

Chronic inflammation and the hallmarks of aging



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ABSTRACT

Background: Recently, the hallmarks of aging were updated to include dysbiosis, disabled macroautophagy, and chronic inflammation. In particular, the low-grade chronic inflammation during aging, without overt infection, is defined as “inflammaging,” which is associated with increased morbidity and mortality in the aging population. Emerging evidence suggests a bidirectional and cyclical relationship between chronic inflammation and the development of age-related conditions, such as cardiovascular diseases, neurodegeneration, cancer, and frailty. How the crosstalk between chronic inflammation and other hallmarks of aging underlies biological mechanisms of aging and age-related disease is thus of particular interest to the current geroscience research.

Scope of review: This review integrates the cellular and molecular mechanisms of age-associated chronic inflammation with the other eleven hallmarks of aging. Extra discussion is dedicated to the hallmark of “altered nutrient sensing,” given the scope of *Molecular Metabolism*. The deregulation of hallmark processes during aging disrupts the delicate balance between pro-inflammatory and anti-inflammatory signaling, leading to a persistent inflammatory state. The resultant chronic inflammation, in turn, further aggravates the dysfunction of each hallmark, thereby driving the progression of aging and age-related diseases.

Main conclusions: The crosstalk between chronic inflammation and other hallmarks of aging results in a vicious cycle that exacerbates the decline in cellular functions and promotes aging. Understanding this complex interplay will provide new insights into the mechanisms of aging and the development of potential anti-aging interventions. Given their interconnectedness and ability to accentuate the primary elements of aging, drivers of chronic inflammation may be an ideal target with high translational potential to address the pathological conditions associated with aging.

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Keywords Aging; Ageing; Inflammation; Inflammaging; Nutrient sensing; Senescence

1. INTRODUCTION

Aging is a complex process that affects humans at the molecular, cellular, tissue, and systemic levels. It results, in part, from the compounding accumulation of damage and waning repair mechanisms at each hierarchical level, altogether contributing to the development of age-related diseases (ARDs) later in life. Therefore, to no surprise, aging is the single most important risk factor for many chronic diseases contributing to morbidity and mortality. While aging predisposes older populations to more severe infection resulting from communicable diseases, such as influenza and severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) [1,2], this review will focus on the interplay of inflammaging and aging hallmarks associated with non-communicable diseases such as cardiovascular diseases (CVDs), cancer, osteoarthritis, type 2 diabetes (T2D), and

neurodegenerative diseases such as Alzheimer’s disease (AD) and Parkinson’s disease (PD) [3,4].

Despite many proposed theories, none alone comprehensively captures the aging process. Age-related changes were traditionally depicted by nine cellular and molecular hallmarks of aging that include 1) genomic instability, 2) telomere attrition, 3) epigenetic alterations, 4) loss of proteostasis, 5) deregulated nutrient sensing, 6) mitochondrial dysfunction, 7) cellular senescence, 8) stem cell exhaustion and 9) altered intercellular communication [5] (Figure 1). More recently, several concepts have emerged in light of advances in high throughput multi-omics analysis, such as transcriptomics, proteomics, and epigenomics. In a recent review, López-Otin and colleagues expanded on the original nine hallmarks of aging, including disabled macroautophagy, dysbiosis, and chronic inflammation [6]. Each hallmark not only manifests during normal aging but the corresponding aging

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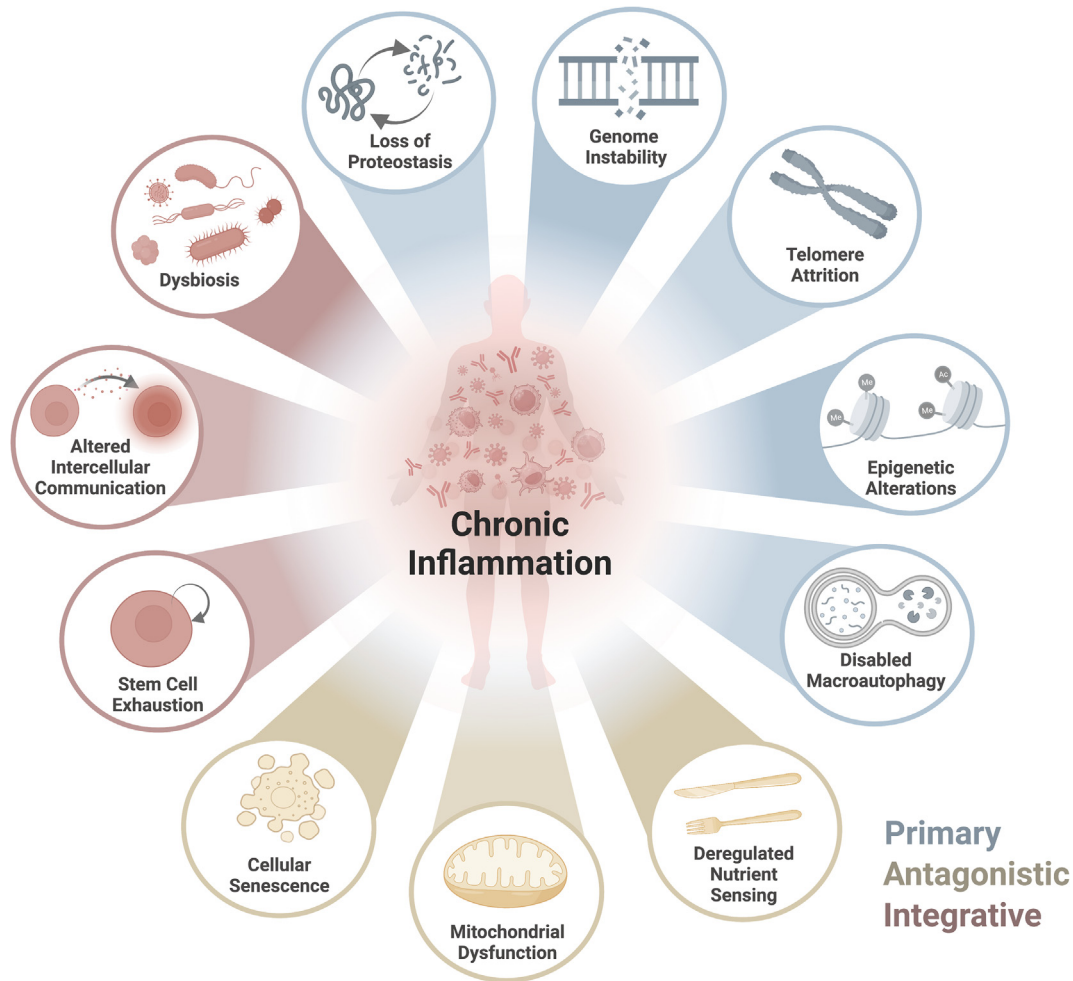


Figure 1: Chronic Inflammation and the Hallmarks of Aging: Relationships of chronic inflammation and other hallmarks of aging: genomic instability, telomere attrition, epigenetic changes, loss of proteostasis, and disabled autophagy grouped as the primary hallmarks; deregulated nutrient sensing, mitochondrial dysfunction, and cellular senescence grouped as the antagonistic hallmarks; altered intercellular communication, stem cell exhaustion, and dysbiosis grouped as the integrative hallmarks (López-Otín, 2023).

pathology can be exacerbated or reduced through experimental aggravation or amelioration of the hallmark [5,6]. The large degree of mechanistic interplay between the hallmarks makes the hallmark relationships best characterized as a web of bidirectional loop interactions rather than stand-alone pillars.

