INTERNATIONAL JOURNAL OF ADVANCES IN PHARMACY, BIOLOGY AND CHEMISTRY

Research Article

Effect of endomycorrhizae on decline of the coffee

plants (Coffea arabica) caused by Fusarium solani

Abdul Hakim Noman Ali Al-Areqi^{1,2}, Mohamed Chliyeh¹, Jihane Touati¹, Ali

Outcoumit, Fadoua Sghir¹, Amina Ouazzani Touhami¹, Rachid Benkirane¹

and Allal Douira¹.

¹Laboratoire de Botanique et de Protection des Plantes, Université Ibn Tofaïl, Faculté des

Sciences, B.P. 133, Kénitra, Maroc (Morocco).

² Ministry of Agriculture and Irrigation, Yemen.

ABSTRACT

Fusarium solani causes a wilt of coffee accompanied by a dry root rot. Endomycorrhizal treatment had a positive effect on the length of coffee plants (17.433 cm) relative to control (10.75 cm). Inoculation of the coffee plants with endomycorrhizae after their inoculation with *F. solani* had increased the length of the plants (14.133 cm). The inoculation of the coffee plants with *F. solani* and endomycorrhizae had no effect on the stem diameter. Endomycorrhizae treatment had a positive effect on the Fresh weight of aerial and root parts (4.033 g and 4.744g) relative to control (1.420g and 1.02g). Isolation of endomycorrhizal spores from the soil of the rhizosphere of the treated coffee plants revealed the presence of 14 species belonging to 2 genera (*Glomus* and *Acaulospora*) (Figure 3 and 4). The highest appearance frequency was recorded on the species of *Acaulospora rehmii* (42), followed by *A. dilatata* (14%) and *Glomus margarita* (7%).

Keywords: Fusarium solani, Endomycorrhizae, Cofee plants, Glomus, Acaulospora.

INTRODUCTION

Botanically, coffee belongs to the family Rubiacea and is classified taxonomically under the genus Coffea which includes at least 64 species grouped into four sections¹. Coffee production relies mainly on two species - Coffea arabica L. and C. canephora Pierre. Higher quality is associated with C. arabica, which contributes 70% of world coffee production². Historical evidence indicates that these base populations all descended from the few trees that survived various efforts to spread arabica coffee from Southern Arabia, now Yemen, into the main coffee producing areas in Latin America, East Africa and Asia. Arabica coffee was introduced for cultivation in Yemen from Ethiopia in earlier time by the Arabs³. The coffee trees from Yemen gave rise to two distinct botanical types⁴: 1) C. arabica var. typica Cramer, which was the earliest grown coffee in Asia and Latin America, and 2) C. arabica var. bourbon (B. Rodr.) Choussy, which came to

South America through the island of La Reunion, formerly called Bourbon⁵.

A coffee tree is susceptible to different fungal pathogen. Among these species, *Colletotrichum coffeanum*⁶, *Hemileia vastatrix*⁷. *Fusarium spp*⁸, *Rhizoctonia solani*⁹. *Cercospora coffeicola*¹⁰. *Fusarium solani* causes a wilt of coffee accompanied by a dry root rot¹¹. Also, in some parts of Africa, wilting and death of coffee tree results from root infection by *Fusarium solani*¹².

F. solani was reported in Taiz¹³; on phyllospher and phylloplane of qat, in Sana'a^{14,15}; on Banana Dwarf Cavendishi & on potatoes var Desiree, Diamond and Baraka, in Al-Huta Market at Lahej and central Market of Aden¹⁶; in houses dust, in the five main quarters (Al-Twahi, Al-Ma'alla, Crater, Khormakser and Dar Saad) at Aden City-Yemen¹⁷; on coffee fruits, in Sana'a, Ibb, Taiz, Amran, Yafea and Lahej¹⁸; on human eyes infections: in Sana'a city¹⁹;

on raw sewage, secondary effluent and dewatered sewage (manure), in Ibb sewage treatment $plant^{20}$; on the roots, the soil and stems of *Cupresus sempervirens* var. *horizontalis* and *Cupresus sempervirens* var. *pyramydales*, in three locations of Sanaa city, Al-Gameah, Al-Hasabah and Haddah²¹; in soils, collected from different localities of Yemen²²; on some spices in Taiz governorate²³; in Ibb sewage treatment plant²⁰; on Zea mays seeds, in local market of Dhmar Governorate ²⁴. The pathogenicity of *F. solani* isolated from coffee trees cultivated in Yemen on the *coffea arabica* plants was never performed in Yemen.

