex of the elytra (Scydmaeninae) or to the abdominal apex (other subfamilies), the length of the forebody from the anterior margin of the mandibles (in resting position) to the posterior margin of the elytra, head length along the middle from the anterior margin of the clypeus (without ante-clypeus) (Aleocharinae) or from the anterior margin of the frons to the posterior constriction of the head (other subfamilies), elytral length at the suture from the apex of the scutellum to the posterior margin of the elytra (if not indicated otherwise), and the length of (the median lobe of) the aedeagus from the apex of the ventral process to the base of the aedeagal capsule. The "parameral" side (i.e., the side where the sperm duct enters) is referred to as the ventral, the opposite side as the dorsal aspect.
The maps were created using Map Creator 2.0 (primap) software. Zoogeographic terminology is primarily based on Lattin (1967).
Labels of type material are cited in the original spelling, except that geographic and other data given in Cyrillic script are transliterated. Different labels are separated by slashes.
The taxonomic parts on Omaliinae (section 3.6.1), Pselaphinae (section 3.6.2) and Scydmaeninae (section 3.6.7) are exclusively authored by Adriano Zanetti, Volker Brachat, and Heinrich Meybohm, respectively, those on Tachyporinae and Oxytelinae (sections 3.6.4, 3.6.6) by the second author, and the remainder (sections 3.6.2, 3.6.5, 3.6.8, 3.6.9) by the first author.

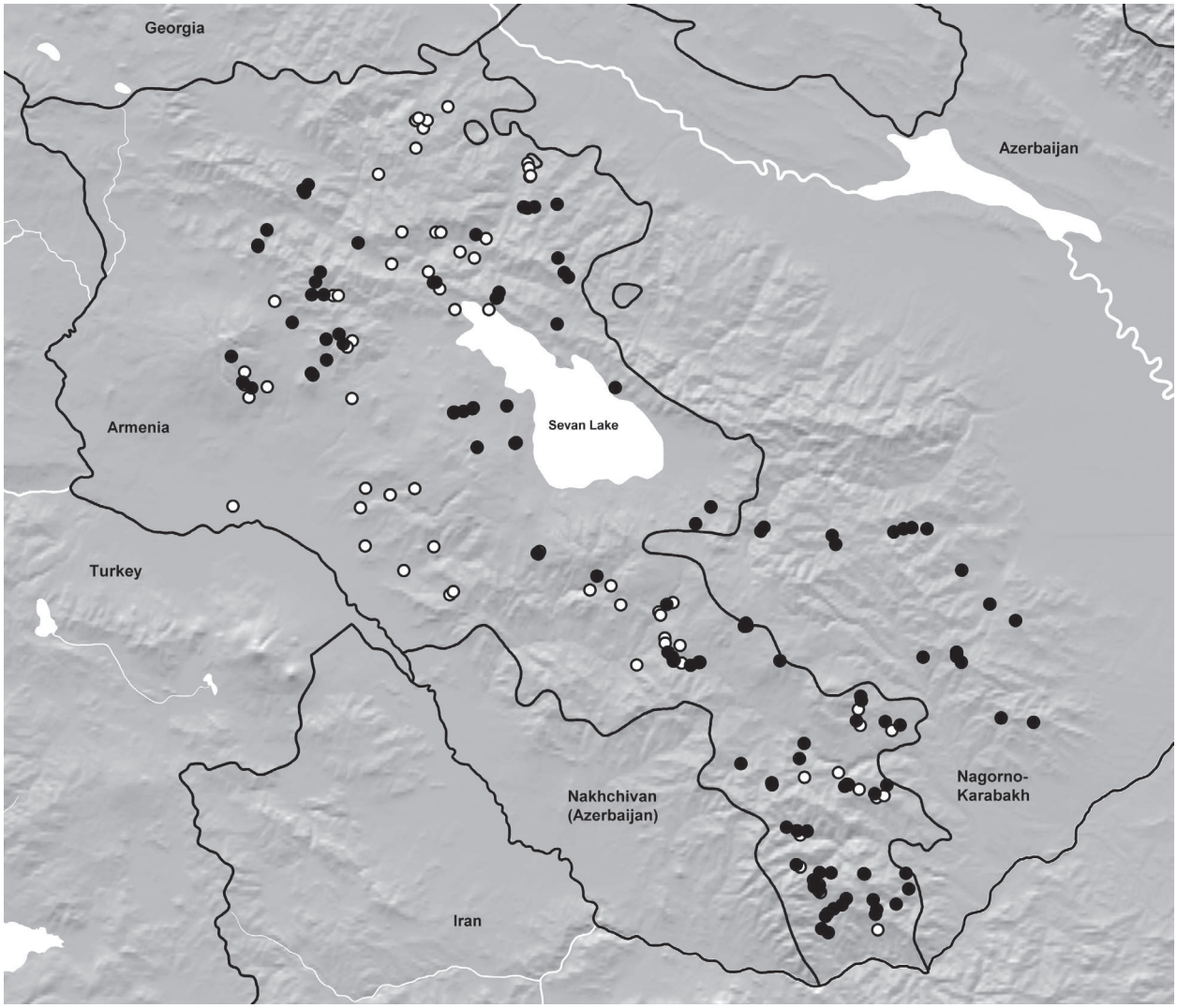
3 Results and discussion

3.1 Diversity

During the three field trips to Armenia and Nagorno-Karabakh conducted by the authors in 2016–2018, as many as 28,927 specimens of Staphylinidae were collected, 20,016 specimens in Armenia and 8,911 in Nagorno-Karabakh. Significant additional material from Armenia, approximately 2,000 specimens (or data of specimens), was contributed by Harald Schillhammer (hand-collected), Jörg Müller (hand-collected), Matúš Kocian (hand-collected), Mark Kalashian, Tigran Ghrejyan, and Gayane Karagyan (pitfall traps). Finally, some specimens collected by various collectors and deposited in several museums and private collections was studied. Hence, the present monograph is based on a total of approximately 31,000 specimens from numerous localities distributed across practically the whole of Armenia (Map 1).

In total, 525 named species were identified at the species level (Tab. 1). (To allow for better readability, the two subspecies of *Bolitobius castaneus* are treated as species in this section.) Approximately 1,900 specimens of *Mocyta* Mulsant & Rey, 1874 (currently under revision) and some 150 other Staphylinidae representing an unknown number of additional species were not named (see Tab. 2).

Disregarding doubtful or erroneous literature records of 99 species (see Tab. 3), 689 species in 15 subfamilies are currently known from Armenia and Nagorno-Karabakh, 675 from Armenia and 198 from Nagorno-Karabakh. As many as 262 species (39 % of total diversity) are reported from Armenia and 184 (92 %) from Nagorno-Karabakh for the first time (Tab. 1). *Atheta aequicollis*, a species previously known only from Mongolia, is even newly reported from the West Palaearctic region. Given the situation in 2016, the number of first records would be even significantly greater. However, the material of several taxa (*Mycetoporus* Mannerheim, 1830; *Sepedo-*



Map 1: Sampling localities in Armenia and Nagorno-Karabakh. Black circles: authors´ field trips (localities 1–131); white circles: localities 132–213 (see Tab. 1).

philus Gistel, 1856; *Atheta* Thomson, 1858; *Geostiba* Thomson, 1858; *Leptusa* Kraatz, 1856; *Liogluta* Thomson, 1858; *Myllaena* Erichson, 1837; *Oxypoda* Mannerheim, 1830; *Lobrathium* Mulsant & Rey, 1878; *Quedius* Stephens, 1829) has already been treated and published in separate articles, including the descriptions of 28 new species, the vast majority of them in the genera *Geostiba* (nine species), *Atheta* (seven species), and *Oxypoda* (six species) (Assing 2016b, c, 2017a–c, e, 2018a, c, g, h, Assing & Vogel 2017, Schülke 2019a, b). Fourteen additional species – five of Scydmaeninae, four of Aleocharinae, two of Pselaphinae, and one each of Omaliinae, Proteininae, and Oxytelinae – are newly described in the present paper (see section 3.6), and the description of one species of Staphylininae is in preparation (Schillhammer in prep.).

With a total of 260 species (38 %), the Aleocharinae are, by far, the subfamily with the greatest diversity in the study region; in Armenia, they alone account for 254 (38 %) species and for 123 (47 %) first records, in Nagorno-Karabakh for 78 (39 %) species and 73 (40 %) first records. Other highly diverse subfamilies are the Staphylininae with 98 species in the study region (Armenia: 95; Nagorno-Karabakh: 30), followed by the Tachyporinae (total: 59 species; Armenia: 59; Nagorno-Karabakh: 28), the Oxytelinae (total: 59; Armenia: 58; Nagorno-Karabakh: 9), the Steninae (total: 50; Armenia: 50; Nagorno-Karabakh: 12), the Paederinae (total: 47; Armenia: 47; Nagorno-Karabakh: 8), the Omaliinae (total: 44; Armenia: 43; Nagorno-Karabakh: 12), and the Pselaphinae (total: 41; Armenia: 41; Nagorno-Karabakh: 7). The relative representation of first records in these subfamilies is primarily related to how well the respective groups and regions have been studied. In the Staphylininae, the new records account for 23 % of the diversity in Armenia and 97 % in Nagorno-Karabakh, in the Tachyporinae for 42 % in Armenia and 89 % in Nagorno-Karabakh, in the Oxytelinae for 42 % in Armenia and 78 % in Nagorno-Karabakh, in the Steninae for 18 % in Armenia and 75 % in Nagorno-Karabakh, in the Paederinae for 21 % in Armenia and 100 % in Nagorno-Karabakh, in the Omaliinae for 47 % in Armenia and 83 % in Nagorno-Karabakh, and in the Pselaphinae for 20 % in Armenia and 86 % in Nagorno-Karabakh.

The evident differences between the figures for Armenia and Nagorno-Karabakh are explained by the fact that the latter region is significantly smaller, less diverse, that next to nothing was known about its Staphylinidae fauna previously, and that it was sampled for only ten days in summer 2018.

A comparison of the total diversity and the systematic composition of the currently known Staphylinidae faunas of Armenia and Nagorno-Karabakh with those of the neighbouring or geographically close Georgia, the Russian South European territory, and Iran reveals some unsurprising similarities and differences, but also conspicuous discrepancies. Considering the much smaller territory, it was to be expected that Armenia and particularly Nagorno-Karabakh host fewer Staphylinidae species than any of the other regions. On the other hand, the data show that the fauna of Georgia and especially that of Iran have been very poorly studied. Based on preliminary studies and on revisions of certain genera such as *Geostiba*, *Leptusa*, and *Ischnosoma* (Assing 2016b, 2017b, c, Assing & Schülke 2017), the western parts of Georgia are rich in endemic species, and there is no reason to assume that even the eastern parts are less diverse than Armenia. Moreover, the area of Georgia exceeds that of Armenia by a factor of approximately 2.3. Based on these observations, it can be inferred that the difference in diversity between Georgia and Armenia should be significantly greater than that indicated in Tab. 4. And Iran, with its vast territory of about 55 times the size of Armenia and 144 times the size of Nagorno-Karabakh and with its topological, geological, and climatic diversity should be expected to host at least twice as many species as Armenia and Nagorno-Karabakh combined. The high proportion of paederines and the low percentage of omaliines in Iran compared to the figures for the other countries and regions is explained with its more southern geographic position; paederines tend to be more, and omaliines less speciose in warmer climates.

Another remarkable observation is that the proportions of aleocharine species in Armenia and Nagorno-Karabakh are about 1.5 times as great as that in Georgia and about twice as great as those in the Russian South European territory and in Iran. The most speciose and at the same time believed to be the most difficult to identify and consequently the least studied of all subfamilies (with the lowest number of active specialized taxonomists), the Aleocharinae of Armenia and Nagorno-Karabakh can now be considered to be better known than those of other countries of the Caucasus region sensu lato. With 67 identified and an unknown number of unidentified species, the genus *Atheta* is particularly diverse in Armenia and alone accounts for approximately 10 % of the total diversity.

A general conclusion that can be drawn from the data in Tab. 4 is that the known species inventories of all Caucasian countries and regions are far from complete. This is revealed when the species numbers are compared to those of small countries like Denmark (area only slightly greater than that of Armenia; 1045 species) and Belgium (area even smaller than that of Armenia; 1303 species) without endemics, with a much lower habitat diversity (e.g., without mountains), and with a history of glacial devastation on the one hand, but a long entomological tradition on the other. Long-term studies, including thorough sampling of certain habitats such as dead wood, decaying matter of all kinds (compost, dung, etc.), riparian and other wetland habitats, nests of ants, mammals, and birds, and especially also using methods suitable for collecting flying insects (car-nets, window traps) or for recording species with short epigeic dispersal activity (pitfall traps) will undoubtedly result in a

significant increase of species numbers. This is particularly true of the fauna of Nagorno-Karabakh, whose fauna is still characterized by pronounced gaps in all subfamilies, particularly in the diversities of Oxytelinae and Pselaphinae.

3.2 Checklist

The below checklist (Tab. 1) summarizes the results of various field trips to Armenia and one to Nagorno-Karabakh, pitfall and window trap studies in Armenia, identification of additional material from other collections, as well as previous literature records. In the localities column, the number of specimens is given in parentheses behind the locality/sample number (or letter). For most of the species that had been recorded from Armenia and Nagorno-Karabakh prior to the present study, primary records are indicated in the references column. Secondary catalogue records (e.g., Herman 2001, Löbl & Besuchet 2004, Schülke & Smetana 2015) or general records provided in keys are included only when primary records were not found. In some cases (especially Steninae and Pselaphinae), such catalogue records are in fact the primary records. Literature records that are at least likely to be based on misidentification, old records of species that are reliably identified only based on an examination of the primary sexual characters, records (most likely) erroneously attributed to the territory of modern Armenia, and records that are doubtful for other reasons are discarded and listed in Tab. 3. Some doubtful literature records were included, however, when the presence of a species in Armenia and/or Nagorno-Karabakh was confirmed based on recently collected material. Recently collected material that could not be reliably identified at the species level for various reasons is listed separately in Tab. 2.

Tab. 1: Checklist of the Staphylinidae recorded from Armenia and Nagorno-Karabakh. For footnotes and details on the localities, samples, and references see the respective sections below the checklist.
Symbols and abbreviations: ●: first record(s) from Armenia or Nagorno-Karabakh, respectively; ○: previously recorded from Armenia or Nagorno-Karabakh, respectively; #: recorded only from Armenia, but taxonomic and zoogeographic status doubtful; *: recorded only from Armenia, but probably more widespread; **: regionally endemic (distribution most likely confined to particular regions or mountain ranges within Armenia and/or Nagorno-Karabakh; *** locally endemic to individual mountains. AR = Armenia; NK = Nagorno-Karabakh.

Species	AR	NK	Localities/Samples	References
Omliinae				
Anthobium atrocephalum (Gyllenhal, 1827)	●	●	22(9), 33(1), 39(2), 63(1), 66b(1), 70(4), 72(2), 89b(2), 100(1)105(2), 108(4), 110(2), 112(3), 113(5), 114(27), 121a(1), 122(30), 126(3), 133(1), 135(1), 136(1), 151(1)	
Anthobium fusculum (Erichson, 1839)	○		122(105), 165(3)	A05a, IK61 [6)]
Anthobium hamatum (Luze, 1905)	○	●	70(1), 114(1)	L05
Anthobium tenue (Eppelsheim, 1881)	●		170(6)	
Anthobium unicolor (Marsham, 1802)	●		181(1)	
Arpedium brachypterum (Gravenhorst, 1802)	●		67(31)	
Dialycera minuta Luze, 1906	○			IK59 [60)]
Dropephylla caucasica (Kolenati, 1846)	○	●	42(3), 48(20), 49(2), 49a(3), 53(1), 116(2)	ATHW0, IK64b
Dropephylla elisabethae Jászay & Hlaváč, 2006	○			JH06
Dropephylla vilis (Erichson, 1840)	○		30(1), 121a(1)	J08
Eusphalerum celsum (Luze, 1910) [1)]	●		8(1), 68(1), 68a(1)	
Eusphalerum fidele (Luze, 1910) [1)]	●		5a(6), 6(2), 7(8), 11(2), 53(1), 63(2), 75(1), 97(1)	
Eusphalerum primulae (Stephens, 1834)	●		135(1), 136(2)	

Species	AR	NK	Localities/Samples	References
Eusphalerum sareptanum (Eppelsheim, 1878)	○	○	114(1)	Z93
Eusphalerum sorbi (Gyllenhal, 1810)	●		6(3), 7(2), 8(2), 69(1)	
Eusphalerum transcaucasicum (Bernhauer, 1902)	○	●	30(1), 31(3), 33(11), 53(3), 58(2), 59(6), 66(3), 66a(1), 71(1), 96(1), 100(2), 101(2), 102(2), 105(8), 106(4), 119(4)	Z93
#*Geodromicus armeniacus* Iablokoff-Khnzorian, 1989	○			IK89
Geodromicus brevicollis Fauvel, 1857	●		18(3), 45(2), 46(10), 49(25), 67(18), 85(25), 85a(24), 85c(8), 86(3), 94(1), 128(5), j(2), k(1), l(1)	ASpp
Geodromicus constricticollis Eppelsheim, 1890	○			IK62
Geodromicus convexus Iablokoff-Khnzorian, 1962	○			IK62
Geodromicus major (Motschulsky, 1860)	●		m(1)	
#*Geodromicus rivularis* Iablokoff-Khnzorian, 1962	○			IK62
Geodromicus striatus Iablokoff-Khnzorian, 1962	○			IK62
Lesteva longoelytrata longoelytrata (Goeze, 1777) [1)]	○		18(7), 49(15), 63(1), 67(1), 76(2), 77(1), 139(14)	
Lesteva monticola Kiesenwetter, 1847	●		204a(1), 213(3)	
Lesteva punctata Erichson, 1839	●	●	20a(1), 63(5), 66a(8), 73(1), 107(3), 112(8), 130(7)	
Mannerheimia brevipennis (Motschulsky, 1860)	○		11b(1)	A05a, IK56 [7)]
**Olophrum aragatzense* Iablokoff-Khnzorian, 1962	○		49(2), 67(82)	A05a, IK62
Olophrum caucasicum Fauvel, 1875	●	●	7(2), 8(1), 9(1), 22(7), 36(2), 66(1), 98(4), 112(1), 113(1), 114(1), 115(1), 116(8), 121a(1), 126(1)	
Olophrum puncticolle Eppelsheim, 1880	○			A05a [2)], A17a, IK64a [2)]
Omalium caesum Gravenhorst, 1806	○	●	8(2), 9(37), 19(1), 41(1), 42(5), 48(38), 49(13), 50(7), 63(1), 67(3), 76(13), 77(2), 81(7), 81a(8), 82(1), 82a(1), 83(1), 105(6), 114(16), 118(1), 126(6), 136(2), 149a(1), 165(1)	E30
**Omalium kociani* Zanetti, spec. nov.	●		165(9)	
Omalium littorale Kraatz, 1857	●	○	4(2), 7(2), 9(85), 9a(9), 16(1), 19(1), 22(83), 23(1), 30(27), 31(7), 33(1), 35(1), 39(6), 41(1), 42(3), 48(80), 49(17), 49a(44), 54(33), 55(1), 57(17), 60(2), 64(1), 67(3), 70(7), 76(100), 81(1), 88(2), 89(8), 89b(21), 91(1), 95(1), 107(1), 108(1), 113(3), 114(38), 116(24), 117(1), 118a(1), 133(2), 135(1), 136(3), 170(5), 172(2)	Za02

Species	AR	NK	Localities/Samples	References
Omalium rivulare (Paykull, 1789)	●	●	1(1), 8(1), 9(8), 11(2), 11b(2), 12(1), 13(1), 33(3), 39(4), 41(1), 42(1), 48(13), 49(5), 63(6), 67(2), 74(2), 81(3), 81a(2), 82(5), 93(1), 114(3), 116(3), 133(3), 135(10), 136(23), 148(1), 149a(3), 149b(1), 150(13), 151(44), 155(3), 167(1), 170(2), 172(2)	
Omalium rugatum Mulsant & Rey, 1880		●	105(6)	
Omalium turcicum Smetana, 1967	●		4(1), 7(10), 13(3), 15(1), 33(3), 39(9), 82(2), 121a(2), 126(8), 131(5), 165(4)	
Omalium wunderlei Zanetti, 2002	●		122(22)	
Phloeonomus minimus (Erichson, 1839)	●		148d(1)	
Phloeonomus punctipennis Thomson, 1867	●		68a(1)	
Phloeostiba plana (Paykull, 1792)	○		135(11), 148b(6), 148c(1), 149b(3), 150(2), 156(4)	ATHW00
Phyllodrepa floralis (Paykull, 1789)	○	●	50(1), 114(2)	E30
Phyllodrepa nigra (Gravenhorst, 1806)	●		68(1)	
Xylodromus affinis (Gerhardt, 1877)	○		9a(1), 22(7), 48(1), 54(1), 64(1), 68(1), 76(6), 82a(1), 135(3)	ATHW00
Xylodromus depressus (Gravenhorst, 1802)	●		54(1)	
Proteininae				
Megarthrus depressus (Paykull, 1789)	●	●	63(1), 105(10)	
Megarthrus stercorarius Mulsant & Rey, 1878	●	●	11b(1), 88(2), 63(6), 105(14)	
Metopsia similis Zerche, 1998	●	●	35(8), 66(4), 98(1), 112(2), 113(1)	
Proteinus atomarius Erichson, 1840	●	●	63(1), 69(1), 105(28)	
Proteinus baculatus Assing, spec. nov.	●		8(1), 118(1)	
Proteinus brachypterus (Fabricius, 1792)		●	114(1)	
Micropeplinae				
Micropeplus marietti Jacquelin du Val, 1857		●	114(4), 116(1)	
Dasycerinae				
Dasycerus crenatus Motschulsky, 1839	●		70(1)	
Pselaphinae				
Batrisodes delaporti (Aubé, 1833)	○			E30
Batrisodes ruprechtii (Kolenati, 1846)	○	●	66b(2), 98(3)	LB04
Bibloplectus pusillus (Denny, 1825)	○			LB04

Species	AR	NK	Localities/Samples	References
Bibloporus variicolor Reitter, 1882	●		53(1)	
Brachygluta araxidis (Reitter, 1889)	○			SBBB04
Brachygluta fossulata (Reichenbach, 1816)	○			LB04
Brachygluta furcata (Motschulsky, 1835)	○			SBBB04
Brachygluta nodosa (Motschulsky, 1835)	○	●	36(22), 42(1), 66(4), 66a(4), 90(8), 90a(4), 103(22), 104(3), 109(36), 110(3), 114(1), 121a(5)	SBBB04
Brachygluta spinicoxis spinicoxis (Motschulsky, 1835)	○			LB04
Brachygluta sursicauda (Ganglbauer, 1900)	●		b(1)	
***Bryaxis armeniacus* Brachat, spec. nov.**	●		4(21)	
Bryaxis bulbifer (Reichenbach, 1816)	○	●	6(4), 7(4), 9(23), 11(1), 11b(1), 15(7), 19(8), 20(7), 22(30), 31(12), 33(3), 36(10), 39(31), 40(7), 48(4), 51(2), 52(1), 53(26), 55(15), 56(2), 57(4), 61(4), 62(11), 63(19), 66(5), 66a(6), 66b(9), 68(15), 68a(4), 69(6), 70(4), 72(8), 75(1), 76(6), 81(4), 81a(10), 82(2), 89(32), 89b(45), 90(12), 97(2), 100(11), 101(10), 103(5), 105(17), 106(2), 107(2), 109(3), 110(1), 112(2), 117(9), 119(38), 121(1), 125(7), 126(27), 127(8), 130(6), 131(18), 199a(7)	BK07
Bryaxis clavipes (Motschulsky, 1851)	○	○	5(1), 55(10), 63(1), 66a(22), 66b(12), 70(2), 72(2), 81(5), 81a(1), 88(3), 98(6), 100(8), 101(7), 102(11), 105(25), 108(2), 109(4), 110(74), 112(2), 113(8), 114(6), 117(4)	BK07
Bryaxis longipalpis (Motschulsky, 1835)	○		5(1), 6(1), 51(1), 58(26), 66b(3), 70(6), 72(1)	BK07
***Bryaxis meghruicus* Brachat, spec. nov.**	●		120(1)	
Bryaxis rostratus (Motschulsky, 1845)	○			BK07
***Bryaxis seductus* Besuchet & Kurbatov, 2007**	○		30(25), 31(1), 121(1)	BK07
Bythinus gracilis Motschulsky, 1851	○	●	22(1), 33(13), 36(3), 39(8), 40(2), 50(2), 52(3), 53(3), 61(6), 63(14), 66a(5), 66b(6), 70(3), 81(1), 81a(3), 88(21), 89(3), 89b(1), 90(1), 103(1), 105(4), 109(2), 110(8), 199a(7)	LB04
Chennium prometheus Saulcy, 1875	○			LB04
Claviger colchicus Motschulsky, 1837	○			LB04
Ctenistes marthae Reitter, 1891	○			LB04
Ctenistes palpalis Reichenbach, 1816	○			LB04
Enoptostomus globulicornis (Motschulsky, 1851)	○			E30

Species	AR	NK	Localities/Samples	References
Euplectus piceus piceus Motschulsky, 1835	○			LB04
Plectophloeus nubigena nubigena (Reitter, 1877)	●		31(1), 70(1)	
Pselaphus acuminatus Motschulsky, 1835	○		121(3), 121a(9), 212(1)	LB04
Pselaphus caspicus Reitter, 1882	●	●	66(1), 66a(1), 98(1), 100(1)	
Pselaphus caucasicus Motschulsky, 1845	○			LB04
Rybaxis longicornis (Leach, 1817)	○	●	66(1), 66a(5), 100(1), 105(1)	LB04
Trimium caucasicum Kolenati, 1846	○		53(2), 66(2), 66b(4), 70(2)	LB04
Trissemus antennatus serricornis Schmidt-Göbel, 1838	●		66a(1)	
Trissemus melinus (Solsky, 1869)	○		177(1)	LB04
Trissemus montanus (Saulcy, 1876)	○		130(1)	LB04
Tychus armeniacus Saulcy, 1878	●		66(1)	
Tychus colchicus Saulcy, 1878	○		9(1), 19(2), 81a(1)	LB04
Tychus dubius Sabella & Kurbatov, 2002	●		88(1)	
Tychus guillebeaui Raffray, 1904	○		54(1), 89(2)	LB04
Tychus lederi Saulcy, 1878	○			LB04
***Tychus milvus* Sabella & Kurbatov, 2002	○			SKu02
Tychus remaudierei Besuchet, 1969	○			SKu02
**Zibus yunonae* Kalashian, 1990	○			Kn90
Tachyporinae				
Bolitobius castaneus boreomontanicus Schülke, 2010	●	●	82(1), 108(1), 112(1), 148d(1), 148f(1), 152(1)	
Bolitobius castaneus castaneus (Stephens, 1832)	●		11(1), 11b(1), 47(5), 68(1)	
Bryophacis rugipennis (Pandellé, 1869)	●		o(1)	
Bryoporus cernuus (Gravenhorst, 1806)	●		59(1), 97(1), 150c(2), 152(1), p(1)	
Bryoporus multipunctus Hampe, 1867	●		148d(1), q(1)	
Cilea silphoides (Linnaeus, 1767)	○			Sm04
Ischnosoma longicorne (Mäklin, 1847)	○	●	1(1), 2(3), 3(5), 3a(1), 6(4), 7(5), 11(1), 22(6), 42(1), 47(2), 50(3), 51(2), 52(3), 53(1), 64(1), 66b(1), 72(1), 75(8), 77(1), 88(10), 89(7), 90(1), 91(3), 98(6), 100(3), 110(4), 112(25), 114(1), 118(2), 119(1), 125(1), 127(18), 129(1), 131(1), 148b(1), 148d(1), 148f(2), 152(1), 154(1), 164(4), 167(1), 170(1), 199a(4)	Sm04

Species	AR	NK	Localities/Samples	References
Ischnosoma splendidum (Gravenhorst, 1806)	●	●	1(3), 4(5), 6(2), 7(4), 9(9), 19(2), 36(3), 39(4), 48(8), 54(1), 67(1), 76(2), 78(3), 82(1), 89(1), 90a(2), 92(2), 103(1), 104(1), 117(1), 118(2), 119(1), 125(1), 126(22), 130(7), 150d(1), 152(1), 165(1), 199a(4)	
Lamprinodes haematopterus (Kraatz, 1857)	●		126(1)	
Lamprinus erythropterus (Panzer, 1796)	○			Sc08
Lordithon bimaculatus (Schrank, 1798)	●		60(1), r(1)	
Lordithon rostratus (Motschulsky, 1860)	●		55(1), 57(29), 58(1), 60(!), 69(4), 135(1), 148d(1), 153(1), s(1)	
Lordithon thoracicus thoracicus (Fabricius, 1777)	●	●	4(7), 6(8), 7(3), 11(2), 11a(1), 11b(3), 19(1), 30(1), 31(1), 36(2), 39(8), 50(2), 51(1), 53(22), 55(28), 57(6), 60(1), 61(2), 62(1), 63(2), 66a(2), 66b(1), 67(1), 68(2), 68a(1), 69(3), 70(7), 74(1), 72(1), 76(3), 82(3), 89(4), 92(3), 100(2), 101a(1), 103(1), 105(1), 112(2), 116(1), 118(3), 118a(1), 119(10), 121a(2), 121a(2), 125(6), 126(36), 142(1), 143(1), 148b(1), 148d(2), 148e(1), 150(2), 153(1), 155(2), 206(1), t(1)	
Lordithon trimaculatus (Fabricius, 1792)	●		136(4), u(1)	
Mycetoporus bimaculatus Lacordaire, 1835	○		20(1)	Am00 [3)], Sm04
Mycetoporus corpulentus Luze, 1901	●		4(4), 7(1), 11(1), 52(1), 57(1), 90(1), 131(1)	
Mycetoporus eppelsheimianus Fagel, 1968	○	●	1(1), 11(4), 11a(4), 11b(2), 12(11), 13(1), 76(2), 77(1), 81(2), 82(1), 92(1), 116(5), 118(1), 119(1)	Am00 [4)]
Mycetoporus forticornis Fauvel, 1875	●		88(1), 172(1)	
Mycetoporus glaber glaber (Sperk, 1835)	○		49a(1), 50(7), 52(1), 73(2), 76(4), 78(2), 151c(1)	SK00
Mycetoporus imperialis Bernhauer, 1902	●	●	32(1), 33(1), 39(1), 112(2), 118(1), 121(1), 126(6), 130(1), 148d(1), w(1)	
Mycetoporus lepidus (Gravenhorst, 1806)	○			Sm04
Mycetoporus liliputanus Luze, 1901	●	●	26(1), 35(2), 77(1), 78(3), 79(51), 81(3), 81a(2), 98(1), 107(1), 110(1), 112(1), 114(1), 116(2)	
Mycetoporus longulus Mannerheim, 1830	○		19(3)	Sc08
Mycetoporus monticoloides Schülke, 2019	●	●	7(1), 63(2), 103(1), 118a(1), 121a(1)	Sc19a
Mycetoporus phaedrus (Kolenati, 1846)	●	○	4(1), 9(5), 10(2), 12(2), 13(16), 19(1), 22(4), 23(2), 30(1), 31(1), 32(1), 39(19), 40(1), 41(1), 47(1), 48(10), 49(2), 49a(2), 50(4), 57(2), 70(3), 71(2), 74(5), 76(2), 77(1), 82(1), 83(3), 88(6), 89(10), 89b(8), 101(1), 103(2), 107(2), 108(1), 110(3), 114(3), 116(26), 118(2), 118a(1), 119(1), 126(16), 129(1)	Ko46, Sc19a

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Mycetoporus praetextoides Schülke, 2019	●	●	4(1), 13(1), 32(66), 39(25), 74(2), 78(2), 79(2), 89b(2), 103(3), 109(1), 110(1), 114(7), 118(1), 118a(2), 121(1), 126(11)	Sc19a
Mycetoporus punctus (Gravenhorst, 1806)	○		4(2), 11(1), 30(2), 48(1), 88(2), 126(1), 127(1), 159(1), x(1)	Sc08
Mycetoporus reichei (Pandellé, 1869)	○	●	3a(3), 11(8), 11a(1), 11b(34), 12(11), 13(3), 15(4), 22(8), 22a(5), 26(2), 30(5), 31(3), 32(36), 33(7), 34(3), 39(8), 45(1), 50(7), 50a(1), 53(8), 66(1), 66b(2), 67(1), 68(2), 70(2), 73(2), 75(2), 75(1), 76(2), 77(17), 78(8), 79(17), 81(1), 81a(1), 82a(2), 83(1), 88(5), 89(2), 89b(7), 91(4), 95(4), 96(1), 97(3), 103(1), 105(2), 108(1), 110(1), 112(9), 113(1), 116(2), 121(1), 118(5), 118a(1), 119(1), 121(1), 124(2), 125(5), 126(6), 127(5), 129(8), 131(6), 146a(2), 164(4), 165(1), 167(2), 172(2), y(1)	Sm04
Mycetoporus silvaticus Iablokoff-Khnzorian, 1962	○	●	32(1), 42(1), 50(1), 66(1), 67(2), 74(1), 75(1), 76(3), 77(1), 89(3), 89b(3), 96(1), 109(2), 110(1), 112(3), 113(2), 118a(3), 119(2), 121(1), 122(1), 126(1), 129(1), 180(1), v(1)	IK62
Parabolitobius inclinans (Gravenhorst, 1806)	○	●	114(1), 155(2)	Sm04
Sepedophilus immaculatus (Stephens, 1832)	●	●	6(1), 7(4), 11a(1), 60(1), 62(1), 66a(1), 68(1), 75(1), 89(1), 91(1), 96(1), 98(1), 105(1), 113(1), 119(2), 127(3), 131(1), 150b(2), 153(1), 163(2)	
Sepedophilus obtusus Luze, 1902	●	●	1(1), 3a(3), 6(3), 7(1), 11(3), 11a(5), 11b(4), 13(1), 26(35), 32(8), 39(1), 40(1), 41(1), 47(1), 53(9), 57(1), 58(1), 66(1), 77(1), 79(1), 81(1), 88(15), 89(1), 91(1), 92(7), 93(1), 95(3), 96(6), 97(1), 98(5), 100(2), 101(2), 103(2), 104(1), 106(1), 112(2), 118(2), 118(6), 119(1), 121a(3), 125(1), 126(21), 129(2), 130(1), 131(1), 135(1), 138(1), 150(3), 150c(2), 151d(1), 162(2), 163(5), 164(8), 167(2), 168(3), 170(1), 180(3)	
Sepedophilus rufulus (Hochhuth, 1849)	○	●	4(1), 9(1), 11b(2), 20(1), 22(12), 22a(1), 30(1), 35(1), 39(1), 43(1), 50a(1), 64(3), 76(1), 78(1), 79(1), 80b(1), 81a(1), 88(2), 112(2), 116(1), 118a(1), 121(1), 121a(1), 126(12), z(1), aa(1)	Sm04
Sepedophilus testaceus (Fabricius, 1792)	○		66b(1), 68a(1)	E30
Sepedophilus transcaucasicus Schülke, 2019	●	●	12(4), 26(1), 30(2), 32(10), 36(1), 53(2), 66(1), 78(2), 79(1), 88(2), 91(1), 92(3), 98(8), 100(1), 101(1), 103(3), 105(1), 109(1), 112(18), 113(1), 121(1), 126(15), 180(1), 196a(1), 199a(1)	
Tachinus caucasicus Kolenati, 1846	○	○	135(1), 136(1), 148a(2), 148b(3), 148c(2), 154(1), ab(1)	Sm04, U75 [64)]
Tachinus cingulatus Solsky, 1864	○			Sc89
Tachinus corticinus Gravenhorst, 1802	○	●	4(1), 6(1), 7(1), 16(1), 15(6), 19(5), 20(2), 22a(1), 41(5), 43(3), 49(7), 49a(1), 53(24), 54(2), 59(2), 70(2), 77(5), 78(2), 82(2), 82a(5), 89(1), 93(1), 98(1), 101(2), 103(2), 118(1), 121a(3), 142(1), 145(1), 160(1), 180(2)	Am99a

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Tachinus discoideus Erichson, 1839	○		204a(1)	Sm04
Tachinus elongatus Gyllenhal, 1810	○		16(1), 48(1), 49(1), 49a(1), 87(1)	U75
Tachinus fimetarius Gravenhorst, 1802	○		30(2), 39(2), 41(5), 48(22), 50(1), 76(3), 82(1)	IK59 [12)]
Tachinus humeralis marginicollis Kolenati, 1846	○	○		H01 [5)], Sc04
Tachinus laticollis Gravenhorst, 1802	○	●	4(2), 6(1), 7(3), 51(1), 53(3), 60(3), 62(1), 63(33), 68(2), 68a(4), 105(10), 148b(2), 148c(2), 156(1), 181(1), 196a(1), 199a(1)	Sc04
Tachinus nigerrimus Solsky, 1864	○		44(7)	U75
Tachinus pallipes Gravenhorst, 1806	○			Am99a
Tachinus rufipes (Linnaeus, 1758)	○	●	1(2), 3(1), 4(1), 6(1), 11(1), 11b(1), 19(1), 31(1), 39(1), 41(2), 45(3), 46(1), 49(3), 51(2), 63(2), 67(26), 68(2), 68a(1), 69(3), 77(1), 81(1), 105(1), 123(1), 126(3), 128(5),	Am99a
Tachinus rufitarsis Hochhuth, 1849	○		46(1), 48(1), 76(2), 126(1)	Am99a
Tachinus schneideri Luze, 1900	○			U75
Tachinus subterraneus (Linnaeus, 1758)	○		ab(2)	Am99a
Tachyporus atriceps Stephens, 1832	●	●	13(1), 16(2), 77(1), 78(5), 79(1), 88(2), 89b(2), 98(5), 104(1), 167(1)	
Tachyporus chrysomelinus (Linnaeus, 1758)	○		43(1), 47(1), 50(1), 64(10), 67(32), 76(3), 77(1), 82(1), 82a(2), 87a(1), 88(1), 168(1), 182(1), 186(1), 194(1)	Sm04
Tachyporus dispar (Paykull, 1789)	●		13(1), 16(2), 165(1)	
Tachyporus fascipennis Reitter, 1883	○			ATHW00
Tachyporus hypnorum (Fabricius, 1775)	○	●	22(232), 22a(9), 23(1), 30(104), 31(43), 32(8), 33(1), 34(13), 36(7), 39(21), 40(1), 55(4), 57(10), 58(24), 60(1), 66(1), 66a(1), 66b(1), 70(14), 71(7), 72(3), 76(1), 78(106), 79(7), 81(24), 81a(13), 82(1), 88(186), 89(92), 89b(173), 90(3), 90a(1), 91(16), 98(20), 101(1), 107(9), 108(29), 109(14), 110(43), 112(5), 113(7), 114(48), 115(6), 116(3), 117(2), 118a(1), 119(1), 121(2), 121a(1), 124(2), 125(4), 126(4), 127(16), 133(5), 135(1), 206(1), c(1), d(1)	Ko46 [58)]
Tachyporus nitidulus (Fabricius, 1781)	○	●	3a(1), 4(17), 9(4), 11b(1), 13(2), 15(3), 19(24), 20(3), 22(118), 22a(29), 30(13), 31(3), 32(1), 39(225), 40(9), 47(1), 48(1), 54(1), 55(1), 58(2), 59(4), 60(1), 61(1), 64(6), 69(1), 70(4), 71(4), 72(2), 76(6), 77(4), 78(17), 79(1), 81(5), 81a(14), 82(10), 82a(1), 88(52), 89(9), 89a(1), 89b(15), 90(1). 90a(1), 91(1), 94(1), 98(7), 100(1), 101(1), 102(1), 103(1), 104(3), 105(1), 107(1), 108(7), 109(4), 110(10), 112(5), 113(6), 114(41), 115(5), 116(16), 117(2), 119(14), 121(13), 121a(4), 124(1), 125(1), 126(117), 127(2), 130(5), 131(5), 133(3), 134a(1), 199a(1), 203a(2), 207(3)	ATHW00, E30