Under the geroscience hypothesis, failure in this “network of aging” and their homeostatic mechanisms accelerates the pace of aging and susceptibility to ARDs. An integrated and holistic approach to aging biology emphasizes the significance of influential factors across the network of hallmarks, including diet, exercise, and other lifestyle factors. The human immune system, specifically inflammation, is also largely considered one of these ubiquitous factors that, to some degree, impacts each of these processes [7]. Inflammation is a broad term referring to the defense mechanisms that evolved to protect an organism from infection and injury. While acute inflammation is a cascade of steps in response to infection or injury that ultimately clears invading pathogens and incites wound healing, chronic inflammation is a potentially pathologic process arising from the perpetuity of the initial trigger or the dysregulation of signaling pathways that is harmful to health. “Inflammaging” describes this non-resolving, low-grade, and chronic inflammation process that progresses with age [8,9].

This review emphasizes the broad overview of bidirectional relationships between chronic inflammation and the other 11 hallmarks of aging, which contribute to “inflammaging.” We will dedicate additional discussion on altered nutrient sensing.

2. PRIMARY HALLMARKS

The primary hallmarks, either individually or through synergy, can lead to damage on a molecular and cellular level. The cumulative effects of this damage are universally negative and believed to be involved in the instigation and progression of ARDs.

2.1. Genomic instability

Genomic instability refers to DNA alterations, including changes in nucleic acid sequences and aneuploidies [10]. These events can irreversibly alter the content of the genome, contributing to detrimental changes in gene expression, cell division, senescence, and death. Altogether, these cell outcomes conspire towards functional impairment at the tissue level and ultimately manifest accelerated aging pathology and ARDs as cells accumulate nuclear and mitochondrial genomic damage with age. Maintenance of genomic stability is

essential for cellular integrity to prevent errors from DNA replication by endogenous and exogenous challenges. Each day, endogenous insults - including replication errors and free radical damage during normal metabolism - result in as many as 10^4 – 10^5 DNA lesions per normal cell [11,12]. While aberration from exogenous exposures (such as ultraviolet light (UV), ionizing radiation, mutagenic chemicals, etc.) are largely unique to each individual's biography of exposures, the relatively high baseline rate of DNA damage suggests that the stability of the genome largely hinges on effective DNA repair processes which diminish with age rather than avoiding damage altogether. This decline in repair capacity is thought to contribute to the accumulation of genomic insults and accelerated aging pathology later in life, as well as the stepwise increase in cancer with age [12,13]. Chronic inflammation and oxidative stress are interconnected pathological processes that lead to genomic damage and instability (Figure 2).

This process is most keenly characterized in the inflammation-mediated initiation and progression of cancers, where chronic inflammation has been shown to account for 20–25% of cancers [14] and influence mitogenicity [15]. However, the inflammation-induced genomic instability that potentiates the fitness of cancer cells can also result in the cellular and tissue debility characteristic of aging decline [16]. Therefore, the relationship between chronic inflammation and genomic instability is multifaceted and bidirectional.

2.1.1. Genomic instability leads to inflammation

Chronic inflammation can also be caused by genomic damage and instability by several mechanisms generating a bidirectional feedback loop. Firstly, reactive oxygen species (ROS) induced single-stranded breaks, which can be detected by the DNA-break sensor, poly (adenosine diphosphate-ribose) polymerase 1 (PARP1) [17]. Persistent PARP1