Howard²⁵ valued "the presence of an effective mycorrhizal symbiosis as an essential prerequisite for plant health". Data have been acquired that AM plants are more resistant to some root pathogens but more susceptible to shoot pathogens and viruses^{26, 27}. Localized morphological (lignifications of endoderm cell walls) and biochemical (antifungal chitinase) alterations in AM roots were suggested to increase the resistance against wilt diseases in tomato and cucumber ^{28, 26}. Furthermore, many details are known about the physiological and biochemical changes in plants due to symbiosis²⁹.

The aim of this work was to study the pathogenicity of *F. solani* on the coffee plants and to evaluate the effect of endomycorrhizae on the decline of the coffee plants caused by *F. solani*.

MATERIALS AND METHODS Pathogen isolation:

Fusarium sp. was isolated from the roots of the coffee plants showing the decline symptoms. These roots were washed with water, disinfected with alcohol for five minutes, put on sterile distilled water and then dried with sterile filter paper. Then, they were put on PSA agar plates (Potato Sucrose Agar: 200 g potato, 20 g sucrose, 15 g Agar-agar, and 1000 ml distilled water) and incubated on darkness at 28 degree. The morphological studies were performed after 7, 15, 25 days of incubation at 25±1°C. The culture was stained with 0.1% lacto phenol cotton blue and observed for the micro conidia, macro conidia and chlamydospores using a compound microscope. The pathogen isolates were mainly identified on the basis of cultural and morphological characters as Fusarium solani (Mart) Sacc.³⁰. The pure cultures of different isolates of F. solani were maintained on PDA slants.

Seed germination

Germination tests were undertaken at temperate of 20°C, in darkness. The complete seeds or the extracted embryos were placed into 9 cm diameter Petri dishes, on cotton humidified with distilled

water. Germination was regularly observed during 30 to 60 days. An embryo or a seed was considered having germinated when the radicle had reached a length of a few mm (for an embryo), or had broken through the seed coats. Germinated seeds were transplanted in sterile Mamora soil, watered every 3 days.

Substrate preparation

Two types of substrate were used in this experiment: Substrate 1: Used for the coffee plants inoculated with *Fusarium solani* and plants used as a control; composed by Sterile Mamora soil.

Substrate 2: Used for the coffee plants inoculated with endomycorrhizae (Endomycorrhizae; *F. solani* + endomycorrhizae).

Inoculums preparation and inoculation

a- Endomycorrhizae inoculums

A composite endomycorrhizal inoculum was selected from the soil and the root samples taken from rhizosphere the olive trees grown in different Moroccan olive groves. Barley seeds were disinfected with Sodium hypochlorite (5%) for two minutes; they were rinsed with the tap water and sown in pots containing mycorrhizal soil and roots fragments of the olive trees. These pots were brought to the greenhouse and sprayed regularly with distilled water and received 100 mL of a nutritive solution every two weeks³¹.

The inoculation with the endomycorrhizal inoculums was performed by planting coffee plants (4 leaves stage) in the substrate 2.

b- Fusarium inoculums

F. solani conidial suspension was prepared by scraping the conidia developing on this fungus culture aged at the age of 7 days with water distilled water. This suspension was adjusted to a concentration of 10^{6} conidia/mL.

Inoculation with *F. solani* was performed by dipping the roots of the coffee plants in the conidial suspension for 6 hours after their artificial injury. Plants inoculated with *F. solani* and endomycorrhizae were inoculated first with *F. solani* and planted in the substrate 2.

Measured parameters

10 months after inoculation, coffee plants were cut in the level of the collar. The roots were washed with a tap water and dried on absorbent paper overnight under ambient laboratory conditions. The height of the vegetative part was measured with a meter. Fresh weights of vegetative biomass and root biomass were measured using a digital scale and the leaves number was counted on the vegetative part. The mycorrhizal frequency and intensity were quantified using the technique of Phillips and Hayman³², as modified by Koske and Gemma³³. Spores were extracted following the wet sieving method described by Gerdemann and Nicolson³⁴.

Analysis of the variance and of the mean comparisons using the LSD test (p = 5%) were performed using the software STATISTICA program.

RESULTS

F. solani culture was hyaline with the appearance of a clear purple in the center of the PSA culture, exactly after 10 days of incubation (Figure 1A).