Species	AR	NK	Localities/Samples	References
Tachyporus obtusus (Linnaeus, 1767)	○			Sm04
Tachyporus pusillus Gravenhorst, 1806	●	●	36(36), 66(1), 81(2), 88(1), 98(1), 121a(8), 191a(1)	
Tachyporus scitulus Erichson, 1839	●	●	9(11), 9a(1), 12(1), 13(4), 15(1), 19(3), 21(1), 21a(1), 22(1), 35(1), 36(1), 39(6), 41(4), 45(1), 47(3), 48(7), 50(47), 50a(1), 53(5), 55(1), 64(37), 65(1), 66(2), 66a(1), 73(2), 74(2), 75(4), 76(7), 78(1), 80(1), 81(87), 81a(13), 82(2), 82a(3), 83(3), 84(2), 85a(30), 85c(4), 87(1), 87a(2), 88(15), 89(26), 89b(40), 90(1), 91(2), 95(6), 103(1), 105(1), 121(1), 121a(23), 126(23), 129(2), 130(5), 158(1), 160(2), 179(1), 204a(1)	
Tachyporus solutus Erichson, 1839	○	●	13(2), 22a(1), 30(3), 33(1), 34(1), 36(1), 48(1), 50(2), 66(1), 66b(1), 74(1), 75(1), 78(1), 81(1), 82(5), 88(2), 89(1), 92(1), 98(1), 104(1), 105(1), 112(3), 121(1), 126(3), 129 (3), 135(1), 186(1)	ATHW00
Habrocerinae				
Habrocerus capillaricornis (Gravenhorst, 1806)		●	98(5)	
Habrocerus simulans Assing & Wunderle, 1995	●	●	34(2), 35(2), 66(11), 66a(5), 66b(3), 88(11), 102(1), 112(11), 113(2), 121(2), 130(10), 163(1), 165(1), 167(3), 170(1), 199a(1)	
Aleocharinae				
Acrotona benicki (Allen, 1940)	●		66a(1), 74(2), 79(4), 80b(2), 81(10), 81a(27), 82(1), 89(5), 89b(3), 121a(1), 126(1)	
Acrotona concamerata Assing, 2004	●		76(30), 77(1)	
Acrotona exigua (Erichson, 1837)	●		19(22), 22(1)	
Acrotona muscorum (Brisout, 1860)	○	●	13(1), 19(1), 22(5), 22a(2), 30(1), 31(2), 35(1), 36(1), 39(2), 50(1), 63(1), 70(1), 72(1), 80b(3), 81(8), 81a(2), 88(2), 89b(1), 92(1), 95(1), 97(3), 98(3), 102(6), 103(5), 105(3), 107(1), 108(2), 109(1), 110(28), 112(1), 113(1), 114(22), 115(1), 116(11), 118(1), 119(2), 121a(2), 125(1), 126(5), 127(3), 130(2)	Be74
Acrotona obfuscata (Gravenhorst, 1802)	●	●	5(1), 33(4), 39(6), 100(17), 126(22), 180(1), 199(2), 199a(3)	
Acrotona parvula (Mannerheim, 1830)	●		44(10), 66(1)	
#Aleochara armeniaca Likovský, 1971	○			ATHW00, Li71
Aleochara bilineata Gyllenhal, 1810	○			ATHW00
Aleochara bipustulata (Linnaeus, 1760)	●	●	13(1), 18(1), 21(60), 21a(2), 24(1), 28a(1), 33(1), 77(1), 81(7), 81a(4), 82(1), 105(1), 106(1), 107(16), 109(1), 110(2), 113(1), 114(3), 120(1), 121(1), 122(7), 123(19), 124(1), 126(17), 212(1)	
Aleochara brevipennis Gravenhorst, 1806	○		77(1)	A18a

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Aleochara cephalotes Bernhauer, 1901	○			Li71
Aleochara conviva Eppelsheim, 1878	○		32(1), 165(5)	A18a
Aleochara cuniculorum Kraatz, 1858	○			ATHW00, Li71 [51)]
Aleochara curtula (Goeze, 1777)	●		121a(1), 151b(1), 155(55)	
Aleochara erythroptera Gravenhorst, 1806	○		89a(1)	A18a
Aleochara falcata Assing, 2009	○		17(18), 87(2)	A18a
#*Aleochara fugax* Iablokoff-Khnzorian, 1962	○			ATHW00, IK62, IK64b, Li71 [10)]
Aleochara grandeguttata Assing, 2009	○		7(3), 7a(1), 85b(1), 187(2)	A09a, A18a
Aleochara haematoptera Kraatz, 1858	○		8b(2), 36a(1), 120(5), 189(2), 210(1)	A17a, A18a, Am00 [8)], Li71 [9)]
Aleochara inconspicua Aubé, 1850	○	●	22(28), 22a(2), 26(1), 30(1), 31(3), 36(1), 39(7), 41(2), 69(1), 70(3), 81(4), 81a(1), 88(10), 89(17), 89b(23), 91(5), 110(3), 114(2), 116(4), 126(6), 165(1), 172(1)	A18a
Aleochara intricata Mannerheim, 1830	○		4(1), 8(1), 148(1), 151b(1)	SS15
Aleochara lanuginosa Gravenhorst, 1802	○	●	88(1), 115(5), 116(1)	A18a
Aleochara lata Gravenhorst, 1802	○			IK64b
Aleochara laticornis Kraatz, 1856	○		151a(1), 155(2), 157(1)	Li71
Aleochara maculata Brisout, 1863	○		16(1), 45(4), 58(1), 81a(1)	A18a
Aleochara maculipennis Baudi di Selve, 1857	○			Li71
Aleochara milleri Kraatz, 1862	○			E30
Aleochara moerens Gyllenhal, 1827	○		23(b), 148b(1)	A18a
Aleochara moesta Gravenhorst, 1802	○			E30
Aleochara morion (Gravenhorst, 1802)	●		8(1)	
Aleochara peeziana Lohse, 1961	○			Li71
Aleochara plicelytrata Assing, 2018	○	●	20a(2), 22(1), 31(1), 39(12), 53(2), 57(2), 58(1), 59(2), 75(1), 89(12), 89b(4), 90(1), 91(8), 100(16), 104(1), 105(4), 125(1), 126(10), 127(1), 156(2), 157(1)	A18a
**Aleochara polychroma* Iablokoff-Khnzorian, 1966	○			A17a, IK66
Aleochara sparsa Heer, 1839	○		54(1), 63(1), 75(1), 133(1), 135(23), 136(1), 145(17), 145a(36), 145b(66), 146(39), 146a(186), 148(3), 148b(17), 148c(3), 149(12), 149a(5), 149b(9), 150(6), 151(11), 155(22), 156(18)	ATHW00, Li71

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Aleochara subtumida (Hochhuth, 1849)	○	●	9(1), 30(1), 31(3), 33(2), 36(3), 39(1), 90a(1), 114(2), 115(1), 119(3), 125(3), 126(14), 155(1), 156(1), 170(1)	A18a, Am99b, IK60 [53)]
Aleochara tristis (Gravenhorst, 1806)	○			Ep78
Aleochara vagepunctata Kraatz, 1856	○			ATHW00, Li71
Aleochara verna Say, 1833	●		44(35), 49(1)	
Aleochara villosa Mannerheim, 1830	○			Li71
Alevonota gracilenta (Erichson, 1839)	○			AW08
Alevonota hepatica (Erichson, 1839) [65)]	●		6(1), 68(1), 133(1), 135(1)	
Alevonota libanotica (Fagel, 1965)	●		4(1), 58(1)	
Aloconota cambrica (Wollaston, 1855)	○		190(1), 199(1), 204a(1)	Be74
#Aloconota differens (Benick, 1974)	○			Be74
Aloconota gregaria (Erichson, 1839)	○		67(1), 161(1)	Sm04
Aloconota insecta (Thomson, 1856)	●	●	39(1), 100(1), 104(1), 112(1), 121a(2), 126(1), 130(3), 199a(17)	
Aloconota mediterranea (Thomson, 1856)	●		67(1)	
Amidobia talpa (Heer, 1841)	●		89a(1)	
Amischa analis (Gravenhorst, 1802)	●		5(1), 8a(1), 9(1), 16(1), 19(1), 20(10), 45(1), 46(1), 49(3), 54(2), 60(1), 67(1), 203a(1)	
Amischa bifoveolata (Mannerheim, 1830)	●		49(10), 49a(1), 54(3), 67(1), 73(1), 85a(1), 203a(1), 204a(1)	
Amischa decipiens (Sharp, 1869)	●		176(1), 201(1)	
Amischa forcipata Mulsant & Rey, 1873	●		196(1)	
Anaulacaspis caucasica (Fagel, 1969)	○		121(1), 121a(55), 130(32), 212(5)	A16a
Anaulacaspis nigra (Gravenhorst, 1802)	○	●	36(4, 66a(4), 99(1), 100(2), 103(2), 117(1)	A16a, E30
Anaulacaspis persica (Fagel, 1969)	○			A16a
**Atheta abscisa* Assing & Vogel, 2017	○	●	9(8), 10(2), 11b(1), 19(2), 20(1), 20a(1), 39(36), 67(1), 76(33), 77(6), 81(12), 81a(4), 83(1), 104(2), 126(268), 131(1)	AV17
Atheta aeneicollis (Sharp, 1869)		●	114(1), 115(1)	
Atheta aequicollis Benick, 1978		●	104(1)	
Atheta albomontis Assing, 2009	●		77(1)	
Atheta araxis Benick, 1943	○		5(2), 8(2), 9(3), 16(1), 18(3), 21(19), 21a(1), 32(1), 39(3), 45(11), 49(2), 76(4), 85c(1), 127(1), 139(1)	AV17 [67)], Be74 [67)]
Atheta atramentaria (Gyllenhal, 1810)	●	●	63(1), 82(1), 116(3), 177(1)	

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Atheta atricolor (Sharp, 1869)	●		44(1)	
Atheta benickiella Brundin, 1948	●	●	11a(5), 55(2), 60(1), 63(34), 89(1), 96(4), 114(1)	
Atheta bispinosa Assing & Vogel, 2017	○	●	1(1), 2(2), 3(1), 4(325), 5(33), 6(8), 7(20), 8(2), 11(4), 11a(2), 11b(18), 12(1), 16(1), 23(3), 28(1), 43(6), 51(19), 55(1), 57(3), 58(1), 63(10), 68a(3), 69(7), 74(9), 75(3), 76(13), 82(4), 82a(1), 83(1), 114(3), 118(3), 119(1), 125(4), 138(2), 165(4), 170(1), 172(1), 196a(1), 199a(5)	AV17
Atheta brevapicalis Assing & Vogel, 2017	○	●	15(1), 39(3), 82(5), 76(1), 77(2), 81(1), 104(1), 105(1), 119(1), 126(1), 130(4)	A18b, AV17
**Atheta brevitheca* Assing & Vogel, 2017	○		5(4), 9(12), 39(4), 47(2), 48(10), 50(1), 73(3), 74(24), 76(41), 82a(1), 83(1), 86(1), 93(13), 94a(1), 123(5), 126(5), 139(1), 165(1), 204a(1)	AV17
Atheta britanniae Bernhauer & Scheerpeltz, 1926	●		57(1), 69(1), 70(2), 133(3)	
Atheta castanoptera (Mannerheim, 1830)	○	●	6(4), 53(3), 55(2), 57(10), 62(1), 63(4), 66a(1), 69(15), 70(12), 92(1), 100(1), 102(1), 105(49), 106(2), 108(2), 112(4), 117(2), 118(1), 125(4), 135(7), 136(3), 144(3), 148(3), 148b(4), 148c(3), 149(3), 151(16), 170(8)	Ep78
Atheta cauta (Erichson, 1837)	●	●	44(12), 50(1), 81(1), 88(1), 108(1)	
Atheta chefsurica (Eppelsheim, 1880)	○	●	6(3), 60(1), 63(3), 105(15), 125(1)	A05a, IK60
Atheta crassicornis (Fabricius, 1792)	●	●	6(2), 11b(2), 39(3), 48(1), 53(2), 55(2), 64(9), 72(2), 98(1), 103(3), 118(1), 126(1), 135(1), 146(6), 148(2), 148c(7), 149(32), 149a(4), 149b(4), 150a(1), 151(2), 155(4)	
Atheta dadopora Thomson, 1867	●		6(1), 53(1), 63(2), 72(1), 135(2)	
Atheta debilis (Erichson, 1837)	○		9(1)	E30
Atheta dissimulans Assing, 2007	●		75(5)	
Atheta ebenina (Mulsant & Rey, 1873)	●	●	105(1), 126(1), 165(1)	
Atheta epirotica Benick, 1981	●		76(1), 82(4), 82a(1)	
#*Atheta erevanensis* Benick, 1985	○			Be74, Be85
Atheta fimorum (Brisout, 1860)	○			Be74
Atheta fuscicolor Benick, 1943	●	●	114(1), 126(1)	
Atheta gagatina (Baudi di Selve, 1848)	●	●	6(10), 98(1), 100(3)	
**Atheta hamulata* Assing & Vogel, 2017	○		9(39), 9a(6), 20a(1), 33(1), 39(2), 48(6), 67(2), 76(36), 77(4), 82(16), 82a(1), 93(2), 126(2)	AV17
Atheta hansseni Strand, 1943	○		11a(1), 11b(1)	AV17
Atheta harwoodi Williams, 1930	●		135(1)	
Atheta heymesi Hubenthal, 1913	●	●	89(1), 89b(1), 105(2), 118(6), 118a(2), 125(1)	
Atheta hybrida Sharp, 1869	●		135(32)	
Atheta indubia (Sharp, 1869)	●		66a(1)	
Atheta ischnocera Thomson, 1870	●		44(1)	
Atheta judith (Saulcy, 1865)	○		39(6), 126(5), 131(1), 162(7), 165(1), 167(5)	Be74

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Atheta knabli Benick, 1938	●		9(7), 39(1), 42(3), 48(2), 126(2)	
Atheta laevana (Mulsant & Rey, 1852)	●	●	103(4), 110(1), 126(1)	
Atheta laevicauda Sahlberg, 1867	●	●	4(3), 9(6), 10(1), 11(1), 13(2), 15(6), 20a(1), 21a(10), 22(1), 30(1), 31(1), 39(1), 43(1), 47(2), 48(1), 49(3), 50(4), 52(1), 74(2), 75(3), 76(2), 77(1), 78(1), 81a(1), 82a(1), 83(1), 88(3), 89(3), 89b(14), 90(1), 90a(1), 91(4), 103(2), 105(1), 116(1), 118(9), 118a(2), 119(3), 126(4), 127(2), 129(2), 131(3), 183(1)	
Atheta laevigata (Hochhuth, 1849)	●	●	32(4), 49(16), 60(1), 73(2), 76(2), 77(1), 82(1), 85c(1), 118(2), 125(3), 128(2), 181(1), 184(1), 203a(1)	
Atheta laticollis (Stephens, 1832)	○	●	36(1), 110(1)	E30
Atheta liturata (Stephens, 1832)	●		69(93), 70(1)	
Atheta longicornis (Gravenhorst, 1802)	●	●	22(3), 30(1), 39(1), 44(1), 70(1), 80b(4), 101(1), 107(1), 109(3), 113(1), 114(4), 116(1), 119(1), 121a(1), 126(19)	
Atheta luctuosa (Mulsant & Rey, 1853)	●	●	32(1), 39(2), 40(1), 114(7), 121(2), 126(2)	
Atheta lyciana Assing, 2003	●		11(1), 46(1), 49(3), 76(34), 77(2), 82(2), 83(1), 85c(3), 93(1), 128(54), 165(1)	
Atheta malleiformis Benick, 1975		●	114(2)	
***Atheta meghruica* Assing, spec. nov.	●		123(1), 126(2)	ASpp
Atheta melanaria (Mannerheim, 1830)	●	○	89b(1), 109(1), 116(1)	Ko46 [35)]
Atheta melanocera (Thomson, 1856)	●		16(1), 21(10), 85c(13)	
Atheta membranata Benick, 1974	●		53(1)	
Atheta monstruosa Assing, 2011	●		9(7), 10(1), 73(3), 76(4), 82(1), 127(1)	
Atheta nigra (Kraatz, 1856)	●		80b(1), 82(1), 126(1)	
Atheta nigritula (Gravenhorst, 1802)	●		6(2), 11(2), 57(3), 148b(9)	
Atheta occulta (Erichson, 1837)	○			Be74
Atheta pechlaneri Scheerpeltz, 1933	○		21a(2), 67(2), 86(6), 125(1)	AV17
Atheta putrida (Kraatz, 1856)	●		39(1), 118(1)	
Atheta ravilla (Erichson, 1839)	●		53(1), 60(8), 63(12), 64(2)	
Atheta scabens Assing, 2011	●		45(1)	
Atheta schneideri (Eppelsheim, 1889)	●		21(1), 39(3), 45(2), 73(12), 74(3), 81(2), 81a(1), 118(3), 122(7), 123(3), 126(5)	
**Atheta semialba* Assing & Vogel, 2017	○		5(1), 8(6), 20a(1), 21(8), 28a(1), 39(3), 45(9), 83(5), 94(1), 123(1), 126(1), 145b(1), 155(5)	AV17
**Atheta senticollis* Assing & Vogel, 2017	○		63(2), 144(3)	AV17
Atheta serrata Benick, 1938	●		118(1)	
Atheta singularis Bernhauer, 1912	○			Be74

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Atheta sodalis (Erichson, 1837)	●		6(1), 39(2), 75(1), 118(1), 121(8), 121a(2), 125(8), 126(1), 127(2), 153(1)	
Atheta subglabra (Sharp, 1869)	●		127(1)	
Atheta subsinuata (Erichson, 1839)	●		44(8)	
Atheta taxiceroides Munster, 1932	●		53(1), 72(1), 88(1)	
Atheta testaceipes (Heer, 1839)	●		88(1)	
Atheta tibialis (Heer, 1839)	○		16(1), 45(4), 46(10), 49(22), 67(29), 85a(5), 85b(1), 94(1)	Be74
Atheta triangulum (Kraatz, 1856)	○		39(1), 81(1)	Be74
Atheta trinotata (Kraatz, 1856)	●		86(1)	
Atheta volans (Scriba, 1859)	●		19(1), 20(1), 94(1)	
Atheta xanthopus (Thomson, 1856)	●		118(1)	
Autalia rivularis (Gravenhorst, 1802)	●	●	44(1), 105(1)	
****Bellatheta khustupica* Assing, spec. nov.	●		122(6)	
Bellatheta palata (Benick, 1970)	●		50(5)	
Bolitochara bella Märkel, 1844	●	●	98(3), 112(3), 134(1), 181(1)	
Bolitochara obliqua Erichson, 1837	○	●	1(1), 22(2), 39(1), 53(1), 68a(1), 71(2), 75(1), 92(1), 110(2), 112(1), 119(1), 126(5), 127(1), 135(2), 136(9), 138(1), 143(2), 144(2), 148(2), 150(1), 155(1), 156(2)	ATHW00
Bolitochara tecta Assing, 2014	○		68a(2)	A14
Boreophilia eremita (Rye, 1866)	●		7(1), 19(1), 20(1), 21(30), 67(6), 85c(8)	
Brachida exigua (Heer, 1839)	●	●	66(5), 79(3), 103(4)	
Brachyusa concolor (Erichson, 1839)	●		a(1)	
**Calodera alticola* Assing, spec. nov.	●		128(1)	
Cordalia obscura (Gravenhorst, 1902)	●		36(1), 199(1), 199a(15)	
Cousya araxis (Bernhauer, 1902)	○	●	50(4), 70(1), 88(1), 107(1)	A18e
Cousya nigrata (Fairmaire & Laboulbène, 1856)		●	114(1), 116(1)	
Cousya schuelkei Assing, 2007	○		5(1), 48(5), 49a(5), 122(1)	A18e
Cypha laeviuscula (Mannerheim, 1830)	●	●	81a(2), 95(1), 103(2), 118(1)	
Cypha spathulatha Assing, 2007	●	●	3(1), 98(1)	
Cypha takhtajani (Iablokoff-Khnzorian, 1961)	○			A05a, IK61
Cypha tarsalis (Luze, 1902)	●	●	11(2), 13(6), 33(1), 40(1), 76(1), 83(1), 98(1), 105(1), 119(3), 121a(1), 131(1), 165(7)	
Dinaraea aequata Erichson, 1837	●		134(1)	
Dinaraea linearis (Gravenhorst, 1802)	●		68a(2)	

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Dinarda hagensii Wasmann, 1889	●		89a(6)	
Drusilla canaliculata (Fabricius, 1787)	○	●	15(4), 19(29), 20(3), 20a(1), 30(1), 32(3), 33(1), 36(1), 40(2), 53(1), 64(10), 66(2), 73a(1), 78(7), 79(3), 80a(1), 88(10), 90(1), 101(2), 104(2), 109(1), 113(1), 114(2), 117(2), 126(9), 129(3), 130(2), 131(1), 148b(7), 150(1), 151a(2), 151b(3), 165(3), 167(1), 172(1), 183(2), 195(1)	Ca39 [38]
Encephalus complicans Stephens, 1832	●	●	7(1), 105(1)	
Falagria caesa Erichson, 1837	○		53(2), 58(1), 63(1), 177(1), 192(1)	Hh49
Falagria sulcatula (Gravenhorst, 1806)	○		58(1), 150(1), 203a(10)	E30
Falagrioma thoracica (Stephens, 1832)	●	●	33(1), 34(1), 35(8), 36(5), 39(1), 103(1), 112(9), 121(9), 121a(10), 126(8), 127(38), 130(22), 150b(2), 152(1)	
***Geostiba deliqua* Assing, 2018	○		118(3), 118a(1)	A18h
***Geostiba immutata* Assing, 2017	○		89(10), 89b(8)	A17b
***Geostiba kalavanica* Assing, 2017	○		53(13)	A17b
***Geostiba khnzoriani* Pace, 1983	○		22(1), 22a(3), 34(3), 78(1), 88(7), 121a(1), 129(2)	A17b, A18h, Pa83
***Geostiba meghruica* Assing, 2016	○		30(12), 31(1), 124(2)	A16b, A18h
***Geostiba nigrohortensia* Assing, 2018		○	105(96), 110(68), 113(1)	A18h
***Geostiba pambakica* Assing, 2016	○		5(10), 6(8), 8(3), 58(6), 70(39)	A16b, A17b
Geostiba sororcula Assing, 2001	○		1(2), 3a(14), 9(1), 19(68), 23(1), 41(25), 42(24), 43(1), 47(1), 48(22), 54(19), 76(1), 77(10), 80(1), 81a(18), 82(17), 121a(2)	A17b, A18h
***Geostiba tigrani* Assing, 2017	○		50(50), 51(32), 52(10), 57(3), 68(15), 69(16)	A17b
***Geostiba unicuneata* Assing, 2016	○	○	11(5), 11b(1), 81(1), 81a(2), 88(32), 105(16), 119(9)	A16b, A17b, A18h
***Geostiba unituber* Assing, 2016	○		4(23), 57(6), 62(1), 63(15),	A16b, A17b
Gnypeta carbonaria (Mannerheim,1830)	○			Sm04
#*Gnypeta gracilior* Benick, 1974	○			Be74
Gnypeta rubrior Tottenham, 1939	●		7a(1), 187(1), 190(1)	
Gyrophaena affinis Mannerheim, 1830	●	●	10(4), 11(5), 11a(10), 11b(136), 12(1), 22(23), 39(8), 53(7), 55(43), 62(1), 66(5), 66a(2), 69(8), 72(10), 75(2), 92(6), 101(1), 101a(1), 103(1), 105(177), 106(1), 108(1), 109(1), 110(3), 113(1), 119(7), 125(2), 126(2), 127(1)	
Gyrophaena bihamata Thomson, 1867	●	●	1(3), 4(2), 33(2), 51(2), 55(1), 58(3), 62(4), 66(3), 66a(2), 66b(6), 70(2), 98(2), 100(1), 191a(2), 100(1), 103(1), 105(113), 106(1), 109(1), 110(3), 112(1), 113(1), 117(2), 126(3)	
Gyrophaena boleti (Linnaeus, 1758)	○			E30

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Gyrophaena caucasica Strand, 1939	○		6(44), 34(1), 58(1), 66a(2), 142(2)	ES16
Gyrophaena gentilis Erichson, 1839	○	●	36(1), 50(1), 55(4), 60(8), 63(1), 105(3)	ES16
Gyrophaena hanseni Strand, 1946	●		70(5)	
Gyrophaena joyi Wendeler, 1924	○	●	62(1), 101(1), 102(1), 105(16)	ES16
Gyrophaena joyioides Wüsthoff, 1937	●		66a(3)	
Gyrophaena korbi Strand, 1939		●	98(1), 101a(2), 102(1)	
Gyrophaena manca Erichson, 1839	●		66a(1), 68a(6), 69(7)	
Gyrophaena nana (Paykull, 1800)	●	●	55(17), 66(1), 70(1), 97(4), 105(6), 136(1)	
Gyrophaena orientalis Strand, 1938	●		6(2)	
Haploglossa gentilis (Märkel, 1844)	●		148(1), 149a(2)	
Haploglossa picipennis (Gyllenhal, 1827)	○			ATHW00, IK64b
Haploglossa villosula (Stephens, 1832)	○		135(3)	ATHW00, IK57a, IK64b
#Hydrosmecta salsolae Benick, 1974	○			Be74
Ischnoglossa obscura Wunderle, 1990	●		59(1)	
Ischnopoda umbratica (Erichson, 1837)	●		190(1), 196(3), 199(4), 199a(1), 206(1)	
**[52]*Leptusa armeniaca* Pace, 1989	○		22(41), 30(113), 31(50), 32(14), 33(24), 35(11), 40(3), 85a(1), 118(5), 124(21), 125(8), 127(2)	A17c, Pa89
Leptusa fuliginosa (Aubé, 1850)	○	●	60(1), 68(1), 68a(1), 69(1), 98(1)	A17c, Pa89
Leptusa laeviuscula (Hochhuth, 1849)	○			Pa89
Leptusa venusta (Hochhuth, 1849)	○		6(2), 7(4), 52(1), 66b(2), 68a(1), 72(2), 135(1), 144(1), 148b(1)	A17c, Sz66a
**Liogluta armeniaca* Assing & Vogel, 2017	○	●	9(1), 22(5), 23(10), 28(9), 28a(1), 31(1), 32(1), 33(1), 39(12), 40(1), 46(7), 48(2), 49(1), 67(17), 68(5), 69(17), 70(1), 71(3), 73(3), 74(9), 81(1), 83(1), 110(2), 113(1), 114(1), 118(1), 119(4), 126(4)	AV17
Liogluta granigera (Kiesenwetter, 1859)	○		48(2), 51(1), 53(4), 55(58), 58(9), 60(13), 63(10), 68(8), 60(19), 70(3), 71(1), 72(1), 135(5), 136(1), 148(5), 148c(5), 149a(4), 199a(2)	AV17, Be74
Liogluta longiuscula (Gravenhorst, 1802)	●	●	57(3), 58(1), 90a(1), 106(1), 110(1), 126(1), 133(2), 148(1), 149(1)	
Liogluta microptera Thomson, 1867	●	●	4(1), 5(1), 6(1), 7(1), 48(2), 61(1), 62(1), 91(1), 105(1)	
Liogluta roettgeni (Bernhauer, 1903)	○			Be74
Lomechusa paradoxa Gravenhorst, 1806	○			ATHW00

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Lomechusoides strumosus strumosus (Fabricius, 1792)	○			ATHW00
Lomechusoides teres (Eppelsheim, 1884)	○			Sm04
Lyprocorrhe anceps (Erichson, 1837)	●		8a(2)	
Meotica exilis (Gravenhorst, 1806)	●	●	39(1), 73(1), 100(1)	
Meotica arasensis Smetana, 2004	●		40(1), 130(1)	
Mocyta fungi (Gravenhorst, 1806) [63]	●		1(8), 5(1), 9(2), 19(15), 21(1), 39(1), 76(1)	
Mocyta orbata (Erichson, 1837) [63]	●		9(1), 34(7), 35(18), 36(5), 39(11)	
**Myllaena ambulans* Assing, 2018	○		19(7), 20(17), 73(2), 76(1)	A18g
Myllaena caucasica Eppelsheim, 1880	○	○	7(1), 20(12), 20a(1), 23(1), 39(1), 63(4), 73(6), 76(1), 90a(1), 100(1), 112(17), 121(7), 121a(5), 125(23), 126(2), 127(5), 130(5), 181(1), 199a(1)	A18g
Myllaena infuscata Kraatz, 1853	○	○	63(1), 66a(3), 100(5), 112(1), 121a(1)	A18g
Myllaena intermedia Erichson, 1837	○		8b(7), 36(2), 180(2), 190(2), 199(1)	A18g
Myrmoecia reitteri (Eppelsheim, 1881)	○			A17a, IK60
Myrmoecia urartu (Iablokoff-Khnzorian, 1962)	○			A19a, IK62
Nehemitropia lividipennis (Mannerheim, 1830)	○	●	36(4), 103(1), 106(2)	E30
Ocalea alutacea Eppelsheim, 1878	●	●	68(16), 101a(1), 105(1)	
Ocalea badia Erichson, 1837	●		73(1)	
Ocalea dubia Motschulsky, 1860 [62]	●	●	58(4), 93(1), 100(4), 101(1), 101a(8), 105(7), 109(2), 110(14), 112(82), 113(75), 114(120), 115(10), 117(10), 148(1)	
Ocalea minor Eppelsheim, 1878	●	●	9(24), 11a(1), 19(1), 20(1), 22(1), 23(4), 32(18), 39(12), 45(1), 48(115), 49(12), 50(8), 73(17), 75(2), 76(50), 77(41), 81(3), 81a(11), 82(5), 83(2), 91(1), 100(1), 104(3), 105(19), 106(1), 109(1), 110(1), 112(2), 114(14), 116(2), 118(3), 119(2), 121(8), 121a(4), 123(1), 125(11), 126(17), 129(1)	
Oligota pumilio Kiesenwetter, 1858	●		48(2), 50(1), 70(1), 81a(4)	
Oligota pusillima (Gravenhorst, 1806)	●		1(1), 11a(1), 19(1), 39(8)	
Oxypoda abdominalis (Mannerheim, 1832)	○		1(1), 28(1), 36(1), 166(4)	A18c
Oxypoda alternans (Gravenhorst, 1802)	○	●	22(1), 55(2), 57(18), 69(4), 114(2), 153(1), 170(4)	A18c, Ko46
Oxypoda articollis Assing, 2016	○		63(1), 69(2)	A18c

Species	AR	NK	Localities/Samples	References
Oxypoda brevicornis (Stephens, 1832)	○	●	4(1), 7(1), 8(1), 9(2), 11b(2), 13(1), 22(1), 32(1), 39(7), 40(1), 45(3), 46(5), 48(4), 49(16), 54(4), 63(4), 64(1), 66a(1), 67(12), 68(3), 70(1), 73(1), 74(4), 75(1), 76(6), 77(1), 80a(1), 81(6), 81a(5), 90(1), 90a(8), 92(1), 105(7), 106(1), 110(3), 113(3), 114(5), 118(6), 121(1), 121a(3), 123(1), 125(4), 126(13), 127(11), 181(2), 206(1)	A18c
Oxypoda caucasica Bernhauer, 1902	●	●	6(1), 11(6), 11a(3), 11b(1), 12(9), 22(1), 31(1), 32(1), 33(3), 53(6), 58(2), 60(2), 62(2), 63(7), 66a(3), 66b(8), 68(7), 68a(4), 70(1), 92(1), 95(4), 101(2), 102(3), 105(13), 106(3), 110(2), 112(2), 113(1), 118(24), 118a(2), 119(5), 125(1), 149(3), 166(3), 170(2), 172(1), 196a(1), 199a(2)	
Oxypoda ferruginea Erichson, 1839	○		8a(1), 39(8), 49(2), 53(3), 54(2), 64(1), 77(2)	A18c
Oxypoda flavicornis Kraatz, 1856	○		11a(5), 92(2)	A18c
**Oxypoda flexa* Assing, 2016	○		9(14(, 9a(4), 54(1), 61(2), 76(4)	A16c, A18c
Oxypoda formosa Kraatz, 1856	○		57(1), 70(3)	A18c
***Oxypoda grandecristata* Assing, 2016	○		11(22), 11a(11), 11b(4)	A16c
Oxypoda haemorrhoa (Mannerheim, 1830)	○	●	20(1), 63(8), 89a(6), 109a(2), 191a(1)	A18c, E30
Oxypoda ignorata Zerche, 1996	○	●	22(4), 39(2), 49(1), 54(2), 63(1), 64(1), 67(3), 80b(1), 86(3), 112(1), 114(68), 116(1), 119(1), 123(1), l135(7), 162(2)	A18c
**Oxypoda infissoides* Assing, 2016	○	●	9(2), 11b(1), 12(13), 13(1), 22(1), 39(121), 50(29), 89b(1), 95(1), 96(2), 97(1), 114(5), 118(2), 118a(4), 121(1), 121a(5), 126(34), 129(6), 131(2), 167(1)	A16c, A18c
Oxypoda lentula Erichson, 1837	●		16(1), 85c(2), 128(3)	
**Oxypoda levipunctata* Assing, 2016	○		21(1)	A16c
Oxypoda longipes Mulsant & Rey, 1861	○		42(2), 46(1), 48(4), 49(1), 67(1), 148(12), 149(1)	A18c
Oxypoda obscuricollis Assing, 2007	●		134b(1)	
Oxypoda recta Assing, 2006	○	●	3a(2), 10(1), 13(1), 16(1), 41(1), 46(1), 47(2), 49a(1), 64(24), 65(1), 73(5), 80b(1), 87(1), 103(1), 104(1), 194(4), 200(1)	A18c
Oxypoda soror Thomson, 1855	○		6(1), 47(40), 75(1)	A18c
Oxypoda spaethi Bernhauer, 1901	○		167(1)	
**Oxypoda subplicata* Assing, 2016	○		30(100), 31(73), 33(88), 70(1), 77(2), 81(8), 90a(1), 118(24), 123(2), 124(10), 125(8)	A16c, A18c
**Oxypoda verminata* Assing, 2018	○		48(7), 49(1), 50(4)	A18c
Oxypoda vicina Kraatz, 1858	○	●	1(1), 9(41), 9a(2), 19(9), 20(1), 39(11), 48(7), 50(46), 54(2), 76(6), 80a(1), 82(1), 113(1), 126(30)	A18c
Oxypoda vittata Märkel, 1842	○	●	8(1), 9(1), 11(1), 31(1), 48(1), 57(1), 60(1), 61(2), 69(1), 114(4), 126(1), 133(1), 136(2), 148b(1), 149(2), 151(3), 155(4), 156(7), 172(1)	ATHW00, A18c