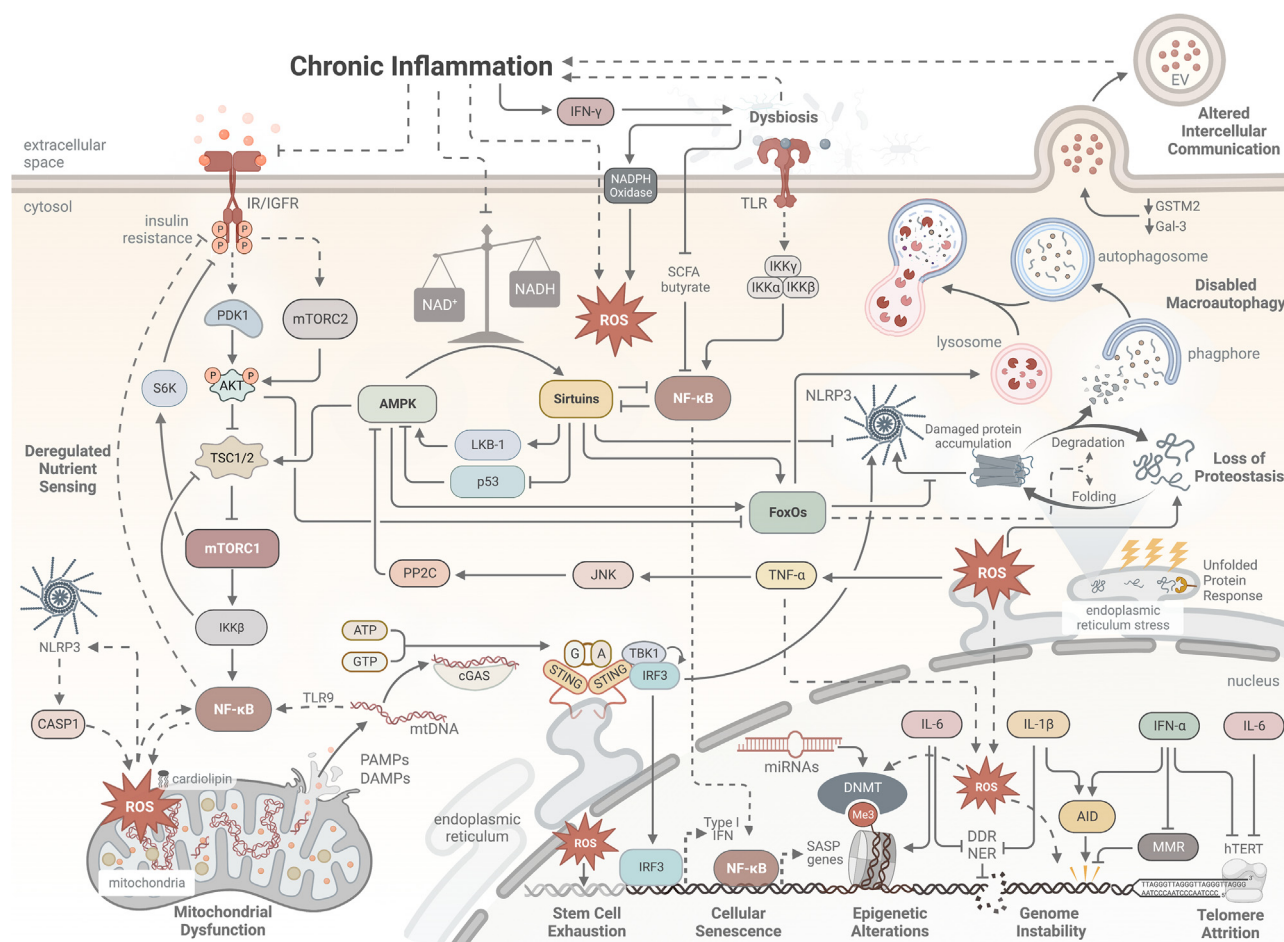


Figure 2: Proposed Mechanisms of Relationships Underlying Chronic Inflammation and the Hallmarks of Aging: Chronic inflammation interacts with other hallmarks of aging via a complex network of intra- and extracellular signalings, including but not limited to nutrient-sensing, danger-sensing, and autophagy pathways, acting as both a cause and a result of aging. A = adenine, AID = activation-induced cytidine deaminase, AKT = Protein kinase B, AMPK = 5'-adenosine monophosphate-activated protein kinase, ATP = adenosine triphosphate, CASP1 = caspase-1, cGAS = cyclic GMP-AMP synthase (cGAS), DAMPs = deoxyribonucleic acid damage-associated molecular patterns, DDR = deoxyribonucleic acid damage response, DNMT = methyltransferase, EV = extracellular vesicle, FoxO = forkhead box protein O, G = guanine, GSTM2 = Glutathione S-transferase Mu 2, GTP = guanine triphosphate, hTERT = human telomerase reverse transcriptase, IFN = interferon, IGF1R = insulin-like growth factor receptor, IKK = inhibitor of nuclear factor- κ B (I κ B) kinase, IL = interleukin, IR = insulin receptor, IRF = Interferon regulatory factors, JNK = jun n-terminal kinase, LKB-1 = Liver Kinase B1, Me3 = methyl, MMR = mismatch repair, mtDNA = mitochondrial deoxyribonucleic acid, mTORC = mechanistic target of rapamycin complex, NAD/NADH = nicotinamide adenine dinucleotide, NER = nucleotide excision repair, NF- κ B = nuclear factor kappa B, NLRP3 = nod-like receptor family pyrin domain-containing 3, PAMPs = pathogen-associated molecular pattern molecules, PDK1 = 3-phosphoinositide-dependent protein kinase-1, PP2C = Protein phosphatase 2C, ROS = reactive oxygen species, S6K = Ribosomal protein S6 kinase, SASP = senescence-associated secretory phenotype, SCFA = short chain fatty acid, STING = stimulator of interferon genes, TBK1 = tank-binding kinase 1, TLR = toll-like receptor, TNF = tumor necrosis factor, TSC = tuberous sclerosis complex.

activation, however, depletes NAD⁺, which dampens downstream sirtuin (Sirt) activities [18,19]. The resultant Sirt3 and Sirt5 depletion can mediate the accrual of mitochondrial DNA damages, leading to the pro-inflammatory cytokine profile of mitochondrial dysfunction-associated senescence [20]. Impaired functions of other Sirts, including Sirt2, also lead to higher nod-like receptor family pyrin domain-containing 3 (NLRP3) inflammasome activity, which is associated with accelerated aging [21] and altered Sirt-dependent nutrient-sensing pathways [22]. These intertwined impairments fuel accelerated aging pathology by promoting inflammation and senescence through mechanisms introduced in the following sections (Inflammation & Cellular Senescence).

DNA damage can also directly trigger inflammation through leakage of DNA into the cytoplasm, which can occur through ruptured micronuclei or DNA end resection [23]. Cytoplasmic DNA can prompt the cyclic GMP-AMP synthase (cGAS) stimulator of interferon gene (STING) pathway to activate tank-binding kinase 1 (TBK1) and interferon regulatory factor 3 (IRF3) to induce a type I interferon (IFN) response and nuclear factor kappa B (NF-κB) cascade. DNA damage response to single and double-stranded breaks also recruits ataxia telangiectasia and Rad3-related protein (ATR) and ataxia-telangiectasia mutated (ATM), which activate NF-κB through GATA4 stabilization [24,25] or directly activate tumor necrosis factor receptor-associated factor 6 (TRAF6) or STING [26,27]. Other events associated with DNA damage, including DNA repair deficiencies, activation of transposons, cellular senescence, R-loop formation, and changes in chromatin structure that link DNA damage to inflammation and aging, have been recently reviewed in detail [24,28].

Finally, if not corrected, genomic instability can lead to neoantigens in cells, which predispose cells to be targeted by the immune system and contribute to inflammation. This phenomenon is exemplified in the susceptibility of microsatellite-unstable tumors, which are mismatch repair (MMR)-deficient, to treatment with immunotherapies, such as immune checkpoint inhibitors [29].

2.1.2. Inflammation causes genomic instability

Chronic inflammation has been shown to induce genomic instability through multiple mechanisms. Among pro-inflammatory cytokines, tumor necrosis factor-α (TNF-α) is commonly elevated with age [30] and induces ROS through cell-specific and common mechanisms. Specifically, binding of TNF-α to its receptors induces downstream phosphorylation of p47phox (phox: phagocyte oxidase), which recruits TRAF4. This complex then translocates to the plasma membrane, facilitating nicotinamide adenine dinucleotide phosphate hydrogen (NADPH) oxidase activity [31,32]. Despite the effectiveness of ROS production in pathogen defense and cell signaling, ROS can directly cause damage to macromolecules within cells, and DNA is one of the most susceptible biological targets of oxidative stress due to its limited structural stability [33]. ROS-mediated deleterious effects are thought to contribute to human age-related degenerative conditions, including AD [34] and PD [35], CVD [36], pulmonary fibrosis [37], nonalcoholic fatty liver disease (NAFLD) [38], and reproductive decline [39,40]. A more comprehensive review of the impact of inflammation-induced ROS and reactive nitrogen species (RNS) on DNA base damage, as well as strand breaks, has been published recently [41].

Chronic inflammation also downregulates MMR proteins, such as MutL homolog 1 (MLH1), through various mechanisms attributable to TNF-α, interleukin (IL)-1β, and Prostaglandin E1 (PGE1) signaling (Figure 2) [42]. Mutations or epigenetic silencing of MMR members is associated with increased genetic instability, termed microsatellite instability (MSI)

which accentuates the accumulation of DNA replication errors throughout the genome [43]. Additionally, IL-6, a marker of chronic inflammation, appears to negatively impact the nucleotide excision repair pathway, which is responsible for repairing a wide variety of DNA lesions caused by UV radiation, mutagenic chemicals, and chemotherapeutic agents [44]. The underlying mechanisms, however, remain unclear.

Furthermore, activation-induced cytidine deaminase (AID) is a DNA mutator enzyme induced by inflammation [45]. Evolutionarily, AID functions in activated B cells to generate immune diversity by inducing somatic hypermutation and class-switch recombination in immunoglobulin genes [46]. Under chronic inflammation, however, AID can be aberrantly expressed in epithelial cells and induce somatic mutations and chromosomal aberrations [46]. The expression of AID is induced by age-associated inflammatory cytokines, including NF-κB, IL-4, and TGF-β (transforming growth factor-beta) [47,48] and overexpressed in several inflammatory-mediated cancers and non-cancer-related inflammatory diseases such as chronic gastritis [49], ulcerative colitis [50], and pancreatitis [51].