Micro conidia: $6-7 \ \mu m$ in size, sickle shaped with blunt ends was (Figure 1B), macro conidia ware round to oval with a size of $23-26 \ \mu m$ (Figure 1C). *Fusarium solani* was able to form intercalary and terminal chlamydospores (Figure 1E and 1D).

F. solani has proven able to induce decline symptoms on coffee plants, the endomycorrhizae treatments had a positive effect on the growth of coffee plants inoculated with *F. solani* (Figure 1).

The data on table 1 showed that the treatment with endomycorrhizae showed a highest leaves number (16.088) relative to the control (9.166). The inoculation with *F. solani* showed the lowest leaves number (6). In fact, the treatment with endomycorrhizae has increased the leaves number of the coffee plants inoculated with *F. solani* (11.44).

As the same, endomycorrhizae treatment had a positive effect on the length of coffee plants (17.433 cm) relative to control (10.75 cm). Inoculation of the coffee plants with endomycorrhizae after their inoculation with *F. solani* had increased the length of the plants (14.133 cm). The inoculation of the coffee plants with *F. solani* and endomycorrhizae had no effect on the stem diameter (Table 1). Endomycorrhizae treatment had a positive effect on the Fresh weight of aerial and root parts (4.033 g and 4.744g) relative to control (1.420g and 1.02g) (Table 1).

The roots observation of the inoculated coffee plants showed that endomycorrhizae has been introduced and fixed inside the roots forming different structures; Vesicles and arbuscular (Figure 2). Arbuscular content was 31.5% in the roots of the coffee plants inoculated only with endomycorrhizae. Thus, it was 29.5% in the group of plants inoculated with *F. solani* and endomycorrhizae (Table 2). Vesicular content was respectively 33.76 and 27% in the group of the coffee plants inoculated only with endomycorrhizae and with *F. solani* and endomycorrhizae.

Isolation of endomycorrhizal spores from the soil of the rhizosphere of the treated coffee plants revealed the presence of 14 species belonging to 2 genera (*Glomus* and *Acaulospora*) (Figure 3 and 4). The highest appearance frequency was recorded on the species of *Acaulospora rehmii* (42%), followed by *A. dilatata* (14%) and *Glomus margarita* (7%) (Figure 4).

The highest spore's number was isolated from the rhizosphere of the coffee plants inoculated with F. *solani* and endomycorrhizae (126 spores) compared to those inoculated only with endomycorrhizae (72 spores) (Table 2).

DISCUSSION AND CONCLUSION

The reduction of wilt disease infestation of the pathogen and loss of shoot fresh weight by AM depended on the resistance level of the cultivars. Toth et al.³⁵ reported for maize inbreeds that high levels of genetic resistance against fungal pathogens can be associated with a lower susceptibility against AMF, reducing the benefits, these plants could form the symbiosis. Arbuscular mycorrhizal fungi have shown a positive effect on different plant species; boxthorn ³⁶ and Palm date ³⁷. However, this is not true regarding wilt resistance of Linum, since all cultivars presented were highly infected by AMF In cultivars of winter barley, too, levels of AMF colonization were not correlated to the beneficial AM effect against soil borne fungal pathogens³⁸. Comparing different AMF colonization densities within one tomato cultivar Caron *et al.*³⁹ found the same. According to Morandi⁴⁰, this resistance is due to the

fact that mycorrhizal fungi cause an accumulation of phenolics, in particular phytoalexins and associated flavonoids and isoflavonoids, in the roots of their host plants. Similar results on the growth of wild strawberries have been reported by Mark and Cassells⁴¹. Where it was observed that the beneficial effect of the endomycorrhizal fungus Glomus fistumosum prevailed over the pathogenic fungi *Phytophthora fragariae*. Trotta *et al.*⁴² working with wild strawberry and tomato plants found that the effect of endomycorrhizae offset the negative effect of the pathogenic fungi P. fragariae and P. nicotianae respectively. In all relevant literature, the improved P uptake by the mycorrhizal plants is emphasized⁴³. The main contribution of AMF to the host is to reach and translocate phosphate through their extracortical hyphae, which can extend up to 9 cm in the soil⁴⁴. Root colonization in both the plant species is reduced by nearly 50 % following the inoculation (Verticillium double dahliae + Mycorrhizae), compared to AM inoculation only⁴⁵.