Species	AR	NK	Localities/Samples	References
Parocyusa rubicunda (Erichson, 1837) [36]	●	●	7(1), 9(5), 19(6), 33(2), 39(12), 41(4), 67(8), 73(12), 75(4), 76(2), 101(1), 103(2), 125(7), 126(4), 130(1), 199(6), 199a(26)	
Pella funesta (Gravenhorst, 1806)	○		156(3)	ATHW00
Pella laeviceps (Eppelsheim, 1880)	○	●	2(4), 7(3), 50(2), 63(8), 70(1), 71(1), 74(2), 92(1), 97(1), 100(1), 105(1), 133(14), 135(4), 136(7), 149a(5)	ATHW00, Ma06
Pella lugens (Gravenhorst, 1802)	○	●	100(1), 135(1), 148(4), 156(2)	ATHW00, Ma06
Pella similis (Märkel, 1844)	●		39(1)	
Piochardia reitteri (Wasmann, 1894)	○			A99, ATHW00
Poromniusa procidua (Erichson, 1837)	●		21(1)	
Pronomaea araxicola Reitter, 1898	●			A07b, IK57b [13]
Pronomaea flavirostris Semenov, 2003	●		121(2), 121a(19), 130(24)	
Pronomaea khnzoriani Semenov, 2003	○			A07b, Se03
Pycnota paradoxa (Mulsant & Rey, 1861)	○			ATHW00 [14]
Rhopalocerina clavigera (Scriba, 1859)	●	●	53(5), 58(2), 88(1), 92(1), 96(19), 97(3), 105(2), 130(3)	
Tachyusa impressa Eppelsheim, 1878	●		99(7), 161(9), 178(1), 198(1), 199(1)	
**Tachyusa unguis* Assing, spec. nov.	●		206(5)	ASpp
Thamiaraea cinnamomea (Gravenhorst, 1802)	●		148b(1), 150(2)	
Thiasophila angulata (Erichson, 1837)	●		8a(6)	
Thinonoma atra (Gravenhorst, 1806)	●		85c(10), 198a(1), 199(1)	
Trichiusa immigrata Lohse, 1984	●		80b(1)	
Tropimenelytron mirabile (Eppelsheim, 1884)	●		41(4), 89(1), 121(4), 121a(2), 125(1)	
Tropimenelytron tuberiventre (Eppelsheim, 1880)	○		1(1), 6(1), 7(2), 8(1), 51(3), 53(2), 55(2), 58(1), 59(1), 62(4), 63(13), 66b(2), 68(2), 68a(1), 69(1), 70(2), 75(9), 92(1), 180(1)	Be74 [15]
Zyras collaris (Paykull, 1800)	●	●	66a(2), 78(2), 88(6), 105(1), 112(7), 127(4)	
Zyras fulgidus (Gravenhorst, 1806)	●		135(1), 183(1), 205(1), c(1)	
Zyras haworthi (Stephens, 1832)	○	●	11(3), 66a(1), 81a(1), 88(1), 100(1), 114(2)	Dv84, ATHW00
Scaphidiinae				
Scaphisoma agaricinum (Linnaeus, 1758)	○			IK57a

Species	AR	NK	Localities/Samples	References
Scaphisoma balcanicum Tamanini, 1954	○			Lö70
Scaphisoma simillimum Löbl, 1970	○			IK85
Scaphisoma subalpinum subalpinum Reitter, 1880	○			IK57a
Oxytelinae				
Anotylus clypeonitens (Pandellé, 1867)	●	●	36(1), 63(2), 100(2), 105(3), 110(1), 171(6), ac(1)	
Anotylus fairmairei (Pandellé, 1867)	●		44(6), 45(3), ad(1)	
Anotylus gibbulus (Eppelsheim, 1878)	○			HMM79
**Anotylus hamatoides* Schülke, spec. nov.	●		136(1), 148c(2), 150(2), 151a(1)	
Anotylus hammondi Schülke, 2009	●		4(1), ae(1), af(1)	
Anotylus intricatus (Erichson, 1840)	●		ag(1)	
Anotylus inustus (Gravenhorst, 1806)	●		8(1)	
Anotylus khachikovi Gildenkov, 2017	●	●	64(1), 74(3), 81(1), 82(3), 105(52), 109(1), 126(5)	
Anotylus mutator (Lohse, 1963)	●		7(3), 15(2), 63(1), 135(4), 136(6), 148c(5), 149a(9), 150(3), 150a(21), 151(3), 151a(9), 151b(6)	
Anotylus nitidulus (Gravenhorst, 1802)	○	●	8a(1), 47(1), 49(3), 49a(1), 104(1), 186(1), 204a(2)	Sm04
Anotylus pumilus (Erichson, 1839)	○			E30
Anotylus rugosus (Fabricius, 1775)	○		20(6), 36(1), 199(1), 199a(1)	Sm04
Anotylus sculpturatus (Gravenhorst, 1806)	○	●	22a(1), 30(2), 32(4), 33(1), 53(4), 61(1), 66(5), 66a(1), 66b(1), 81(4), 81a(14), 89(6), 90(1), 191a(1), 105(31), 109(3), 110(1), 112(1), 113(4), 114(4), 115(2), 118(1), 125(5), 126(16), 127(3), 148a(12), 148b(28), 149b(9), 150a(1), 151a(1), 152(4)	Sm04
Anotylus strigifrons (Hochhuth, 1849)	●		53(1), 63(2), 67(5), 73(1), 74(1), 77(1), ah(1), ai(1), aj(1), ak(2)	
Anotylus tetracarinatus (Block, 1799)	○		45(1), 58(1), 81a(1), 96(1)	E30
Anotylus tetratoma (Czwalina, 1871)	●		44(3), 81a(1)	
Aploderus caelatus (Gravenhorst, 1802)	●		53(1)	
Bledius gallicus (Gravenhorst, 1806)	○			E30[16)]
Bledius debilis Erichson, 1840		○		Hh49
Bledius hinnulus Erichson, 1840	○			E30
Bledius minniensis Herman, 1986	○			H86

Species	AR	NK	Localities/Samples	References
Bledius roettgeni Bernhauer, 1928	○			Sm04
Bledius spectabilis Kraatz, 1857	○		a(4)	E30
Bledius tibialis Heer, 1839	●	○	al(1)	Ko46 [37]
Bledius tricornis (Herbst, 1784)	●		a(2), d(1)	
Carpelimus bilineatus Stephens, 1834	○		177(2), 199(1), 199a(1), a(1), b(1), d(1)	Sm04
Carpelimus corticinus (Gravenhorst, 1806)	○		16(1), 20(1), 66a(3), 85c(2), 128(1), 130(1), 177(6)	Gi15
Carpelimus elongatulus elongatulus (Erichson, 1839)	○			Sm04
Carpelimus erichsoni (Sharp, 1871)	○		196(1)	
Carpelimus exiguus (Erichson, 1839)	○			Sm04
Carpelimus fuliginosus (Gravenhorst, 1802)	○			Sm04
Carpelimus gracilis (Mannerheim, 1830)	○			Gi01
Carpelimus gusarovi Gildenkov, 1997	●		g(1)	
Carpelimus heydeni (Klima, 1904)	●		212(12)	
Carpelimus impressus (Lacordaire, 1835)	○			Sm04
Carpelimus manchuricus subtilicornis (Roubal, 1946)	●		53(4), 199a(2)	
Carpelimus rivularis (Motschulsky, 1860)	○		66a(1)	Sm04
Carpelimus similis (Smetana, 1967)	○			G01, G09
Coprophilus pennifer (Motschulsky, 1845)	○		e(1)	ATHW00
Coprophilus striatulus (Fabricius, 1792)	●		181(1)	
Deleaster dichrous (Gravenhorst, 1802)	○		134(1), f(1)	Fa71 [17]
Oxytelus fulvipes Erichson, 1839	○			E30
Oxytelus laqueatus (Marsham, 1802)	○		8(3), 59(1), 148a(1)	Sm04
Oxytelus piceus (Linnaeus, 1767)	○		44(1)	Sm04
Oxytelus sculptus Gravenhorst, 1806	○			Sm04
Platystethus alutaceus Thomson, 1861	●		21(1), 128(106)	
Platystethus capito Heer, 1839	●	●	49(1), 54(1), 58(3), 63(12), 66a(5), 67(2), 72(1), 73(3), 76(3), 81a(1), 88(1), 93a(38), 94(1), 99(41), 100(6), 127(2), 131(1), 195(1), 197(1), 199a(4), 202(1), 204a(2), 210(11), ah(1), am(1), an(1), ao(1), ap(1), aq(1)	

Species	AR	NK	Localities/Samples	References
Platystethus cornutus (Gravenhorst, 1802)	○		177(4), 190(1)	Sm04
Platystethus gildenkovi Khachikov, 2013	●		44(1)	
Platystethus nitens (C. R. Sahlberg, 1832)	○	●	41(2), 45(3), 49(1), 67(1), 100(1), 112(1), 177(6), 184(4), 206(1)	Sm04
Platystethus rufospinus Hochhuth, 1851	●		184(2), 186(1)	
Platystethus rugifrons Bernhauer, 1900	○			Sm04
Platystethus spinosus Erichson, 1840	○			E30
Syntomium aeneum (Müller, 1821)	●		52(3), 153(1),	
Thinodromus dilaticollis (Eppelsheim, 1884)	○	●	39(1), 100(2), 109(1), 117(3), 121a(2), 126(1), 188(4), 205(1), 206(6)	Gi00
Thinodromus kiesenwetterii (Hochhuth, 1851)	○			Gi00
Thinodromus mannerheimii (Kolenati, 1846)	○			Gi00
Thinodromus motschulskyi (Hochhuth, 1860)	●		199(3), 199a(3)	
Thinodromus transversalis (Wollaston, 1857)	●		212(1)	
Oxyporinae				
Oxyporus rufus rufus (Linnaeus, 1758)	●		53(1), 70(4), 133(1)	
Steninae				
Dianous coerulescens elegans Iablokoff-Khnzorian, 1957	○		209(1)	IK57b
Dianous ponticus Fagel, 1963	○		181(1)	P16
Stenus arabicus Saulcy, 1865	●		183(2)	
Stenus araxis Ryvkin, 1990	○			Ry90
Stenus argus Gravenhorst, 1806	○		19(1), 201(2), 202(1)	Sm04 [50)]
Stenus armeniacus Puthz, 1967	○			Pu67
Stenus ater Mannerheim, 1830	○			Ry90
Stenus atratulus Erichson, 1839	○	○	50(2), 61(1), 66(1), 66a(2), 73(1), 76(3), 103(1), 106(1), 107(1), 126(4)	Ry90
Stenus biguttatus Linnaeus, 1758	○			Ry90
Stenus bimaculatus Gyllenhal, 1810	○			Ry90
Stenus binotatus Ljungh, 1804	○			Sm04
Stenus callidus Baudi di Selve, 1848	○		19(1), 20(4), 210(1)	Hr90 [47)], P08, Ry90
Stenus caspius Puthz, 1972	○		43(1)	Ry90
Stenus cautus Erichson, 1839	●		192(1)	

Species	AR	NK	Localities/Samples	References
Stenus cephallenicus Bernhauer, 1913	●		9(3), 20(5), 40(1), 67(1), 76(5), 77(9), 82(19), 82a(8), 93(5), 119(1), 126(16), 130(7), 131(2)	
Stenus cicindeloides (Schaller, 1783)	○		206(3)	Sm04 [50)]
Stenus claritarsis Puthz, 1971	○		183(2)	Ry90
Stenus clavicornis Scopoli, 1763	○	○	47(14), 64(3), 66a(1), 67(1), 78(3), 79(2), 89(1), 89b(1), 98(1), 100(1), 101(2), 101a(1), 107(2), 109(1), 127(1), 130(4), 183(3)	Ry90
Stenus coarcticollis coarcticollis Eppelsheim, 1890	○	●	1(4), 6(1), 7(7), 11(25), 11a(11), 11b(18), 12(5), 15(10), 30(1), 33(6), 39(4), 50(15), 51(5), 53(6), 58(2), 60(1), 63(1), 66(4), 66b(1), 68(1), 71(6), 72(6), 75(2), 82(7), 82a(17), 83(5), 88(1), 89(18), 89b(21), 90a(1), 91(1), 92(4), 95(1), 96(2), 97(11), 103(7), 105(2), 113(1), 118(12), 118(5), 119(6), 125(17), 126(50), 131(13), 135(2), 136(1), 151a(1), 180(1)	Ry90
Stenus comma comma LeConte, 1863	○	○	14(1)	Ry90
Stenus erythrocnemus Eppelsheim, 1884	○	○	66a(1), 68(1), 112(1), 126(1)	A06, Ry90
Stenus ganglbaueri Bernhauer, 1905	○		n(1)	Sm04 [50)]
Stenus humilis Erichson, 1839	○	○	3a(4), 6(5), 41(4), 42(3), 43(1), 45(1), 47(1), 48(1), 51(1), 53(11), 54(6), 57(3), 58(1), 63(1), 66(3), 66a(5), 67(4), 68(2), 68a(1), 70(2), 88(11), 98(18), 100(3), 101(12), 102(2), 105(5), 106(4), 107(2), 109(1), 117(3), 136(1), 180(1)	P83, Ry90
Stenus ignotus Eppelsheim, 1890	○			Ry90
Stenus juno (Paykull, 1789)	○		41(3), 198(1), 201(4)	Sm04 [50)]
Stenus machulkai Hromádka, 1977	○	●	19(3), 20(11), 22(7), 39(2), 77(30), 81(749), 81a(677), 82(4), 88(9), 89(7), 89b(9), 90(6), 90a(6), 91(11), 103(1), 105(3), 106(2), 107(4), 109(27), 110(6), 112(2), 113(1), 114(7), 117(1), 118(3), 121a(1), 126(65), 127(13), 128(2), 129(1), 130(33), 131(1)	Ry90
Stenus medus Puthz, 1981	○	●	50(1), 76(2), 114(38), 115(2), 118(1), 121(1), 121a(20), 125(1), 130(46)	P81, Ry90
Stenus morio Gravenhorst, 1806	○		63(1)	Ep78
Stenus nanus Stephens, 1833	●		46(1), 54(2), 67(1), 73(1), 94(1)	
Stenus nigritulus Gyllenhal, 1827	○	●	103(2), 104(1)	P12
Stenus nodipes Puthz, 1972	○			P09
Stenus ochropus Kiesenwetter, 1858	○	○	22(83), 22a(4), 26(57), 30(23), 31(2), 32(50), 34(2), 35(26), 39(10), 71(1), 78(6), 79(7), 81(140), 81a(51), 88(46), 89(3), 89b(1), 97(2), 98(51), 100(26), 101(23), 101a(8), 102(6), 103(5), 109(1), 112(105), 114(260), 115(1), 116(24), 117(7), 118(14), 118a(7), 119(7), 121(6), 121a(23), 124(2), 126(24), 127(27), 135(1), 136(2), 137(2), 149(1), 149a(1), 150a(1), 155(1), 167(1), 170(1)	Ry90
Stenus peripherus Korge, 1971	○			P08

Species	AR	NK	Localities/Samples	References
Stenus piscator Saulcy, 1865	○			Ry90
Stenus prometheus Puthz, 1967	○		4(1), 11(3), 11b(2), 13(5), 16(15), 45(1), 49(1), 49a(1), 70(1), 83(3), 119(1), 123(1)	P67, P03, Ry90
Stenus proprius Benick, 1921	●		31(1)	
Stenus providus providus Erichson, 1839	○		20(1), 81a(1), 130(6)	Ry90
Stenus similis (Herbst, 1784)	○		6(1), 30(1), 47(2), 76(1), 81a(1), 82(1), 82a(1), 88(2), 125(1)	Ry90
Stenus skoraszewskyi Korge, 1971	○		85a(1), 85c(14), 94a(2), 128(1), 201(6)	Sm04 [50)]
Stenus stigmula Erichson, 1840	○			P81
Stenus subditus Benick, 1920	○			P83
Stenus turk Puthz, 1972	○	○	90a(1), 114(381), 130(1)	P72b [18)], Ry90
Scydmaeninae				
Cephennium perispinctum perispinctum Kolenati, 1846	○			VB04
***Euconnus karabakhus* Meybohm, spec. nov.		●	98(2), 101(3), 102(1)	
Euconnus lalvarensis Iablokoff-Khnzorian, 1964	○	●	3a(1), 51(1), 53(1), 55(1), 66(1), 75(1), 88(1), 112(1)	IK64a
***Euconnus longilaminatus* Meybohm, spec. nov.	●		70(11)	
***Euconnus tavushus* Meybohm, spec. nov.	●		66a(20)	
Euconnus wetterhallii (Gyllenhal, 1813)	●		66a(1)	
Neuraphes angulatus (Müller & Kunze, 1822)	●	●	7(1), 8(1), 11b(1), 92(1), 105(5)	
***Neuraphes gomarantsus* Meybohm, spec. nov.	●		30(3), 31(1)	
***Neuraphes syunikus* Meybohm, spec. nov.	●		81a(1), 118(14), 118a(2), 119(1)	
Scydmaenus cornutus Motschulsky, 1844	○			E30
Scydmoraphes helvolus (Schaum, 1844)	●		81a(1), 118(1)	
Scydmoraphes yermolowi (Saulcy, 1878)	○		66(5), 66b(2), 68(3), 68a(1)	Da04
Stenichnus poweri (Fowler, 1884)	●		53(3), 57(3), 63(1), 66(4), 66b(7), 77(1)	
Stenichnus pusillus pusillus (Müller & Kunze, 1822)	●		66(3)	
Stenichnus scutellaris (Müller & Kunze, 1822)	●	●	9(1), 22(1), 103(2)	
Stenichnus subseriatus Franz, 1960 [59)]	●	●	6(3), 50(3), 51(3), 52(1), 55(2), 57(1), 58(3), 59(2), 61(1), 62(4), 63(8), 66(2), 68(5), 69(4), 71(2), 98(1), 100(2), 101(1), 105(3), 114(1), 117(1)	
Paederinae				
Achenium caucasicum Laporte, 1835	○			AKÖ11

Species	AR	NK	Localities/Samples	References
Achenium debile Erichson, 1840	○			Kh37
Achenium humile (Nicolai, 1822)	○			A17a
Achenium quadriceps Eppelsheim, 1889	●		177(4), a(2), i(1)	
Astenus bimaculatus bimaculatus (Erichson, 1840)	○			E30
Astenus lyonessius (Joy, 1908)	●	●	22a(5), 23a(1), 36(1), 38(1), 78(6), 79(15), 80(1), 88(14), 89b(1), 114(10), 115(1), 121a(1), 126(5), 194(1)	
Astenus procerus (Gravenhorst, 1806)	○			A17a, Am00 [19], IK61 [20]
Astenus rufopacus Reitter, 1909	○		79(1)	An17, C60 [39]
Homaeotarsus chaudoirii Hochhuth, 1851	●		b(1)	
Lathrobium brunnipes (Fabricius, 1792)	○			A17a
Lathrobium fulvipenne (Gravenhorst, 1806)	○		183(1)	A17a
Lathrobium laevipenne Heer, 1839	●		204a(4)	
Lathrobium longulum Gravenhorst, 1802	○		9(1)	A17a
Lathrobium permutatum Assing, 2009	●	●	33(11), 36(6), 39(2), 50a(1), 56(1), 66(1), 101(1), 104(1), 112(1), 126(11), 199a(3)	
Lathrobium tichomirovae Coiffait, 1981	●	●	42(1), 43(1), 50a(1), 54(1), 67(3), 81a(1), 82(10), 90(2), 107(1), 179(3), 206(3)	
Leptobium gracile (Gravenhorst, 1802)	○			A05b
**Lobrathium ancoriferum* Assing, 2017	○			A17d
Lobrathium rugipenne (Hochhuth, 1851)	○		179(1), 181(1), 204a(1)	A17d, Hh51
Luzea graeca (Kraatz, 1857)	○			IK [41]
Medon dilutus pythonissa (Saulcy, 1865)	○			A18d, C70c [40]
Medon exquisitus Kirshenblat, 1951 [42]	○			A13a, IK64b, Ki51
Medon fusculoides Coiffait, 1970	○	●	112(1), 121(10), 121a(50), 130(1)	C70d
Medon maronitus (Saulcy, 1865)	○			A18d
Medon paradisiacus Assing, 2004	○		26(2), 34(5)	A18d
**Medon sequax* Assing, 2004	○		188(3)	A04a
Ochthephilum egregium (Reitter, 1884)	●		36(11), 66(5), 66a(3)	
Ochthephilum turkestanicum (Korge, 1968)	●		190(1)	
Paederidus rubrothoracicus (Goeze, 1777)	○			Ki32, Sz57 [49]

Species	AR	NK	Localities/Samples	References
Paederidus ruficollis (Fabricius, 1777)	●		199(5)	
Paederus fuscipes fuscipes Curtis, 1826	○		177(5), 190(1), 203a(3), d(2)	An18, Ki32
Paederus littoralis Gravenhorst, 1802	○	●	7a(8), 13(1), 37(1), 63(1), 66a(1), 78(1), 79(1), 81a(1), 88(1), 99(1), 105(1), 109(2), 111(1), 112(2), 134a(1), 137(1), 141(1), 203a(1), h(1)	An18
Paederus riparius (Linnaeus, 1758)	○			Ki32
Rugilus armeniacus (Coiffait, 1970)	○	●	101(1), 121a(1), 199a(1)	A17a, C70a
Rugilus longicollis (Fauvel, 1900)	●	●	15(2), 30(6), 32(13), 36(2), 63(1), 66(1), 66a(2), 80b(22), 88(1), 89(1), 98(2), 101(1), 103(1), 109(2), 112(3), 117(1), 118(1), 118a(1), 119(1), 121a(1), 125(1), 127(1), 130(6), 131(2), 150(2), 151b(1), 155(2), 164(1), 165(2), 167(1), 172(2), 195(1), 211(1)	
Rugilus orbiculatus (Paykull, 1789)	○		41(1), 77(1), 126(93)	Sm04
Rugilus similis (Erichson, 1839)	○	●	20(1), 41(2), 47(3), 81(1), 81a(1), 101(3), 126(1), 183(1)	Sm04
Scopaeus bicolor Baudi di Selve, 1848	○			E30
Scopaeus cameroni Coiffait, 1968	○	●	112(11), 121a(7), 130(9)	C68 [44)], Fr10
Scopaeus chalcodactylus (Kolenati, 1846)	○	○	204a(3)	Fr97, Fr07a, Ko46
Scopaeus debilis Hochhuth, 1851	○		212(1)	Hh51, Fr12
Scopaeus gracilis (Sperk, 1835)	○		199(1)	Fr97, Fr07b
Scopaeus khnzoriani Coiffait, 1968	○			C68, Fr09
Scopaeus laevigatus (Gyllenhal, 1827)	○		92(1), 177(9), 212(1)	Fr10
Scymbalium anale (Nordmann, 1837)	○			Sm04
Sunius fulgocephalus (Coiffait, 1970)	○			A17g, C70d
Sunius khnzoriani (Coiffait, 1970)	○	●	9(2), 12(1), 25(2), 26(22), 32(2), 76(1), 98(1)	A08c, A17g
Sunius melanocephalus (Fabricius, 1792)	○			C70d [43)]
Tetartopeus quadratus (Paykull, 1789)	●		85a(1)	
Tetartopeus stylifer (Reitter, 1909)	○			A17a
Staphylininae				
Acylophorus glaberrimus (Herbst, 1784)	●		190(4)	
Astrapaeus ulmi (Rossi, 1790)	○			IK57a

Species	AR	NK	Localities/Samples	References
Atanygnathus terminalis (Erichson, 1839)	●		ar(1)	
Bisnius cephalotes (Gravenhorst, 1802)	○			ATHW00, IK64b
Bisnius fimetarius (Gravenhorst, 1802)	○	●	39(6), 67(6), 68(1), 68a(3), 80b(11), 81(3), 81a(4), 90a(2), 105(13), 106(2), 108(1), 109(2), 110(12), 113(8), 114(44), 121(2), 126(3), 144(4)	Sm04
#Bisnius microtophilus (Coiffait, 1967)	○			ATHW00, C67a
Bisnius pentheri (Ganglbauer, 1905)	○			ATHW00, C67a, IK64b
Bisnius piochardi (Fauvel, 1875)	○		122(1)	C69 [48], Sr04
Bisnius reitteri (Eppelsheim, 1889)	●		195a(3)	
Bisnius sordidus (Gravenhorst, 1802)	○	●	22(1), 70(2), 80b(3), 81(1), 81a(1), 105(2), 107(3), 110(3), 114(32), 116(11), 118a(1), 122(315), 123(11), 124(2), 126(1)	Sm04
**Bisnius karkarensis* Schillhammer, in prep. [66]	●		67(1), 82(1), 86(9)	
Creophilus maxillosus maxillosus (Linnaeus, 1758)	○			IK64b
Dinothenarus flavocephalus flavocephalus (Goeze, 1777)	○	●	72(1), 112(1), 136(3), 148b(4), 148c(1),	Sz66b
Dinothenarus pubescens (De Geer, 1774)	●		86(1)	
Emus hirtus (Linnaeus, 1758)	●		44b(2)	
Erichsonius cinerascens (Gravenhorst, 1802)	○		19(1), 20(2)	Pe80
Erichsonius dux Smetana, 1967	○			Pe80
Erichsonius rivularis rivularis (Kiesenwetter, 1858)	○		66a(1)	C65 [21]
Erichsonius subopacus (Hochhuth, 1851)	○	●	36(2), 66a(1), 100(2), 103(1), 175(1), 183(1), 185(1), 202(1)	Br44
#Gabrius armeniacus Coiffait, 1966	○			C66a, Sr03
Gabrius astutoides (Strand, 1946)	●	●	100(2), 113(1), 119(1), 125(1), 195(1), 199a(1)	
Gabrius femoralis (Hochhuth, 1851)	●	●	12(3), 19(1), 22(4), 22a(1), 39(1), 50(2), 50a(1), 59(1), 73(1), 74(2), 77(1), 78(2), 79(1), 80(1), 80b(2), 81(4), 81a(4), 82a(1), 83(1), 88(4), 89(6), 89b(10), 90(1), 110(1), 112(2), 118(4), 118a(1), 119(1), 121(12), 121a(6), 126(31), 179(1)	
Gabrius latro Joy, 1913	○		177(1), 190(1)	C74 [22]
Gabrius muelleri Gridelli, 1928	●	●	41(2), 46(1), 53(2), 66(1), 66b(1), 88(1), 93(1), 98(4), 101(2), 103(1)	
Gabrius nigritulus (Gravenhorst, 1802)	○	●	100(11), 179(2), 192(2), 194(1), 201(1)	Sm04
Gabrius ravasinii Gridelli, 1920	●		127(4), 204a(1)	
Gabrius robustus Smetana, 1953	●		16(1), 67(2), 77(3), 82(2), 85a(4), 85c(11), 128(27), 128a(1)	

Species	AR	NK	Localities/Samples	References
Gabrius sacerdotalis Joy, 1913	●		66a(2), 73(3), 93a(1)	
Gabrius subnigritulus Joy, 1913	●	●	103(4), 202(1)	
Gabrius suffragani Joy, 1913	●		180(2), 181(4)	
Gabrius toxotes Joy, 1913	●		179(1)	
Gabrius trossuliformis Schillhammer, 1999	○		18(1), 21(15), 44(1), 45(9), 46(3), 49(1), 67(1), 77(2), 82(2), 85a(4), 85c(3), 86(1), 94(7), 128(14)	ATHW00 [23)], C66a [23)]
Gauropterus fulgidus (Fabricius, 1787)	○			Sm04
Gauropterus sanguinipennis (Kolenati, 1846)	○		178(1)	Ko46, Bo11b
Gyrohypnus angustatus Stephens, 1833	○	●	19(1), 20(2), 32(1), 33(2), 39(9), 41(3), 67(1), 80b(1), 82(3), 103(1), 121a(4), 126(9), 130(1), 143(1), 179(2), 181(1), 186(1), 199a(1)	Sm04
Gyrohypnus fracticornis (O. Müller, 1776)	○	●	22(1), 39(2), 44(1), 45(1), 54(1), 80b(4), 86(2), 110(3), 114(58), 115(1), 116(4), 123(5), 139(1)	Sm04
Gyrohypnus punctulatus (Paykull, 1789)	●		8(1), 80b(2), 81a(1), 86(3)	
Heterothops balthasari Smetana, 1967	○			Sm04
Heterothops dissimilis (Gravenhorst, 1802)	○		41(6), 43(1), 46(1), 47(3), 67(1), 76(1), 80b(2), 92(3), 95(1), 126(1), 130(1)	ASpp, ATHW00, C77 [56)], Mo60 [24)]
Heterothops laeticolor Reitter, 1891	○			ASpp, ATHW00
#Heterothops macrops Coiffait, 1977	○			C77
#Heterothops microtophilus Coiffait, 1977	○			ASpp, ATHW00, C77
Heterothops praevius Erichson, 1839	○	●	22(35), 22a(5), 30(19), 31(6), 54(1), 57(5), 70(2), 71(3), 72(2), 80a(3), 81a(2), 89(6), 89b(7), 114(14), 116(1), 118(1), 122(3), 123(10)	ASpp, ATHW00, IK66 [57)]
Jurecekia asphaltina (Erichson, 1840)	○			ATHW00, Sr04
#Leptacinus armeniacus Coiffait, 1966	○			A17f, C66b
Leptacinus batychrus (Gyllenhal, 1827)	○			E30
Leptacinus nigerrimus Coiffait, 1971	○			Bo11a
Leptacinus pusillus (Stephens, 1833)	○			Sm04
Leptacinus sulcifrons (Stephens, 1833)	○		67(1), 86(1)	Sm04
Neobisnius prolixus (Erichson, 1840)	○		8b(4), 14(1), 120(4), 185(4), 190(1), 199(4)	Hh49
Nudobius lentus (Gravenhorst, 1806)	●		136(1)	
Nudobius umbratus (Motschulsky, 1860)	○			ATHW00, IK57a

Species	AR	NK	Localities/Samples	References
Ocypus curtipennis (Motschulsky, 1849)	○		36(1), 185(1)	E30
Ocypus forficularius (Motschulsky, 1860)	●		6(1), 7(4), 135(1)	
Ocypus fulvipennis Erichson, 1840	●		9(1), 40(1), 41(1), 74(1), 88(1), 126(3), 140(1), 179(1)	
Ocypus hochhuthi Eppelsheim, 1878	●		22(1), 156(2)	
Ocypus mus (Brullé, 1832)	●		26(7), 35(1)	
Ocypus nitens nitens (Schrank, 1781)	○			Sm04
Ocypus ophthalmicus ophthalmicus (Scopoli, 1763)	○			Hh49 [25)]
Ocypus orientis Smetana & Davies, 2000	○			E30 [26)]
Ocypus picipennis caucasicus (Müller, 1926)	○	●	26(4), 65(9), 81b(1), 115(1), 116(1), 139(2), 183(4), 185(1), 193(2), 202(1), 204a(1)	H01
Ocypus sericeicollis (Ménétriés, 1832)	○	●	22(1), 88(5), 89b(1), 114(2), 116(1)	Sm04
Ocypus syriacus primigenius (Müller, 1923)	○			Mü23
**Ontholestes chalcopygus* (Hochhuth, 1849)	○			C74
Ontholestes murinus (Linnaeus, 1758)	○	●	105(1), 180(1)	Hh49
Othius grandis Hochhuth, 1849	○	●	58(1), 72(1), 114(3)	A03
Othius lapidicola Märkel & Kiesenwetter, 1848	○		2(4), 4(16), 5(1), 6(6), 7(6), 11(20), 11a(20), 11b(33), 30(11), 31(4), 33(4), 51(8), 55(3), 58(4), 59(1), 60(1), 62(1), 63(12), 69(4), 68a(1), 69(4), 70(2), 71(1), 72(1), 74(1), 75(10), 81(1), 82a(1), 88(21), 89(6), 89b(2), 90a(1), 91(1), 92(4), 97(1), 118(20), 119(4), 124(4), 125(5), 126(4)	A97, A03, A15
Othius punctulatus (Goeze, 1777)	○	●	1(1), 2(2), 3(3), 4(5), 6(1), 7(3), 11(8), 11a(3), 11b(4), 12(2), 51(2), 55(1), 62(1), 69(3), 75(5), 95(1), 96(1), 110(1), 135(1), 148b(1)	A97, A03
Othius stenocephalus Eppelsheim, 1881	●		6(2), 7(2), 59(2), 68(1), 69(2)	
Phacophallus parumpunctatus (Gyllenhal, 1827)	○			Sm04
Philonthus aculeatus Coiffait, 1963	○	●	81a(2), 110(2), 112(3), 121a(2), 126(2), 127(1)	Am00
Philonthus alberti Schillhammer, 2000	○	●	36(1), 68(1), 90(1), 103(4), 105(2), 110(7), 112(9), 113(2), 114(2), 125(1), 127(3), 195a(1)	Sm04
Philonthus alpinus Eppelsheim, 1875	●		44(59)	
Philonthus atratus (Gravenhorst, 1802)	○		6(1), 76(1), 120(6), 179(2)	E30

Species	AR	NK	Localities/Samples	References
Philonthus carbonarius (Gravenhorst, 1802)	●	●	14(1), 29(1), 39(1), 70(1), 88(1), 89b(1), 95(1), 106(1), 108(1), 114(1), 116(1), 127(1), 128(1), 147(4), 156(4)	
Philonthus caucasicus Nordmann, 1837	○			Sm04
Philonthus cochleatus Scheerpeltz, 1937	○			C74
Philonthus cognatus Stephens, 1832	●	●	75(1), 89(2), 110(1), 114(4), 115(1), 116(1), 120(1), 126(1), 130(1), 183(1)	
Philonthus concinnus (Gravenhorst, 1802)	○	●	14(3), 17(1), 21(2), 22a(1), 36(10), 80b(4), 81(2), 81a(3), 85(1), 85c(1), 90a(1), 94(1), 99(1), 109(1), 114(3), 120(2), 121a(1), 123(4), 126(9), 128(2)	Sm04
Philonthus corruscus (Gravenhorst, 1802)	○		76(1), 157(1)	IK64b
Philonthus cruentatus (Gmelin, 1790)	○	●	44(12), 107(1), 122(1), 126(1), 179(1)	H01
Philonthus debilis (Gravenhorst, 1802)	○			Sm04
Philonthus dimidiatipennis Erichson, 1840	○			Hh49
Philonthus discoideus (Gravenhorst, 1802)	○			Sm04
Philonthus ebeninus (Gravenhorst, 1802)	○	●	44(1), 78(1), 81(1), 116(1)	Sm04
Philonthus eriwanensis Bernhauer & Schubert, 1914	○			ATHW00, C74
Philonthus fumarius (Gravenhorst, 1806)	●		66a(1)	
Philonthus intermedius (Lacordaire, 1835)	○	●	82(1), 115(1)	Sm04
Philonthus juvenilis Peyron, 1858	●		185(1)	
Philonthus laminatus (Creutzer, 1799)	○	●	107(1), 114(1), 115(1), 116(1), 122(1), 126(1)	Sm04
Philonthus laxatus Fauvel, 1875	○			Hh62[27)]
Philonthus lindbergi Scheerpeltz, 1958	●		183(1), 201(1)	
Philonthus longicornis Stephens, 1832	○			Sm04
Philonthus nigrita (Gravenhorst, 1806)	●		201(1)	
Philonthus nitidicollis (Lacordaire, 1835)	○		21a(1), 86(1), 94a(1), 139(3)	Sm04
Philonthus parvicornis (Gravenhorst, 1802)	○		44(7), 177(1)	Sm04
Philonthus picimanus (Ménétriés, 1832)	○		33(4), 39(1), 46(1), 49(2), 53(1), 66(1), 67(10), 90(1), 94(1), 118(1), 126(8), 127(1), 135(4), 156(1), 179(2), 181(1)	Sm04
Philonthus politus (Linnaeus, 1758)	○		148a(1), 148b(1), 149b(1), 179(1)	Sm04

Species	AR	NK	Localities/Samples	References
Philonthus punctus punctus (Gravenhorst, 1802)	○			Sm04
Philonthus quisquiliarius quisquiliarius (Gyllenhal, 1810)	○		183(3), 190(3), 201(4), 203a(2), a(38)	Sm04
Philonthus rectangulus Sharp, 1874	○		86(1)	ATHW00
Philonthus rubripennis Stephens, 1832	○	●	36a(1), 100(1), 120(2), 179(5), 185(3), 199b(1), 204a(1)	Sm04
Philonthus rufimanus Erichson, 1840	●		8b(11), 14(6), 120(13)	
Philonthus rufimargo Reitter, 1909	○			R09
Philonthus sanguinolentus (Gravenhorst, 1802)	○			C74
Philonthus spinipes kabardensis (Bolov & Kryzhanovskij, 1969)	○			H01
Philonthus splendens (Fabricius, 1792)	●		46(1)	
Philonthus succicola Thomson, 1860	○		148b(3)	Sm04
Philonthus svanetiensis Coiffait, 1974	●		4(2), 45(22), 45a(1), 46(14), 49a(2)	
Philonthus tenuicornis Mulsant & Rey, 1853	○	●	41(1), 44(3), 62(5), 70(1), 75(2), 86(2), 103(2), 133(3), 135(1), 136(4), 183(1)	Sm04
Philonthus varians (Paykull, 1789)	○		7(1), 15(1), 23a(1)	IK64b
Philonthus velatipennis Solsky, 1869	○		a(11)	Pe80
Philonthus ventralis (Gravenhorst, 1802)	○			Sm04
Philonthus viridipennis Fauvel, 1875	○			Br01 [28)]
Physetops tataricus (Pallas, 1773)	○			ATHW00, IK64b [54)], SG05
Platydracus chalcocephalus (Fabricius, 1801)	●	●	8(1), 106(1), 112(2), 127(1), 148b(1), 151a(2), 156(2), 157(8), 199a(2)	
Platydracus flavopunctatus (Latreille, 1804)	○		120(1)	Hh49 [29)]
Platydracus meridionalis (Rosenhauer, 1847)	○			Fa74
Platydracus stercorarius stercorarius (Olivier, 1795)	○			Sm04
Platyprosopus bagdadensis Stierlin, 1867	○			Sm04
Platyprosopus elongatus Mannerheim, 1830	○			Mm30
Quedius acuminatus Hochhuth, 1849	○			H01