Finally, inducible nitric oxide synthase (NOS2), which produces NO, and whose expression is associated with inflammation, was recently shown to reduce DNA methyltransferase 1 (DNMT1), resulting in long interspersed nuclear element 1 (LINE1) retrotransposon hypomethylation, expression, and DNA damage [52]. In addition, LINE1 induction can drive type I IFN secretion in senescent murine cells, thereby contributing to the formation of a cycle where inflammation further facilitates DNA damage, maintenance of senescence secretory products and inflammaging [53]. The interplay between epigenetics, inflammation and DNA damage is discussed further in subsequent sections (Inflammation & Epigenetic Changes).

2.2. Telomere attrition

Somatic cells, including B cells and T cells, have a finite capacity to divide due to the inability of DNA polymerase to completely synthesize the distal ends, or telomeres, of eukaryotic DNA [54]. Thus, after a certain number of cell cycles known as the “Hayflick Limit” [55], telomere length decreases to a critical length, after which the cell ceases to replicate and instead diverts to cellular senescence or apoptosis through DNA damage response (DDR) signaling [56]. In humans, the lack of somatic expression of telomerase, the enzyme necessary for the maintenance of telomere length, in most cells accelerates the progression of telomere erosion, which has been shown to contribute to a pro-inflammatory milieu, the development of chronic diseases, and shorter life expectancy [57]. Evidence suggests that age-associated inflammation also worsens telomeric loss [58,59]. Shortened telomeres or altered telomere structures can ultimately lead to apoptosis [60], cellular senescence [61], and a state of extensive genomic instability known as “telomere crisis,” linking telomere attrition to other hallmarks of aging [62,63]. This section will briefly discuss links between telomere attrition and chronic inflammation in aging.

2.2.1. Telomere attrition promotes inflammation

Inflammation can be triggered and maintained on several levels by telomere dysfunction. For example, the telomere-mitochondria axis has recently been identified as a novel contributor to inflammation [64]. In addition, during aging, DNA damage and telomere dysfunction activate p53, which suppresses peroxisome proliferator-activated receptor gamma coactivator 1 alpha (PGC-1α) and mitochondrial Sirts (Sirt3/4/5) [65,66], thus impairing mitochondrial function and leading to overproduction of ROS [67]. The resultant oxidative stress promotes

inflammation and further damages telomere structures [68]. Additionally, in macrophages from late-generation telomerase-deficient mice, downregulation of PGC-1 α induces activation of the NLRP3 inflammasome, resulting in an amplified inflammatory response upon bacterial challenge [69]. Increased caspase-1 from NLRP3 exacerbates inflammation by facilitating the conversion of pro-IL-18 to mature IL-18, which is sourced from telomere attrition-induced ATM/cABL activation and yes-associated protein 1 (YAP1) nuclear translocation [62]. Furthermore, telomere dysfunction resulting from shelterin loss also triggers autophagic cell death by activating the cGAS-STING pathway [70], which can initiate an inflammatory cascade via IRF3-TBK1-mediated upregulation of Type I interferons, leading to enhanced cross-priming activity and recruitment of IFN γ -producing CD8⁺ T cells [71].

Aging is also associated with telomere shortening in T cells [72], which is associated with a loss of naive T cell pool, lower T cell receptor diversity, and accumulation of antigen-experienced and even senescent T cells exhibiting a pro-inflammatory profile, such as increased IL-6 and TNF- α [73–75]. Overall, these changes lead to impaired functional responsiveness of the aged immune systems to emerging antigens, such as SARS-CoV-2 [76,77]. Interestingly, some CD4⁺ (especially naive and central memory) T cells can elongate their telomeres by acquiring telomere-containing vesicles from antigen-presenting cells (APCs), thus gaining protection from immunosenescence and conferring long-lasting immune response [78]. However, it remains to be studied if the vesicle-mediated telomere acquisition can be harnessed for its therapeutic potential to ameliorate systemic inflammaging.

2.2.2. Inflammation drives telomere attrition

In multiple pathological conditions of aging, inflammation can also contribute to telomere attrition. On the tissue level, studies have demonstrated a direct link between local inflammation and telomere attrition, including gastritis [79], NAFLD [80], and chronic obstructive pulmonary disease (COPD) [81]. Chronic inflammation markers, such as TNF- α and IL-6, are associated with shortened telomeres in non-cancerous cells [82], though this association in malignant cells is less understood [83]. TNF- α , specifically, has been shown to exhibit a causal role in downregulating telomerase activity through downstream phosphorylation of ATF7 by p38, resulting in accelerated telomere shortening associated with aging in mice [84]. The oxidative stress resulting from age-associated mitochondrial dysfunction and ROS production can also accelerate telomere shortening by generating irreparable single-strand breaks in telomere regions [85,86], which in itself can incite more inflammation. Similarly, ROS produced by neutrophils when fighting pathogens facilitates the spreading of telomere attrition and senescence to neighboring non-immune cells through paracrine signaling [58]. In addition, age-associated inflammation depletes cellular NAD⁺ levels, thus limiting the availability of Sirt6 [87–89]. Reduced NAD⁺ levels may downregulate Sirt6 activity, lowering telomere stability and increasing attrition [65,66,90]; however, further research is needed to further characterize this mechanism as direct or indirect.

Type I IFNs, pro-inflammatory cytokines secreted during viral infection, are drivers for T cell senescence [91] and have also been shown to inhibit telomerase activity and promote telomere erosion in stimulated CD8⁺ T cells [92] and CD4⁺ T cells [93]. Specifically, IFN- α is suggested to inhibit human telomerase reverse transcriptase (hTERT) activity by decreasing activation of NF- κ B that can activate hTERT, despite its pro-inflammatory potential [92] (Figure 2). Persistent activation of NF- κ B, however, contributes to telomere shortening in certain

cell types, including muscle stem cells (MuSCs) and hepatocytes [94,95].

2.3. Epigenetic alterations

The interconnection between epigenetics and inflammaging is an important area in contemporary aging research. Age-associated epigenetic modulations are increasingly shown to favor the formation of a pro-inflammatory environment, which can, in turn, remodel epigenetic patterns [96]. Changes in DNA methylation and histone modifications during aging can lead to inflammatory consequences, while age-dependent alterations in non-coding RNA (ncRNA) and retrotransposons (RTPs) [97] can regulate genes involved in inflammaging, exhibiting potentials for novel therapeutic targets for ARDs [98].

2.3.1. DNA methylation

Aging has been associated with abnormal patterns of DNA hypo- and hypermethylation at promoter CpG islands [96,99]. The DNA hypomethylation during aging can be partially attributable to the decreased activity of DNMTs, such as DNMT-1 and DNMT-3B [100] (Figure 2). During aging, loss of DNA methylation usually activates genes involved in inflammatory responses, such as NF- κ B signaling, macrophage activation, and IFN signaling [101]. Age-associated DNA hypomethylation, caused by tissue necrosis and incomplete clearance of apoptotic cells, also impacts cell-free DNA (cfDNA) [102]. The unmethylated cfDNA resembles microbial DNA, as sensed by DNA-sensing receptors, such as cGAS and toll-like receptor (TLR)-9, leading to inflammatory responses [103].