	••	1
XX/XX/XX/	119n	bc.com
** ** **	Jap	DC.COM
	. J . L	

Effect of <i>Fusarium solani</i> and endomycorrhizae on different agronomical parameters of coffee plants							
	Control	F	Мус	F + Myc			
Leaves number	9.166 ^b	6 ^c	16.088 ^a	11.444 ^b			
Length (cm)	10.75 °	7.837 ^d	17.433 ª	14.133 ^b			
Stem diameter (cm)	0.316 ª	0.237 ^a	0.5222 ª	0.4111 ^a			
F.A.W. (g)	1.420 ^b	0.960 °	4.033 ^a	1.888 ^b			
F.R.W. (g)	1.020 ^b	0.86 ^c	4.744 ^a	1.255 ^b			

Table 1

The results of the same line followed by different letters differ significantly at 5%. F: *F. solani* Myc: Mycorrhizal F.A.W.: Fresh aerial weight F.R.W.: Fresh root weight

 Table 2

 Mycorrhizal parameters of coffee plants inoculated with *F. solani* and endomycorrhizal inoculums.

	Control	F	Мус	F + Myc
Spores number	0 c	0 c	72 b	126 a
Arbuscular content	0 b	0 b	31.5 a	29.5 a
Vesicular content	0 a	0 a	33.76 a	27 a

The results of the same line followed by different letters differ significantly at 5%.

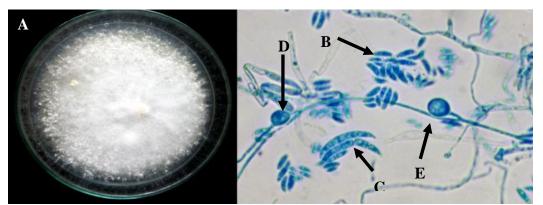
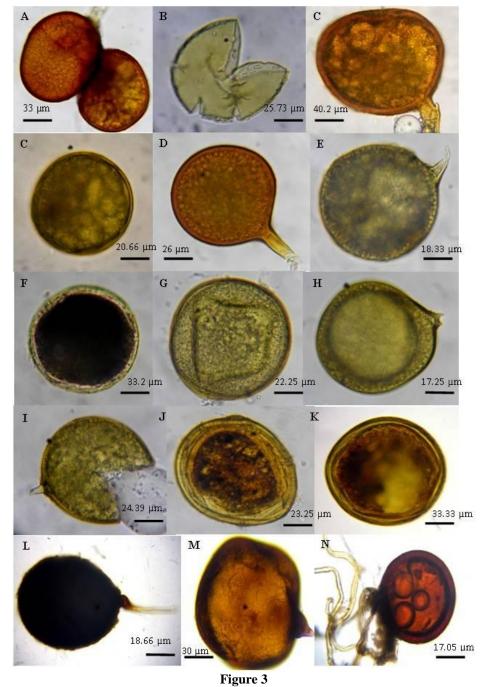


Figure 1

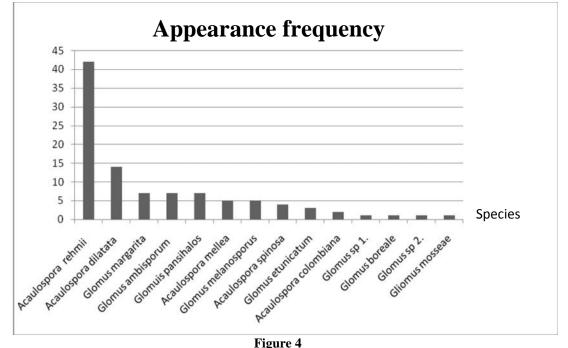
Fusarium solani in the age of 10 days PSA media. Different organs of *F. solani*; micro conidia (B); macro conidia (C); *intercalary chlamydospore* (D); terminal chlamydospore (E).



Figure 2 Coffee plants inoculated with different inoculums; Control (A); Endomycorrhizae (B); F. solani (C); Endomycorrhizae + F. solani (D).



Acaulospora rehmii (A) ; A. dilatata (B) ; Glomus margarita (C) ; G. ambisporum (D) ; G. pansihalos (E); A. mellea (F) : A. spinosa (G) ; G. melanosporus (H) ; Glomus etunicatum (I); A. colombiana (J) ; Glomus sp 1. (K), Glomus boreale (L); Glomus sp 2. (M); G. mosseae (N).



Appearance frequency of the endomycorrhizal species isolated from the rhizosphere of the coffee plants inoculated with endomycorrhizae.