Species	AR	NK	Localities/Samples	References
Quedius boops (Gravenhorst, 1802)	●	●	7(1), 9(24), 11(1), 11a(2), 11b(3), 12(1), 13(31), 15(2), 19(18), 20(2), 22(25), 22a(16), 23(1), 30(23), 31(2), 32(1), 33(1), 36(4), 39(43), 40(6), 41(2), 46(4), 47(2), 48(2), 49a(1), 50(19), 50a(1), 51(1), 54(3), 55(1), 57(1), 59(1), 60(5), 61(1), 63(1), 64(3), 67(3), 68(3), 69(2), 70(14), 71(2), 72(1), 73(2), 74(6), 75(9), 76(24), 77(6), 79(1), 81(34), 81a(20), 82(27), 82a(26), 83(17), 85a(1), 88(46), 89(142), 89a(3), 89b(221), 90a(2), 91(10), 92(3), 93(3), 95(29), 100(2), 104(1), 105(1), 107(7), 108(20), 109(4), 110(65), 112(4), 113(1), 114(38), 115(2), 116(42), 118(9), 118a(7), 119(8), 121(13), 121a(1), 123(2), 124(3), 126(258), 127(1), 128(1), 129(2), 131(30), 135(5), 184(1)	
Quedius brevalatus Assing, 2017	○		88(21)	A17e
Quedius brevicornis (Thomson, 1860)	●		151a(1)	
Quedius cinctus (Paykull, 1790)	○	●	22(6), 55(1), 68(2), 80b(1), 81a(1), 82(1), 114(12), 118(2), 123(3)	Sm04
Quedius cohaesus Eppelsheim, 1888	●	●	22(39), 30(34), 31(1), 32(37), 33(4), 36(1), 39(19), 40(4), 68(2), 81(15), 81a(7), 82(2), 89(4), 89b(30), 90(8), 90a(18), 91(14), 100(11), 101(2), 101a(1), 105(3), 107(10), 108(3), 109(16), 110(14), 112(109), 113(12), 114(210), 115(75), 116(49), 117(4), 121(21), 121a(52), 123(1), 125(9), 126(50), 127(19), 130(1), 131(3), 145a(1)	
Quedius cruentus (Olivier, 1795)	○		132(4), 134(2)	A19b, IK61 [61)]
Quedius curtipennis Bernhauer, 1908	○	●	66(1), 69(1), 101(1), 109(2), 115(1), 130(3), 149(1), 156(1)	Sm04
Quedius edmundi Coiffait, 1969	●		68a(1)	
Quedius fuliginosus (Gravenhorst, 1802)	●	●	2(1), 3(1), 4(1), 7(2), 19(3), 20a(1), 55(3), 62(1), 63(1), 66b(1), 69(1), 72(1), 73(1), 75(1), 78(1), 81(1), 89(1), 90(1), 105(1), 106(1), 107(1)	
Quedius gridellii Scheerpeltz, 1933	●		30(1), 31(2), 39(1)	
Quedius humeralis Stephens, 1832	○			E30
Quedius invreae Gridelli, 1924	●		155(1), 157(3)	
Quedius korgeanus Fagel, 1968	○		40(1), 49(2), 123(4), 124(2), 125(1)	So04
#*Quedius latus* Hochhuth, 1851	○			Hh51
Quedius levicollis (Brullé, 1832)	●	●	114(1), 136(3)	

Species	AR	NK	Localities/Samples	References
Quedius limbatus (Heer, 1839)	○	●	3(1), 3a(1), 4(4), 6(3), 7(3), 9(20), 9a(3), 10(1), 11(28), 11a(20), 11b(16), 12(7), 13(6), 15(5), 16(6), 19(2), 20a(1), 21(5), 21a(2), 22(2), 28(2), 30(12), 31(7), 32(23), 39(47), 40(1), 43(1), 46(3), 49a(5), 50(45), 50a(6), 52(1), 53(14), 58(2), 63(1), 64(3), 66(4), 66a(8), 66b(1), 67(19), 69(1), 70(1), 71(2), 72(12), 74(2), 75(3), 76(23), 77(2), 82(1), 83(2), 84(1), 85(1), 85a(22), 85c(6), 87(1), 88(8), 89(4), 89b(3), 91(4), 92(4), 93(2), 97(2), 98(8), 100(2), 102(1), 103(5), 104(1), 105(5), 106(1), 108(1), 112(29), 113(1), 115(1), 116(2), 118(16), 118a(3), 119(6), 121(9), 121a(19), 124(3), 125(2), 126(354), 127(5), 129(1), 131(2), 135(4), 136(6); 145(2), 148(3), 149a(1)	So02
Quedius nitipennis (Stephens, 1833)	○		20(1), 23(1), 41(1), 67(30), 73(3), 76(1), 77(3), 78(1), 89(1)	C67b [46], So04
Quedius ochripennis (Ménétriés, 1832)	○	●	22(8), 88(1), 114(25), 116(1),135(3)	IK64b
Quedius orientalis Korge, 1971		●	112(5)	
Quedius scintillans (Gravenhorst, 1806)		●	114(22)	
Quedius semiobscurus (Marsham, 1802)	●	●	22a(2), 78(2), 79(4), 88(2), 114(7), 116(1), 118(1)	
Quedius suramensis Eppelsheim, 1880	○			So02
Quedius suturalis suturalis Kiesenwetter, 1845	○			So02
Quedius tetrapunctatus Coiffait, 1977	○			C77, IK61 [45]
**Quedius transcaucasicus* Iablokoff-Khnzorian, 1961	○			IK61
Quedius umbrinus Erichson, 1839	○	●	1(3), 6(2), 9(4), 13(1), 19(2), 20(10), 33(4), 39(6), 41(4), 49(1), 54(3), 63(2), 67(9), 75(2), 76(2), 77(5), 78(1), 81(4), 81a(5), 82(4), 89(2), 89b(2), 90(2), 90a(3), 91(5), 95(2), 97(4), 100(4), 101(2), 103(1), 105(1), 106(2), 107(3), 112(11), 114(1), 116(1), 117(3), 121(1), 121a(5), 125(3), 126(29), 130(3)	A18f, So02
Quedius vexans Eppelsheim, 1881	○			ATHW00
Quedius vulneratus Gemminger & Harold, 1868	○		4(7)	So02
Rabigus abauriae (Gridelli, 1924)	○		27(1)	C74
Rabigus pullus (Nordmann, 1837)	○		126(3)	C74
Staphylinus caesareus caesareus (Cederhjelm, 1798)	○		61(2), 141(1), 185(1), 186(1), 194(2), 195(1)	Hn65
Stenistoderus cephalotes cephalotes (Kraatz, 1858)	○			A17f, C66b [30]
#Stenistoderus nanus (Iablokoff-Khnzorian, 1961)	○			IK61
Tasgius eppelsheimianus (Jakobson, 1909)	●	●	4(1), 58(1), 80b(1), 105(2), 136(2)	

Species	AR	NK	Localities/Samples	References
Tasgius gracilicornis (Hochhuth, 1849)	○	●	103(1), 157(1), 185(1), 194(2)	AM00 [31)]
Tasgius limbifrons (Hochhuth, 1849)	○			Hh49
Tasgius pedator pedator (Gravenhorst, 1802)	○			Sm04
Tasgius protensus (Ménétriés, 1832)	○			Sm04
**Xantholinus adustus* Ushakov, 1986	○		126(1)	Uv86
Xantholinus araxis Reitter, 1898 [32)]	○		26(3)	A17f
Xantholinus audrasi Coiffait, 1956	○	●	26(1), 35(1), 36(2), 64(1), 66a(1), 73(1), 78(3), 79(4), 88(4), 89b(1), 91(1), 100(1), 103(1), 112(3), 113(1), 114(1), 121a(1), 125(1), 126(10), 179(1), 183(1), 187(1), 203a(2)	A17f, ATHW00 [33)], Bo17, C56 [33)], C70b [34)]
Xantholinus fortepunctatus Motschulsky, 1860	○		130(3)	A17f, R08
Xantholinus haematodes Kolenati, 1846		○		Ko46
Xantholinus khnzoriani Coiffait, 1966	○		7(1), 7a(3), 55(3), 68(2), 199a(2)	A17f, C66b
**Xantholinus kirschenblati* Bordoni, 1975	○	●	6(2), 12(1), 22(1), 26(1), 30(2), 32(2), 33(1), 43(2), 47(1), 51(1), 52(2), 53(2), 63(2), 64(2), 66(1), 66a(1), 68(1), 75(1), 82(1), 82a(2), 83(1), 105(1), 112(1), 124(1), 133(1), 135(2),	A17f, Bo75
Zeteotomus scripticollis (Hochhuth, 1849)	○			ATHW00, IK57a

Localities/samples:
1–40: **field trip Armenia, 2016, leg. Assing & Schülke**: **1**: N Yerevan, W Hrazdan, 40°31'13"N, 44°30'38"E, 2020 m, grassy slope with scattered oak, litter and roots sifted, 25.VI.2016; **2**: N Yerevan, W Hrazdan, 40°32'02"N, 44°33'16"E 2130 m, secondary forest margin, litter and roots sifted, 25.VI.2016; **3**: N Yerevan, W Hrazdan, 40°30'28"N, 44°34'12"E, 1870 m, mixed deciduous forest margin, litter sifted, 25.VI.2016; **3a**: same data as 3, but roots of grass and herbs sifted; **4**: N Yerevan, NW Hrazdan, 40°41'40"N, 44°29'16"E, 2500 m, grassy W-slope with scattered *Salix*, litter and roots of grass sifted 26.VI.2016; **5**: N Yerevan, NW Hrazdan, 40°40'07"N, 44°28'22"E, 2100 m, grassy W-slope with bushes, litter and grass roots sifted 26.VI.2016; **5a**: same data as 5, but W-slope near stream, swept from vegetation; **6**: N Yerevan, NW Hrazdan, 40°38'07"N, 44°30'05"E, 2010 m, mixed deciduous forest, litter and grass roots sifted, 27.VI.2016; **7**: N Yerevan, NW Hrazdan, 40°38'06"N, 44°27'37"E, 2110 m, stream valley, mixed deciduous forest, litter and grass roots sifted, 28.VI.2016; **7a**: same data as 7, but moist margin of mountain track; **8**: N Yerevan, NW Hrazdan, 40°33'45"N, 44°23'41"E, 2000 m, mixed deciduous forest margin, litter sifted, 28.VI.2016; **8a**: same data as 8, but pasture, nests of *Formica exsecta* and *Lasius flavus* sifted; **8b**: same data as 8, but stream bank with gravel; **9**: S Martuni, Sulema Pass, 39°57'58"N, 45°14'13"E, 2340 m, slope with small stream and scattered bushes, litter and roots near stream sifted, 29.VI.2016; **9a**: same data as 9, but rocky slope with grass, moss, and fern, roots and moss sifted; **10**: S Martuni, Sulema Pass, 39°58'N, 45°14'E, 2340 m, road margin with bushes, litter and gravel sifted, 29.VI.2016; **11**: ca. 50 km NW Sisian, Jermuk, 39°50'02"N, 45°40'21"E, 2110 m, oak forest and forest margin, litter and roots sifted, 30.VI.2016; **11a**: same data as 11, but 3.VII.2016; **11b**: same data as 11, but 12.VII.2016; **12**: 40 km NW Sisian, W-side of Vorotan Pass, 39°42'36"N, 45°40'30"E 1960 m, dry oak forest, litter and roots sifted, 30.VI.2016; **13**: 20 km S Sisian, Dastakert, 39°21'50"N, 46°01'27"E, 2080 m, grassy slope with scattered bushes, litter and roots beneath bushes sifted, 1.VII.2016; **14**: 20 km S Sisian, Dastakert, 39°22'10"N, 46°01'24"E, 2070 m, stream bank, in gravel and under stones, 1.VII.2016; **15**: 20 km SW Sisian, 39°25'09"N, 45°55'09"E, 1860 m, grassy slope with bushes, litter and roots sifted 1.VII.2016; **16**: 30 km NW Sisian, 39°46'33"N, 45°56'05"E, 2960 m, grassy slope with rocks, roots and debris

sifted, 2.VII.2016; **17**: 30 km NW Sisian, 39°46'36"N, 45°56'03"E, 2970 m, road margin and pasture, under stones, 2.VII.2016; **18**: 30 km NW Sisian, 39°46'53"N, 45°56'20"E, 3000 m, snowfields, under stones and debris sifted, 2.VII.2016; **19**: 35 km NW Sisian, 39°40'59"N, 45°46'50"E, 2070 m, stream valley, litter beneath bushes near stream sifted 3.VII.2016; **20**: 35 km NW Sisian, 39°41'01"N, 45°46'46"E, 2080 m, swampy meadow, debris sifted, 3.VII.2016; **20a**: same data, but litter and roots beneath bushes near small stream sifted; **21**: N Sisian, near Mt. Tsugh, 39°41'10"N, 46°03'13"E, 3300 m, soil and debris near snowfields sifted, 4.VII.2016; **21a**: same data, but meadows, partly near snowfields, under stones; **22**: 20 km SSE Goris, Shurnukh, 39°21'38"N, 46°24'33"E, 1720 m, *Quercus* and *Carpinus* forest, litter and dead wood sifted, 5.VII.2016; **22a**: same data as 22, but grassland near forest margin, litter beneath bushes sifted; **23**: WSW Kapan, Meghri Pass, 39°07'00"N, 46°09'38"E, 2520 m, moist grassy slope with scattered bushes, litter and grass roots sifted, 6.VII.2016; **23a**: same data as 23, but grassy slope, under stones; **23b**: same data as 23, but grassy slope, in mushroom; **24**: WSW Kapan, S Meghri Pass, 39°05'56"N, 46°09'47"E, 2090 m, stream valley, under stones, 6.VII.2016; **25**: SW Kapan, 15 km N Meghri, 39°01'44"N, 46°12'24"E, 1680 m, road margin, under stones, 6.VII.2016; **26**: SW Kapan, 10 km N Meghri, 38°59'22"N, 46°11'11"E, 1350 m, slope with oak, other trees, and bushes, litter sifted, 6.VII.2016; **27**: SW Kapan, 10 km N Meghri, 38°58'45"N, 46°12'32"E, 1130 m, pasture near stream, under stones, 6.VII.2016; **28**: WSW Kapan, S Meghri Pass, 39°06'10"N, 46°10'47"E. 2310 m, oak forest near tree line, oak litter sifted, 6.VII.2016; **28a**: same data as 28, but road margin, in *Formica* nest; **29**: 25 km S Kapan, Gomarants Pass, 39°01'17"N, 46°22'01"E, 2230 m, under stones, 7.VII.2016; **30**: 25 km S Kapan, N Gomarants Pass 39°01'32"N, 46°21'59"E, 2190 m, oak forest, litter and dead wood sifted, 7.VII.2016; **31**: 25 km S Kapan, N Gomarants Pass, 39°02'15"N, 46°22'13"E, 2050 m, oak forest with *Acer*, *Carpinus*, and fern undergrowth, litter and dead wood sifted, 7.VII.2016; **32**: WSW Kapan, S Meghri Pass, 39°05'20"N, 46°10'35"E, 2170 m, oak forest margin, litter (partly moist litter under bushes) sifted, 8.VII.2016; **33**: WSW Kapan, S Meghri Pass, 39°05'56"N, 46°09'47"E, 2090 m, stream valley, litter near stream sifted, 8.VII.2016; **34**: 10 km S Kapan, SE Chakaten, 39°07'50"N, 46°28'14"E, 990 m, mixed deciduous forest, litter and debris sifted, 9.VII.2016; **35**: 15 km S Kapan, SE Chakaten, 39°05'25"N, 46°28'41"E, 1010 m, mixed deciduous forest, litter between stones sifted, 9.VII.2016; **36**: 20 km Kapan, W Tsav, 39°03'04"N, 46°26'12"E, 1170 m, stream valley, litter and flood debris near stream sifted, 9.VII.2016; **36a**: same data as 36, but collected from gravel; **37**: WSW Kapan, N Meghri Pass, 39°08'08"N, 46°10'55"E, 2030 m, road margin, 10.VII.2016; **38**: 25 km SW Kapan, 39°02'26"N, 46°13'36"E, 1750 m, margin of pista, under stone, 10.VII.2016; **39**: 25 km SW Kapan, 39°04'01"N, 46°16'10"E, 2150 m, near stream, litter of *Salix* and debris sifted, 10.VII.2016; **40**: 25 km SW Kapan, 39°03'04"N, 46°15'16"E, 1890 m, stream valley, litter of *Salix* and other trees sifted, 10.VII.2016.

41–91: **field trip Armenia, 2017, leg. Assing & Schülke**: **41**: Gavar, 40°21'01"N, 45°07'45"E, 1950 m, old poplar trees near river, litter and roots of herbs sifted, 25.VI.2017; **42**: mountain range W Gavar, 40°20'31"N, 45°00'44"E, 2330 m, grassy slope, litter and roots sifted, 26.VI.2017; **43**: mountain range W Gavar, 40°20'06"N, 44°58'51"E, 2490 m, grassy slope, litter and roots sifted, 26.VI.2017; **44**: mountain range W Gavar, 40°20'N, 44°57–58'E, 2600–2700 m, horse dung sifted, 26.VI.2017; **44a**: same data as 44, but under stones; **44b**: same data as 44, but cow dung; **45**: mountain range W Gavar, 40°19'53"N, 44°56'51"E, 2700 m, snow field, debris sifted, 26.VI.2017; **45a**: same data as 45, but snow field, under stones; **46**: mountain range W Gavar, 40°19'59"N, 44°56'53"E, 2700 m, debris near snow field and moist litter on slope sifted, 26.VI.2017; **47**: mountain range W Gavar, 40°20'40"N, 45°00'52"E, 2250 m, grassy slope, tall herbs and roses, roots and debris sifted, 26.VI.2017; **48**: Vardenis mountain range SE Vardenis, 40°05'14"N, 45°49'22"E, 2330 m, stream valley with *Salix*, litter and roots beneath *Salix* sifted, 27.VI.2017; **49**: Vardenis mountain range SE Vardenis, 40°02'38"N, 45°46'12"E, 2730 m, slope below snowfields, stream bank, debris sifted, 27.VI.2017; **49a**: same data as 49, but grass and herb roots in shade of large rocks sifted; **50**: Sevani mountain range E Sevan lake, Tsapatagh env., 40°23'51"N, 45°29'53"E, 2025 m, oak forest margin, litter and roots sifted, 28.VI.2017; **50a**: stream valley, litter and roots beneath bush at stream bank sifted; **51**: SSE Dilijan, NW Semyonovka, 40°40'09"N, 44°52'35"E, 2050 m, *Sorbus* forest, litter sifted, 29.VI.2017; **52**: SSE Dilijan, NW Semyonovka, 40°40'12"N, 44°53'05"E, 1900 m, stream valley, forest margin and bushes, sifted, 29.VI.2017; **53**: WSW Dilijan, Kalavan, 40°38'40"N, 45°06'04"E, 1700 m, calcareous S-slope with stream valley and oak forest, litter and roots sifted, 29.VI.2017; **54**: NE Sevan Lake, Karmir pass, 40°33'48"N, 45°18'01"E, 2150 m, debris, grass, and roots beneath scattered *Sorbus* sifted, 30.VI.2017; **55**: E Dilijan, road Ttujur–Berd, 40°41'05"N, 45°20'18"E, 1900 m, moist mixed deciduous forest with *Fagus orientalis*, *Sorbus*, etc., litter and roots sifted, 30.VI.2017; **56**: E Dilijan, road Ttujur–Berd, 40°41'47"N, 45°19'23"E, 2040 m, litter near small stream sifted, 30.VI.2017; **57**: E Dilijan, road Ttujur–Berd 40°44'04"N, 45°18'10"E, 1930 m, slope with beech and bushes, litter, roots, and mushrooms sifted, 30.VI.2017; **58**: road Berd–Ijevan, 40°52'26"N, 45°18'00"E, 1350 m, beech forest margin, litter and roots sifted, 30.VI.2017; **59**: road Dilijan–Vanadzor, 40°46'15"N, 44°37'02"E, 1790 m, birch forest, litter sifted, 1.VII.2017; **60**: N Vanadzor, N Pushkin pass, 40°55'15"N, 44°26'38"E, 1900 m, mixed forest (oak, pine, etc.), litter sifted, 1.VII.2017; **61**: N Vanadzor, S Pushkin pass, 40°54'26"N, 44°25'36"E, 1880 m, N-slope with scattered bushes, debris and grass roots sifted, 1.VII.2017; **62**: N Vanadzor, S Pushkin pass, 40°54'15"N, 44°25'46"E, 1850 m, forest with *Quercus*, *Fagus*, *Sorbus*, etc., litter and roots sifted, 1.VII.2017; **63**: N Vanadzor, S Pushkin pass, 40°54'00"N, 44°25'55"E, 1780 m, forest margin with *Quercus*, *Fagus*, *Sorbus*, etc., litter and roots near small stream sifted, 1.VII.2017; **64**: SW Gavar, 40°15'18"N, 45°09'40"E, 2170 m, N-slope with scattered bushes, litter and roots roots sifted, 2.VII.2017; **65**: SW Gavar, 40°15'10"N,

45°09'25"E, 2180 m, grassy N-slope near lake, under stones, 2.VII.2017; **66**: ENE Dilijan, Hovk, 1290 m, 40°47'39"N, 45°01'17"E, stream valley, litter beneath *Acer* sifted, 3.VII.2017; **66a**: same data as 66, but stream valley, moist litter near stream sifted; **66b**: same data as 66, but forest (*Quercus*, *Fagus*, *Acer*, etc.) margin, litter and dead wood beneath old *Fagus* sifted; **67**: SW Gavar, 40°14'31"N, 45°01'41"E, 2570 m, stream valley, moist litter and roots near stream sifted, 4.VII.2017; **68**: WSW Dilijan, Kalavan, 40°37'52"N, 45°05'46"E, 1960 m, forest with *Quercus*, *Betula*, and *Carpinus*, litter sifted, 5.VII.2017; **68a**: same data as 68, but bark of dead oak sifted; **69**: WSW Dilijan, Kalavan, 40°37'45"N, 45°05'31"E, 2100 m, forest at timber line with oak and birch, litter and large fungus on oak sifted, 5.VII.2017; **70**: pass road E Ijevan, 40°52'00"N, 45°13'19"E, 1790 m, forest with old *Quercus* and *Carpinus*, litter and roots sifted, 6.VII.2017; **71**: pass road E Ijevan, 40°51'51"N, 45°11'52"E, 1630 m, oak and beech forest, litter and roots sifted, 6.VII.2017; **72**: pass road E Ijevan, 40°52'02"N, 45°11'03"E, 1400 m, young beech forest, litter and dead wood sifted, 6.VII.2017; **73**: S Spitak, 40°45'45"N, 44°16'23"E, 2000 m, deforested stream valley, bank of small stream, roots and moss sifted, 7.VII.2017; **73a**: same data as 73, but under stones, 7.VII.2017; **74**: S Spitak, 40°45'34"N, 44°16'23"E, 2070 m, deforested stream valley with scattered trees and bushes, litter and roots beneath small trees and bushes sifted, 7.VII.2017; **75**: SE Spitak, 40°48'07"N, 44°18'11"E, 1840 m, oak forest margin, litter and roots sifted, 7.VII.2017; **76**: S Martuni, Sulema Pass, 39°57'58"N, 45°14'13"E, 2340 m, slope with small stream and scattered bushes, litter and roots near stream sifted, 8.VII.2017; **77**: 40 km NW Sisian, Vorotan Pass, 39°41'48"N, 45°41'28"E, 2090 m, grassy slope with scatteres bushes, litter, debris, and moss sifted, 8.VII.2017; **78**: E Goris, Khndzoresk env., 39°31'32"N, 46°24'21"E, 1590 m, ruderal grassy slope with *Rosa* bushes and *Rubus*, litter and roots sifted, 9.VII.2017; **79**: E Goris, Khndzoresk env., 39°30'58"N, 46°27'21"E, 1350 m, slope with dense small trees, litter and grass roots sifted, 9.VII.2017; **80**: pass N Goris, 39°34'53"N, 46°19'39"E, 2110 m, grassy slope with stones, under stones, 9.VII.2017; **80a**: same data as 80, but mouse nest under stone sifted; **80b**: same data as 80, but small straw heap sifted; **81**: pass N Goris, 39°35'34"N, 46°19'29"E, 1990 m, N-slope with small stream valleys with and without water, litter, debris, and moss sifted, 9.VII.2017; **81a**: same data as 81, but N-slope with small stream valleys with water, litter, debris, and moss sifted, 10.VII.2017; **81b**: same data as 81a, but grassy N-slope, under stone; **82**: 40 km NW Sisian, Vorotan Pass, 39°40'33"N, 45°45'07"E, 2140 m, stream valley with *Salix*, litter and roots sifted, 10.VII.2017; **82a**: same data as 82, but stream valley, rose bushes, litter and roots sifted; **83**: 40 km NW Sisian, Vorotan Pass, 39°41'19'N, 45°41'48"E, 2160 m, stream valley, rose bushes, litter and roots sifted, 10.VII.2017; **84**: 30 km NW Sisian, Mt. Karkar, 39°46'38"N, 45°56'04"E, 2970 m, pasture, mouse nest under stone sifted, 11.VII.2017; **85**: 30 km NW Sisian, Mt. Karkar, 39°47'05"N, 45°56'22"E, 3000 m, grassland, under stones, 11.VII.2017; **85a**: same data as 85, but mouse nests sifted; **85b**: same data as 85, but horse dung sifted; **85c**: same data as 85, but wetland, moist debris near water sifted; **86**: 30 km NW Sisian, Mt. Karkar, 39°46'51"N, 45°56'30"E, 2990 m, straw manure sifted, 11.VII.2017; **87**: 30 km NW Sisian, Mt. Karkar, 39°46'38"N, 45°56'34"E, 3000 m, N-slope, debris between stones and under plants sifted, 11.VII.2017; **87a**: same data as 87, but grassy N-slope with rocks, under stones; **88**: NW Goris, W Verishen, 39°31'43"N, 46°18'33"E, 1670 m, margin of oak forest, litter, moss, and roots beneath *Quercus* and *Rosa* sifted, 12.VII.2017; **89**: SW Goris, ESE Tatev, 39°21'47"N, 46°16'52"E, 1950 m, margin of mixed deciduous forest (*Quercus*, *Carpinus*, *Acer*, etc.), litter and roots sifted, 13.VII.2017; **89a**: same data as 89, but nests of *Formica exsecta* sifted; **89b**: same data as 89, but 14.VII.2017; **90**: SW Goris, ESE Tatev, 39°21'32"N, 46°16'05"E 1730 m, margin of mixed deciduous forest (*Quercus*, *Carpinus*, *Acer*, etc.), litter and debris in or near small ditch near road sifted, 13.VII.2017; **90a**: same data as 90, but mixed deciduous forest (*Quercus*, *Carpinus*, *Acer*, etc.), stream valley, moist litter near small stream sifted; **91**: SW Goris, ESE Tatev, 39°21'50"N, 46°16'32"E 1820 m, margin of mixed deciduous forest (*Quercus*, *Carpinus*, *Acer*, etc.), litter and debris between rocks sifted, 14.VII.2017.

92–131: **field trip Armenia and Nagorno-Karabakh, 2018, leg. Assing & Schülke [92–97, 118–131**: **Armenia; 98–117**: **Nagorno-Karabakh**]: **92**: NW Yerevan, Aragats, 40°23'28"N, 44°15'28"E, 2110 m, oak forest, litter sifted, 29.VI.2018; **93**: NW Yerevan, Aragats, 40°24'22"N, 44°13'40"E, 2200 m, moist slope, sifted, 29.VI.2018; **93a**: same data as 93, but near stream, in gravel near puddle; **94**: NW Yerevan, Aragats, 40°28'20"N, 44°11'14"E, 3150 m, debris below snow field sifted, 29.VI.2018; **94a**: same data as 94, but meadow near lake and slope below snowfield, under stones; **95**: N Yerevan, Mt. Arayilor, 40°25'54"N, 44°27'41"E, 2010 m, oak forest, litter sifted, 30.VI.2018; **96**: N Yerevan, Mt. Arayilor, 40°25'35"N, 44°27'58"E, 2090 m, oak forest, litter sifted, 30.VI.2018; **97**: N Yerevan, Buzhakan, 40°27'58"N, 44°30'47"E, 1930 m, oak forest, litter sifted, 30.VI.2018; **98**: ca 12 km NW Stepanakert, 39°54'22"N, 46°39'12"E, 870 m, mixed forest with Fagus, *Acer*, *Tilia*, *Corylus*, etc., litter sifted, 2.VII.2018; **99**: ca. 12 km NW Stepanakert, 39°54'58"N, 46°40'14"E, 810 m, stream bank, floated from gravel, 2.VII.2018; **100**: ca. 40 NW Stepanakert, W Vank, 40°01'04"N, 46°26'32"E, 1420 m, S-slope with mixed deciduous forest, moist litter near small stream sifted, 3.VII.2018; **101**: ca. 40 km NW Stepanakert, W Vank, 40°01'34"N, 46°28'33"E, 1210 m, deciduous forest on river bank, litter sifted, 3.VII.2018; **101a**: same data as 101, but deciduous forest, debris and litter near small waterfall sifted; **102**: ca. 40 km NW Stepanakert, W Vank, 40°01'44"N, 46°30'15"E, 1080 m, mixed deciduous forest, dry leaf litter sifted, 3.VII.2018; **103**: ca. 75 km WNW Stepanakert, S Karvachar, 40°02'02"N, 46°00'10"E, 1630 m, degraded mixed deciduous forest with small stream, litter sifted, 4.VII.2018; **104**: ca. 75 km WNW Stepanakert, S Karvachar, 40°01'24"N, 45°59'28"E, 1680 m, river bank, *Salix* litter sifted, 4.VII.2018; **105**: ca. 50 km WNW Stepanakert, S Dadivank, 39°59'13"N, 46°14'45"E, 1610 m, mixed deciduous forest margin, litter and roots sifted, 5.VII.2018; **106**: ca. 50 km WNW Stepanakert, S Dadivank, 40°00'40"N, 46°14'00"E,

1560 m, mixed deciduous forest with small stream, moist litter and debris near stream sifted, 5.VII.2018; **107**: pass 20 km SW Stepanakert, 39°40'40"N, 46°39'56"E, 1940 m, stream bank, litter and roots sifted, 6.VII.2018; **108**: pass 20 km SW Stepanakert, 39°41'41"N, 46°39'02"E, 1760 m, secondary mixed forest with pine and oak, 6.VII.2018; **109**: pass 20 km SW Stepanakert, 39°41'35"N, 46°39'00"E, 1760 m, stream bank with *Salix* and herbs, litter and roots sifted, 6.VII.2018; **109a**: same data as 109, but *Formica exsecta* nest sifted; **110**: pass 20 km SW Stepanakert, 39°41'50"N, 46°39'00"E, 1730 m, stream valley with mixed deciduous forest, predominantly oak, litter sifted, 6.VII.2018; **111**: Stepanakert, hotel, 39°49'38"N, 46°45'54"E, 800 m, at light, 6.VII.2018; **112**: 8 km N Berdzor, 39°41'31"N 46°32'10"E, 1220 m, N-slope with mixed deciduous forest and small stream, litter and debris near stream sifted, 7.VII.2018; **113**: pass 20 km SW Stepanakert, 39°42'13"N 46°39'02"E, 1700 m, stream valley with deciduous trees, litter near stream and near rocks sifted, 7.VII.2018; **114**: 13 km W Hadrut, 39°31'05"N, 46°54'19"E, 1760 m, mixed deciduous forest with *Quercus*, *Acer*, *Carpinus*, *Corylus*, litter in and near moist stream bed sifted, 8.VII.2018; **115**: 8 km W Hadrut, 39°31'52"N 46°57'50"E, 1480 m, stream valley with mixed deciduous forest, moist litter near stream sifted, 8.VII.2018; **116**: mountain ca. 10 km SE Stepanakert, 39°47'03"N, 46°51'02"E, 1520 m, mixed deciduous forest, litter sifted, 9.VII.2018; **117**: ca. 30 km NW Stepanakert, SE Vank, 40°01'30"N, 46°33'21"E, 1050 m, deciduous forest with stream, litter and debris near stream sifted, 9.VII.2018; **118**: WSW Kapan, W Kajaran, 39°09'22"N, 46°06'13"E, 2050 m, steep N-slope with mixed deciduous forest, leaf litter and grass roots sifted, 10.VII.2018; **118a**: same data as 118, but 13.VII.2018; **119**: ca. 30 km W Kapan, 39°15'13"N, 46°04'16"E, 2040 m, N-slope, margin of mixed deciduous forest, litter and grass roots sifted, 11.VII.2018; **120**: ca. 30 km W Kapan, 39°14'40"N, 46°06'24"E, 1780 m, river bank, floated from gravel, 11.VII.2018; **121**: ca. 30 km W Kapan, 39°14'36"N, 46°08'28"E, 1680 m, N-slope with trickling water, moist litter and debris sifted, 11.VII.2018; **121a**: same data as 121, but river bank, litter and debris sifted; **122**: SW Kapan, Mt. Khustup, 39°07'52"N, 46°19'43"E, 3060 m, N-slope, soil and roots of *Rumex* sp. near large rocks sifted, 12.VII.2018; **123**: SW Kapan, Mt. Khustup, 39°07'50"N, 46°19'55"E, 2980 m, litter near spring sifted, 12.VII.2018; **124**: SSW Kapan, forest above Shishkert, 39°03'48"N, 46°21'36"E, 2040 m, roots, litter, and stony soil sifted, 12.VII.2018; **125**: WSW Kapan, S Lernadzor, 39°08'02"N, 46°13'07"E, 2000 m, dark stream valley, litter and roots near stream sifted, 13.VII.2018; **126**: 25 km SW Kapan, Vank, 39°04'01"N, 46°16'10"E, 2150 m, near stream, litter of *Salix*, roots, and debris sifted, 14.VII.2018; **127**: 25 km S Goris, ESE Tatev, 39°20'18"N, 46°22'06"E, 1590 m, mixed deciduous forest with small stream, moist litter sifted, 15.VII.2018; **128**: 30 km NW Sisian, Mt. Karkar, 39°47'05"N, 45°56'22"E, 3000 m, wetland, wet litter near shore of lake sifted, 15.VII.2018; **129**: 20 km SW Sisian, Darbas, 39°25'55"N, 46°06'57"E, 1680 m, margin of secondary deciduous forest, litter and soil sifted, 16.VII.2018; **130**: reservoir 15 km SW Sisian, 39°28'18"N, 46°07'57"E, 1340 m, shore of reservoir, *Salix* litter, moss, and roots sifted, 16.VII.2018; **131**: 35 km SSE Martuni, Arates, 39°54'28"N, 45°26'07"E, 2040 m, margin of small forest patch with small stream, litter and roots sifted, 17.VII.2018.

132–144: **field trip Armenia, 2016, leg. A. & J. Müller**: **132**: Ditavan env., 40°58'49"N, 45°11'57"E, 610 m, 21.V.2016; **133**: Ditavan env., 40°56'45"N, 45°12'20"E, 1240 m, 22.V.2016; **134**: Ditavan env., 40°58'07"N, 45°12'08"E, 740 m, 22.V.2016; **134a**: same data as 134, but 23.V.2016; **134b**: Ditavan env., 40.94750°N, 45.20793°E, 1290 m, 28.IX.2016; **135**: Ayrum env., Zikatar Environmental Center, 41°07'34"N, 44°55'21"E, 1250 m, 19.V.2016; **136**: Teghut env. (Akhtala), 41.091°N, 44.813°E, 1020 m, 20.V.2016; **137**: Teghut env. (Dilijan), 41°05'26"N, 44°48'51"E, 1340 m, 17.V.2016; **138**: Aragazotn, Byurakan env., 40°23'42"N, 44°18'40"E, 1930 m, 26.V.2016; **139**: Aragazotn, Mt. Aragats, 40°25'57"N, 44°14'01"E, 2680 m, 26.V.2016; **140**: Aragazotn, Byurakan env., 40°24'21"N, 44°13'39"E, 2190 m, 26.V.2016; **141**: Tsovagyugh env., 40.653°N, 44.898°E, 2010 m, 24.V.2016; **142**: Hovk env. (Dilijan), 40°47'05"N, 45°03'22"E, 850 m, 24.V.2016; **143**: Dilijan env., 40°41'50"N, 44°51'29"E, 1670 m, 17.V.2016; **144**: Dsegh env. (Tsover Lake), 40.950°N, 44.685°E, 1390 m, 23.V.2016.

145–160: **Armenia, 2013–2017, leg. Kalashian, Grejyan & Karagyan**: **145**: Vayotsdzor prov., Amulsar mine E of Jermuk and Gndevaz, 39.74646°N, 45.66483°E, 2040 m, 28.VI.–18.VII.2017; **145a**: same data as 145, but 18.VII.–7.VIII.2017; **145b**: same data as 145, but 7.VIII.–23.IX.2017; **146**: Vayotsdzor prov., Amulsar mine E of Jermuk and Gndevaz, 39.7335°N, 45.6664°E, 1985 m, 7.VIII.–23.IX.2017; **146a**: same data as 146, but 18.VII.– 7.VIII.2017; **147**: Vayotsdzor prov., Amulsar mine E of Jermuk and Gndevaz, 39.72721°N, 45.71643°E, 2975 m, 18.VII.– 7.VIII.2017; **148**: Lori prov., Teghut mine, near artificial pond, 41.0907°N, 44.8117°E, 990 m, soil traps, 28.IV.–25.V.2017; **148a**: same data as 148, but 25.V.–23.VII.2017; **148b**: same data as 148, but 23.VII.–13.VIII.2017; **148c**: same data as 148, but 13.VIII.–13.X.2017; **148d**: same data as 148, but 19.VI.–23.VII.2015; **148e**: same data as 148, but 23.VII.–23.VIII.2015; **148f**: same data as 148, but 1.–24.V.2015; **148g**: same data as 148, but 23.VIII.–23.IX.2015; **149**: Lori prov., Teghut mine, near refuse heap, 41.0705°N, 44.8378°E, 1210 m, soil traps, 28.IV.–25.V.2017; **149a**: same data as 149, but 23.VII.–13.VIII.2017; **149b**: same data as 149, but 13.VIII.–13.X.2017; **150**: Lori prov., Teghut mine, Dukanadzor gorge, 41.0910°N, 44.8514°E, 890 m, soil traps, 28.IV.–25.V.2017; **150a**: same data as 150, but 25.V.–23.VII.2017; **150b**: same data as 150, but 19.VI.–23.VII.2015; **150c**: same data as 150, but 1.–24.V.2014; **150d**: same data as 150, but 23.VIII.–23.IX.2015; **151**: Lori prov., Teghut mine, Dukanadzor gorge, Kharatanots gorge, 41.0964°N, 44.8219°E, 910 m, soil traps, 28.IV.–25.V.2017; **151a**: same data as 151, but 25.V.–23.VII.2017; **151b**: same data as 151, but 23.VII.–13.VIII.2017; **151c**: same data as 151, but 19.VI.–23.VII.2015; **151d**: same data as 151, but 920 m, 1.–4.V.2014; **152**: Lori prov., Teghut mine, 920 m, 20.VI.–1.VIII.2013; **153**: Lori prov., Teghut mine, 1100 m, 22.VIII.–1.X.2013; **154**: Lori prov., Teghut mine, tailing

dump, 1100 m, soil traps, 1.–24.V.2014; **155**: Syunik prov., ~5 km N Shvanidzor, 38.9851°N, 46.3746°E, 1335 m, soil traps, 9.VI.–8.VIII.2017; **156**: Syunik prov., between Tandzaver and Aghvani, 39.3499°N, 46.3164°E, 1670 m, 5–30. VII.2015; **157**: Syunik prov., N Verin Khotanan, 39.3284°N, 46.3751°E, 1660 m, 5–30.VII.2015; **158**: Syunik Province, S Dastakert, 1930 m, 29.VII.2011; **159**: Vayotsdzor Province, E Gndevaz, 1980 m, soil traps, 5–22.VI.2014; **160**: Vayotsdzor Province, E Gndevaz, 2190 m, soil traps, 5.–22.VI.2014.