However, persistent inflammation can also contribute to aberrant DNA methylation. Recently, a large multi-ethnic epigenome-wide association study suggests that the altered CpG methylation patterns can result from elevated C-reactive protein (CRP) during chronic inflammation. This inflammatory methylation signature can significantly increase the risk for cardiometabolic diseases and COPD [104]. At the molecular level, oxidative stress triggered by chronic inflammation can relocate DNMTs to guanine and cytosine-rich regions, causing DNA methylation in potentially undesirable genes [105]. For example, DNA hypermethylation at the promoter regions of autophagy-related 5 (ATG5) or microtubule-associated protein 1A/1B-light chain 3 (LC3-B) can downregulate autophagy in aging murine macrophages [106]. Inflammaging can also alter epigenetic patterns in several chronic diseases, including neurodegeneration [107], ulcerative colitis [108], *Helicobacter pylori* infection [109], and hepatitis [110].

2.3.2. Histone modification

The N terminal tails of the four core histones (H2A, H2B, H3, and H4) are subjected to post-translational modifications, including acetylation, methylation, and phosphorylation, and depending on histones residues and modification types, gene transcription can be activated or repressed [100]. The activated, accessible euchromatin is associated with increased acetylation and trimethylation of histone H3 lysine (H3K)-4, H3K-36, and H3K-79, whereas transcriptionally inactive heterochromatin has low acetylation and increased methylation of H3K-9, H3K-27, and H4K-20 [111]. Specifically, the global increase of H3K-27me3 in multiple tissues of various organisms has been newly identified as a common epigenetic signature of aging [112]. Generally, inflammatory genes are activated and repressed by histone acetyltransferases (HATs) and histone deacetylases (HDACs), respectively [113]. Correspondingly, cell senescence can be delayed via the inhibition of HATs and induced by inhibiting HDACs [114–116].

During aging and cell senescence, profound chromatin rearrangement can occur and cause global heterochromatin loss [96,117]. Compromised chromatin architecture at specific domains, including centromere, telomeres, and retrotransposons, results in chromatin relaxation, reactivation of deleterious genes, and genomic instability that fuel a pro-inflammatory environment [118,119]. For example, Sirt6 shows HDAC activity at H3K-9 and H3K-56, while Sirt6-deficient mice exhibited not only genomic instability and shortened lifespan but also accumulation of LINE1 cDNA, which triggers type I interferon response via cGAS-STING [90,120]. Aging is also associated with a global reduction in histone trimethylation at H3K-9me3, which is implicated in suppressing the reactivation of human endogenous retrovirus (HERV) [121,122]. Indeed, human endogenous retrovirus-K (HERV-K) is found to increase in patients with AD and cause subsequent neurodegeneration and microglial accumulation through TLR-8 activation [123,124]. Notably, HERV-K can also drive cell senescence through cGAS-STING activation [125], further emphasizing the causal role of DNA hypomethylation in inflammaging.

2.3.3. microRNAs (miRNAs)/non-coding RNA

Micro-ribonucleic acids (miRNAs) are small, non-coding RNAs that bind to the 3' untranslated region of messenger RNA (mRNA) and regulate gene expression [126]. miRNAs can act as either repressors or activators of gene expression, depending on the target mRNA and the miRNA's corresponding seed sequence [126]. By doing so, miRNAs regulate aging by altering the expression of genes involved in the aging process, such as those involved in DNA damage repair, cell cycle control, and apoptosis. miRNAs can also regulate inflammation by targeting cytokines and other molecules involved in inflammatory pathways. This miRNA subset has been coined "inflammamiRs" owing to their ability to influence NF- κ B/NLRP3 and IL-6 inflammatory pathways (particularly miR-21-5p and miR-146a-5p) [127]. In addition, miRNAs can interact with epigenetic machinery, such as DNA methylation and histone modification, to modulate gene expression and aging-related processes. For example, miR-17 mediates DNMT-1 downregulation in neurotoxin-induced PD and leads to an aberrant DNA methylation pattern in PD [128]. Therefore, miRNA may serve as a promising therapeutic target for ARDs, including neurodegenerative disease [129], CVD [130], degenerative musculoskeletal disease [131], metabolic disease [132], and ovarian aging [133].

2.4. Loss of proteostasis

Protein homeostasis, or "proteostasis," is a network of intra- and extracellular interactions that ensure the proper translation and folding of newly synthesized proteins, as well as the refolding and degradation of misfolded proteins [134]. Derangement of proteostasis, such as by proteasomal deregulation [135], is a primary manifestation of aging, underlying several ARDs [134]. A growing body of evidence indicates that immune reactions are induced by proteostatic stress, and excessive inflammation may contribute to the age-related decline in proteostasis [136], indicating that proteostasis and inflammation are mutually influenced [137]. With aging, both aspects of proteostasis—folding and degradation—are reduced, resulting in an accumulation of damaged proteins and organelles, termed "garb-aging," which can exacerbate inflammaging (Figure 2) [138].

2.4.1. Reduced proteostasis promotes inflammation

During aging, the robustness of the proteostasis network is challenged by continuous exposure to mutagens that cause misfolded proteins and damaged organelles, such as mitochondria [139]. The perturbed proteostasis can promote inflammation as misfolded proteins form proteotoxic aggregates, which initiate excessive unfolded protein response

(UPR) through activation of the inositol-requiring transmembrane kinase/endoribonuclease 1 α (IRE1 α), the activating transcription factor 6 (ATF6), and the protein kinase R-like endoplasmic reticulum kinase (PERK), eventually leading to NF- κ B activation [140,141]. The dysregulation of UPR in immune cells thus contributes to multiple pathologies, including autoimmunity and metabolic disorders [142]. Proteotoxic aggregates also can be recognized, as damage-associated molecular patterns (DAMPs), by pattern-recognition receptors (PRRs) like TLRs and nod-like receptors (NLRs), leading to the assembly of the NLRP3 inflammasome, increased IL-18 and IL-1 β secretion by caspase-1 cleavage [143], and eventually pyroptosis-mediated cell death [144]. These deleterious effects can be amplified in postmitotic cells, such as neurons and cardiomyocytes, which are not actively dividing to dilute cellular debris into daughter cells, increasing inflammaging [145]. Proteostasis depends on the proper functioning of proteolytic systems, namely, the ubiquitin-proteolytic system (UPS) and the autophagy-lysosome system (ALS) [134]. However, as cells age, damaged cellular content accumulates and overwhelms the cellular proteolytic systems, contributing to diverse types of tissue inflammation, such as the neuroinflammation in PD [146] and senescence-associated secretory phenotype (SASP)-induced inflammation [147]. Reduced proteasome function also can potentially lead to proteasome-associated auto-inflammatory syndrome (PRAAS), characterized by prominent type I IFN gene signatures [148]. In mice with T cell-specific knockout of Rpn13, a ubiquitin receptor critical for proteasome induction, there is a marked increase in programmed cell death protein 1 (PD-1)⁺CD44^{high}CD4⁺ T cell frequency upon TCR stimulation, accompanied by profound production of pro-inflammatory cytokines and chemokines [149]. Besides UPS dysfunction, impaired ALS function, as seen in age-dependent downregulation of autophagy genes [106,150], can also contribute to inflammaging (discussed in depth in Disabled Macroautophagy).