According to McAllister et al.46, pathogenic fungi reduced to AM inoculation fungi were established on a root system. Chliveh et al.⁴⁷ had mentioned a benific effect of AMF on the growth of tomato plants and a benefic effect against Verticillium dahliae.

After inoculating with endomycorrhizae, plants inoculated with F. solani have shown a high number of endomycorrhizal spores in their rhizosphere. So the infection by pathogen may promote a host to form a symbiotic relation with endomycorrhizae.

The results presented here and in literature show that AM plants react as a whole - besides the non infected parts of the root system. However, the type of this reaction depends on the pathogen and on the plant part infected. To allow generalization about mechanisms of interactions between plants, AMF, and pathogens, it would be ideal to have results regarding all relevant physiological and morphological parameters measured all in combinations of symbionts and pathogens. So, the mycorrhization of plants would be a benefical way to protect coffee tree against F. solani and other fungi in Yemen.

CONCLUSION

Fusarium solani, fungal pathogen of plants wilting, was able to induce wilt, dwarfing and leaves alteration on plants of coffee plants 'Coffea arabica'. Plants of this species inoculated with this pathogen and planted in a soil which contains composite endomycorrhizal inoculums have shown an amelioration of the growth parameters: growth in height, leaves number, vegetative and root fresh weight. This increase seems to be the results of root mycorrhization of plants infected with F. solani. This mycorrhization of roots was illustrated by the isolation of endomycorrhizal species from the rhizosphere of plants inoculated with F. solani developing in soil containing mycorrhizae.

REFERENCES

- 1. Carvalho, A. and Monaco. The breeding of Arabica coffee, in F.P. Ferwerda and F. Wit (eds) Outlines of Perennial Crop Breeding in the Veenman and Zonen Tropics, NV. Wageningen, 1969; pp: 198-216.
- 2. Lashermes P, Combes MC, Robert J, Trouslot P. D'Hont A, Anthony F, Charrier A. Molecular characterisation and origin of the Coffea arabica L. genome. Molecular and General Genetics MGG, 1999; 261 (2): 259-266.
- 3. Smith FR. A history of coffee. In : M N . Clifford & KC . Wilson (Eds) Coffee, Botany, Biochemistry and production of beans and beverage. Croom Helm, London & Sydney, 1985; pp. 1-12.
- 4. Krug CA, JET Mendes & A Carvalho. Taxonomia de Coffea arabica L. Descripao das variedas e formas encontradas no estado de Sao

Paulo . Boletim Tecnico 62, Instituto Agronomico do Estado (Campinas, Brazil), 1939; 57 p.

- Lashermes P, M C Combes, J Cros, P Trouslot, E Anthony & A. Charrier. Origin and genetic diversity of *Coffea arabica* L. based on DNA molecular markers. 16th Conference of ASIC, Kyoto (Japan), in press, 1995.
- Nutman FJ, Roberts FM. Investigations on a disease of *Coffea arabica* caused by a form of *Colletotrichum coffeanum* Noack: I. Some factors affecting infection by the pathogen, 1960; 43 (3): 489–505.
- Fernandez D, Santos P, Agostini C, Bon MC, Petitot AS, Silva MC. Leonor Guerra-Guimarães, Ribeiro A, Argout X. and Nicole M. Coffee (*Coffea arabica* L.) genes early expressed during infection by the rust fungus (*Hemileia vastatrix*), 2004 ; 5 (6) : 527–536.
- Muleta D, Assefa F. and Granhall U. In vitro Antagonism of Rhizobacteria Isolated from *Coffea arabica* L. against Emerging Fungal Coffee Pathogens. Engineering in Life Sciences, 2007; 7 (6): 577–586.
- Priyatmojo A, Escopalao VE, Tangonan NG, Pascual CB, Suga H, Kageyama K and Hyakumachi M. Characterization of a New Subgroup of *Rhizoctonia solani* Anastomosis Group 1 (AG-1-ID), Causal Agent of a Necrotic Leaf Spot on Coffee, 2001; 91 (11): 1054-1061.
- Staver C, Guharay F, Monterroso D, Muschler RG. Designing pest-suppressive multistrata perennial crop systems: shade-grown coffee in Central America, 2001, 53 (2): 151-170.
- 11. Waller JM, Bigger M, Hillocks RJ. Coffee Pests, Diseases and Their Management. Wallingford : CABI, 2007; 333 pp.
- 12. Varnam AH, Sutherland JP, Beverages : technology, chemistry and microbiology. Gaithersburg, Maryland: Aspen, 1999; 439pp.
- Kamal, M, Al Agbari, AA. Manual of plant diseases in the Yemen Arab Republic. London (United Kingdom), Precision Press, 1985; 144 p.
- 14.Alhubaishi AAA, Abdel-Kader MIA. Phyllosphere and phylloplane fungi of qat in Sana'a, Yemen Arab Republic. Journal of Basic Microbiology, 1991; 31(2): 83–89.
- 15.Mahmoud ALE. Mycotoxin-producing potential of fungi associated with qat (*Catha edulis*) leaves in yemen. Folia Microbiologica,1996; 45 (5): 452-456.
- 16. Shatarh MM. Pot-harvest diseases of some prioritized perishables and methods of control. Thesis submitted in a partial fulfillment for the