161–174: **2015, Armenia, leg. Kocian**: **161**: Dashtakar env., Vedi river, 39.920756°N, 44.782199°E, 1000 m, river bank, 18.V.2015; **162**: Lanjanist env., 39.859084°N, 44.937384°E, 1560 m, leaf litter and dead wood sifted, 18.V.2015; **163**: Jermuk env., 39.814797°N, 45.644753°E, 2060 m, W slope, deciduous forest, sifted, 19.V.2015; **164**: Yeghegis env., 39.871366°N, 45.411616°E, 1700 m, N slope, deciduous forest, sifted, 120.V.2015; **165**: above Jermuk, 39.839053°N, 45.693496°E, 2400 m, plant debris near snow fields sifted, 21.V.2015; **166**: Jermuk env., 39.806823°N, 45.650697°E, 2000 m, 19.V.2015; **167**: Artavan valley, 39.676737°N, 45.569628°E, 1530 m, river bank, leaf litter sifted, 22.V.2015; **168**: Vorotan pass, 39.681390°N, 45.720964°E, 2250 m, grass and debris sifted, 23.V.2015; **169**: Tatev env., Vorotan valley, 39.394394°N, 46.247672°E, 1300 m, macchia, sifted, 24.V.2015; **170**: Goris, 39.516611°N, 46.322847°E, 1700 m, E slope, deciduous forest, leaf litter sifted, 25.V.2015; **171**: Goris env. Khndzoresk, old cemetery, 39.502770°N, 46.428540°E, 1400 m, macchia, sifted, 26.V.2015; **172**: Goris env., above Verishen, 39.558515°N, 46.316772°E, 1880 m, sifted, 26.V.2015; **173**: Vorotan pass, 39.686492°N, 45.693326°E, 2160 m, grass and plant debris sifted, 27.V.2015; **174**: 3 km S Nshkhark, 39.971887°N, 45.241237°E, 2280 m, grass and plant sifted, 27.V.2015.

175–213: **2001, Armenia, leg. Shaverdo & Schillhammer**: **175**: N Yerevan, SW Carencavan, SE Lusakert, Hrazdan river, ~1280 m, 30.IV.2001; **176**: E Yerevan, approximately halfway between Yerevan and Garni, below Atsavan, ~ 1300 m, lake with reed, 1.V.2001; **177**: SE Yerevan, Azat reservoir near Bartsrashen, close to inflow of river, ~ 1040 m, 3.V.2001; **178**: E Yerevan, Garni gorge near Garni, 40°07'N, 44°44'E, ~ 1200 m, confluence of Gekhard and Azat rivers, 5.V.2001; **179**: Mt. Aragats, S-slope, NW Byurakan, near Amberd, 40°24'N, 44°14'E, ~ 2100 m, small river, 6.V.2001; **180**: N Yerevan, NW Charentsavan, Arzakan env., near Agveran resort, 40°31'N, 44°36'E, ~ 1850 m, leaf litter along bank of forest stream sifted, 7.V.2001; **181**: N Yerevan, NW Charentsavan, Arzakan env., 40°30'N, 44°35'E, ~ 1850 m, very wet leaf litter along bank of small tributary to "Arzakan" river sifted, 7.V.2001; **182**: N Yerevan, 24 km NW Hrazdan, below Ankavan, Marmaryk river, 40°38'N, 44°32'E, ~ 1970 m, 9.V.2001; **183**: 21 km NW Hrazdan, 40°38'N, 44°33'E, ~ 1850 m, flooded grassland near Marmaryk river, 9.V.2001; **184**: N Aparan, 40°37'N, 44°20'E, ~ 1950 m, Kasagh river, 10.V.2001; **185**: Mt. Aragats, S-slope, Amberd river above Byurakan, 40°22'N, 44°15'E, ~ 1580 m, near river bank, 10.V.2001; **186**: SE Yerevan, E Yeghegnadzor, 7 km E Bazarchay, Vorotan river above Sisian reservoir, 39°41'N, 45°47'E, ~ 2050 m, 12.V.2001; **187**: SE Yerevan, E Yeghegnadzor, 2 km W Karmrashen, 39°50'N, 45°31'E, ~ 2050 m, stream flowing through alpine plain, 13.V.2001; **188**: SE Yerevan, NE Yeghegnadzor, E Gyullidus, Gyadikvank env., Yeghegis river, 39°53'N, 45°29'E, ~ 1850 m, river bank with *Salix*, 13.V.2001; **189**: SE Yerevan, NE Areni, W Lusashogh, 39°52'N, 44°57'E, ~ 1700 m, pond, 14.V.2001; **190**: W Yerevan, SW Echmiadzin, Yerasghaun env., 40°05'N, 44°12'E, ~ 850 m, 15.V.2001; **191**: SE Yerevan, Khosrov Nature Reserve, 39°59'N, 44°53'E, 1300–1500 m, 16–17.V.2001; **191a**: same as 191, but ~ 1320 m, leaf litter sifted; **192**: E Yerevan, Gekhard env., Gekhard river, 40°08'N, 44°49'E, ~ 1600 m, 18.V.2001; **193**: E Yerevan, Gekhard env., between Gekhard and Goght, 1600–1800 m, 18.V.2001; **194**: 32 km SE Vanadzor, above Fioletovo, 40°43'N, 44°44'E, ~ 1730 m, 20.V.2001; **195**: NE Dilijan, near Haghartsin monastery, 40°48'N, 44°53'E, ~ 1450 m, along riverbanks and under stones on slopes, 21.V.2001; **195a**: same as 195, but sifted from dripping wet leaf litter along river bank; **196**: NE Dilijan, road to Haghartsin, 40°48'N, 44°46'E, ~ 1250 m, 21.V.2001; **196a**: same as 196, but leaf litter sifted; **197**: same as 196, but 40°48'N, 44°54'E, ~ 1300 m, deep car tracks; **198**: E Dilijan, Parzlich Lake, 40°45'N, 44°58'E, ~ 1330 m, puddle in forest, 22.V.2001; **199**: E Dilijan, below Gosh, 40°44'N, 45°01'E, ~ 1000 m, bank of small stream, 22.V.2001; **199a**: same as 199, but sifted from very wet leaf litter at base of slope along bank of stream; **199b**: same as 199, but under stones near stream; **200**: NE-shore of Sevan Lake, 40°36'N, 45°04'E, ~ 1940 m, from vegetation; **201**: ca 10 km N Sevan, Tzovagyugh env., 40°36'N, 44°57'E, ~ 1940 m, large swampy area and flooded meadow along channels, 22.V.2001; **202**: same as 182, but 27.V.2001; **203**: same as 183, but 27.V.2001; **203a**: same as 203, but under stones; **204**: same as 179, but 28.V.2001; **204a**: same as 204, but terrestrial; **205**: 25 km N Kapan, Nor Arajadzor env., SW-shore of water reservoir "Davidbeksoe", ~ 1400 m, lakeshore, 30.V.2001; **206**: 25 km N Kapan, Nor Arajadzor env., Khashuni river bank, 30.V.2001; **207**: between Kapan and Kajaran, Darmanadzor (Aramazd) gorge, ~ 1700 m, under stones, 30.V.2001; **208**: 3 km W Kadzharan, "Old Kajaran" env., ~ 2000 m, puddles on the banks of Kajaran river, 31.V.2001; **209**: 16 km SW Kajaran, ~ 2130 m, stream, 31.V.2001; **210**: same as 209, but puddle near larger stream; **211**: SSE Sisian, Lernashen env., ~ 1870 m, under stones, 1.VI.2001; **212**: SE Yerevan, ca 10 km E Artashat, foot of Mt. Otsasar, ~ 1050 m, semi-desert, 2.VI.2001; **213**: E Yerevan, Gekhard river near Garni, 1200 m, 29.IV.2001.

Additional records, Armenia: **a**: 12 km SE Armavir, Arazap env., Araratskaya Koshenil Reserve, salt marsh, 40°04'N, 44°08'E, 850 m, 13.VI.2017, leg. Šumpich (NMP, cAss); **b**: 3.2 km NW Meghri, Artsvakar, Arevik National Park, 38°55'N, 46°16'E, 750 m, rocky steppe, gorge, 6–7.VI.2017, leg. Šumpich (NMP); **c**: Vedi env., Goravan, Goravan Sands Reserve, 960 m, 38°55'N, 44°44'E, sandy steppe, 31.VI.2017, leg. Šumpich (NMP); **d**: Shvanidzor env., Arevik National Park, 38°57'N, 46°23'E, 780 m, steppe, 8.VI.2017, leg. Šumpich (NMP); **e**: Areni env., Noravank monastery, 39°42'N, 45°13'E, 1330 m, 10.VI.2017, leg. Šumpich (NMP); **f**: 2.3 km NW Aygedzor, Lichtkvaz, Arevik National Park, 38°59'N,

46°11'E, 1335 m, mountain steppe, 4.VI.2017, leg. Šumpich (NMP); **g**: 3 km NW Meghri, Lehvaz env., Arevik National Park, 38°55'N, 46°13'E, 840 m, rocky steppe, 5.VI.2017, leg. Šumpich (NMP); **h**: 5.5 km N Shvanidzor, 4 km S Gomarants pass, Arevik National Park, 38°59'N, 46°22'E, 1330 m, 9.VI.2017, leg. Šumpich (NMP); **i**: 13 km SE Yerevan, Hatsavan env., Azat Reservoir, 40°05'N, 44°37'E, steppe, 11.VI.2017, leg. Šumpich (NMP); **j**: Aragats Mt., Lake Karilitsh, 3300 m, 21.IX.1982, leg. Merkl et Ronkay (MNB); **k**: Tsakhkadzor, 2800 m, under stones, 1.X.1982, leg. Merkl & Ronkay (MNB); l: Aragats Mt., 3200 m, 27.VIII.1976, leg. Vásárhelyi (HNHM); **m**: Taakhkadzor, 1800–2300 m, 1.X.1982, leg. Merkl & Ronkay (HNHM); **n**: Yerevan, 11.V.1951, leg. Khnzorian (cKhn); **o**: Kotayk province, Tsakhkadzor, 17.IV.1978, leg. Khnzorian (cKhn); **p**: Lori province, "Kirowakan, Chagali" [= Vanadzor], 2.VI.1948, leg. Khnzorian (cKhn); **q**: Lori province, Stepanavan, "Yagan", 17.VI.1955, leg. Khnzorian (cKhn); **r**: Kotayk province, Tsakhkadzor, 10.V.1949, leg. Khnzorian (cKhn); **s**: Lori province, Vanadzor, 2.VI.1949, leg. Khnzorian (cKhn); **t**: Goris, "Shurnukhi", 9.VIII.1950, leg. Khnzorian (cKhn); **u**: Tavush province, Ijevan, 24.V.1951, leg. Khnzorian (cKhn); **v**: Yerevan, Jrvezh, 9.VI.1952, leg. Khnzorian (cKhn); **w**: Tavush province, Alaverdi, Akhtala, 6.VI.1940, leg. Khnzorian (cKhn); **x**: "Basargenar, Mazr. Kochewki", 22.VII.1958, leg. Khnzorian (cKhn); **y**: Syunik province, Kapan, 18.VII.1952, leg. Khnzorian (cKhn); **z**: Yerevan, zoo, 16.XI.1952, leg. Khnzorian (cKhn); **aa**: Yerevan, Jrvezh, 9.VI.1952, leg. Khnzorian (cKhn); **ab**: Tavush province, Dilijan, Haghartsin, 10.V.1975, leg. Khnzorian (cKhn); **ab**: Tavush province, Dilijan, Haghartsin, 10.V.1975, leg. Khnzorian (cKhn); **ac**: Lori province, Vanadzor, "Magaly", 1.VI.1949, leg. Khnzorian (cKhn); **ad**: Kotayk province, Yerevan, Geghard, 3.V.1950, leg. Khnzorian (cKhn); **ae**: Yerevan, "Bushakan", 25.VI.1959, leg. Khnzorian (cKhn); **af**: Aragats, "Karilik", 29.VIII.1948, leg. Khnzorian (cKhn); **ag**: Alaverdi, Sanahin, 5.VI.1949, leg. Khnzorian (cKhn); **ah**: Lori province, Stepanavan, Gyulagarak, 18.VII.1954, leg. Khnzorian (cKhn); **ai**: Sevan, "Khitapalar", 1.VIII.1948, leg. Khnzorian (cKhn); **aj**: Gegharkunik province, Sevan lake, Chambarak, 2700 m, 11.VI.1949, leg. Khnzorian (cKhn); **ak**: Argatsotn province, Aragats, K'ari Lich, 29.VIII.1948, leg. Khnzorian (cKhn); **al**: Vayots Dzor province, Yeghegnadzor, Arpa river, 20.VII.1949, leg. Khnzorian (cKhn); **am**: Syunik province, Kapan, Vachagan [39°11'N 46°23'E], 16.VII.1952, leg. Khnzorian (cKhn); **an**: Vayots Dzor province, Yeghegnadzor, Shatin [39°50'N, 45°18'E], 16.VII.1950, leg. Khnzorian (cKhn); **ao**: Lori province, Vanadzor, 2.VI.1949, leg. Khnzorian (cKhn); **ap**: Lori province, Vanadzor, "Vartanidzor", 20.V.1950, leg. Khnzorian (cKhn); **aq**: Lori province, Vanadzor, Pambak range, Mt. Maimekh, 21.V.1950, leg. Khnzorian (cKhn); **ar**: Yerevan, "Digrliuz", 14.IV.1950, leg. Khnzorian (cKhn).

References:

A97 = Assing (1997); A99 = Assing (1999); A03 = Assing (2003); A04a = Assing (2004a); A05a = Assing (2005a); A05b = Assing (2005b); A06 = Assing (2006); A07b = Assing (2007b); A08a = Assing (2008a); A08b = Assing (2008b); A08c = Assing (2008c); A09a = Assing (2009a); A10a = Assing (2010a); A10b = Assing (2010b); A13a = Assing (2013a); A13b = Assing (2013b); A14 = Assing (2014); A15 = Assing (2015); A16a = Assing (2016a); A16b = Assing (2016b); A16c = Assing (2016c); A17a = Assing (2017a); A17b = Assing (2017b); A17c = Assing (2017c); A17d = Assing (2017d); A17e = Assing (2017e); A17f = Assing (2017f); A17g = Assing (2017g); A18a = Assing (2018a); A18b = Assing (2018b); A18c = Assing (2018c); A18d = Assing (2018d); A18e = Assing (2018e); A18f = Assing (2018f); A18g = Assing (2018g); A18h = Assing (2018h); A19a = Assing (2019a); A19b = Assing (2019b); AKÖ11 = Anlaş et al. (2011); Am99 = Amiryan (1999a); Am99b = Amiryan (1999b); Am00 = Amiryan (2000); An17 = Anlaş (2017); An18 = Anlaş (2018); ASpp = Assing & Schülke (present paper); ASo98 = Assing & Solodovnikov (1998); ATHW00 = Amiryan et al. (2000); AV17 = Assing & Vogel (2017); AW08 = Assing & Wunderle (2008); Be74 = Benick (1974); BK07 = Besuchet & Kurbatov (2007); Bo84 = Bordoni (1984); Bo11a = Bordoni (2011a); Bo11b = Bordoni (2011b); Bo17 = Bordoni (2017); Br01 = Bernhauer (1901); Br08 = Bernhauer (1908); Br44 = Bernhauer (1944); BrS10 = Bernhauer & Schubert (1910); BrS12 = Bernhauer & Schubert (1912), BrS14 = Bernhauer & Schubert (1914); BrSz26 = Bernhauer & Scheerpeltz (1926); BS99 = Besuchet & Sabella (1999); C56 = Coiffait (1956); C60 = Coiffait (1960); C65 = Coiffait (1965); C66a = Coiffait (1966a); C66b = Coiffait (1966b); C67a = Coiffait (1967a); C67b = Coiffait (1967b); C68 = Coiffait (1968); C69 = Coiffait (1969); C70a = Coiffait (1970a); C70b = Coiffait (1970b); C70c = Coiffait (1970c); C70d = Coiffait (1970d); C72a = Coiffait (1972a); C72b = Coiffait (1972b); C74 = Coiffait (1974); C77 = Coiffait (1977); Ca39 = Cameron (1939); Da04 = Davies (2004); Dv84 = Dvořák (1984); E30 = Eichler (1930); Ep78 = Eppelsheim (1878); Ep90b = Eppelsheim (1890b); ES16 = Enushchenko & Semenov (2016); Fa71 = Fauvel (1871); Fa72 = Fauvel (1872); Fa74 = Fauvel (1874); Fr97 = Frisch (1997); Fr07a = Frisch (2007a); Fr07b = Frisch (2007b); Fr09 = Frisch (2009); Fr10 = Frisch (2010); Fr12 = Frisch (2012); G00 = Gildenkov (2000); G01 = Gildenkov (2001); G09 = Gildenkov (2009); G15 = Gildenkov (2015); H01 = Herman (2001); H86 = Herman (1986); Hh49 = Hochhuth (1849); Hh51 = Hochhuth (1851); Hh62 = Hochhuth (1872); HMM79 = Hammond et al. (1979); Hn65 = Horion (1965); Hr90 = Hromádka (1990); IK56 = Iablokoff-Khnzorian (1956); IK57a = Iablokoff-Khnzorian (1957a); IK57b = Iablokoff-Khnzorian (1957b); IK57c = Iablokoff-Khnzorian (1957c); IK59 = Iablokoff-Khnzorian (1959); IK60 = Iablokoff-Khnzorian (1960); IK61 = Iablokoff-Khnzorian (1961); IK62 = Iablokoff-Khnzorian (1962); IK64a = Iablokoff-Khnzorian (1964a); IK64b = Iablokoff-Khnzorian (1964b); IK66 = Iablokoff-Khnzorian (1966); IK85 = Iablokoff-Khnzorian (1985); IK89 = Iablokoff-Khnzorian (1989); J08 = Jakobson (1908);

JH06 = Jaszay & Hlaváč (2006); Ka40 = Karaman (1940); Ka74 = Karaman (1974); Kh37 = Koch (1937); Ki32 = Kirshenblat (1932); Ki51 = Kirshenblat (1951); Ko46 = Kolenati (1846); Kn90 = Kalashian (1990); L03 = Luze (1903); L05 =Luze (1905); LB04 = Löbl & Besuchet (2004); Li71 = Likovský (1971); Li81 = Likovský (1981); Lö70 = Löbl (1970); Lu01 = Luze (1901); Lu02 = Luze (1902); Ma06 = Maruyama (2006); Mk14 = Makranczy (2014); Mm30 = Mannerheim (1830); Mo60 = Motschulsky (1860); Mü23 = Müller (1923); P67 = Puthz (1967); P72a = Puthz (1972a); P72b = Puthz (1972b); P81 = Puthz (1981); P83 = Puthz (1983); P03 = Puthz (2003); P08 = Puthz (2008); P09 = Puthz (2009); P12 = Puthz (2012); P16 = Puthz (2016); Pa83 = Pace (1983); Pa89 = Pace (1989); Pe80 = Petrenko (1980); Ps06 = Paśnik (2006); R89 = Reitter (1889); R05a = Reitter (1905a); R05b = Reitter (1905b); R08 = Reitter (1908); R09 = Reitter (1909); Ry1990 = Ryvkin (1990); SBBB04 = Sabella & al. (2004); Sc89 = Schülke (1989); Sc04 = Schülke (2004); Sc08 = Schülke (2008); Sc19a = Schülke (2019a); Sc19b = Schülke (2019b); Se03 = Semenov (2003); SG05 = Solodovnikov & Grebennikov (2005); SK00 = Schülke & Kocian (2000); SKu02 = Sabella & Kurbatov (2002); Sm04 = Smetana (2004); So02 = Solodovnikov (2002); So04 = Solodovnikov (2004); So05 = Solodovnikov (2005); Sr03 = Schillhammer (2003); Sr04 = Schillhammer (2004); SS15 = Schülke & Smetana (2015); Sz57 = Scheerpeltz (1957); Sz58 = Scheerpeltz (1958); Sz66a = Scheerpeltz (1966a); Sz66b = Scheerpeltz (1966b); U75 = Ullrich (1975); Uv86 = Ushakov (1986); VB04 = Vít & Besuchet (2004); Za93 = Zanetti (1993); Za02 = Zanetti (2002).

Footnotes: [1)] Previous literature records based on erroneously attributing the locality "Araxesthal" (or equivalent) to Armenia rather than Azerbaijan. [2)] Reported as *O. erevanicum* Iablokoff-Khnzorian, 1964 (synonym). [3)] Reported as *Mycetoporus ruficornis* Kraatz, 1857 (synonym). [4)] The record of *Mycetoporus brucki* (Pandellé, 1869) probably refers to this species. [5)] The record of *Tachinus humeralis humeralis* Gravenhorst, 1802 probably refers to this subspecies. [6)] Reported as *Lathrimaeum melanochromum* Iablokoff-Khnzorian, 1961 (synonym). [7)] Reported as *M. kirschenblatti* Iablokoff-Khnzorian, 1956 (synonym). [8)] Reported as *A. crassicornis* Iablokoff-Khnzorian, 1835 (synonym). [9)] Reported as *A. nobilis* Likovský, 1965 (misidentification). [10)] The record of *A. bobaci* Krása, 1931, now a synonym of *A. parvicornis* Fauvel, 1900, most likely refers to the similar *A. fugax*. [11)] Reported as *Callicerus velox* Iablokoff-Khnzorian, 1960 (synonym). [12)] Reported as *Tachinus kirschenblatti* Iablokoff-Khnzorian, 1959 (synonym). [13)] Reported as *Pronomaea subterranea* Iablokoff-Khnzorian, 1957 (synonym). [14)] Reported as *Atheta nidorum* (Thomson, 1868) (synonym). [15)] Reported as *Atheta sipaliformis* Benick, 1974 (synonym). [16)] Reported as *B. fracticornis* (Paykull, 1790) (synonym). [17)] Locality not specified. [18)] Reported as *S.* "*cordatoides* A". [19)] Reported as *A. filiformis* (Latreille, 1806 (synonym). [20)] Reported as *A. silvicola* Iablokoff-Khnzorian, 1961 (synonym). [21)] Partly reported as *E. armeniacus* Coiffait, 1965, according to Uhlig (pers. comm.) a synonym of *E. rivularis*. [22)] Reported as *G. latroides* Coiffait, 1963 (synonym). [23)] Reported as *G. trossuloides* Coiffait, 1966 (synonym). [24)] Reported as *Heterothops flavolimbatus* Motschulsky, 1860 (synonym). [25)] Reported as *Ocypus olens* (O. Müller, 1764) (misidentification). [25)] Reported as *O. cyanochloris* Hochhuth, 1849 (synonym). [26)] Reported as *O. tomentosus* Baudi, 1870 (synonym). [27)] Reported as *P. lucens* (Mannerheim, 1830) (misidentification). [28)] Reported as *Philonthus armeniacus* Bernhauer, 1901 (synonym). [29)] Reported as *P. lutarius* (Gravenhorst, 1806) (synonym). [30)] Reported as *S. armeniacus* (Coiffait, 1966) (synonym). [31)] Reported as *T. ensifer* (J. Müller, 1932) (synonym). [32)] The record in Coiffait (1972b) is based on the type material from "Araxesthal" and can consequently not be attributed to Armenia. [33)] Reported as *Xantholinus armeniacus* Coiffait, 1956 (synonym). [34)] Reported as *Xantholinus microtophilus* Coiffait, 1970 (synonym). [35)] Reported as *A. taeniata* (Kolenati, 1846) (synonym). [36)] According to Schülke & Smetana (2015) currently assigned to *Tetralaucopora* Bernhauer, 1928, which is most likely a junior synonym of *Parocyusa* Bernhauer, 1902 (Assing in prep.). [37)] Reported as *B. pubescens* (Kolenati, 1846) (synonym). [38)] Reported as *D. canaliculata armeniaca* (Cameron, 1939) (synonym). [39)] Reported as *A. baali* Coiffait, 1960 (synonym). [40)] Reported as *M. erevanensis* Coiffait, 1970 (synonym) (Assing 2004). [41)] Reported as *Medon praecursor* Iablokoff-Khnzorian, 1861 (synonym). [42)] Species dubia. [43)] Reported as *S. armeniacus* Coiffait, 1970 (synonym). [44)] Reported as *Scopaeus armeniacus* Coiffait, 1968 (synonym). [45)] Reported as *Quedius quadripunctatus* Iablokoff-Khnzorian, 1961 (synonym). [46)] Reported as *Quedius acuminatus khnzoriani* Coiffait, 1967 (synonym). [47)] Reported as *S. libertas* Hromádka, 1990 (synonym). [48)] Reported as *Jurecekia asphaltina* (misidentification); see Schillhammer (2004). [49)] Reported as *Paederidus rubrothoracicus caucasicus* Scheerpeltz, 1957 (synonym) from "Ost-Armenien", i.e., not in modern Armenia. [50)] Record from Armenia in Sm04 and SS15 based on Puthz (unpublished) (Puthz, pers. comm.); see also additional records section. [51)] Reported as *A. peusi* Wagner, 1949 (synonym). [52)] Distribution extending into North Iran. [53)] Reported as *Zyras stenocephalus* Iablokoff-Khnzorian, 1960 (synonym). [54)] Reported as *Physetops giganteus* Semenov, 1906 (synonym). [55)] Reported as *Aleochara khnzoriani* Amiryan, 1999 (synonym). [56)] Reported as *Heterothops armeniacus* Coiffait, 1977 (synonym). [57)] Reported as *Heterothops montanus* Iablokoff-Khnzorian, 1966 (synonym). [58)] Reported as *Tachyporus armeniacus* Kolenati, 1846 (synonym) from "Armeniae montuosae sylvis". [59)] Previously regarded as subspecies of *S. collaris* (Müller & Kunze, 1822). [60)] Reported as *Phyllodrepa armena* Iablokoff-Khnzorian, 1959 (synonym). [61)] Reported as *Quedius xanthurus* Iablokoff-Khnzorian, 1961 (synonym). [62)] Previously listed as a junior synonym of *O. badia*. [63)] Only material identified by Vladimir Gusarov listed; the remainder is given as *Mocyta* spp. in Tab. 2.

Mocyta is listed as a subgenus of *Atheta* in Schülke & Smetana (2015). However, since V. Gusarov (pers. comm.) is planning to treat this taxon as a distinct genus in his revision (in prep.), this concept is adopted here, too. [64] Reported as *Tachinus laciniatus* Eppelsheim, 1890 (synonym). [65] Previously assigned to the genus *Enalodroma* Thomson, 1859, now a synonym of *Alevonota* Thomson, 1859 (Assing 2019c). [66] Similar to *Bisnius zhuk* Gusarov, 1995; to be described together with other species of the *B. fimetarius* group by Schillhammer (in prep.). [67] Recorded as *Atheta altiviva* Benick, 1974, a name to be synonymized with *A. araxis* by Vogel (in prep.) (Vogel pers. comm.); note that *A. araxis* is listed in the genus *Liogluta* Thomson, 1858 by Schülke & Smetana (2015).

3.3 Unidentified and unnamed species

A reliable identification at the species level was not possible for some of the recently collected material because it was represented by females only, because the respective taxon is currently in a state of taxonomic confusion (*Mocyta* Mulsant & Rey, 1874, *Homoeusa* Kraatz, 1856), or for other reasons. Some of the unidentified species may even be undescribed.

Tab. 2: Unidentified and unnamed species. As in Tab. 1, the number of specimens is given in parentheses behind the locality number.

Species	Localities/samples
Omaliinae	
Lesteva sp. (female)	76(1)
Proteininae	
Proteinus spp. (females)	39(1), 68(1), 114(3), 119(1), 151(3)
Pselaphinae	
Bryaxis spp.	15(1), 39(1), 52(1), 77(1), 81a(2), 88(1), 89(2),
Euplectus sp.	68a(1)
Tychus spp.	22(1), 121(1), 121a(1)
Tachyporinae	
Mycetoporus spp.	7(1), 9(1), 26(1), 30(1), 35(1), 36(1), 46(1), 123(1)
Aleocharinae	
Acrotona spp.	8(1), 127(3)
Aleochara spp.	4(1), 9(6), 13(2), 70(2), 88(1), 155(1)
Aloconota sp.	135(1)
Amischa spp.	46(1), 67(1), 139(1)
Atheta (*Atheta*) sp.	11b(1)
Atheta (*Dimetota*) spp.	11(1), 11b(1), 13(1)
Atheta (*Dralica*) sp.	19(1)
Atheta (*Microdota*) spp.	11a(1), 59(1), 76(4), 81(1), 114(1), 118(1), 126(1)
Atheta (*Parameotica*) sp.	77(1)
Atheta (*Philhygra*) spp. (females)	16(2), 67(5), 168(1)
Atheta spp.	9(1), 12(1), 87(1), 112(1), 199(1), 199a(2), 203a(1)
Gyrophaena spp. (females)	30(1), 62(1), 66a(4), 92(1), 102(1), 112(2), 127(2), 131(1)
Homoeusa sp.	63(1)
Liogluta sp.	105(2)
Mocyta spp.	1(12), 4(5), 5(2), 6(1), 8(3), 9(2), 11(10), 11a(6), 11b(31), 12(1), 13(1), 16(11), 19(21), 20(4), 21(44), 21a(1), 22(328), 22a(97), 26(1), 30(98), 31(8), 32(1), 33(2), 34(24), 35(41), 36(32), 39(48), 41(19), 42(10), 43(5), 45(1), 46(15), 47(5), 48(76), 49(9), 50(7), 50a(1), 53(8), 58(1), 60(2), 61(1), 62(2), 63(6), 64(1), 66(4), 66a(7), 66b(2), 67(71), 68(2), 70(35), 71(7), 72(3), 75(1), 76(6), 77(5), 78(33), 79(69), 81(24), 81a(7), 82(22), 82a(46), 83(9), 85c(16), 86(2), 88(33), 89(32), 89b(45), 90(1), 90a(1), 91(4), 92(9), 96(1), 98(8), 101(3), 102(8), 103(9), 105(12), 107(1), 108(6), 109(1), 110(14), 112(12), 113(1), 114(66), 115(3), 116(99), 117(1), 118(3), 118a(1), 119(65), 121(1), 121a(2), 123(3), 124(1), 125(11), 126(95), 127(4), 129(1), 130(4), 131(2), 133(9), 135(1), 136(13), 148(11), 150(2), 151a(2), 159(3), 170(3), 171(1), 172(1), 186(1), 191a(1), 196(4), 199(2), 199a(1)
Ocalea sp.	117(1), 121(2)
Oxypoda spp.	5(1), 48(1)
Tachyusa sp. (female)	161(1)
Scaphidiinae	
Scaphisoma cf. *agaricinum* (female)	98(1)

Species	Localities/samples
Oxytelinae	
Anotylus sp. (female)	31(1), 39(1), 48(1), 62(1), 82(1), 86(1), 182(1)
Carpelimus (*Boopinus*) sp. (female)	212(1)
Carpelimus (*Trogophloeus*) sp.	190(7), 203(1)
Platystethus sp. (female)	184(1)
Steninae	
Stenus cf. *piscator* (female)	199a(1)
Scydmaeninae	
Euconnus spp.	58(1), 63(2)
Nanophthalmus sp.	30(1)
Neuraphes sp.	89b(1), 110(1)
Staphylininae	
Bisnius sp. (female)	94(1)
Gabrius (*nigritulus* group) spp. (females)	19(1), 20(1), 36(2), 82(1), 103(1), 114(1), 117(1)
Heterothops sp.	114(1), 118(1)
Leptacinus spp.	177(1), 183(1)
Quedius sp. (female)	135(1)

3.4 Erroneous and doubtful records

Numerous species listed for Armenia in recent catalogues (Herman 2001, Schülke & Smetana 2015, Smetana 2004) and/or reported in other works are based on erroneous or doubtful records. They were either wrongly or doubtfully attributed to (modern) Armenia, or they are at least likely to be based on misidentification.

Many of these records are from localities that are not situated in Armenia (any more), but in Azerbaijan, Georgia, or East Turkey (today). Some of these localities have been misinterpreted even in more recent papers. This is particularly true of "Aresch" (or Aresh, or Geok Tapa), which has been interpreted as Verin (Yerevan), but which is in fact a historical fortress built by the Osmanian empire in the 16th century and located near today´s Khaldan, Evlakh district, Azerbaijan. Another frequently mis-assigned locality is "Araxesthal" or "Araxestal"; most of the material from this locality was collected near Ordubad (Azerbaijan). Moreover, a number of previous records from Armenia is based on specimens labelled "Armenisches Gebirge" (or similar), which includes the mountain ranges from Georgia and East Turkey to North Iran. It is unclear which of this material originated from the territory of modern Armenia, if any. In fact, it can be inferred from the records of species with more restricted distributions (e.g., *Quedius walteri*) that they are most likely based on material from East Turkey and/or the Lesser Caucasus in Georgia. Finally, the "Armenia" of 19th-century authors (e.g., Hochhuth 1849, 1851) included vast territories of what is Georgia, East Turkey, and Azerbaijan today. Consequently, if no localities were specified, such records had to be discarded.

Previous records were considered doubtful owing to unreliable or erroneous identification under the following conditions:

a) a revision of reference material proved the record to be erroneous;

b) records of species with restricted distributions whose presence in Armenia would be unlikely;

c) records of species of taxonomically difficult groups that can be identified only based on an examination of the genitalia (e.g., *Acrotona* spp., *Atheta* spp.), which was not common practice at the time of the publication of a record, a criterion particularly applying to numerous records reported by Eichler (1930);

d) records of species belonging to groups in which additional similar species were described after the publication of a record (e.g., *Oxypoda opaca*);

e) records of species of taxonomically difficult groups (especially Aleocharinae) whose reliable identification would have required the expertise of a specialist of the respective taxon.

Tab. 3: Species erroneously or doubtfully recorded from Armenia and Nagorno-Karabakh.