2.4.2. Inflammation impairs proteostasis

Systemic and chronic inflammation during aging can deregulate proteostasis via multiple mechanisms, undermining both protein folding and degradation. The inflammation-induced ER stress can activate UPR, thus altering the protein synthesis/folding arm of proteostasis by 1) PERK-dependent phosphorylation of eukaryotic translation initiation factor 2 (eIF2 α) to hinder translation of misfolded protein; and 2) decreasing influx of translated proteins into the endoplasmic reticulum (ER) by degrading ER membrane-associated mRNAs with regulated IRE1 α -dependent decay (RIDD) [151]. Inflammation in itself can lead to increased levels of ROS and oxidative stress [137,152], which impair protein folding by exposing hydrophobic cores that facilitate the formation of misfolded, cytotoxic, and degradation-resistant protein aggregates [153,154]. In addition, "oxi-inflamm-aging," a newly coined aging pathology [155], describes that oxidative stress elicited by inflammation promotes the accumulation of a variety of proteotoxic cellular products, including lipofuscins, advanced glycation end-products (AGEs), Tau protein aggregates, α -synuclein fibrils, and β -amyloid, all contributing to deregulated proteostasis [138]. Specifically, lipofuscin, a type of highly oxidized and cross-linked protein or lipid aggregate, contributes to the failure of lysosomal enzymes and inefficient clearance of autophagosomes [134], leading to enhanced inflammation, ROS production, and even senescence in postmitotic cells, such as cardiomyocytes [156].

In the presence of oxidative stress or chronic inflammation, TNF- α and IFN- γ can promote the conversion of normal proteasome into an inducible isoform, or "immunoproteasome" (IP) [157]. IP is constitutively expressed in hematopoietic cells and participates in antigen processing for the presentation by major histocompatibility complex I

(MHC I) molecules. It can degrade oxidized proteins more efficiently, thus playing a critical role in cytokine signaling and proteostasis [148]. However, the IP can also promote pro-inflammatory T helper 1 (Th1) and T helper 17 (Th17) cells and suppress the Tregs cells [157], while malfunctioning of the IP is involved in the development of autoimmune diseases [158,159] and accelerated aging [160,161], indicating that inflammation and derailed proteostasis is interdependently contributing to the pathogenesis of aging.

2.5. Disabled macroautophagy

Due to its increasingly broad importance in aging, macroautophagy has emerged as its own hallmark of aging, despite its strong connection to proteostasis [6]. With autophagosome-lysosome fusion, macroautophagy processes not only protein cargos but also non-protein macromolecules (including lipid vesicles and glycogen) and organelles (such as “mitophagy” for mitochondria, “ribophagy” for ribosomes) [162], thus controlling cytoplasmic quality, cellular metabolism, as well as innate and adaptive immunity [163]. However, a general decline in autophagy function, which has different subtypes such as macroautophagy, microautophagy, and chaperone-mediated autophagy (CMA), is observed in older individuals to accelerate aging by promoting chronic inflammation [164]. Therefore, this section focuses on how diverse crosstalk, beyond the scope of macroautophagy, between chronic inflammation and autophagy can contribute to inflammaging (Figure 2).

2.5.1. Autophagy removes sources of chronic inflammation

The reduction of autophagic flux can contribute to poor cytosolic quality through the accumulation of protein aggregates, cytosolic DNA, dysfunctional organelles, and reduced elimination of pathogens — all of which contribute to inflammation signaling [165]. Thus, a failure of adequate autophagy often manifests as dysregulated inflammation in animal models and human diseases [166] through the accumulation of cytosolic protein aggregates and condensates [167], which act as DAMPs [168]. Given that microbes, damaged organelles, and organic or inorganic crystals are all sources of inflammatory signals (PAMPs and DAMPs), the cytoplasmic clean-up function of autophagy is typically anti-inflammatory in cells capable of activating cell-autonomous inflammatory responses [166]. For instance, CMA is recently found to assist in removing palmitoylated NLRP3, thus preventing sustained inflammation in human and mouse macrophage models [169]. The age-related decline in CMA function is associated with increased NLRP3 activation and IL-1 β secretion, leading to vascular inflammation and accelerated atherosclerosis progression [170].

A declining capacity to remove damaged organelles also contributes to inflammaging. For instance, “MitophAging” refers to the age-related pathogenesis resulting from the loss of mitophagy [171]. Aging human CD4⁺ T cells are found to be burdened with declining mitophagy, shown as accumulating autophagosomes consisting of undigested and damaged mitochondria, which stimulate ROS generation and the NF- κ B pathway, triggering chronic inflammation, immunosenescence, and dysfunctional adaptive immunity in older individuals [172]. As well, immunosenescence can be ameliorated by autophagy during aging. In old human B cells, spermidine-induced synthesis of the autophagosomal and lysosomal master regulator, transcription factor EB (TFEB), leads to enhanced autophagy and reversed B cell senescence [173]. However, although autophagy plays a crucial role in removing senescent cells [174], it is worth noting that autophagy may also provide building blocks for SASP during some forms of senescence, thus complicating the relationship between autophagy and age-related senescence [175].

Notably, caloric restriction-mimetic therapies, such as metformin [176], rilmenidine [177], rapamycin, and “rapalogs” [178], as well as senolytic drugs [179,180], are proposed to exert anti-inflammatory effect by activating autophagic processes, including mitophagy and CMA, thus promoting the proper disposal of damaged cellular components, which can otherwise trigger a cascade of inflammatory pathways [138]. The mechanistic target of rapamycin 1 (mTORC1), located on lysosomes, is of particular interest as a modulator linking metabolism, autophagy, inflammation, and aging biology (see the section on *Deregulated nutrient sensing*).

2.5.2. Chronic inflammation impairs autophagy

Chronic inflammation is also understood to impair autophagy. This effect is often seen in neurodegenerative disorders such as PD [181], AD [182], Huntington’s Disease [183], and Amyotrophic Lateral Sclerosis [184]. The role of neuroinflammation in impairing efficient autophagy was demonstrated as lipopolysaccharide (LPS)-induced inflammatory stress substantially increased cytokine production (IL-1 β , TNF- α , and IL-6) and decreased the levels of autophagy markers (Beclin-1, p62, and LC3 II) [185]. Similarly, TNF- α can inhibit microglial autophagy via the mTOR pathway, while enhanced autophagy can promote microglial M2 polarization, promoting the resolution of inflammation in PD [186]. Recent studies also suggest that microglia-induced neuroinflammation can promote the accumulation and transmission of α -syn and Tau proteins to aggravate PD and AD, respectively [187]. Specifically, constitutive microglial activation, for example, by LPS, can increase α -syn accumulation and transmission to the substantia nigra and striatum, leading to increased autophagy burden and worsened PD [188–190]. In a murine AD model, constitutive activation of microglial NF- κ B exacerbates Tau seeding and spreading in young PS19 mice, while NF- κ B inactivation rescues microglial autophagy [191]. Taken together, chronic inflammation impacts autophagy and may be particularly relevant in the proteinopathies conferred by aging.

3. ANTAGONISTIC HALLMARKS

The antagonistic hallmarks are a response to damages caused by the primary hallmarks. Initially, the response helps mitigate the damage and has benefits. However, incidentally or with prolonged pathway stimulation, the mechanisms become harmful, contributing to the aging process.