requirements of the Degree of Master of Science in Agriculture. Faculty of Nasser of Agricultural Sciences. University of Aden, 1999.

- 17. Dughaish ZH. Fungi of the house Dust in Aden-Yemen. Master thesis. Faculty of education department of biological sciences and geology. Ain Shams, 2002.
- Al-Kolaibe AMG. Myobiota and Mycotoxins of Coffee Fruits in Yemen. The degree of M. Sc. Botany (Microbiology-fungi), 2002; 150pp.
- 19. Alghalibi SMS, Shater ARM. Mycoflora and mycotoxin contamination of some dried fruits in Yemen Republic. Association of University Bulletin on Environmental Research, 2004;7: 19–27.
- 20. Al-Zubeiry AHS. Microflora inhabiting raw sewage, secondary effluent and dewatered sludge in Ibb, Yemen republic. Ass. Univ. Bull. Environ. Res., 2005; 8 (1): 1-16.
- 21. Al-Bori EAM. Studies on Cypress wilt disease in Sana'a-Yemen. Thesis submitted in a partial fulfillment for the requirements of the Degree of Master of Science in Agriculture (Plant pathology). Faculty of Agriculture, Sana'a University, 2006; 160pp.
- 22. EL-Said AHM. Keratinophilic fungi in soils of Yemen Arab Republic. Journal of Islamic Academy of Sciences, 1995; 8:4, 151-154.
- 23. Gazem MAH. Mycological and aflatoxins status of some species commonly consumed in Taiz Governorate, Republic of Yemen. A thesis submitted in partial fulfillment of the Requirements for the degree of M. Sc. In Microbiology. Faculty of Science, Taiz University, 2008;160 pp.
- 24. Al Qufaily SAS . Detection of fungi, Aflatoxin B1 and in Seeds of Zea mays in local market and the study of its Biological effects. AThesis Submitted to the College of Education-University of Thamar, 2010; 54 pp.
- 25. Howard A .An Agricultural Testament, Oxford University Press, Geoffrey Cumberlege, Londen, UK. 1943.
- Dehne H W. Zur Bedeutung der vesikulararbuskul~iren (VA) Mykorrhiza ftir die Pflanzengesundheit. Habil. Univ, Haanover, Germany,1987a.
- Schonbeck F ;Endomycorrhiza in relation to plant diseases. In Soil-Borne Plant Pathogens. Eds. B Schippers and W Gains,1979 ;pp 272-280. Acadamic Press, London, UK.
- Dehne H W und Schtinbeck F. Untersuchungen zum Einflub der endotrophen Mykorrhiza auf Pflanzenkrankheiten. II. Phenolstoffwechsel und

Liginifizierung. Phytopatol Z. 5, 1979 ; 210-216.