Species	References
Omalinae	
Anthophagus schneideri Eppelsheim, 1878	Ep78 [27)], H01 [27)], Lu02 [27)]
Eusphalerum zolotarevi (Reitter, 1909)	Am00 [1)]
Geodromicus globulicollis Mannerheim, 1830	L03 [1)]
Geodromicus luzianus Bordoni, 1984	Bo84 [2)]
Olophrum assimile (Paykull, 1800)	IK64a [1)], ATHW00
Omalium cribriceps Fauvel, 1900	Am00 [1)]
Proteininae	
Proteinus planicollis Reitter, 1905	R05a [3)], BrS10 [3)], Sm04 [3)]
Proteinus reflexicollis Reitter, 1905	R05b [3)], BrS10 [3)], Sm04 [3)]
Pselaphinae	
Afropselaphus pentagonus (Saulcy, 1878)	Ka40, LB04 [6)]

Species	References
Batrisus erivanus Motschulsky, 1845	LB04 [21)]
Bryaxis crassicornis (Motschulsky, 1835)	BK07 15)
Claviger araxidis Reitter, 1890	LB04 [3)]
Claviger raffrayi Reitter, 1893	LB04 [3)]
Euplectus thomlini Joy, 1906 [4)]	E30
Tychus armeniacus Saulcy, 1878	BS99
Tychus onobrychidis (Iablokoff-Khnzorian, 1957)	IK57c [32)], LB04 [32)]
Tychus serricornis Motschulsky, 1845	LB04 [21)]
Tachyporinae	
Ischnosoma myops (Eppelsheim, 1880)	K97 [6)]
Mycetoporus ambiguus Luze, 1901	Am00 [1)]
Mycetoporus baudueri Mulsant & Rey, 1875	Lu01 [1)]
Sepedophilus marshami (Stephens, 1832)	ATHW00 [1)]
Sepedophilus pedicularius (Gravenhorst, 1802)	E30 [1)]
Tachinus fauveli Pandellé, 1869	L00 [6)]
Tachinus punctipennis J. Sahlberg, 1876	U75 [6)]
Aleocharinae	
Acrotona pygmaea (Gravenhorst, 1802)	E30 [1)]
Aleochara adusta Eppelsheim, 1890	Sm04 [1)]
Aleochara ignipennis Fauvel, 1900	Li71 [26)]
Aleochara laevigata Gyllenhal,1810	E30 [1)]
Aleochara nobilis Likovský, 1965	Li71 [8)]
Aleochara parvicornis Fauvel, 1900	Li71 [34)]
Aleochara rambouseki Likovský, 1964	Li81 [7)], SS15 [1)]
Aleochara spadicea (Erichson, 1837)	ATHW00, Li71

Species	References
Aleochara spissicornis Erichson, 1839	E30 [1)]
Amarochara forticornis (Lacordaire, 1835)	BrSz26 [20)], Sm04 [20)]
Anaulacaspis gratilla (Erichson, 1839)	Am00 [1)]
Atheta amicula (Stephens, 1832)	E30 [1)]
Atheta elongatula (Gravenhorst, 1802)	E30 [1)]
Atheta euryptera (Stephens, 1832)	E30 [1)]
Atheta filicornis Eppelsheim, 1890	Ep90b [6)]
Atheta orphana (Erichson, 1837)	Hh49 [1)]
Brundinia concolor (Eppelsheim, 1892)	Be74 [1)]
Euryalea murina (Erichson, 1839)	E30 [1)]
Gyrophaena lucidula Erichson, 1837	Ko46 [1)]
Ischnoglossa prolixa (Gravenhorst, 1802)	Ko46 [38)]
Leptusa pulchella (Mannerheim, 1830)	Sm04 [17)]
Oreusa araxis (Reitter, 1898)	Sm04 [3)]
Oxypoda opaca (Gravenhorst, 1802)	E30 [1)]
Tachyusa balteata Erichson, 1839	E30 [35)]
Tachyusa coarctata Erichson, 1837	SS15 [19)]
Scaphidiinae	
Scaphisoma laeviusculum Reitter, 1898	Lö70 [6)]
Oxytelinae	
Carpelimus pusillus (Gravenhorst, 1802)	Gi15 [6)]
Carpelimus transversicollis (Scheerpeltz, 1947)	Gi15 [6)]
Ochthephilus emarginatus (Fauvel, 1871)	Mk14 [3)]
Ochthephilus sericinus (Solsky, 1874)	Mk14 [3)]
Platystethus arenarius (Geoffroy, 1785)	Am00 [1)]

Species	References
Platystethus laevis Märkel & Kiesenwetter, 1848	Am00 [1) 10)]
Platystethus rufospinus Hochhuth, 1851	Hh51 [5)]
S t e n i n a e	
Stenus aereus Solsky, 1871	Am00 [1)]
Stenus arabicus Saulcy, 1865	Pu72a [9)]
Stenus circularis Gravenhorst, 1802	Sm04 [28)]
Stenus incanus Erichson, 1839	Sm04 [29)]
Stenus melanarius melanarius Stephens, 1833	Sm04 [30)]
Stenus pallitarsis abanticola Puthz, 1972	Sm04 [31)]
S c y d m a e n i n a e	
Euconnus robustus Reitter, 1882	Ka74 [6)]
Stenichnus pelliceus Holdhaus, 1908	Da04 [1)]
P a e d e r i n a e	
Achenium depressum (Gravenhorst, 1802)	SS15 [1)]
Achenium planum Erichson, 1840	A10b [22)]
Achenium quadriceps Eppelsheim, 1889	A10b [22)]
Astenus gracilis (Paykull, 1789)	E30 [1) 11)]
Lathrobium geminum Kraatz, 1857	BrS12 [1)], Sm04 [1)]
Lathrobium pallidipenne Hochhuth, 1851	Hh51 [5)]
Luzea infirma (Erichson, 1840)	A08b [22) 23)], A10a [22)]
Luzea nigritula (Erichson, 1840)	ATHW00 [1)]
Luzea rossica (Bernhauer, 1908)	A10a [22)], Br08 [22)]
Medon apicalis (Kraatz, 1857)	ATHW00 [1)]
Medon fusculus (Mannerheim, 1830)	Sm04 [1)]
Micrillus pallidus (Reitter, 1887)	A08a [22)], A13b [22)]
Micrillus testaceus (Erichson, 1840)	E30 [1)]

Species	References
Platydomene angusticollis (Lacordaire, 1835)	Sm04 [1)]
Platydomene bicolor bicolor (Erichson, 1840)	Sm04 [1)]
Pseudobium angusticolle (Hochhuth, 1851)	Hh51 [5)]
Pseudolathra araxidis Coiffait, 1972	A17a [25)], C72a [34)]
Rugilus angustatus (Geoffroy, 1785)	Sm04 [1)]
Scopaeus minutus Erichson, 1840	Am00 [1) 36)]
S t a p h y l i n i n a e	
Gabrius astutus (Erichson, 1840)	Fa74 [1)], Hh51 [5) 24)]
Gauropterus sanguinipes Reitter, 1889	R89 [26)], C72b [26)]
Ocypus helleni (J. Müller, 1926)	Sz58 [14)]
Othius hebes Assing & Solodovnikov, 1998	ASo98 [6)]
Philonthus armeniacus Hochhuth, 1851	Hh51 [5)]
Philonthus rotundicollis (Ménétriés, 1832)	ATHW00 [1)]
Philonthus salinus Kiesenwetter, 1844	Sz58 [14)]
Quedius walteri Korge, 1971	So05 [13)]
Rabigus formosus (Motschulsky, 1860)	Am00 [1)]
Rabigus tenuis (Fabricius, 1792)	E30 [1)]
Tasgius winkleri (Bernhauer, 1906)	C74 [1) 18)]
Xantholinus linearis (Olivier, 1795)	Sm04 [1)]
Xantholinus longiventris Heer, 1839	Sm04 [1)]
Xantholinus tricolor (Fabricius, 1787)	E30 [1)]
Xantholinus variabilis Hochhuth, 1851	C72b [12)], Hh51 [5)]

Footnotes: [1)] Record most likely based on misidentification. [2)] The type locality "Schawnabad" was erroneously attributed to Armenia rather than Georgia. [3)] Previous literature records based on incorrectly attributing the type locality "Araxesthal", "Araxestal", or similar to

Armenia rather than Azerbaijan. [4] Manuscript name. [5] Record based on type locality "Armenien" in the sense of 1851. [6] The locality "Armenisches Gebirge" (or equivalent) was – most likely erroneously – attributed to Armenia. [7] Reported as *A. addenda* Likovský, 1981 (synonym), whose type locality "Ordubad" was erroneously attributed to Armenia (Assing 2017a). [8] Record based on misidentification (Assing 2017a). [9] Doubtful female-based record. [10] Reported as *Platystethus cephalotes* Eppelsheim, 1878 (identification doubtful). [11] Reported as *A. neglectus* (Märkel, 1844) (synonym). [12] Type locality "Helenendorf" erroneously attributed to Armenia. [13] Record erroneously attributed to Armenia (Assing 2016d). [14] Record from Turkey, erroneously attributed to Armenia. [15] Record(s) from Georgia, erroneously attributed to Armenia. [16] Type locality in Georgia, no Armenian localities reported by Besuchet & Sabella (1999). [17] Record most likely based on misidentification; the species is not indicated from Armenia in Pace (1989). [18] Record based exclusively on females. [19] Record/identification doubtful; not indicated from Armenia in Paśnik (2006). [20] Record based on misidentification and locality erroneously attributed to Armenia (Assing 2002). [21] Species dubia. [22] Record from "Aresch" (or equivalent) [= Geok-Tapa; = Agdash in Azerbaijan] erroneously attributed to Armenia. [23] Reported as *L. caucasica* (Luze, 1912) (synonym). [24] Reported as *Philonthus erythrostomus* Hochhuth, 1851 (synonym). [25] Type locality "Djoulfa" [= Culfa] is in Nakhchivan (Azerbaijan), not in Armenia. [26] The record from "Ordubad" erroneously attributed to Armenia. [27] Erroneous record from Armenia in SS15 probably based on Ep78 (Elisabetpol) and/or Lu02 (Helenendorf), both in Azerbaijan. [28] Confirmed records from Armenia unknown; record possibly referring to *Stenus planifrons robustus* (Puthz, pers. comm.). [29] Confirmed records from Armenia unknown (Puthz, pers. comm.). [30] Record refers to *S. peripherus* (Puthz, pers. comm.). [31] Record most likely refers to *S. claritarsis* (Puthz, pers. comm.). [32] Type locality in Nakhchivan (Azerbaijan), not in Armenia. [33] Reported as *A. peusi* Wagner, 1949 (synonym). [34] The record of *A. bobaci* (synonym of *A. parvicornis*) is probably refers to the similar *A. fugax*. [35] Not indicated from Armenia in Paśnik (2006); record most likely based on misidentified pale-coloured specimens of *T. impressa*. [36] Record probably referring to the similar *S. chalcodactylus* (Frisch pers. comm.). [37] Most likely misidentified; other species similar to *A. spadicea* have been described in the meantime (Assing 2009a). [38] Reported from Nagorno-Karabakh as *I. pubescens* (Kolenati, 1846) (synonym).

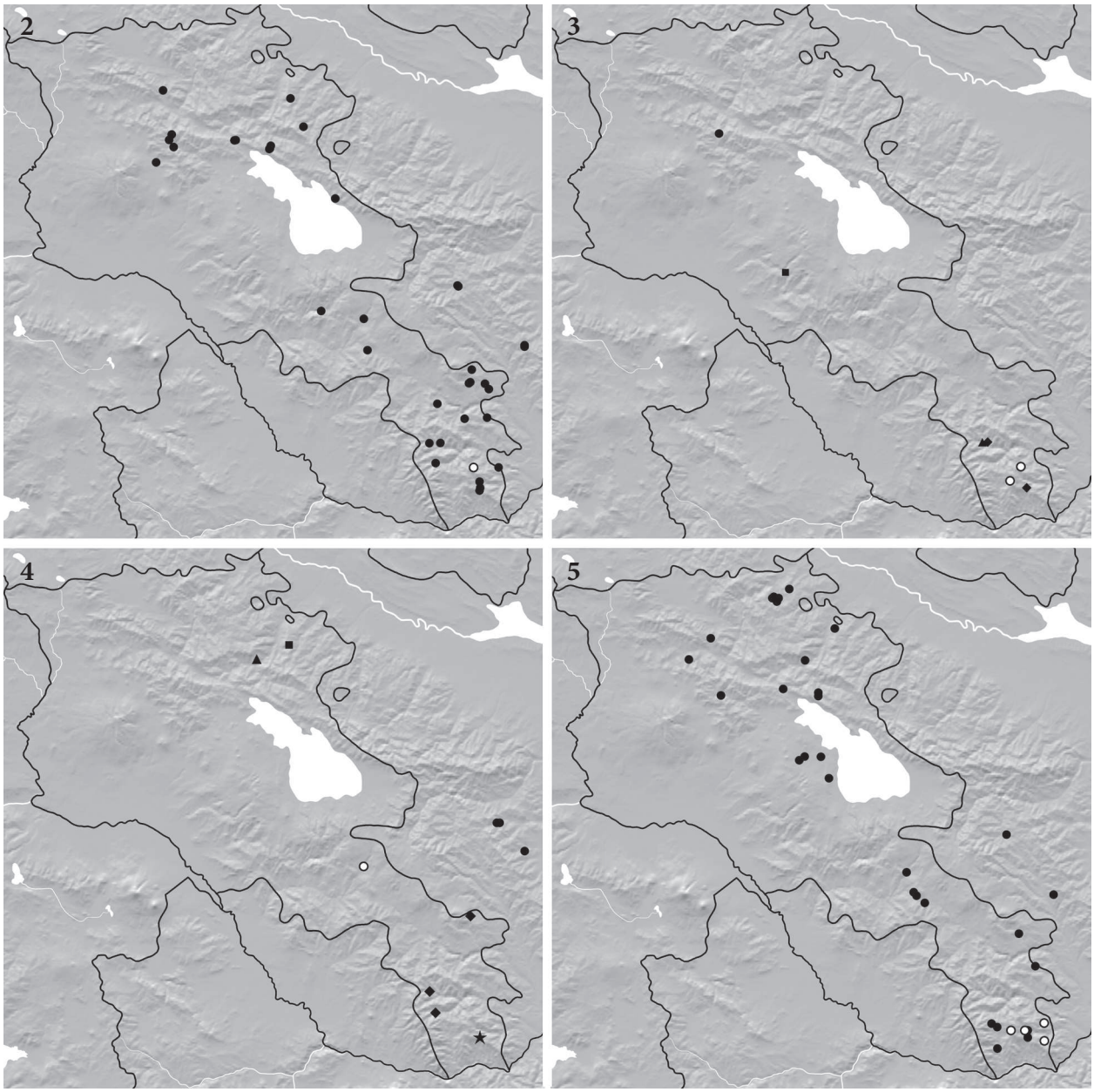
Tab. 4: Diversity and systematic composition of the faunas of Armenia, Nagorno-Karabakh, and neighbouring countries and regions. The table summarizes total species numbers and the species numbers and proportions of the seven most speciose subfamilies of Armenia (AR), Nagorno-Karabakh (NK), Georgia (GG), the Russian South European Territory (ST), and Iran (IN). The data for GG, ST, and IN are based on Schülke & Smetana (2015) and additions up to approximately the end of 2017.

	AR	NK	GG	ST	IN
Species total	675	198	887	1077	784
Aleocharinae	260	78	224	191	151
%	38.51	39.39	25.25	17.73	19.2
Staphylininae	95	30	170	266	141
%	14.07	15.15	19.17	24.70	17.98
Tachyporinae	59	28	71	73	36
%	8.74	14.14	6.88	6.78	4.59
Oxytelinae	58	9	74	120	74
%	8.59	4.55	8.34	11.14	9.44
Steninae	50	12	69	83	66
%	7.41	6.06	7.78	7.71	8.42
Paederinae	47	8	71	105	135
%	6.96	4.04	8.00	9.75	17.22
Omaliinae	43	12	60	73	24
%	6.37	6.06	6.76	6.78	3.06
Pselaphinae	41	7	93	98	57
%	6.07	3.54	10.48	9.10	7.27

3.5 Zoogeography

3.5.1 Endemism

Armenia and Nagorno-Karabakh have a remarkably poor endemic staphylinid fauna, especially when compared to West Georgia and Northeast Turkey. Some species have exclusively been reported from Armenia, but their taxonomic and/or zoogeographic status is unclear: *Aleochara armeniaca*, *A. fugax*, *Aloconota differens*, *Atheta erevanensis*, *Gnypeta gracilior*, *Hydrosmecta salsolae*, *Bisnius microtophilus*, *Gabrius armeniacus*, *Heterothops macrops*, *H. microtophilus*, *Leptacinus armeniacus*, *Quedius latus*, *Q. transcaucasicus*, *Stenistoderus nanus*. Similarly, several species have only been reported from Armenia or Nagorno-Karabakh, but may also be present in neighbouring countries: *Geodromicus armeniacus*, *G. rivularis*, *Olophrum aragatzense*, *Omalium kociani*, *Zibus yunonae*, *Mycetoporus praetextoides, Aleochara polychroma*, *Atheta hamulata*, *A. senticollis*, *Atheta* (*Oreostiba*) *abscisa*, *A.* (*O.*) *brevitheca*, *A.* (*O.*) *semialba*, *Calodera alticola*, *Liogluta armeniaca*, *Myllaena*



Maps 2–5: Distributions of endemic species and species groups. **Map 2** (upper left): regionally endemic *Geostiba* species (all species pooled; *G. sororcula* omitted) (black circles) and the locally endemic *Bellatheta khustupica* (white circle). **Map 3** (upper right): *Atheta meghruica* (white symbols) and regionally endemic Pselaphinae (black symbols): *Bryaxis armeniacus* (black circle), *B. meghruicus* (black triangle), *B. seductus* (black diamonds), *Tychus milvus* (black square). **Map 4** (lower left): *Oxypoda grandecristata* (white circle) and regionally endemic Scydmaeninae (black symbols): *Euconnus karabakhus* (black circles), *E. longilaminatus* (black square), *E. tavushus* (black triangle), *Neuraphes gomarantsus* (black star), *N. syunikus* (black diamonds). **Map 5** (lower right): *Xantholinus adustus* (white circles), a regional endemic of South Armenia, and *X. kirschenblati* (black circles), a species currently known only from Armenia and Nagorno-Karabakh, but probably present also in adjacent regions.

ambulans, *Oxypoda flexa*, *O. infissoides*, *O. levipunctata*, *O. verminata*, *Anotylus hamatoides*, *Lobrathium ancoriferum*, *Medon sequax*, and *Ontholestes chalcopygus*. These species do not possess typical adaptations of endemics (flightlessness, reduced eye size, etc.) and belong to species groups or ecological guilds whose representatives are usually more widespread.

Two of the endemics of South Armenia are relatively common and widespread in this region: *Leptusa armeniaca* (wingless) and *Oxypoda subplicata* (winged). They are (*L. armeniaca*), or may potentially be (*O. subplicata*) present also in adjacent regions of North Iran.

Regionally endemic species of Armenia and Nagorno-Karabakh, with one exception (*Atheta meghruica*) all flightless, belong to the Aleocharinae (twelve species), Pselaphinae (four species), Scydmaeninae (five species), and Staphylininae (one species). These include *Atheta* (*Paralpinia*) *meghruica* (Map 3), *Geostiba* (*Sibiota*) *deliqua*, *G.* (*S.*) *immutata*, *G.* (*S.*) *kalavanica*, *G.* (*S.*) *meghruica*, *G.* (*S.*) *nigrohortensia*, *G.* (*S.*) *pamba-*

kica, *G.* (*S.*) *tigrani*, *G.* (*S.*) *unicuneata*, *G.* (*S.*) *unituber*, *G.* (*Tropogastrosipalia*) *khnzoriani* (Map 2), and *Oxypoda grandecristata* (Map 5) of the Aleocharinae, *Bryaxis armeniacus*, *B. meghruicus*, *B. seductus*, and *Tychus milvus* of the Pselaphinae (Map 3), *Euconnus longilaminatus*, *E. tavushus*, *E. karabakhus*, *Neuraphes gomarantsus*, and *N. syunikus* of the Scydmaeninae (Map 4), and *Xantholinus adustus* (Map 5) of the Staphylininae. *Geostiba nigrohortensia* and *Euconnus karabkhus* (Map 4) are endemic to Nagorno-Karabakh, *Geostiba unicuneata* to adjacent regions in Armenia and Nagorno-Karabakh, and the remainder to Armenia. For maps illustrating the individual distributions of the *Geostiba*, *Leptusa*, and *Oxypoda* species see Assing (2016b, c, 2017b, c, 2018c, h).

Remarkably, the study region is nearly devoid of high-altitude endemics. The only mountain where endemic species associated with habitats above the tree-line were found is Mount Khustup in South Armenia. Based on currently available evidence, one of them, *Bellatheta khustupica*, may actually be locally endemic (Map 2).

The majority of the regional endemics inhabits montane forests. The greatest density of these species was observed in South Armenia.

3.5.2 Other distribution types

The zoogeography of most Staphylinidae species still requires thorough study; many of them are known only from few records. Nevertheless, an analysis of the distributions of the taxa known from Armenia and Nagorno-Karabakh reveals that the majority of the species is widespread. They represent Cosmopolitan, trans-Palaearctic, expansive Holo- or Ponto-Mediterranean, or other elements. In addition, the fauna of the study region is characterized by the presence of the following distribution types.

Species such as *Mycetoporus praetextoides*, *Xantholinus kirschenblati* (Map 5), and *Rugilus armeniacus* are currently known only from Armenia, Nagorno-Karabakh, and (only the latter) from Azerbaijan. The former two are widespread and not uncommon in Armenia and Karabakh and only few records are known of *R. armeniacus*. It appears likely that these species will eventually be recorded also from Georgia and/or North Iran.

The known distributions of another set of species is restricted to Northeast Turkey, Armenia, and partly also Nagorno-Karabakh. Examples are *Proteinus baculatus*, *Mycetoporus phaedrus*, *Geostiba sororcula* (see Assing 2017b: map 1), *Dianous ponticus*, and *Quedius brevalatus*.

A significant proportion of the fauna of Armenia and Nagorno-Karabakh is of the Caspian distribution type, with a refugial centre in Transcaucasia (see Lattin 1967). Most of these species have a Caucasian distribution typically including the Greater Caucasus, the Lesser Caucasus, the East Pontic Mountains, Transcaucasia, and North Iran, or parts of this region. Representatives of this distribution type are *Anthobium hamatum*, *A. tenue*, *Dropephylla elisabethae*, *Eusphalerum fidele*, *E. sareptanum*, *E. transcaucasicum*, *Geodromicus brevicollis*, *G. major*, *Olophrum caucasicum*, *Brachygluta araxidis*, *B. nodosa*, *Bryaxis clavipes*, *B. longipalpis*, *B. rostratus*, *Ctenistes marthae*, *Pselaphus caspicus*, *Tychus colchicus*, *T. dubius*, *T. guillebeaui*, *Lordithon rostratus*, *Mycetoporus monticoloides*, *Sepedophilus transcaucasicus*, *Tachyporus fascipennis*, *Tachinus caucasicus*, *T. cingulatus*, *T. nigerrimus*, *Aleochara conviva*, *A. plicelytrata*, *A. subtumida*, *Anaulacaspis caucasica*, *Atheta altiviva*, *A. bispinosa*, *A. brevapicalis*, *A. chefsurica*, *A. laevigata*, *Cousya schuelkei* (Assing 2018e: Fig. 142), *Gyrophaena caucasica*, *Leptusa fuliginosa*, *L. venusta*, *Myllaena caucasica*, *Ocalea alutacea*, *O. dubia*, *O. minor*, *Oxypoda articollis*, *O. caucasica*, *Pella laeviceps*, *Tropimenelytron mirabile*, *Anotylus strigifrons*, *Platystethus gildenkovi*, *Thinodromus dilaticollis*, *T. motschulskyi*, *Stenus araxis*, *S. armeniacus*, *S. caspius*, *S. medus*, *S. peripherus*, *S. skoraszewskyi*, *Cephennium perispinctum perispinctum*, *Euconnus lalvarensis*, *Scydmoraphes yermolowi*, *Lathrobium permutatum*, *L. tichomirovae*, *Leptusa laeviuscula*, *Ochthephilum egregium*, *Rugilus longicollis*, *Scopaeus chalcodactylus*, *S. khnzoriani*, *Sunius fulgocephalus*, *S. khnzoriani*, *Tetartopeus stylifer*, *Bisnius pentheri*, *B. reitteri*, *Erichsonius dux*, *Gabrius sacerdotalis*, *G. trossuliformis*, *Ocypus forficularius*, *O. hochhuthi*, *Othius grandis*, *O. stenocephalus*, *Philonthus picimanus*, *P. svanetiensis*, *Quedius orientalis*, *Q. vulneratus*, *Tasgius eppelsheimianus*, *T. gracilicornis*, *T. limbifrons*, *Xantholinus araxis*, and *X. khnzoriani*.

Iranian elements are typically characterized by an often more or less pronounced circum-Caspian distribution eastwards reaching into Middle Asia. The following species may be assigned to this type: *Dropephylla caucasica*, *Batrisodes ruprechtii*, *Brachygluta spinicoxis spinicoxis*, *Trissemus melinus*, *T. montanus*, *Tychus lederi*, *T. remaudierei*, *Anotylus khachikovi*, *Stenus machulkai*, *S. piscator*, *S. turk*, *Bisnius piochardi*, *Gabrius muelleri*, *Philonthus velatipennis*, *Physetops tataricus*, *Quedius cohaesus*, *Xantholinus fortepunctatus*.

The Syrian distribution type may be represented in the study region by species such as *Cousya araxis* (Assing 2018e: figure 141).

The distinction between Iranian, Caspian, and Syrian elements, however, is sometimes difficult and requires a profound knowledge of the distribution. Examples of doubtful cases are *Atheta brevapicalis* (Iranian or Caspian; see map 1 in Assing (2018b)), *Gyrophaena korbi* (Iranian or Caspian), *Medon fusculoides* and *M. paradisiacus* (Iranian or Caspian), *Myrmoecia urartu* (Iranian or Syrian; see map 1 in Assing (2019a)), *Stenus machulkai* (Caspian, Syrian, or Iranian).

A similar problem is encountered with species like *Tachinus schneideri* and *Gyrophaena orientalis*, which are distributed in the North Palaearctic and in the Caucasus region. They may represent Siberian elements, but too little is known about their distribution in the Northeast Palaearctic region to be certain.

3.5.3 Discontinuous distributions

Several new records of species from Armenia and Nagorno-Karabakh revealed some remarkable range disjunctions. In rarely recorded and poorly known species pronounced distribution gaps may represent artefacts reflecting the lack of data, as is probably the case with, for instance, *Aleochara falcata*, a species currently known only from the Russian Central European territory and Armenia. In general, such discontinuous distributions were observed particularly in species found at high altitudes.

Omalium wunderlei was described based on five specimens from three localities (one of them vague) in Southwest Turkey (Antalya) (Zanetti 2002). The species was subsequently reported also from the Turkish province Muğla (Assing 2004b). In summer 2018, however, a number of specimens was discovered near the peak of Mount Khustup in South Armenia at an altitude of approximately 3,000 m, about 1,500 km from the previously known range (Map 6).

Similar, though less spectacular examples of species previously known from Turkey, partly in West Turkey, with pronounced gaps between the Turkish localities and those in Armenia and Nagorno-Karabakh are *Acrotona concamerata*, *Atheta albomontis*, *A. dissimulans*, *A. lyciana*, *A. monstruosa*, and *Oxypoda recta*. For a map illustrating the currently known distribution of *O. recta* see Assing (2018c: map 2).

Some disjunctions, however, are even more pronounced. The previously known distribution of *Atheta malleiformis* was confined to the Alps and adjacent regions and separated from the record in Nagorno-Karabakh by a distance of some 2,700 km. *Atheta pechlaneri*, *A. serrata*, and *A. taxiceroides* had been recorded only from West, Central, and North Europe (*A taxiceroides* also from the Italian Apennines; see Tagliapietra & Zanetti 2003), as well as from Bulgaria; the distance between the Bulgarian records and those from Armenia amounts to more than 1,600 km. *Bellatheta palata* is currently known only from the Balkans and Armenia, *Ischnoglossa obscura* from North and Central Europe, Turkey, and Armenia, *Oxypoda lentula* from Europe, Siberia, Ukraine, and from Armenian localities at altitudes of approximately 3,000 m, *Gabrius robustus* was previously recorded only from few mountains in Macedonia and Turkey and is now known also from Armenia, with most of the Armenian records at altitudes of approximately 3,000 m. Another spectacular example of a huge distribution gap is *Oxypoda spaethi*, a nidicolous species associated with gopher burrows (previously known from Austria and Hungary (Assing 2012); gap distance 2,200 m) (Map 7). Based on the maps and concepts provided by Lattin (1967), this species may represent a Caspian element with postglacial range disjunction.

In the case of *Pronomaea flavirostris*, the gap is not between the West or Northwest and the study region, but between Middle Asia (Tajikistan, Afghanistan) and Armenia, a distance of approximately 1,700 km (Map 8).

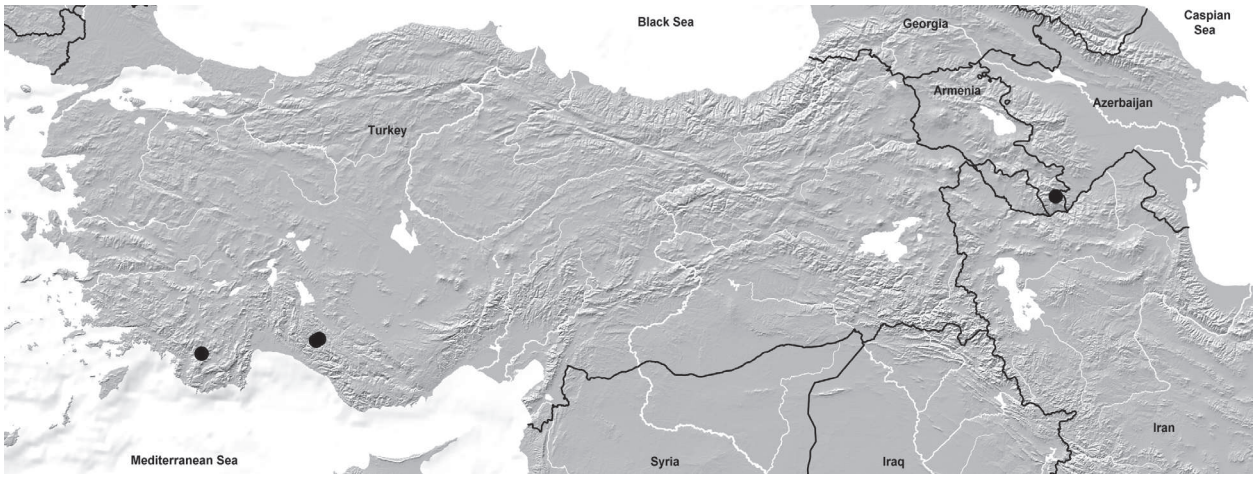
Ecology

Staphylinidae are associated with an enormous variety of terrestrial and semiaquatic habitats and can be found practically everywhere. They represent the full range between extreme specialization and extreme generalization. The field trips conducted by the authors were primarily targeted at the native and endemic fauna of natural and semi-natural habitats at intermediate to high altitudes (forests, wetlands, alpine habitats). The fauna associated with such habitats, including the endemic fauna, can now be considered relatively well studied, at least as far as Armenia is concerned. Sampling in Nagorno-Karabakh was confined to a relatively short period in only one year and almost exclusively focused on forests; high-altitude environments were inaccessible and habitats such as decaying dung, river banks, and other wetlands were sampled only rarely. Remarkably, the Armenian fauna above the tree-line is largely composed of widespread species. The only true local endemic in such an environment is *Bellatheta khustupica* in Mount Khustup, South Armenia.

Habitats that may host numerous additional species to be recorded from the study region are, above all, various kind of decaying matter (dung, compost, rotting wood, bark, carrion, etc.), nests of mammals, birds, and ants, banks of rivers and streams, and other wetlands. Species associated with such habitats are still poorly represented in the fauna of Armenia and Nagorno-Karabakh. For instance, not a single species of *Thinobius* Kiesenwetter, 1844 (mostly ripicolous), *Platydomene* Ganglbauer, 1895 (ripicolous), only one of doubtful status of *Hydrosmecta* Thomson, 1858 (mostly ripicolous), and relatively few of *Carpelimus* Leach, 1819 (various wetlands) are known from the region.

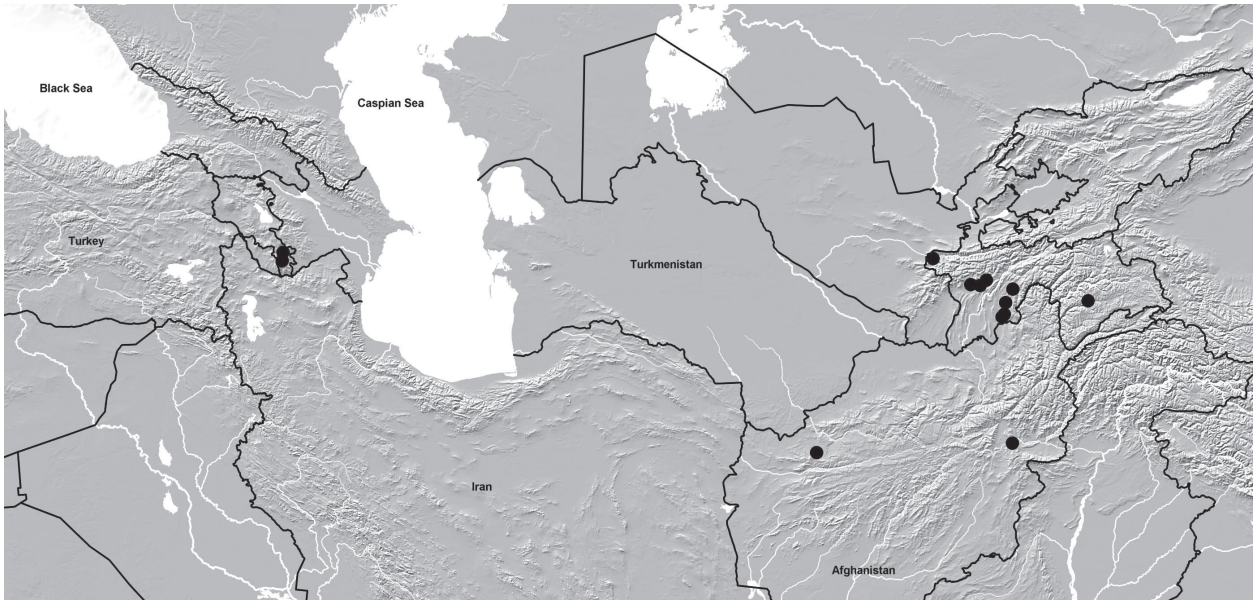
Sampling subterranean, endogean or hypogean, fauna has not been attempted. Based on experiences with such a fauna in the East Mediterranean, it does not seem unlikely that at least endogean fauna may be present particularly in South Armenia.

The primary methods used for recording species during the authors´ field trips were sifting, floating (river banks), and turning stones (at high altitudes, mostly near snowfields). Applying other methods such as pitfall traps and especially car-netting and window traps (for flying insects) would undoubtedly result in recording species whose habitats are essentially unknown (of which there are numerous examples in Staphylinidae) and in significantly increasing species numbers in the study region.



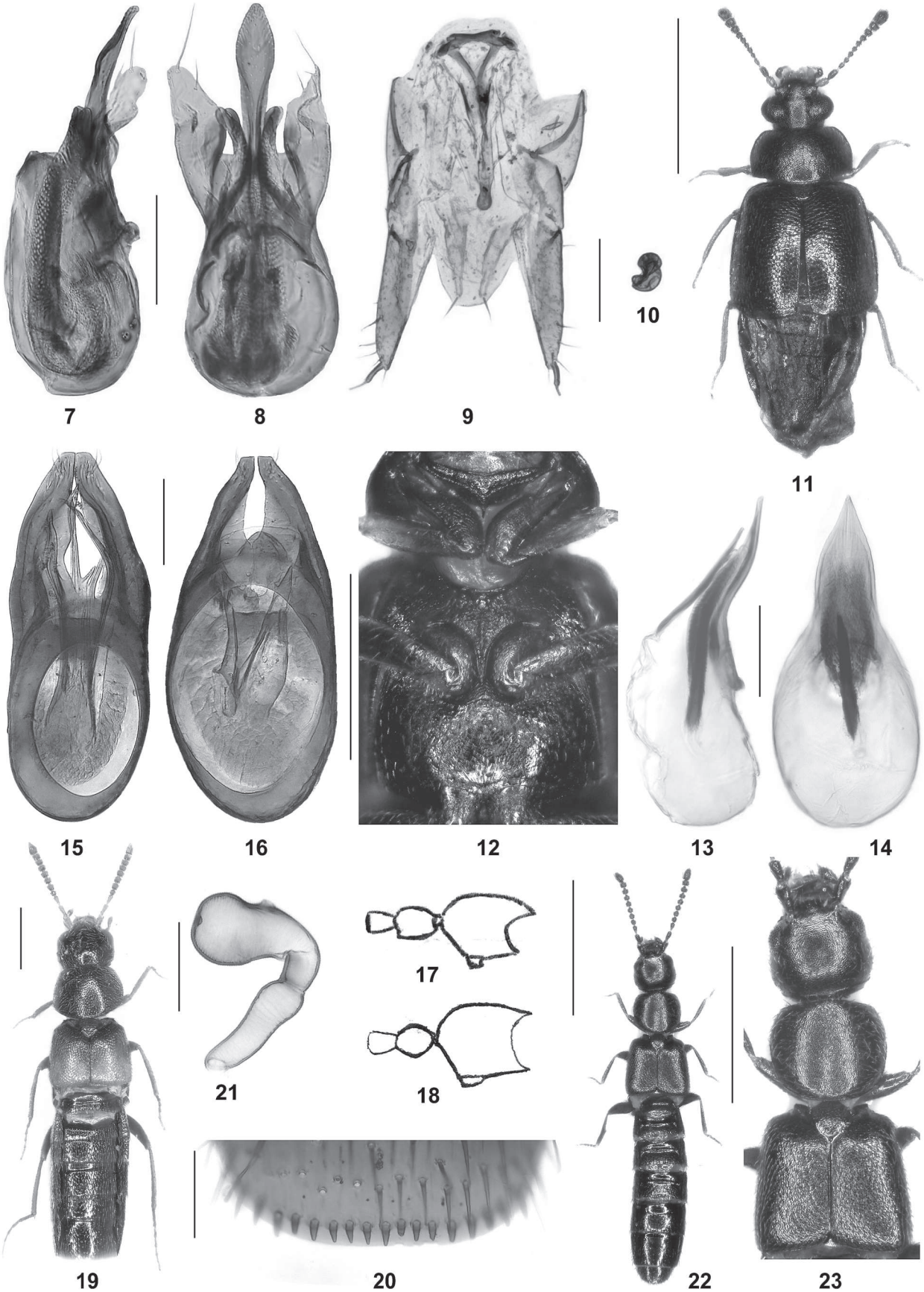
Map 6: Distribution of *Omalium wunderlei*.

Map 7: Distribution of *Oxypoda spaethi* (records based on material in cAss and on Horion (1967)).

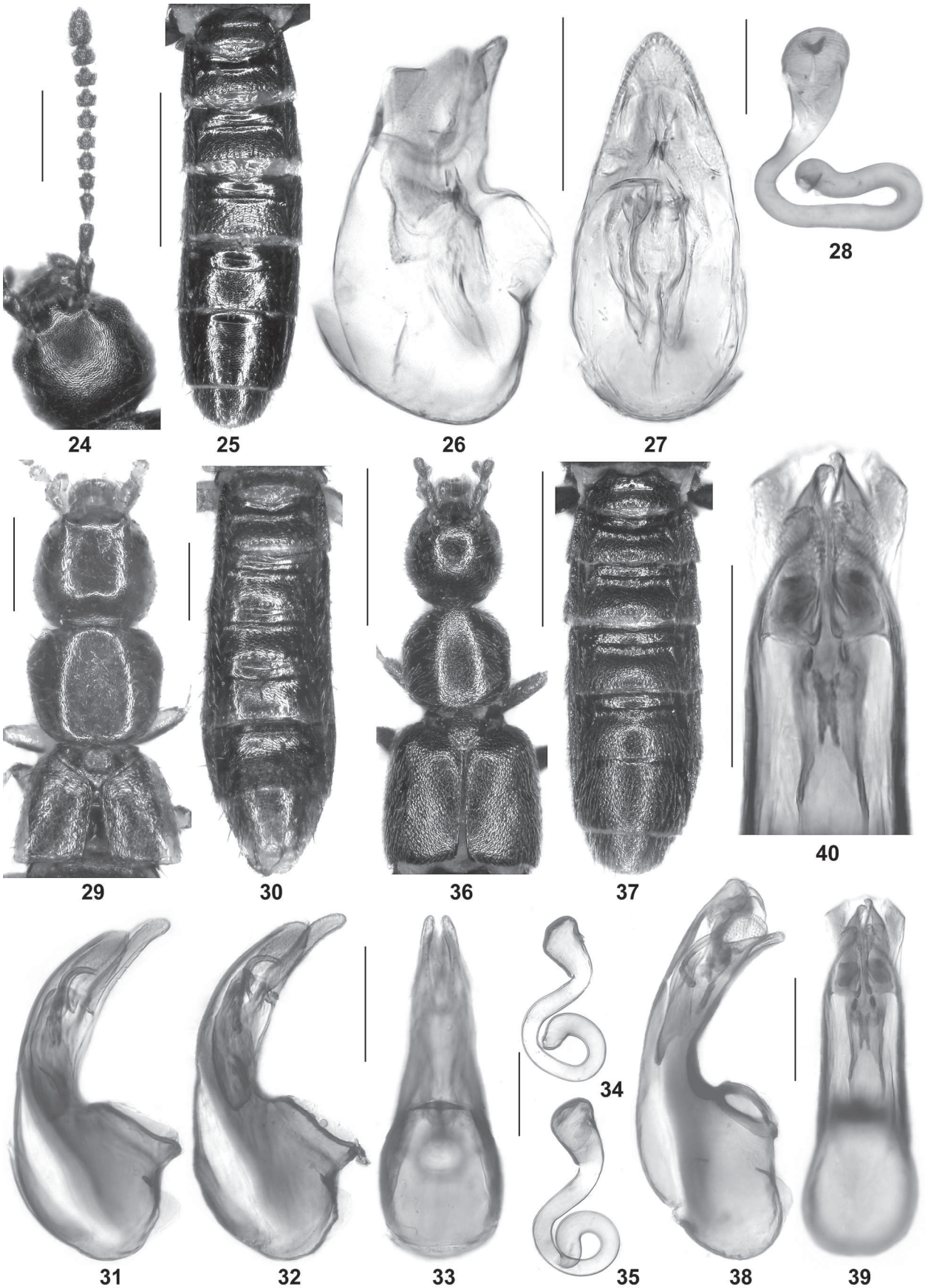


Map 8: Distribution of *Pronomaea flavirostris* (previous records based on Assing (2007b)).