3.1. Deregulated nutrient sensing

Nutrient sensing is essential in regulating the aging process. Current evidence suggests prolonged overnutrition accelerates aging, while caloric restriction favors longevity [192]. In addition, nutrient sensing can be deregulated under pathogenic conditions, including multiple metabolic illnesses associated with chronic, low-grade inflammation, such as T2D and atherosclerosis, common chronic diseases of age. Classical pathways that regulate nutrient sensing include the insulin and the insulin-like growth factor 1 (IGF-1) signaling pathways (collectively known as the IIS pathway), the mTOR pathway, the AMPK pathway, and the Sirts. This section will focus on the bidirectional relationships between the deregulation of these pathways and the chronic inflammation that compounds the aging process and related pathology (Figure 2).

3.1.1. IIS deregulation induces inflammation

During aging, the IIS pathway can be deregulated and mediate chronic inflammation. Aging is usually associated with declined insulin

clearance, leading to hyperinsulinemia and insulin resistance (IR) in peripheral metabolic tissues [193,194]. IR also induces dyslipidemia, represented as high plasma levels of triglycerides and free fatty acids (FFAs) [195]. Triglycerides and FFAs can trigger macrophage inflammatory responses by inducing NF- κ B, culminating in a pro-inflammatory environment that worsens IR [196]. Chronic hyperinsulinemia and elevated FFA also support chronic mTOR activation [197], a major driver of glycolysis in immune cells [198]. Increased glycolysis in innate immune cells can facilitate M1-like macrophage polarization with cytokines linked to inflammaging, like TNF, IL-1, and IL-6 [199]. In addition, insulin inhibits forkhead box protein O1 (FoxO1) activity, while FoxO1 deficiency impairs proteostasis in aged T cells and causes pro-inflammatory, senescence-like phenotypes [200].

Insulin itself can also directly act on the immune system to fuel inflammaging. For instance, insulin receptor signaling has been shown to acutely promote murine immune cell functions, including T cell effector responses, through supporting nutrient uptake and glycolytic capacities, key metabolic necessities of inflammation [201,202]. In addition, mice deficient in insulin receptors in T regulatory (Tregs) cells are protected from age-induced glucose intolerance and adipose tissue inflammation, highlighting an inflammatory role for insulin in altering Treg responses [203]. Interestingly, age-associated B cells (ABCs), a T-bet + memory-like B cell subset linked with increased autoimmune diseases and aging in mice [204,205], can promote inflammatory T cells potentially through increased glycolysis [206,207], though it remains unknown if insulin/IGFs is a necessary input. Thus, the immune homeostasis supported by proper IIS signaling is likely essential, especially in the elderly, who have a higher susceptibility to infectious diseases and lower capacity to generate adaptive immune responses against pathogens, such as SARS-CoV-2 and vaccination [2,208]. Altogether, IIS deregulation may form a vicious loop that fuels both inflammatory processes and potential changes linked to immunosenescence, exacerbating ARDs.

3.1.2. mTOR pathway deregulation induces inflammation

The mTOR kinase is essential in maintaining metabolic homeostasis by sensing amino acid levels and modulating insulin signaling [209]. mTOR in mammals exists within the mTOR complex 1 (mTORC1) and 2 (mTORC2). mTORC1 responds to IIS signaling through Akt (protein kinase B), senses amino acid abundance, and controls cell growth and senescence [210]. At the same time, mTORC2 can be activated through insulin-like growth factor receptor (IGFR)- and insulin receptor substrate (IRS)-mediated signaling, controlling metabolism and actin organization [211]. When triggered by prolonged IIS signaling, mTORC1 can be hyperactivated, while several ARDs, including T2D, have mTORC1 hyperactivation as a risk factor [212,213], which fuels inflammation and accelerates aging progression [214]. The chronic, low-grade inflammation induced by mTORC1 hyperfunction and its resulting inhibitor of nuclear factor- κ B (I κ B) kinase β (IKK β) overactivation (Figure 2) [215,216] is associated with decreased lifespan [217]. Reducing mTORC1 signaling and its associated tissue inflammation can also benefit tissue healthspan in aging mice [218], whereas mice with constitutive mTORC1 overactivation showed accelerated aging that can be rescued by rapamycin treatment [219]. The mTOR signaling also impacts aging progression by modulating inflammation through cellular senescence and immune cell responses [209,220]. mTORC1 inhibition with rapamycin can blunt the production of pro-inflammatory and pro-oxidant products from senescent cells exhibiting SASP [221,222]. In murine macrophages, reduction of mTORC1 through Raptor (regulatory-associated protein of TOR) knockdown decreases inflammatory gene expression, and overexpression of mTORC1

signaling through knockdown of mTORC1 inhibitor, TSC1 (Tuberous Sclerosis 1), leads to elevated glycolysis and shift to pro-inflammatory M1-like macrophages [223,224]. Intriguingly, in healthy aging individuals, mTORC1 blockade exhibited enhanced antiviral immunity toward influenza infection [225]. This data is especially significant under the ongoing pandemic of coronavirus disease 2019 (COVID-19), which in itself increases mTORC1 activity [226]. Thus, manipulation of mTOR signaling may provide a novel intervention to enhance the adaptive immunity in aging individuals and reduce mortality from viral infections, thus providing a potential axis to improve human longevity [227].

3.1.3. AMPK-sirtuin pathway deregulation induces inflammation

By acting in the opposite direction to the IIS and the mTOR pathways, the AMPK-sirtuin pathway signals nutrient scarcity and catabolism. The AMPK pathway is activated by an increased AMP:ATP ratio during starvation or caloric restriction [228], while sirtuins are a family of (NAD⁺)-dependent deacetylases [229]. Mechanistically, AMPK activation can increase the cellular NAD⁺ concentration, leading to the expression of Sirt1 that deacetylates the liver kinase B1 (LKB-1), an upstream activator of AMPK [230], forming a positive feedback loop (Figure 2). Functionally, AMPK activation can inhibit mTORC1 by activating TSC1/2 and stimulate Sirts to deacetylate FoxOs or autophagy-related proteins (ATG5 and ATG7) [231,232], leading to 1) enhanced expression of PGC-1 α for better mitochondrial biogenesis [233] and 2) inhibition of cytoplasmic p53 for autophagy [234,235]. Recently, AMPK is also found to directly phosphorylate folliculin-interacting protein 1 (FNIP1), which forms a complex with folliculin to suppress the complex's function; through this interaction, AMPK is able to promote TFEB-mediated autophagy and induce lysosomal and mitochondrial biogenesis [236]. Of note, metformin, an AMPK activator, normalizes mitochondrial function and alleviates age- and senescence-associated inflammation by enhancing T cell autophagy [237]. Therefore, the AMPK-Sirt pathway plays an important role in regulating chronic inflammation.