- 29. Smith S E and Gianinazzi-Pearson V Physiological interactions between symbionts in vesicular-arbuscular mycorrhizal plants. Annu. Rev. Plant Physiol. MoI. Biol, 1988; 39, 211-244.
- Subramanian C V. Hyphomycetes. Indian Council of Agricultural Research, New Delhi, 1971; 930 Pp.
- 31. Chliyeh M, Ouazzani Touhami A, Filali-Maltouf A, El Modafar C, Moukhli A, Oukabli A, Benkirane R and Douira A. Effect of a composite endomycorrhizal inoculum on the growth of olive trees under nurseries conditions in Morocco. International Journal of Pure & Applied Bioscience, 2014; 2 (2): 1-14.
- 32. Phillips, JM, Hayman, DS. Improved procedures for clearing roots and staining parasitic and vesicular- arbuscular mycorrhizal fungi for rapid assessment of infection. Trans. Br. Mycol. Soc, 1970; 55, 158-161.
- Koske I, and Gemma JN.A modified procedure for staining roots to detect VA mycorrhizas. Mycol. Res, 1980; 92: 486-505.
- Gerdemann, J W and T H,Nicolson . Spores of mycorrhizal Endogone extracted from soil by wet sieving and decanting. Trans. Brit. Mycol, 1963 ;Soc. 46.235-244.
- 35. Toth R, Toth D, Starke D and Smith D R . Vesicular-arbuscular mycorrhizal colonization in Zea mays affected by breeding for resistance to fungal pathogens. Can. J. Bot, 1990; 68, 1039-1044.
- 36. Touati J, Chliyeh M, Ouazzani Touhami A, Benkirane R. and Douira A. Effect of arbuscular mycorrhizal fungi on the growth and root development of the boxthorn tree (*Lycium Europaeum*) under a greenhouse conditions. Int. J. Pure App. Biosci, 2014; 2 (6): 84-91.
- 37. Sghir F, Chliyeh M, Touati J, Mouria B, A. Ouazzani Touhami, Filali-Maltouf A. El Modafar C, Moukhli A, Benkirane R, and Douira A. Effect of a dual inoculation with endomycorrhizae and *Trichoderma harzianum* on the growth of date palm seedlings. Int. J. Pure App. Biosci, 2014 ; 2 (6): 12-26.
- Dehne H W, Schtinbeck Fund Baltruschat H. Untersuchungen zum Einfluf3 der endotrophen Mykorrhiza anf Pflanzenkrankheiten. III. Chitinase-Aktivit~it und Ornithinzyklus. Z. Pflkrankh. Pflschutz, 1978b; 85, 666-678.
- 39. Caron M, Fortin, JA, and Richard, C. Effect of phosphorus concentration and Glomus

intraradices on Fusarium crown and root rot of tomatoes. Phytpathology , 1986; 76, 942-946.

- 40. Morandi D. Occurence of phytoalexins and phenolic compounds on endomycorrhizal interactions, and their potential role in biological control. Plant Soil, 1996; 185: 241–251.
- 41. Mark GI and A C Cassells. Genotypedependence in interaction between *Glomus fistulosum*, *Phytophthora frgariae* and the wild strawberry (*Fragaria vesca*). Plant and soil , 1996;185: 233-239.
- 42. Trotta, A, GC Vanese, E Gnavi, A Fascon, S Sampo and G Berta. Interaction between the soilborne root pathogen *Phytophthora nicotianae* Var *parasitica* and the arbuscular mycorrhizal fungus *Glomus mosseae* in tomato plant. Plant Soil , 1996; 185: 199-209.
- 43. Nurlaeny, N, H Marschner, and E George. Effects of liming and mycorrhizal colonization on soil phosphate depletion and phosphate uptake by maize (*Zea mays* L.) and soybean (*Glycine max* L.) grown in two tropical acid soils. Plant Soil, 1996; 181: 275–285.
- 44. Sylvia, DM and DH Hubbell. Growth and sporulation of vesicular-arbuscular mycorrhizal fungi in aeroponic and membrane systems. Symbiosis, 1986; 1: 259-267.
- 45. Karagiannidis, N, F Bletsos and N Stavropoulos. Effect of Verticillium wilt (*Verticillium dahliae* Kleb.) and mycorrhiza (Glomus mosseae) on root colonization, growth and nutrient uptake in tomato and eggplant seedlings. Scientia Horticulturae, 2002; 94(1-2): 145-156.
- 46.Mc Allister, CB, I Garcia-Romera, AJA. Godeas, Ocampo. Interactions between *Trichoderma koningii*, *Fusarium solani* and *Glomus mosseae*. Effects on plant growth, arbuscular mycorrhizae and the saprophyte inoculants. Soil Biol. Biochem, 1994; 26 (10): 1363-1367.
- 47. Chliyeh M, Ouazzani Chahdi A, Selmaoui K, Ouazzani Touhami A, Filali Maltouf A, El Modafar C, Moukhli A, Oukabli A, Benkirane R. and Douira A. Effect of *Trichoderma harzianum* and arbuscular mycorrhizal fungi against Verticillium wilt of tomato. International Journal of Recent Scientific Research, 2014; 5: (2): 449-459.