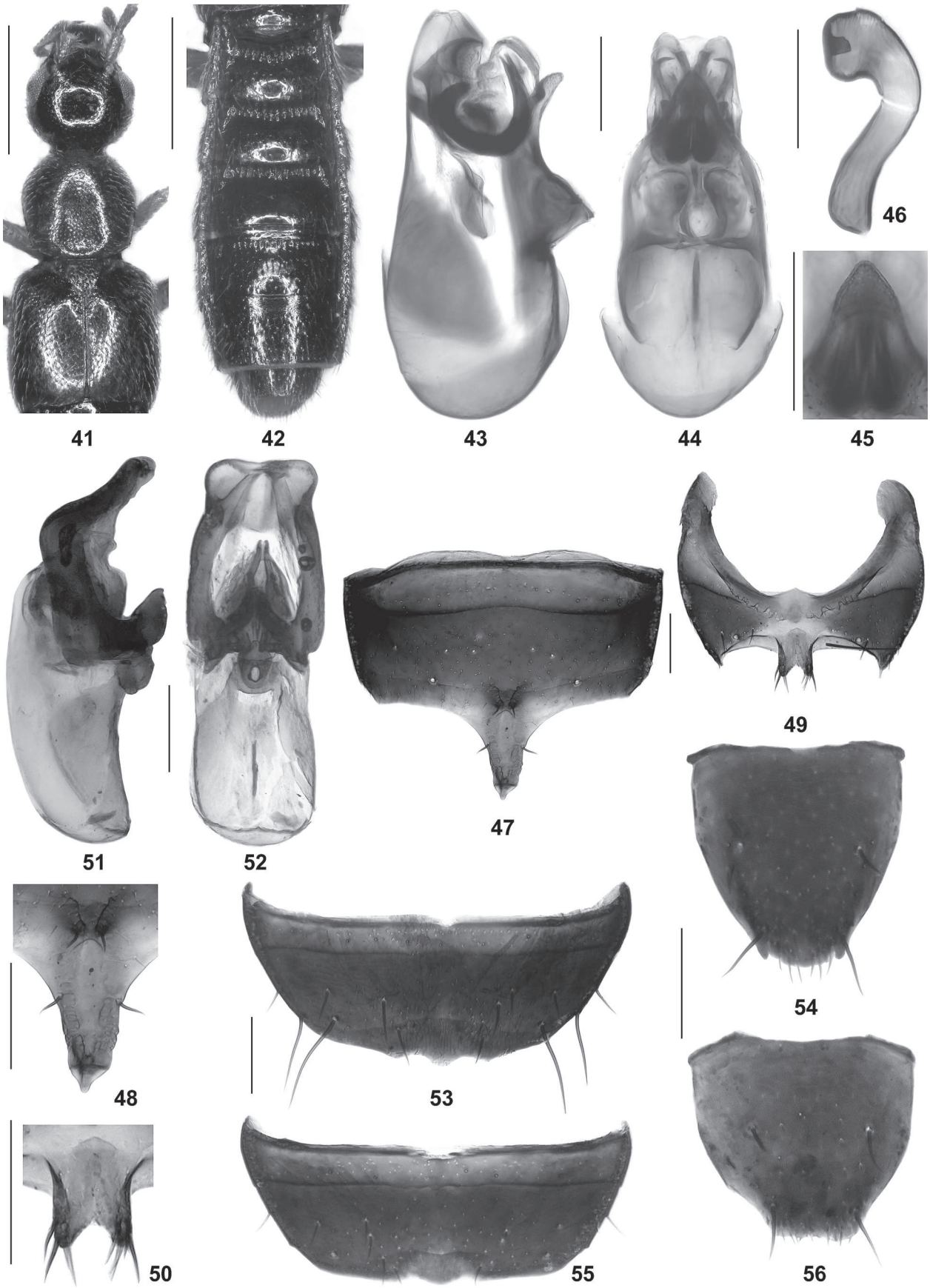
Figs 1–6: Type locality of *Bellatheta khustupica* in the peak region of Mount Khustup (**1**); microhabitat where *B. khustupica* was collected, together with *Omalium wunderlei, Anthobium fusculum,* and other Staphylinidae (**2**); habitus of *Omalium kociani* (**3**), *Mycetoporus silvaticus*, holotype (**4**), *Bellatheta khustupica* (**5**), and *Calodera alticola* (**6**). Scale bars: 1.0 mm.



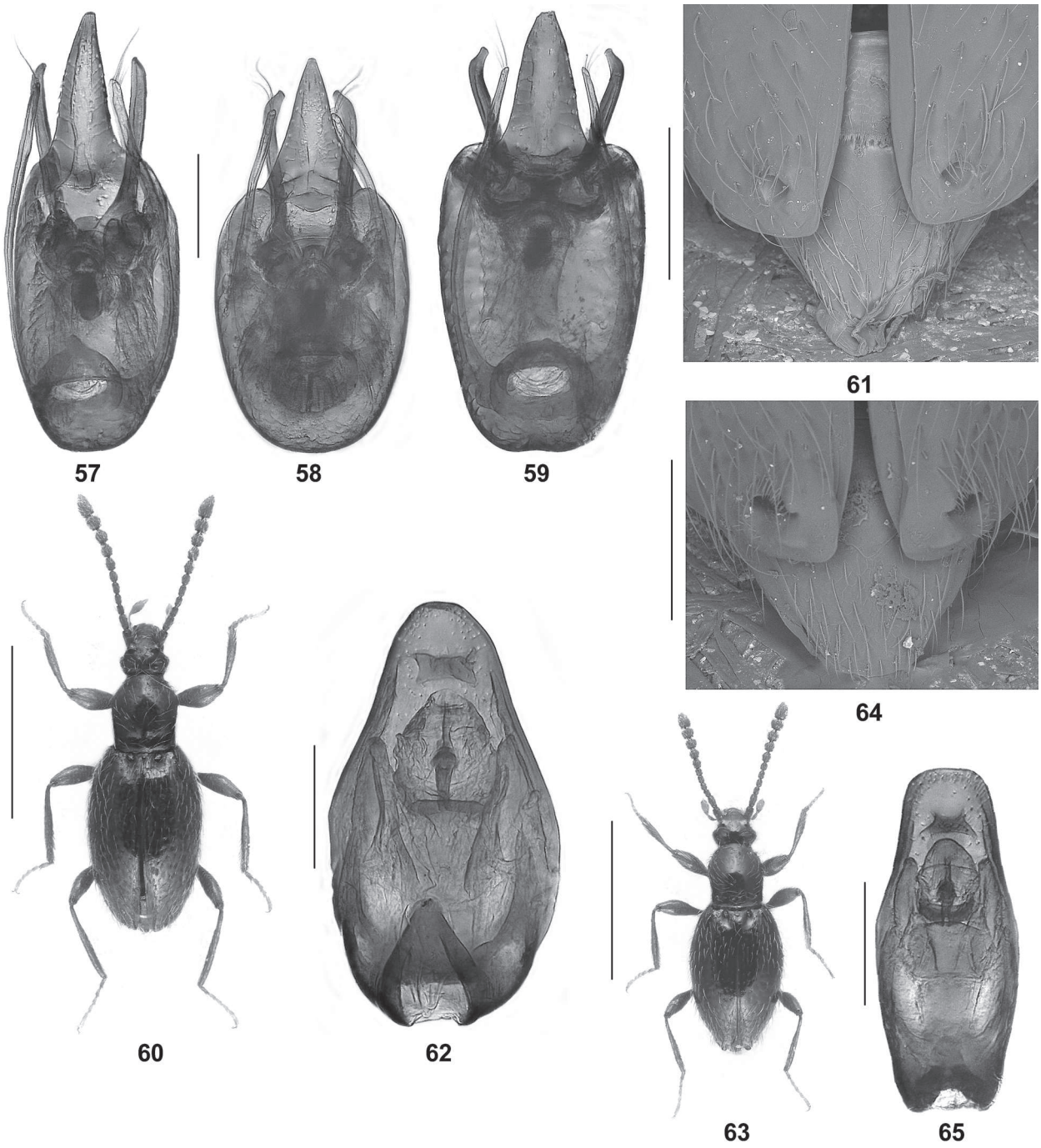
Figs 7–23: *Omalium kociani* (**7–10**), *Proteinus baculatus* (**11–14**), *Bryaxis armeniacus* (**15, 17**), *B. meghruicus* (**16, 18**), *Aleochara fugax* (**19–21**), and *Atheta meghruica* (**22–23**): (median lobe of) aedeagus in lateral and in ventral view (**7–8, 13–16**); female accessory sclerites (**9**); spermatheca (**10, 21**); habitus (**11, 19, 22**); thorax in ventral view (**12**); male antennomeres I–III (**17–18**); posterior margin of female sternite VIII (**20**); forebody (**23**). Scale bars: 11, 19, 22: 1.0 mm; 12, 23: 0.5 mm; 7–10, 13–16, 20–21: 0.1 mm; 17–18: without scale.



Figs 24–40: *Atheta meghruica* (**24–28**), *Bellatheta khustupica* (**29–35**), and *Calodera alticola* (**36–40**): head (**24**); abdomen (**25, 30, 37**); median lobe of aedeagus in lateral and in ventral view (**26–27, 31–33, 38–39**); spermatheca (**28, 34–35**); forebody (**29, 36**); apical portion of median lobe of aedeagus in ventral view (**40**). Scale bars: 25, 36–37: 0.5 mm; 24, 29–30: 0.2 mm; 26–28, 31–35, 38–40: 0.1 mm.



Figs 41–56: *Tachyusa unguis* (**41–46**), *Anotylus hamatoides* (**47–52**), *Platystethus cephalotes* (**53–54**), and *P. laevis* (**55–56**): forebody (**41**); abdomen (**42**); (median lobe of) aedeagus in lateral and in ventral view (**43–44, 51–52**); apex of ventral process of aedeagus in ventral view (**45**); spermatheca (**46**); male sternite VII (**47**); posterior process of male sternite VII (**48**); male sternite VIII (**49, 53, 55**); posterior process of male sternite VIII (**50**); male tergite X (**54, 56**). Scale bars: 41–42: 0.5 mm; 43–56: 0.1 mm.



Figs 57–65: *Euconnus longilaminatus* (**57**), *E. tavushus* (**58**), *E. karabakhus* (**59**), *Neuraphes gomarantsus* (**60–62**), and *N. syunikus* (**63–65**): aedeagus in ventral view (**57–59, 62, 65**); habitus (**60, 63**); posterior portion of elytra (**61, 64**). Scale bars: 60, 63: 1.0 mm; 61, 64: 0.2 mm; 57–59, 62, 65: 0.1 mm.

3.6 Taxonomy

3.6.1 Omaliinae (by Adriano Zanetti)

Omalium kociani Zanetti spec. nov.

urn:lsid:zoobank.org:act:5F24413E-6AB4-4D1C-B567-AB93C38380C9

(Figs 3, 7–10)

Type material: Holotype ♂: "ARMENIA, above Jermuk sifting of plant leavings near snow residues, 2400 m 39,839053N 45,693496E 21.V.2015 M. Kocian lgt. / *Omalium kociani* n. sp. det. A. Zanetti 2016 / Holotypus" (MCSNV). Paratypes: 5 ♂♂, 3 ♀♀: same data as holotype (cKoc, cZan, cAss).

Etymology: This species is dedicated to the collector, Matúš Kocian (Praha), specialist of Tachyporinae.

Description: Measurements (n = 9): body length (with extended abdomen) 2.4–3.1 mm; length of forebody from anterior margin of clypeus to posterior margin of elytra 1.53–1.88 mm; head width 0.46–0.51 mm; head length from clypeus to neck 0.35–0.42 mm; width of pronotum 0.61–0.70 mm; length of pronotum 0.42–0.49 mm; length of elytra from humeral angles to apex 0.70–0.83 mm; length of elytral suture 0.61–0.72 mm; width of elytra 0.75–0.90 mm. Habitus as in Fig. 3. Coloration: head usually blackish, sometimes brown; pronotum reddish, sometimes with more or less extensive brownish discal area: elytra yellowish red; abdomen brownish with yellowish-brown paratergites; antennae, legs and mouth parts entirely yellowish.

Head moderately transverse, 1.2–1.3 times as broad as long; temporal angles rounded but marked; two ridges extending from above insertion of antennae to compound eyes; surface without microsculpture, with dense and regular punctation forming some irregular striae between eyes and ante-ocellar pits, punctation anteriorly extending to the anterior margin of clypeus; sub-antennal impression rather wide, tentorial pits in front of ocelli long and linear, posteriorly extending to the rather small ocelli, the latter separated by a distance equal to that between ocellus and eye; space between tentorial pits and eyes elevated; neck clearly separated from the rest of the head, with dense punctation similar to that of head. Eyes small, slightly shorter than temples, without infra-orbital carina. Antenna rather thin and elongate, antennomere I elongate, twice as long as wide, II elongate ovoid, III–VIII longer than wide, gradually incrassate, IX–X subquadrate, XI longer than wide.

Pronotum transverse, on average 1.4 times as broad as long and 1.4 times as broad as head, widest in anterior half; anterior margin wider than posterior margin, anterior and posterior angles marked and slightly obtuse; lateral margins with trace of crenulation, weakly sinuate posteriorly; surface of pronotum irregular with three longitudinal elevations and two pits near lateral margin; punctation dense, similar to that of head, ground surface smooth without microsculpture.

Elytra small, 1.5 times as long (measured from humeral angles) and 1.3 times as broad as pronotum; punctation irregular and dense, coarser than on pronotum, interstices without microsculpture. Legs simple, with some spines in the apical portions of the meso- and metatibiae.

Abdomen with isodiametric microsculpture, decumbent pubescence and small wing-folding patches.

♂: aedeagus as in Figs 7–8, median lobe with narrow base, strongly enlarged subapically; apex weakly curved dorsad in lateral view and without ventral elevations.

♀: accessory sclerites as in Fig. 9; spermatheca as in Fig. 10.

Comparative notes: The new species belongs to the *Omalium caesum* group as defined by Zanetti (1987): ventral process of the aedeagus proximally much narrower than the basal bulb, parameres distinctly dilated and of intricate structure distally, with very long apical setae, tentorial impressions line-shaped. According to Zanetti (2002), the following representatives of the *O. caesum* group are distributed in Turkey, Cyprus and the Caucasus region: *O. caesum* Gravenhorst, 1806, *O. littorale* Kraatz, 1857, *O. wunderlei* Zanetti, 2002, and *O. rugatum* Mulsant & Rey, 1880). They are all clearly distinguished from *O. kociani* by long elytra, rather uniform and dark colour, and by the shape of the median lobe of the aedeagus (ventral process not narrowed at the base). *Omalium turcicum* Smetana, 1867 is very similar in coloration and in general shape, but differs by wider elytra, smaller eyes, more rounded temples and confluent punctation of the head, and an impunctate clypeus. However, the only reliable distinguishing character is the shape of the aedeagus: in *O. turcicum*, the ventral process is gradually narrowed from the base to the obtuse apex (Zanetti 2002: figure 58), whereas in *O. kociani* the ventral process is narrow at the base and enlarged in the middle, and the apex is more acute (Fig. 8). In populations of *O. caesum* with short elytra from the Alps, the ventral process of the aedeagus has a broad base (Zanetti 2002: figure 55). *Omalium cribriceps* Fauvel, 1900, which is distributed in Iran, Iraq, Syria, and Turkey (Schülke & Smetana 2015), is very similar to *O. kociani* in coloration, habitus, and the punctation of the head, but has an aedeagus of completely different shape (Zanetti 2002: figure 48) with wide median lobe and simple, linear parameres.

Distribution and natural history: This species is known only from the type locality in Armenia. The typical series was collected in an alpine meadow by sifting humus and rotting herbs near a small residual snow field.

Dialycera minuta Luze, 1906

Phyllodrepa armena Iablokoff-Khnzorian, 1959: 66; **syn. nov.**

Type material examined: *Dialycera minuta*: Holotype ♂: "Kana Gal. / J. Sahlb / 2177 / Spec.typ. / *minuta* m. det. Luze / Mus. Zool. H:fors spec. typ. No 1093 *Dialycera minuta* Luze." (MZH).

Phyllodrepa armena: Lectotype ♀, present designation: "Dzhrvezh Erevan ASSR 21.5.52" (cKhn).

Comment: This species will be treated in more detail in a revision of *Dialycera* currently in preparation. Iablokoff-Khnzorian (1959) reported minor and irrelevant differences between *D. minuta* and *D. armena* in body size and elytral punctation. Regarding the shape of the highly distinctive aedeagus, however, both are identical. In the original description of *D. armena* four

specimens from the type locality and with the same date (21.V.1952) are mentioned. The sole syntype in the Khnzorian collection is designated as the lectotype. There are four additional specimens, including males, in cKhn. They, too, were collected in the type locality, but on different dates (3.5.1951, 7.5.52, and 30.5.52) not mentioned in the original description.

3.6.2 Proteininae (by Volker Assing)

Proteinus baculatus Assing spec. nov.
urn:lsid:zoobank.org:act:3DDB2D69-A000-4FFA-AAC7-48CE4BAB7020
(Figs 11–14)

Type material: Holotype ♂: "ARMENIA [27] – WSW Kapan, W Kajaran, 39°09'22"N, 46°06'13"E, 2050 m, mixed forest, 10.VII.2018, V. Assing / Holotypus ♂ *Proteinus baculatus* sp. n. det. V. Assing 2018" (cAss). Paratypes: 1 ♂: "ARMENIA [AR16-08] N Yerevan, NW Hrazdan, 40°33'45"N, 44°23'41"E, 2000 m, mixed deciduous forest margin, litter sifted, 28.VI.2016, leg. M. Schülke" (MNB); 1 ♂: "TR. – Erzurum, 40 km NW Tortum, 2100 m, Mescit Dağları, pine forest, ca. 40°36N, 41°23E, 20.VI.1998, Solodovnikov" (cAss).

Etymology: The specific epithet is an adjective derived from the Latin noun baculus (stick, rod) and alludes to the rod-shaped sclerotized structure in the internal sac of the aedeagus.

Description: Body length 2.1–2.4 mm; length of forebody 1.4–1.6 mm. Habitus as in Fig. 11. Coloration: body black; legs dark-yellowish; antennae black with antennomere I pale-reddish.
Head and pronotum with pronounced isodiametric microsculpture and with extremely fine, barely noticeable sparse punctation. Pronotum approximately 1.8 times as broad as long and 1.4 times as broad as head, posterior margin very finely margined (visible only at a magnification of 100 x). Elytra approximately 1.9 times as long as pronotum; interstices without microsculpture. Process of mesoventrite with a moderately pronounced median keel (Fig. 12).
♂: protarsomere I dilated and very long, slightly longer than the combined length of protarsomeres II–V; mesotibia smoothly curved, with few peg setae and somewhat flattened in apical fourth; metatibia unmodified; aedeagus (Figs 13–14) 0.36–0.38 mm long; ventral process apically very acute both in lateral and in ventral view; internal sac with a long and broad black membranous tube and with a rod-shaped sclerotized structure best visible in ventral view.

Comparative notes: Based on the elongated male protarsomere I, *P. baculatus* belongs to the *P. crenulatus* group, which previously included five species in the West Palaearctic region (Assing 2007a). It is distinguished from *P. crenulatus* and other species of this group by a reddish antennomere I and by the male sexual characters, particularly by the shapes of the ventral process and of the internal structures of the aedeagus. For illustrations of the species of the *P. crenulatus* group see Assing (2007a).
Two *Proteinus* species were described from Ordubad in Azerbaijan, close to the border with Armenia: *P. planicollis* Reitter, 1905 and *P. reflexicollis* Reitter, 1905 (type material of both species examined). The new species is distinguished from them by completely black coloration of the body, by a more convex (cross-section) pronotum, and additionally as follows:
from *P. planicollis* by smaller size (*P. planicollis*: length of forebody 1.7-1.8 mm), a less broad habitus, a less transverse pronotum with indistinctly bordered lateral margins (*P. planicollis*: pronotum 1.96-2.00 times as broad as long and 1.53 times as broad as head), shorter elytra (*P. planicollis*: elytra 2.0 times as long as pronotum), an elongate male protarsomere I (unmodified in *P. planicollis*), and an aedeagus of different shape;
from *P. reflexicollis* by indistinctly bordered lateral margins of the pronotum.
The aedeagi of the two male type specimens of *P. reflexicollis* are either missing or deformed and completely bleached.

Distribution and natural history: This species is currently known from two localities in North and South Armenia and from one locality in Northeast Anatolia. The Armenian specimens were sifted from leaf litter and grass roots in mixed deciduous forests. The altitudes range from 2000 to 2100 m. The male from Turkey is slightly teneral.

3.6.3 Pselaphinae (by Volker Brachat)

Bryaxis armeniacus Brachat spec. nov.
urn:lsid:zoobank.org:act:01F595CB-9F3B-4DEE-8A4D-C97F39DE26E1
(Figs 15, 17)

Type material: Holotype ♂: "ARMENIA [4] – N Yerevan, NW Hrazdan, 40°41'40"N, 44°29'16"E, 2500 m, W-slope, sifted, 26.VI.2016, V. Assing / *Bryaxis armeniacus* spec. nov. ♂, det. Brachat 10.2018 / Holotypus" (cBra). Paratypes: 4 ♂♂, 3 ♀♀: same data as holotype (cBra); 6 ♂♂, 6 ♀♀: same data, but leg. Schülke (cSch).

Etymology: The specific epithet is an adjective derived from Armenia, where the species may be endemic.

Description: Body length 1.6–1.8 mm. Coloration: reddish-brown with the antennae and the maxillary palpi slightly paler. Pubescence on average rather long and nearly depressed; head with scattered long and erect setae.

Head weakly transverse, 0.32–0.34 mm long and 0.34–0.36 mm broad, more or less distinctly punctate in anterior half; frontal lobe 0.18 mm broad; vertex with unmodified median keel; postocular region weakly convex in dorsal view. Antennomeres III–XI 0.50 mm long; antennomeres III distinctly oblong, IV–VIII globulous and approximately as long as broad, IX–XI forming a distinct club, X more transverse than IX, and XI longer than combined length of VIII–X. Maxillary palpi rather short; palpomeres II and III with numerous tubercles, especially ventrally; palpomere IV weakly petiolate, 0.13 mm long and 0.05 mm broad.
Pronotum 0.36–0.38 mm long and 0.40–0.42 mm broad, convex in cross-section, broadest in anterior third, glabrous, punctate only near posterior margin.
Elytra 0.60–0.64 mm long and 0.68–0.72 mm broad, slightly more than one-tenth as broad as long; humeral angles marked; punctation distinct, partly coarse.
♂: head ventrally with deep semicircular gular impression, anterior margin of this impression elevated; eyes small, each composed of 12–15 ommatidia; antenna: scapus 0.145 mm long and 0.110 mm broad, dorsally slightly dilated, inner margin with a small tubercle (Fig. 17); pedicel (Fig. 17) oval, 0.08 mm long and 0.06 mm broad in dorsal view; inner margin weakly, outer margin distinctly convex; metaventrite weakly convex in cross-section, posteriorly flat and with triangular impression; legs robust; pro- and metafemora dilated; profemur with depression near base; metatrochanter posteriorly with keel-shaped dilatation; protibia slightly dilated and with deep emargination in apical third; metatibia distinctly dilated, in apical half with projecting tooth, a deep emargination, and a small spine at apical margin; modifications of male legs are subject to intraspecific variation and may be more or less distinctly reduced; aedeagus 0.40–0.43 mm long and shaped as in Fig. 15.
♀: eyes small, composed of 8–12 ommatidia; antenna with subcylindrical scapus (length: 0.14 mm; width: 0.07 mm) and with oval pedicel (length 0.07 mm; width 0.06 mm); metaventrite flattened posteriorly.

Comparative notes: *Bryaxis armeniacus* belongs to the *B. clavipes* group (see Besuchet & Kurbatov 2007). It is similar to *B. nivarius* Besuchet & Kurbatov, 2007 (Northeast Turkey), but distinguished by smaller size and the internal structures of the aedeagus.

Distribution and natural history: This species is currently known only from one locality to the northwest of Hrazdan, North Armenia. The specimens were sifted from litter and grass roots in a grassy west slope with scattered *Salix* at an altitude of 2500 m. For a photo of the type locality see figure 41 in Assing (2016b).

Bryaxis meghruicus Brachat spec. nov.

urn:lsid:zoobank.org:act:AA6E7E2A-4450-48E8-8048-8061C3C6DF5C

(Figs 16, 18)

Type material: Holotype ♂ [left antennomeres X and XI and right hind leg missing]: "ARMENIA [29] – ca. 30 km W Kapan, 39°14'40"N, 46°06'24"E, 1780 m, river bank, 11.VII.2018, V. Assing / *Bryaxis meghruicus* spec. nov. ♂, det. Brachat 10.2018 / Holotypus" (cBra).

Etymology: The specific epithet is an adjective derived from Meghru, the name of the mountain range where the type locality is situated.

Description: Body length 1.6 mm. Coloration: dark reddish-brown with the antennae, the maxillary palpi, and the legs slightly paler. Pubescence rather long and sub-erect; head with scattered long and erect setae.
Head distinctly transverse, 0.26 mm long and 0.34 mm broad, weakly punctate in anterior half; frontal lobe 0.17 mm broad; vertex with unmodified median keel. Eyes large and distinctly projecting, composed of approximately 25 ommatidia, slightly longer than the weakly convex postocular region in dorsal view. Antenna rather short; antennomeres III–XI 0.46 mm long; antennomeres III weakly oblong, IV–VII globulous and approximately as long as broad, VIII weakly transverse, IX–XI forming a distinct club, IX and X distinctly transverse, and XI slightly broader than X and longer than combined length of VII–X. Maxillary palpi rather short; palpomeres II and III with few tubercles; palpomere IV petiolate, 0.22 mm long and 0.09 mm broad.
Pronotum 0.36 mm long and 0.38 mm broad, convex in cross-section, broadest in anterior third.
Elytra 0.60 mm long and 0.64 mm broad, with fine and posteriorly more distinct punctation; humeral angles marked.
♂: head ventrally with short transverse gular impression, anterior margin of this impression elevated; antenna: scapus 0.15 mm long and 0.12 mm broad, dorsally slightly dilated, inner margin with a small flat tubercle (Fig. 18); pedicel (Fig. 18) oval and unmodified, 0.060 mm long and 0.055 mm broad in dorsal view; metaventrite with median sulcus posteriorly; legs robust; pro- and metafemora dilated; profemur with depression near base; protibia dilated and with deep emargination in apical third; metatibia distinctly dilated, in apical half with projecting tooth, a deep emargination, and a pronounced spine at apical margin; aedeagus 0.41 mm long and shaped as in Fig. 16.

Comparative notes: *Bryaxis meghruicus* belongs to the *B. clavipes* group. It is distinguished from other species of this group by the internal structures of the aedeagus and by the combination of other characters.

Distribution and natural history: The type locality is situated some 30 km to the west of Kapan, South Armenia. The holotype was floated from gravel on a river bank at an altitude of 1780 m.

3.6.4 Tachyporinae (by Michael Schülke)

Mycetoporus silvaticus Iablokoff-Khnzorian, 1962
(Fig. 4)

Mycetoporus dispersus Schülke & Kocian, 2000, **syn. nov.**

Type material: Holotype ♂: "Mravyan 18.6.60 Khnz. / *Mycetoporus silvaticus* Khn. Khnzorian det. / Typus / Holotype / *Mycetoporus silvaticus* Iablokoff-Khnzorian det. M. Schülke 2018" (IZAY).

Comment: When the species of the *Mycetoporus nigricollis* group were revised (Schülke & Kocian 2000), no type material of *Mycetoporus silvaticus* was available. The species was described based on four specimens collected in different localities in northern and southern Armenia (Zaghkadsor, Mravyan, Vachagan). In the original description, the author compared this species with *M. corpulentus* Luze, 1902, a species which was unknown from the Caucasus region at that time.
An examination of the male holotype from Mravyan (Fig. 4) revealed that the large internal structures of the aedeagus are identical to those of *M. dispersus*. As *M. silvaticus* is the senior name, it takes priority, rendering *M. dispersus* a junior synonym.

3.6.5 Aleocharinae (by Volker Assing)

Aleochara (*Ceranota*) *subtumida* (Hochhuth, 1849)

Aleochara (Polychara) khnzoriani Amiryan, 1999b: 62 f.; **syn. nov.**

Type material examined: Holotype ♀: "Armenia, Goris, sel. [village] Tatev, pod kamnem [under stones], 21.05.88, c. Kalashian M. / Type / *Aleochara khnzoriani* sp. n. Det. Amiryan A / *Aleochara subtumida* (Hochhuth), det. V. Assing 2018" (IZAY).

Comment: In the original description of *Aleochara khnzoriani*, which is based on a unique female, Amiryan (1999b) attributed the species to the subgenus *Polychara* Mulsant & Rey, 1874, now a synonym of *Xenochara* Mulsant & Rey, 1874. An examination of the holotype, however, revealed that the specimen belongs to the subgenus *Ceranota* Stephens, 1839 and is conspecific with *A. subtumida*. Hence the synonymy proposed above.

Aleochara (*Xenochara*) *fugax* Iablokoff-Khnzorian, 1962
(Figs 19–21)

Aleochara fugax Iablokoff-Khnzorian, 1962: 112 f.

Material examined: 1 ♀: "Erevan, Dzhrvezh, ASSR. 24.5.48" (cKhn).

Comment: *Aleochara fugax*, whose original description is based on a holotype and one paratype from Armenia (Vedi, Vinogradov sandstone cave, 8.IV.1958), is listed as *Aleochara* incertae sedis in Schülke & Smetana (2015). An examination of the above specimen from the Khnzorian collection, evidently not the holotype, revealed that *A. fugax* is highly similar and undoubtedly closely related to *A. parvicornis* Fauvel, 1900 of the subgenus *Xenochara*. It very much resembles *A. parvicornis* in habitus, antennal morphology, coloration, dense punctation, and the derived chaetotaxy of tergite VIII, but is distinguished by less dense punctation of the whole body (particularly so on the abdominal tergites VI and VII) and by a more slender distal portion of the spermathecal capsule. Habitus, posterior margin of tergite VIII, and spermatheca of *A. fugax* are illustrated in Figs 19–21. For illustrations of *A. parvicornis* see Assing (2009a).
The Armenian record of *A. parvicornis* by Likovský (1971) most likely refers to *A. fugax*.

Atheta (*Paralpinia*) *meghruica* Assing spec. nov.
urn:lsid:zoobank.org:act:4673F1AA-F8C1-4524-AFA3-99E4E7F502FF
(Figs 22–28)

Type material: Holotype ♂: "ARMENIA [35] – 25 km SW Kapan, 39°04'01"N, 46°16'10"E, 2150 m, near stream, sifted, 14.VII.2018, V. Assing / Holotypus ♂ *Atheta meghruica* sp. n. det. V. Assing 2018" (cAss). Paratypes: 1 ♂: same data, but leg. Schülke (cSch); 1 ♀: "ARMENIA [32] – SW Kapan, Mt. Khustup, 39°07'50"N, 46°19'55"E, 2980 m, litter near spring, 12.VII.2018, V. Assing" (cAss).

Etymology: The specific epithet is an adjective derived from Meghru, the name of the mountain range where this species was discovered and where it may be endemic.

Description: Body length 2.2–2.6 mm; length of forebody 1.1 mm. Habitus as in Fig. 22. Coloration: body black; legs blackish-brown to blackish, with paler tarsi; antennae black.
Head (Fig. 23) weakly transverse, weakly dilated posteriad; punctation rather sparse and extremely fine; interstices with pronounced microsculpture composed of isodiametric meshes. Eyes distinctly convex, protruding from lateral contours of head, slightly shorter than distance from posterior margin of eye to posterior carina of head in dorsal view. Antenna 0.65 mm long and shaped as in Fig. 24.

Pronotum (Fig. 23) approximately 1.15 times as broad as long and 1.15 times as broad as head, broadest in anterior half; pubescence of midline directed anteriad; punctation and microsculpture similar to those of head.
Elytra (Fig. 23) approximately as long as pronotum; microsculpture pronounced, composed of isodiametric meshes, these meshes larger than those on head and pronotum; punctation extremely fine, barely noticeable even at high magnification (100 x). Hind wings fully developed.
Abdomen (Fig. 25) slender, distinctly narrower than elytra; punctation fine, moderately dense on tergite III, decreasing in density from tergite III to tergite VII; interstices with distinct microsculpture composed of a mix of large isodiametric and short transverse meshes, on tergites VII–VIII predominantly of isodiametric meshes; posterior margin of tergite VII with palisade fringe; posterior margin of tergite VIII without evident sexual dimorphism, posterior margin with shallow concavity in the middle.
♂: sternite VIII with strongly convex posterior margin; median lobe of aedeagus (Figs 26–27) 0.23 mm long, ventral process of broadly triangular shape in ventral view; crista apicalis strongly projecting, but with very narrow membranous portion.
♀: posterior margin of sternite VIII smoothly convex, with long and moderately modified marginal setae; spermatheca shaped as in Fig. 28.

Comparative notes: *Atheta meghruica* is distinguished from *A. schneideri* (Eppelsheim, 1889), the only other representative of the subgenus *Paralpinia* Benick, 1974 known from the Caucasus region and Armenia, by even smaller size, a more slender body, shorter and much finer antennae with more transverse antennomeres V–X, a less convex pronotum (cross-section), a much smaller median lobe of the aedeagus (*A. schneideri*: length of median lobe nearly 0.5 mm), and a spermatheca with a shorter and less slender distal portion and with a shorter proximal portion.

Distribution and natural history: The high altitudes of the localities (2190–2980 m) and the absence of records from other regions suggest that *Atheta meghruica* is probably regionally endemic in the Meghru mountain range in South Armenia. The male specimens were sifted from litter and debris near a stream, the female from roots and debris near a spring. Both males are slightly teneral.

Bellatheta khustupica Assing spec. nov.
urn:lsid:zoobank.org:act:C342B3DE-9EE5-4950-85BD-817DBEE6FE3E
(Figs 1–2, 5, 29–35)

Type material: Holotype ♂: "ARMENIA [31] – SW Kapan, Mt. Khustup, 39°07'52"N, 46°19'43"E, 3060 m, soil near rocks, 12.VII.2018, V. Assing / Holotypus ♂ *Bellatheta khustupica* sp. n. det. V. Assing 2018" (cAss). Paratypes: 1 ♂, 4 ♀♀: same data as holotype (cAss).

Etymology: The specific epithet is an adjective derived from Khustup, the name of the mountain where this species is probably endemic.

Description: Body length 1.9–2.4 mm; length of forebody 0.8–1.0 mm. Habitus as in Fig. 5. Coloration: forebody pale-brown to brown; abdomen blackish-brown with the apex (segments VIII–X and posterior margin of segment VII) reddish-brown; legs yellow; antennae dark-yellow to pale-brown with antennomeres I–II yellow.
Head (Fig. 29) weakly transverse, somewhat dilated posteriad; punctation extremely fine, barely visible even at high magnification (100 x), and sparse; interstices with shallow microreticulation and glossy. Eyes very small, composed of approximately 6–8 ommatidia with pigmentation. Antenna short, approximately 0.5 mm long, and distinctly incrassate apically; antennomeres III transverse, IV–X strongly transverse, approximately three times as broad as long, and gradually increasing in width. Maxillary palpi short; palpomere III weakly oblong.
Pronotum (Fig. 29) approximately 1.15 times as broad as long and 1.10–1.14 times as broad as head, broadest near anterior angles, and distinctly tapering posteriad; punctation similar to that of head; microsculpture slightly more distinct than that of head.
Elytra (Fig. 29) very short, approximately 0.6 times as long as pronotum; punctation similar to that of pronotum; interstices with distinct microreticulation. Hind wings completely reduced.
Abdomen (Fig. 30) broader than elytra, broadest at segment VII; punctation fine and sparse; interstices with distinct, but shallow microreticulation, glossy; posterior margin of tergite VII without palisade fringe; posterior margin of tergite VIII broadly convex.
♂: sternite VIII much longer than tergite VIII; median lobe of aedeagus (Figs 31–33) 0.28–0.30 mm long, strongly curved in lateral view; crista apicalis very long and without distinct membranous portion.
♀: sternite VIII only indistinctly longer than tergite VIII; spermatheca shaped as in Figs 34–35.

Comparative notes: The genus *Bellatheta* Roubal, 1928 is discontinuously distributed across the Palaearctic region. In the West Palaearctic region exclusive of Middle Asia, it is represented by nine species. Except for the winged and moderately widespread *B. fatrica* Roubal, 1928 (distributed in the Balkans, Italy, and Central Europe) and *B. palata* (Benick, 1970) (recorded from Bosnia-Herzegovina, Bulgaria, Greece, and Armenia), the species are micropterous and locally endemic in mountain ranges of Spain (*B. aragonica* (Assing, 2001)), Italy (*B. kappi* (Assing, 2002); *B. rosai* (Pace, 1978)), Lebanon (*B. besucheti* (Pace, 1982)), mainland Greece (*B. renominata* (Likovský, 1984)), and Crete (*B. albimontis* Assing, 2015; *B. idana* Assing, 2015). *Bellatheta khustupica* is

distinguished from all of them by the primary sexual characters. It is readily separated from *B. palata*, the only other species of the genus recorded from Armenia, by much smaller eyes, smaller body size, paler coloration, much shorter elytra, completely reduced hind wings, and the absence of a palisade fringe at the posterior margin of tergite VII alone.

Distribution and natural history: *Bellatheta khustupica* is probably locally endemic to the alpine regions of Mount Khustup in South Armenia and most likely the only truly locally endemic species of Staphylinidae in Armenia. The specimens were sifted from soil and the roots of *Rumex alpinus* at the base of large rocks on a north slope at an altitude of 3060 m. The type locality and the habitat where the type specimens were collected are illustrated in Figs 1–2.

Calodera alticola Assing spec. nov.
urn:lsid:zoobank.org:act:3BF46F7B-FBDB-4F1B-B0D0-89D669B021C2
(Figs 6, 36–40)

Type material: Holotype ♂: "ARMENIA [37] – 30 km NW Sisian, Mt. Karkar, 39°47'05"N, 45°56'22"E, 3000 m, wetland, sifted, 15.VII.2018, V. Assing / Holotypus ♂ *Calodera alticola* sp. n. det. V. Assing 2018" (cAss).

Etymology: The specific epithet is a noun in apposition meaning inhabitant of high altitude. It alludes to the high elevation of the type locality.

Description: Body length 2.7 mm; length of forebody 1.25 mm. Habitus as in Fig. 6. Coloration: body black; legs blackish-brown with paler brown protibiae and tarsi; antennae blackish-brown with antennomeres I–II brown.
Head (Fig. 36) approximately as long as broad, of suborbicular shape; posteriorly without neck, with broad and short constriction; punctation extremely fine, barely visible even at high magnification (100 x), and moderately dense; interstices with distinct microreticulation. Eyes rather large and weakly convex, nearly as long as postocular region in dorsal view. Antenna 0.75 mm long; antennomeres III weakly oblong, IV–X approximately twice as broad as long and gradually increasing in width.
Pronotum (Fig. 36) weakly transverse, approximately 1.05 times as broad as long and approximately 1.15 times as broad as head; punctation and microsculpture similar to those of head.
Elytra (Fig. 36) slightly longer than pronotum; punctation extremely fine and dense; microsculpture distinct, composed of relatively large meshes. Hind wings fully developed.
Abdomen (Fig. 37) narrower than elytra, broadest at segments VI–VII; tergites III–V with deep, tergite VI with slightly shallower anterior impression; punctation extremely fine and dense; interstices with pronounced microreticulation; posterior margin of tergite VII with palisade fringe.
♂: posterior margin of sternite VIII broadly convex; median lobe of aedeagus (Figs 38–40) 0.32 mm long; apical internal structures of distinctive shape in ventral view.
♀: unknown.

Comparative notes: *Calodera alticola* is characterized particularly by the shape of the apical internal structures of the aedeagus in ventral view. In general appearance, *C. alticola* is similar to *C. aethiops* (Gravenhorst, 1802). It additionally differs from this species by even smaller body size, a shorter and broader posterior constriction of the head, a weakly transverse and smaller pronotum (in relation to the head and the pronotum) with finer punctation, shorter elytra, and darker legs. For illustrations of *C. aethiops* and other small West Palaearctic species see Assing (1996).

Distribution and natural history: The type locality is situated in Mount Karkar to the northwest of Sisian, Armenia. The holotype was sifted from moist debris near the shore of a small lake at an altitude of 3000 m.

Tachyusa unguis Assing spec. nov.
urn:lsid:zoobank.org:act:A4B4E5C2-EEB2-4146-B6F2-27D5873D5AEB
(Figs 41–46)

Type material: Holotype ♂: "S – ARMENIA: 25 km N Kapan, nr. Nora rachadzor village, "Davidbeksoe" water res., 13.5.2001, ca. 1400 m, leg. Shaverdo (71A) / Holotypus ♂ *Tachyusa unguis* sp. n. det. V. Assing 2018" (NHMW). Paratypes: 3 ♂♂, 1 ♀ [without head]: same data as holotype (NHMW, cAss).

Etymology: The specific epithet is a noun in apposition meaning claw. It alludes to the large claw-shaped internal structures of the aedeagus.

Description: Body length 3.0–3.4 mm; length of forebody 1.5–1.7 mm. Coloration: head black; pronotum brown to blackish-brown, sometimes with weak bronze hue; elytra reddish-brown with the anterior and lateral portions slightly darker, with or without weak bronze hue; abdomen moderately bicoloured with segments III–V reddish to reddish-brown and the apical segments bicoloured; legs reddish-yellow; antennae dark-brown with the basal 2–3 antennomeres reddish.
Head (Fig. 41) weakly transverse, with very indistinct median impression at most; punctation fine and moderately dense; interstices with very shallow, partly obsolete microreticulation. Eyes nearly as long as distance from posterior margin of eye to posterior constriction of head in dorsal view. Antenna 1.1–1.2 mm long; antennomeres II and III slender and of subequal length, IV approximately 1.5 times as long as broad, V–X of gradually

decreasing length and decreasingly oblong, and X indistinctly oblong or approximately as long as broad.
Pronotum (Fig. 41) indistinctly transverse and slightly broader than head; disc with or without very shallow and broad median impression or depression in posterior portion; lateral margins not sinuate, converging posteriad in straight line; punctation fine and moderately dense, usually slightly less fine than that of head; interstices without microsculpture.
Elytra (Fig. 41) slightly shorter than pronotum; punctation slightly finer and less dense than that of pronotum. Hind wing fully developed. Metatarsomere I longer than II, but shorter than the combined length of II and III.
Abdomen (Fig. 42) slightly narrower than elytra, broadest at segment VI, rather weakly constricted anteriorly; tergite IV (without paratergites) approximately twice as broad as long; tergites III–V with very deep anterior impressions, these impressions with coarse punctation partly separated by irregular longitudinal keels; tergite VI anteriorly with a narrow band of coarse punctation; remainder of tergal surfaces with rather sparse and fine, but distinct punctation; tergites III–V without, tergites VII–VIII with shallow microsculpture (at least posteriorly), tergite VI with or without very shallow traces of microsculpture; pubescence rather long and mostly suberect; posterior margin of tergite VII with palisade fringe; posterior margin of tergite VIII weakly concave.
♂: posterior margin of sternite VIII convex; median lobe of aedeagus (Figs 43–45) approximately 0.4 mm long and of rather robust shape; ventral process short, in ventral view of broadly triangular shape; internal sac with a pair of large claw-shaped sclerotized structures.
♀: posterior margin of sternite VIII concave in the middle; spermatheca shaped as in Fig. 46.

Comparative notes: Based on the shape of the ventral process, on the presence of claw-shaped structures in the internal sac of the aedeagus, and on the characters indicated by Paśnik (2006) (punctation of abdomen; pronotum of trapezoid shape), *T. unguis* is assigned to the *T. impressa* group. It is distinguished from all the species of this group by the shape of the ventral process of the aedeagus and by the larger claw-shaped internal structures of the aedeagus. It differs from geographically close representatives of the *T. impressa* group as follows:
from *T. impressa* (Caucasus region) by a larger and more robust aedeagus with larger claw-shaped structures, with a longer crista apicalis not clearly separated from the aedeagal capsule, and with a shorter and basally much broader ventral process (ventral view), by a usually less distinctly impressed pronotum with finer punctation, and by slightly coarser punctation separated by more distinct carinae in the anterior impressions of tergites III–V;
from *T. nitella* Fauvel, 1895 (widespread in the West Palaearctic region), with which it shares a broadly triangular ventral process (ventral view) and the shape of the crista apicalis of the aedeagus (lateral view), by paler coloration (especially of the legs, basal antennomeres, and anterior segments of the abdomen), sparser and finer punctation of the pronotum, and by a distinctly larger and more robust aedeagus with much larger claw-shaped internal structures (*T. nitella*: median lobe only approximately 0.3 mm long);
from *T. loebli* Paśnik, 2006 (Turkey) by paler coloration (especially of the legs, basal antennomeres, elytra, and anterior segments of the abdomen), finer punctation of the pronotum, the absence of a distinct median impression on the pronotum, and by a more robust aedeagus with a much broader and shorter ventral process, with a crista apicalis of different shape, and by much larger claw-shaped internal structures.
For illustrations of the primary sexual characters of other *Tachyusa* species see Paśnik (2006).

Comment: According to Paśnik (2006), *T. turcica* Paśnik, 2006 from Turkey belongs to the *T. impressa* group, too. However, based on personal observations, this name is most likely a junior synonym of *T. impressa*.

Distribution and natural history: The type locality is situated to the north of Kapan. The specimens were collected on the bank of Khashuni river at an altitude of about 1400 m.

3.6.6 Oxytelinae (by Michael Schülke)

Anotylus hamatoides Schülke spec. nov.
urn:lsid:zoobank.org:act:0167E052-EBAB-421A-BB72-39E33C1EDC97
(Figs 47–52)

Type material: Holotype ♂: "ARMENIA, Lori prov., Teghut mine, near artef. pond 41.0907° 044.8117° (soil traps) 990 m, 23.07.–23.08.2015 Karagyan & Kalashian leg. / +6 / Holotypus-♂ *Anotylus hamatoides* spec. nov. det. M. Schülke 2018" (IZAY). Paratypes: 1 ♀: same data as holotype but "+14" (IZAY); 1 ♂: "ARMENIA, Lori prov., Teghut mine, N41.0967°, E44.8218° 920 m, 22.08.–01.10.2013, Kalashian leg. / (soil traps Ser.2)" (cSch); 1 ♀: "ARMENIA, Lori prov., Teghut mine, near artef. pond 41.0907° 044.8117° (soil traps) 990 m, 23.08.–23.09.2015 Karagyan & Kalashian leg. / +2" (cSch); 1 ♀: "ARMENIA, Lori prov., Teghut mine, N41.0743° E44.8399°, 1105 m, 22.08.–01.10.2013, Kalashian leg. / (soil traps Ser.3) / *Anotylus fairmairei* Pand." (IZAY); 1 ♀: "ARMENIA, Lori prov., Teghut mine, N41.0910° E44.8514°, 887 m, 22.08.–01.10.2013, Kalashian leg. / (soil traps Ser.4)"; 1 ♀: "ARMENIA, Lori prov., Teghut mine, N41.0967° E44.8218°, 920 m, 20.06.–01.08.2013, Kalashian leg. / (soil traps Ser. 2)"; 1 ♂: "ARMENIA – Teghut env. (Akhtala), 41.091°N, 44.813°E, 1020 m, 20.V.2016, leg. A. & J. Müller" (cAss); 1 ♀: "T6 BF Armenia, near Teghut (Akhtala) 1000 m a. s. l., N 41.09099 E 44.81202 26.VIII.2016 leg. J. Müller" (cSch); 1 ♂, 1 ♀: "ARMENIA – Lori prov., Teghut mine, N41.091°N, E044.812°, 990 m, 13.VIII.–13.X.2017, leg. Kalashian et al." (cAss); 1 ♀:

"ARMENIA – Lori prov., Teghut mine, N41.096°N, E044.822°, 910 m, 25.V.–23.VII.2017, leg. Kalashian et al." (cAss); 2 ♀♀: "ARMENIA – Lori prov., Teghut mine, N41.091°N, E044.851°, 890 m, 28.IV.–25.V.2017, leg. Kalashian et al." (cAss); 1 ex.: "Lenkoran Leder (Reitter / Coll. Reitter / *Anotylus hamatus* (Fairm. & Lab.) det. Makranczy, 1997 / misidentification ! *Anotylus* sp. n. det. Makranczy, 1998" (HNHM).

Etymology: The specific epithet alludes to the strong resemblance to *A. hamatus* (Fairmaire & Laboulbène, 1856), most likely the sister taxon.

Description: Body length 1.8–2.2 mm; length of forebody 1.0–1.2 mm. Coloration: whole body black; legs yellow; antennae black; mandibles and maxillary palpi dark brown.
Measurements: males: head width 0.47–0.50 mm, head length 0.32–0.33 mm, eye length 0.14 mm, length of temples 0.13–0.14 mm, pronotum width 0.49–0.51 mm, pronotum length 0.35 mm, sutural length of elytra 0.36–0.38 mm, elytral width 0.61–0.64 mm. females: head width 0.37–0.38 mm, head length 0.24–0.25 mm, eye length 0.11–0.12 mm, length of temples 0.08 mm, pronotum width 0.40–0.42 mm, pronotum length 0,28–0.29 mm, sutural length of elytra 0.32–0.35 mm, elytral width 0.51–0.53 mm.
In size, body shape, punctuation, and microsculpture indistinguishable from *A. hamatus* and similar species.
♂: head distinctly larger, transverse (width/length 1.45–1.57), eyes as long as, or slightly longer than temples (eye length/temple length 1.0–1.1); sternite VII (Figs 47–48) similar to that of *A. hamatus*, with prominent spine and a small tubercle with two larger spines in front of it; sternite VIII highly modified (Figs 49–50), in general respect similar to that of *A. hamatus*, with broad and deep emargination at base, broad emargination at apical border, interrupted in the middle by a short two-lobed process with 4–5 setae at apex of each lobe; aedeagus (Figs 51–52), 0.47 mm long, nearly symmetric, elongate, with the apical flag-like part of the paramere more extended than in *A. hamatus*.
♀: head distinctly smaller, transverse (width/length 1.47–1.59), eyes distinctly longer than temples (eye length/temple length 1.45–1.50).

Comparative notes: *Anotylus hamatoides* is distinguished from all other representatives of the *Anotylus tetracarinatus* group with the outer edge of the front tibiae apically strongly indented only by the male primary and secondary sexual characters. The pronounced similarity to the widespread *A. hamatus* in the shape of the male sternites VII and VIII and in the shape of the aedeagus suggests a close relationship; most likely they represent sister taxa. Both species are distinguished especially by the shape of the apical margin of the male sternite VIII. Differences in the shape of sternite VII and the aedeagus are less pronounced, but constant.

Distribution and natural history: The species is currently known from the environs of Teghut in Lori province, North Armenia, and from the Lenkoran region in Azerbaijan. The Armenian specimens were collected with pitfall traps at elevations between approximately 890 and 1100 m.

Platystethus cephalotes Eppelsheim, 1878, revalidated

(Figs 53–54)

Platystethus oblongopunctatus Roubal, 1911: 2; **syn. nov.**

A specimen close to *Platystethus laevis* Märkel & Kiesenwetter, 1848 with the following labels was found in the Khnzorian collection: "Gegharkunik province, Sevan, Uchtapalar, 3000 m [= Uch-taplar Mt., 40°24'58.1"N, 45°01'41.3"E, highest 2500, not 3000 m], 1.VIII.1948 / F' de Fontainebl. f.[illegible] Gd. Verneur S. & M. 15.5.1937 [overleaf]". The specimen had been identified as *Platystethus cephalotes* by Peter Hammond, who never published this record or notes about the taxonomic status of this species. *Platystethus cephalotes* was described based on material from Michailovo [= Khashuri], a town in the Kura valley in Central Georgia [altitude approximately 750 m, but surrounded by mountain ranges of up to 2300 m]. The species was synonymized with *P. laevis*, a species originally described from the Austrian Alps, by Ganglbauer (1895), who was the first to record *P. laevis* from the Caucasus region. This synonymy was followed by Bernhauer & Schubert (1911), Herman (2001), Smetana (2004), and Schülke & Smetana (2015). Scheerpeltz (1955) and Horion (1963) reported *P. laevis* from the Caucasus region without explicitly referring to *P. cephalotes* as a junior synonym. The identification by Peter Hammond raised the question if specimens of *P. laevis* from the Caucasus region and those from Central Europe were really conspecific.
A comparison of specimens identified as *P. laevis* both from Central Europe (Austria, Italy, Slovenia) and the Caucasus region (Russia: Krasnodar region, Georgia: Abkhasia, Kvemo Svaneti) shows no significant differences in size, body shape, punctation and microsculpture. The same is true of the shape of the aedeagus. The male sternite VIII of Caucasian specimens is slightly more densely punctate in the median subapical region and has longer stout setae on either side of the midline (Fig. 53; for comparison see Fig. 55). The only clear differences between Central European and Caucasian specimens are the shape and especially the apical margin of the male tergite X. In Central European specimens, the tergite is as broad as long, with a more or less truncate and unmodified apical margin (Fig. 56), whereas in Caucasian specimens the tergite is longer than broad, with the apical margin rounded and equipped with a small tooth on either side (Fig. 54). Even though the differences are not pronounced, they are constant, suggesting that the populations from Central Europe and those from

the Caucasus region represent distinct species. Therefore, *P. cephalotes* is revalidated and *P. oblongopunctatus* Roubal, 1911, originally described based on material from Krasnaya Polyana in the Western Caucasus (Russia) and previously a synonym of *P. laevis*, is synonymized with *P. cephalotes*.
The presence of both species in Armenia is doubtful. The specimen examined from the Khnzorian collection belongs to *P. laevis*, not to *P. cephalotes*. It can inferred from the different records on both sides of the locality label, from the absence of *P. laevis* in examined Caucasian material, and finally from the absence of suitable habitats in the surroundings of Paris (France) that the specimen was most likely mislabeled. In consequence, neither of the two species is included in the checklist (Tab. 1).

3.6.7 Scydmaeninae (by Heinrich Meybohm)

Euconnus (*Cladoconnus*) *lalvarensis* Iablokoff-Khnzorian, 1964

Euconnus lalvarensis Iablokoff-Khnzorian, 1964a: 154.
Euconnus pseudorobustus Franz, 1986: 41; **syn. nov.**

Type material examined: Holotype ♂: "Alaergi Shamlug [Lalvar Shamlugh] ASSR 6.6.60 / *Euconnus lalvarensis* Iablokoff-Khnzorian / Holotypus Gen. präp. Meybohm 2018" (cKhn).

Comment: *Euconnus pseudorobustus* was previously known from Georgia and the Turkish province Rize. An examination of the holotype of *E. lalvarensis*, which was collected in Armenia, in a slope of Lalvar toward Shamlugh, in a dense forest, by sweeping [...], 6.VI.1960 (Iablokoff-Khnzorian 1964a), revealed that it is conspecific with *E. pseudorobustus*. The comparison is based on numerous additional specimens from Georgia and Armenia, and on Hlaváč & Stevanović (2013).

Euconnus (*Tetramelus*) *longilaminatus* Meybohm spec. nov.

urn:lsid:zoobank.org:act:96692EB3-A82F-4ADB -A278-EC1B64040C82
(Fig. 57)

Type material: Holotype ♂: "ARMENIA [30] – pass road E Ijevan, 1790 m, 40°52'00"N, 45°13'19"E, forest, litter & roots sift., 6.VII.2017, V. Assing / *Euc.* (*Tetramelus*) *longilaminatus* m. Meybohm 2018 det. / Holotypus" (cMey). Paratypes: 5 ♂♂, 5 ♀♀: same data as holotype (cMey).

Etymology: The specific epithet (adjective) alludes to the long ventral process of the aedeagus.

Description: Body length sexually dimorphic, in ♂ 1.38–1.45 mm and in ♀ 1.48–1.53 mm. Coloration uniformly bright reddish-brown, appendages paler. Pubescence (pronotum excluded) rather sparse, directed posteriad, sub-erect, and bent towards body. Punctation of head and pronotum very fine, that of elytra fine.
Head approximately 0.26 mm broad, broadest across the minute eyes, weakly oblong, slightly more than 1.1 times as long as broad, lateral contours smoothly curved from the eyes to the posterior constriction, postero-medially weakly bulging and projecting beyond posterior constriction. Eyes rudimentary, with only one ommatidium, without pigmentation, not projecting from lateral contours of head. Antenna with distinct four-jointed club, weakly sexually dimorphic: in ♂ antenna approximately 0.76 mm long and 0.9 times as long as elytra, length of club approximately as long as combined length of antennomeres II–VII; ♀ antenna approximately 0.73 mm long and 0.8 times as long as elytra, length of club distinctly shorter than (0.94 x) combined length of antennomeres II–VII; antennomeres II twice as long as broad, little shorter than combined length of III and IV, III–VI elongate, III and IV of equal length, V longer and VI shorter than IV, VII intermediate between VI and VIII, 1.3 times as long as broad, from VIII to XI gradually broader and XI slightly shorter than the combined length of IX and X.
Pronotum 1.1–1.2 times as broad as head and as long as broad or slightly longer, broadest slightly anterior to middle, more strongly tapering anteriad than posteriad; base with four pits, internal pits shallowly delimited, separated from each other by approximately three times their diameter, connected by a shallow transverse impression.
Elytra on average of 0.55 mm (♂) or 0.60 mm (♀) broad, in both sexes about 1.5 times as long as broad, broadest behind anterior two-fifths of elytral length, moderately convex in lateral and in dorsal view; each elytron with two clearly separated basal pits, the external one larger and laterally delimited by a short elevation, the internal one delimited by an oblique edge directed towards suture; setae approximately as long as antennomere II. Hind wings completely reduced. Femora and tibia without sexual dimorphism. Femora dilated in distal halves, profemora more strongly dilated than meso- and metafemora. Tibiae straight. Metaventrite medially flat (♂) or weakly convex in cross-section (♀).
Aedeagus shaped as in Fig. 57.

Comparative notes: See section on *E. karabakhus*.

Distribution and natural history: See section on *E. karabakhus*.

Euconnus (*Tetramelus*) *tavushus* Meybohm spec. nov.

urn:lsid:zoobank.org:act:7F2BBD70-A7F8-4272-8195-02884B90F2DD
(Fig. 58)

Type material: Holotype ♂: "ARMENIA [26a] – ENE Dilijan, Hovk, 1290 m, 40°47'39"N, 45°01'17"E, stream

valley, sifted, 3.VII.2017, V. Assing / *Euc*. (*Tetramelus*) *tavushus* m. Meybohm 2018 det. / Holotypus" (cMey). Paratypes: 2 ♀♀: same data as holotype (cMey); 7 ♂♂,10 ♀♀: "ARMENIA [17-26a] – ENE Dilijan, Hovk, 1290 m, 40°47'39"N, 45°01'17"E, moist litter near stream sifted, 3.VII.2017 Schülke" (cSch).

Etymology: The specific epithet is an adjective derived from Tavush, the name of the province where the type locality is situated.

Description: External characters as in *E. longilaminatus*, except as follows:
Body length 1.40–1.45 mm (♂) and 1.41–1.50 mm (♀).
Head approximately 0.24 mm broad, 1.1 times as long as broad. Eyes slightly projecting from lateral contours of head.
Male antenna 0.78 mm long on average, 0.92 times as long as elytron, club 0.9 times as long as combined length of antennomeres II–VII; female antenna 0.74 mm long on average, 0.86 times as long as elytron, club 0.9 times as long as combined length of antennomeres II–VII; antennomeres III, IV and VI of equal length, V longer than IV, VII 1.5 times as long as broad.
Pronotum 1.2 times as broad as head and 1.1 times as long as broad; base with five pits, the internal ones connected by a shallow impression and separated from each other by approximately half their diameter.
Elytra on average 0.55 mm (♂) or 0.59 mm (♀) broad, in lateral and in dorsal view moderately convex.
Aedeagus shaped as in Fig. 58.

Comparative notes: See section on *E. karabakhus*.

Distribution and natural history: See section on *E. karabakhus*.

Euconnus (*Tetramelus*) *karabakhus* Meybohm spec. nov.

urn:lsid:zoobank.org:act:A60B66A1-20EB-4341-9950-B6485B1A3853
(Fig. 59)

Type material: Holotype ♂: "NAGORNO-KARABAKH – [10], NW Stepanakert, 40°01'34"N, 46°28'33"E, 1210 m, mixed forest, 3.VII.2018, V. Assing / *Euc*. (*Tetramelus*) *karabakhus* m. Meybohm 2018 det. / Holotypus" (cMey). Paratypes: ♀: same data as holotype (cMey); 1 ♀: same data, but leg. Schülke (cSch); 1 ♀: "NAGORNO-KARABAKH – [7], NW Stepanakert, 39°54'22"N, 46°39'12"E, 870 m, mixed forest, 2.VII.2018, V. Assing" (cMey); 1 ♀: same data, but leg. Schülke (cSch); 1 ♀: "NAGORNO-KARABAKH [AR 18-11], NW Stepanakert, W Vank, 40°01'44"N, 46°30'15"E, 1080 m, mixed forest, litter sifted, 3.VII.2018, leg. M. Schülke" (cSch).

Etymology: The specific epithet is an adjective derived from Nagorno-Karabakh.

Description: External characters as in *E. tavushus*, except as follows:
Body length 1.54 mm (♂) and 1.54–1.61 mm (♀). Body unicoloured dark reddish-brown.
Head approximately 0.26 mm broad, slightly more than 1.1 times as long as broad. Eyes slightly projecting from lateral contours of the head.
Antenna 0.79 mm long on average, about 0.85 times as long as elytra, club about 0.85 times as long as combined length of antennomeres II–VII; III, IV and VI of equal length, V longer than IV, VII 1.7 times as long as broad.
Pronotum 1.2 times as broad as head, slightly longer than broad; base with four pits, internal pits distinctly delimited, separated from each other by approximately three times their diameter, connected by a very shallow transverse impression.
Elytra 0.60 mm (male) and 0.65 mm (female) broad on average, in lateral and in dorsal view strongly convex.
Aedeagus shaped as in Fig. 59.

Comparative notes: The three species described above belong to the *Euconnus* (*Tetramelus*) *reitteri* group (Vít & Hlaváč 1998). They differ in body length and the structure of the antennae, the shape of the eyes, the basal pits of the pronotum, and the shape of the aedeagus. The aedeagi of *E. tavushus* and *E. karabakhus* are very similar, though broader in *E. tavushus* than in *E. karabakhus*, but these species differ significantly in external features.

Distribution and natural history: The ten previously described species of the *E. reitteri* group have more or less restricted distributions in Northeast Turkey and in Georgia. The distributions of the three species described above extend the general distribution of this group to the east, south, and southeast. The specimens were sifted from leaf litter at altitudes of 870–1790 m.

Neuraphes (*Paraphes*) *gomarantsus* Meybohm spec. nov.

urn:lsid:zoobank.org:act:102F7EEC-B2A5-458F-B271-E3582970195F
(Figs 60–62)

Type material: Holotype ♂: "ARMENIA [30] – 25 km S Kapan, Gomarants Ps., 39°01'32"N, 46°21'59"E, 2190 m, oak forest, 7.VII.2016, V. Assing / *Neuraphes gomarantsus* m. Meybohm 2018 det. / Holotypus" (cMey). Paratypes: 2 ♀♀: same data as holotype (cMey); 1 ♀: "ARMENIA [31] – 25 km S Kapan, Gomarants Ps., 39°02'15"N, 46°22'13"E, 2050 m, oak forest, 7.VII.2016, V. Assing" (cMey).

Etymology: The specific epithet is an adjective derived from Gomarants Pass, the type locality.

Description: Body length 1.80–1.85 mm. Body unicoloured reddish-brown, glossy. Palpi, antennae, and legs light-brown. Pubescence bright, long, and sub-erect;

setae about as long as antennomere II. Male habitus as in Fig. 60.

Head 0.28 mm long and 0.29 mm broad; occipital constriction about three-fourths as broad as head; vertex convex, with a pair of large, very shallow impressions located between the eyes and delimiting a median trapeziform flat area; supra-antennal tubercles marked, temples pronounced, half as long as diameter of eyes; eyes large, in female weakly convex and coarsely faceted, in male more convex and more distinctly faceted.

Antennae in ♀ 0.84–0.91 mm long, antennomere I twice as long as broad, II distinctly narrower and slightly shorter than I, twice as long as broad, III and IV distinctly narrower than II and much shorter, 1.5 times as long as broad, V slightly broader and longer than III and IV, VI as broad as V, as long as III and IV, 1.3 times as long as broad, VII much broader than VI, 1.2 times as long as broad; VIII–X (measured without apical conical part): VIII broader than VII, as long as broad, IX broader than VIII, slightly broader than long, X broader than IX, 1.4 times as broad as long, XII slightly shorter than IX–X combined. Antennae in male distinctly longer and more slender than in female, 0.98 mm long, III–V 1.7 times as long as broad, VI much broader than V, but of equal length, XII much shorter than IX–X combined.

Pronotum elongate in dorsal view, 0.45 mm long and 0.39 mm broad, broadest at anterior third, lateral margins strongly rounded in anterior third, distinctly converging in posterior half and slightly concave near the almost rectangular hind angles. Posterior margin weakly arcuate; base with a transverse groove, ending laterally in a shallow depression, medially interrupted by a short longitudinal carina, on both sides of the carina with a large impression; punctures very fine.

Elytra oval, broadest at anterior third, length 1.11 mm, width 0.68 mm; humeral calli marked, forming elongate, longitudinal folds; basal pit on each elytron moderately large, closer situated to scutellum than to humerus; apices of elytra separately rounded; punctures in antero-sutural portion fine and similar to those of head, otherwise coarser; interstices slightly narrower than length of setae.

♂: elytra (Fig. 61) subapically near suture with weak longitudinal elevation, this elevation apically with pit of circular shape separated from the elytral posterior margin by its diameter and from the suture by half its diameter; diameter of this pit equal to width of antennomere VI; anterior margin of this pit with sparse short and straight setae; apical portion of elytra with some long setae directed posteriad. Metaventrite medially with a large and oblong oval impression; impressions not delimited by an elevation and with dense, weakly erect pubescence directed posteriad; pubescence of lateral portions sparser, more erect, and directed obliquely postero-laterad.

Aedeagus 0.35 mm long and 0.19 mm broad; shaped as in Fig. 62; vestigial parameres distinct, not reaching middle of median lobe; large sclerites bent dorsad apically in lateral view; the four lobes of distal sclerite of similar size.

Comment: One female not included in the type series, but collected together with the holotype, differs from the type specimens by blackish body, shorter antennae (0.84 mm), shorter elytra (1.04 mm), and the shape of the pronotum. It most likely belongs to a different species.

Comparative notes: See section on *N. syunikus*.

Distribution and natural history: See section on *N. syunikus*.

Neuraphes (*Paraphes*) *syunikus* Meybohm spec. nov.

urn:lsid:zoobank.org:act:2972F28B-0968-4810-B9F5-471E93F97F43

(Figs 63–65)

Type material: Holotype ♂: "ARMENIA [27] – WSW Kapan, W Kajaran, 39°09'22"N, 46°06'13"E, 2050 m, mixed forest, 10.VII.2018, V. Assing / *Neuraphes syunikus* m. Meybohm det 2018 / Holotypus" (cMey). Paratypes: 9 ♀♀: same data as holotype (cMey); 4 ♀♀: same data as holotype, but leg. Schülke (cSch); 1 ♂, 1 ♀: "ARMENIA [AR18-27a] WSW Kapan, W Kajaran, 39°09'22"N, 46°06'13"E, 2050 m, mixed decid. forest, litter/grass roots sifted, 13.VII.2018, leg. M. Schülke" (cSch); 1 ♀: "ARMENIA [28] – ca. 30 km W Kapan, 39°15'13"N, 46°04'16"E, 2040 m, forest margin, 11.VII.2018, V. Assing" (cMey); 1 ♀: "ARMENIA [41a] – pass N Goris, 39°35'34"N, 46°19'29"E, 1900 m, small stream valleys, 10.VII.2017, V. Assing" (cMey).

Etymology: The specific epithet is an adjective derived from Syunik, the name of the province where the type specimens were found.

Description: External characters as in *N. gomarantsus*, except as follows:

Body length 1.50–1.57 mm. Body slightly bicoloured: head and pronotum reddish-brown, glossy; elytra darker, brown or black. Male habitus as in Fig. 63.

Head length 0.25 mm, width 0.28 mm; temples moderately pronounced.

Female antenna 0.75–0.79 mm long, antennomeres I 1.8 times as long as broad, II distinctly narrower and slightly shorter than I, 1.8 times as long as broad, III and IV distinctly narrower and much shorter than II, 1.3 times as long as broad; V as long as III and IV, but slightly broader, 1.2 times as long as broad, VI as broad as V but shorter, as broad as long, VII much larger than VI, as broad as long, VIII–X (measured without apical conical part) each as long as III, VIII much broader than VII, 1.5 times as broad as long, IX broader than VIII, 1.8 times as broad as long, X broader than IX, twice as broad as long, XII as long as, or slightly shorter than

IX–X combined. Male antenna 0.82 mm long, distinctly longer and more slender than in most females.
Pronotum 0.45 mm long and 0.36 mm broad; base with a transverse groove ending laterally in a distinct pit.
Elytra 0.94 mm long and 0.62 mm broad; apices of elytra slightly oblique; setae suberect, about as long as antennomere II.
♂: elytra (Fig. 64) subapically near suture with distinct longitudinal elevation, this elevation posteriorly with a large circular pit separated from apical margin of elytra and from suture by half its diameter; diameter of this pit equal to width of antennomere VII; anterior margin of this impression with dense, very short and straight setae; elytral apices each with an obtuse erect tooth and with conspicuous long setae directed posteriad.
Aedeagus 0.28 mm long and 0.12 mm broad, shaped as in Fig. 65; vestigial parameres distinct, reaching middle of median lobe; large sclerites bent ventrad apically; basal lobes of distal sclerite large, apical lobes small.

Comparative notes: *Neuraphes syunikus* and *N. gomarantsus* clearly differ in body size, coloration, and particularly in the modifications of the male elytra. Regarding the latter, *N. syunikus* is most similar to *N. caudatus* Reitter, 1896.

Distribution and natural history: The genus is represented in the Caucasus region (including North Turkey) by numerous described and undescribed species, most of which have restricted distributions and are confined to higher altitudes. The two species described above were collected at elevations between 1900 and 2190 m and may be endemic to South Armenia.
The specimens were sifted from leaf litter, debris, and grass roots in various types of forest and in montane meadows.

3.6.8 Paederinae (by Volker Assing)

Astenus rufopacus Reitter, 1909

Astenus rufopacus Reitter, 1909: 150.
Astenus baali Coiffait, 1960: 94 f.; **syn. nov.**

The original description of *A. rufopacus* is based on an unspecified number of syntypes from "Araxesthal, Lenkoran" (Reitter 1909), that of *A. baali* on a holotype and several male paratypes from Lebanon, Turkey, and Armenia ("Delizhan" = Dilijan) (Coiffait 1960). Anlaş (2017) studied the type material of *A. rufopacus*, designated a syntype from Lenkoran as the lectotype, and illustrated the habitus and the male sexual characters. The type material of *A. baali* is deposited in the Coiffait collection in the Natural History Museum in Paris and consequently inaccessible for scientific study (see Assing 2018f). However, based on the details indicated in the original description of *A. baali* and the illustrations of the aedeagus (Coiffait 1960: figures 61–62), there is little doubt that the type material of *A. baali* is conspecific with that of *A. rufopacus*.

3.6.9 Staphylininae (by Volker Assing)

Heterothops dissimilis (Gravenhorst, 1802)

Heterothops armeniacus Coiffait, 1977: 142; **syn. nov.**

Type material examined: Holotype ♀: "Erevan, Kamaker, ASSR – 4.11.48 / Holotype / *Heterothops armeniacus* Coiff., H. Coiffait det. 1969 / *Heterothops dissimilis* (Gravenhorst), det. V. Assing 2018" (cKhn).

Comment: The original description of *H. armeniacus* is based on a unique female holotype from "Erevan, Kamaker, Arménie soviétique" (Coiffait 1977). In external characters, this holotype is identical to Armenian material identified as *H. dissimilis*. A comparison of the internal structures of the aedeagus of material from Germany and Armenia yielded no evidence that these populations should represent different species. These internal structures are slightly longer in the Armenian specimens, but otherwise of identical shape. Consequently, these differences are attributed to intraspecific variation and *H. armeniacus* is placed in synonymy with *H. dissimilis*.

Heterothops laeticolor Reitter, 1891

Material examined: **Armenia**: 1 ♂, Echmiadzin, Markara, 11.VII.1963 (cKhn). **Azerbaijan**: 1 ♂, Nakhchivan, Dzhulfa, Araks,, 3.V.1955 (cKhn).

Comment: In external characters, this species is similar to *H. minutus* Wollaston, 1860, from which it differs only by slight, but distinct differences in the shapes of the internal structures of the aedeagus.

Heterothops microtophilus Coiffait, 1977

Material examined: **Armenia**: 2 ♂♂, Gukasyan, 2000 m, *Microtus arvalis*, 3.I.1967 (cKhn).

Comment: In the shape of the head and the blackish coloration, this species somewhat resembles *H. praevius*, from which it differs by smaller body size (length of forebody 1.9–2.0 mm), completely blackish elytra, more or less distinctly paler basal antennomeres, and by internal structures of the aedeagus of completely different shape.

Heterothops praevius Erichson, 1839

Heterothops montanus Iablokoff-Khnzorian, 1966: 174; **syn. nov.**

Type material examined: Paratypes: 1 ♂, 1 ♀: "Aragaz. 3000, Karilch [= K'ari Lich lake], ASSR. 29.8.48 / vue C / Paratypus / *montanus* / *Heterothops praevius* Erichson, det. V. Assing 2018" (cKhn).

Comment: The original description of *H. montanus* is based on several type specimens from Mount Aragats (Iablokoff-Khnzorian 1966). An examination of the aedeagus (including the internal structures) of the male paratype above revealed that it is identical to that of *H. praevius*. Hence the synonymy proposed above.

Rabigus abauriae (Gridelli, 1924)

The Armenian record of *Rabigus tenuis* (Fabricius, 1782) in Eichler (1930) most likely refers to *R. abauriae*. Similarly, a revision of a female from Amasya, which the sole record of *R. tenuis* from Turkey (Assing 2009b) is based on, revealed that it belongs to *R. abauriae*. Consequently, *R. tenuis* is currently unknown from Turkey and Armenia.

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