As seen in the decreased NAD⁺ level and the associated metabolic dysfunction during aging, the declining AMPK-Sirt responsiveness with age is gaining increasing attention for its proinflammatory potential [238–240]. Sustained AMPK activation is essential for the survival and functions of both human and murine *in vitro* induced Treg cells [241]. In line with Tregs' function in inhibiting chronic inflammation induced by self-antigens in aged persons [242], mice with Treg-specific AMPK deletion developed autoimmune and inflammatory liver diseases [243]. In murine macrophages, AMPK α 1 deletion induces M1 hyperpolarization upon LPS challenge, whereas metformin increases IL-10 and reduces IL-6 and IL-12 secretion in wild-type M1 macrophages [244]. Loss of AMPK in other tissues also causes inflammation and age-associated pathologies, such as NAFLD [245] and chronic kidney disease [246]. Similarly, loss of Sirt6 in the aging mouse brain leads to decreased mitochondrial function and increased ROS production that potentiates inflammaging [247]. In contrast, an intact AMPK-Sirt pathway can suppress NLRP3 (Figure 2), thus quenching chronic inflammation and contributing to a healthier aging process [248–250]. In murine hematopoietic stem cells (HSCs) with age-related defects in autophagy, Sirt3 can restore mitophagy, reduce oxidative stress, and slow aging progression [251]. This evidence indicates that the AMPK-Sirt pathway, if deregulated during aging, can cause profound disruption in energy homeostasis and aberrant inflammatory responses.

3.1.4. Inflammation deregulates the IIS pathway

Local and systemic inflammation accompanying aging and ARDs can also deregulate nutrient-sensing pathways [252]. Aging deregulates

the IIS pathway by promoting a pro-inflammatory environment in metabolic tissues, such as adipose tissues [253,254], muscles [255], and digestive tracts [256,257]. This chronic, low-grade metabolic inflammation is characterized by increased FFAs, ROS, and UPR linked to ER stress, which can activate the JNK pathway, mediating the serine phosphorylation of IRS1/2 and thus blockade the IIS pathway [258]. Likewise, older mice express higher levels of inflammatory markers, such as TNF- α and IL-1 β in skeletal muscles [259], leading to activated IKK β /NF- κ B pathway that phosphorylates IRS1 and contributes to IR [260] (Figure 2). Notably, the reduction of inflammation is implicated in the extension of life/health span observed under caloric restriction. For example, methionine restriction (MR) has been found to reduce basal levels of serum IGF1 and glucose, associated with downregulation of multiple pathways with pro-inflammatory potential, such as TNF signaling via NF- κ B, IL-6, and JAK/STAT3 signaling in *Lmna*^{G609G/G609G} mice, a mouse model of Hutchinson–Gilford progeria syndrome (HGPS) [261]. Similarly, inhibiting methionine catabolism by the pluripotency factor NANOG in *Lmna*^{G609G/G609G} mice restores insulin sensitivity, glucose uptake and glycolysis, and significantly enhances the force-generating ability of aged mice [262].

In digestive tracts, especially the gut, recent studies indicate that microbial dysbiosis can increase with aging in the elderly, where pro-inflammatory commensal bacteria are enriched at the expense of beneficial microbes [263,264]. A pro-inflammatory gut environment due to dysbiosis, such as with aging, can worsen IR in mice by increasing gut permeability, facilitating bacterial product leakage into circulation, which contributes to chronic inflammation in tissues, including in the intestine [265,266]. Chronic inflammation, in turn, interferes with the IIS signaling, propelling age-related metabolic diseases [257,267]. A mucosal source of inflammation may also be related to altered IgA responses with age, as IgA is one link governing inflammation associated with the impairment of nutrient-sensing pathways [268,269].

Remarkably, the insulin receptor can also function through multiple non-canonical pathways, such as acting as a transcription factor by binding to gene promoters [270], controlling senescence through a ligand and tyrosine kinase-independent (LYK-I) pathway [271], and modulating hepatocyte and adipocyte functions by forming dynamic clusters on the cell surface [272]. Since all these pathways are interrupted in insulin-resistant cells or by ROS accumulation [272], future work is needed to tease out precise mechanistic connections between their deregulation and metabolic inflammation in aging populations.

3.1.5. Inflammation deregulates the mTOR pathway

Characterized by increased levels of pro-inflammatory cytokines, particularly IL-1 β , IL-6, and TNF- α [273], inflammaging also feeds into the deregulation of the mTOR pathway. Pro-inflammatory cytokines resulting from age-induced ER stress can interact with mTORC1, perturbing its normal function and expediting aging. For example, IKK β , a downstream effector of the TNF- α pathway activated during aging [274], has been shown to phosphorylate and inhibit TSC1 directly, constitutively activating mTORC1 [275]. Consequently, mTORC1/S6K hyperactivation promotes IRS1 degradation and generates a feedback inhibition of the phosphatidylinositol-3-kinase (PI3K)/Akt pathway, leading to peripheral IR (Figure 2) [276,277]. In addition, viral infection and chronic inflammation during aging can activate nucleic acid sensors, such as cGAS, retinoic acid-inducible gene I (RIG-I), and anti-melanoma differentiation-associated gene 5 (MDA5) [278–280], whose downstream target, TBK1, has been shown to activate mTORC1 [281] directly and activate mTORC2 signaling indirectly [282].

The inflammaging-induced deregulation of mTOR signaling is also observed in aged murine peritoneal macrophages (PM). With metabolomics and proteomic profiling, a recent study found a significant upregulation of pro-inflammatory pathways, such as integrin signaling, mTOR, NF- κ B, NO and ROS production, p38 MAPK and TNFR1/2 signaling in old M2a-PM [283]. These changes are accompanied by a downregulation of pathways for inflammatory function of M1 macrophages, such as glycolysis, nuclear factor erythroid 2-related factor 2 (NRF2), and hypoxia-inducible factor 1- α (HIF-1 α) signaling [283], indicating that the dichotomy of pro-inflammatory (M1-like) and pro-resolving (M2-like) macrophages during inflammaging can be metabolically obscure.

Interestingly, sestrins (SESNs), a family of stress-inducible proteins, are emerging as metabolic regulators that inhibit inflammaging mainly through activating AMPK and inhibiting mTORC1 [284,285]. For example, SESN1/2 inhibits liver mTORC1 signaling in a leucine-sensitive manner, thus ameliorating age-induced reduction in ketogenesis [286,287]. However, SESNs themselves also can be a target reduced during inflammation, leading to mTOR hyperactivation [288]. For example, the induction of SESN2 has been shown to require the PI3K/Akt pathway, which is frequently obstructed during inflammaging and obesity due to IRS1 serine phosphorylation [289]. In support of this, SESNs levels are lower in healthy aging men than younger men [290], while SESN1-SESN2-knockout mice showed hyperactivation of mTORC1 in aged knockout mice [291]. However, how inflammation directly alters SESNs expression warrants further research. These findings indicate that inflammation can exert a potent impact on aging physiology through cytokine-mTOR interaction and crosstalk with other components in age-associated stress responses.

3.1.6. Inflammation deregulates AMPK-sirtuin pathway

Although the AMPK pathway integrates the IIS, mTOR, and Sirt pathways, acting as a master regulator of metabolism and inflammation, inflammaging can also reversely impose a profound impact on the AMPK-Sirt pathway by preventing it from maintaining proper energy homeostasis. The activation capacity of this pathway declines under age-induced ER stress, oxidative stress, and chronic inflammation [238,292,293], while high glucose/lipids/amino acid levels, LPS, and pro-inflammatory cytokines, such as TNF- α and NF- κ B, can also inhibit AMPK activity [294–296]. Particularly, LPS can mediate inflammation by dose-dependently facilitating AMPK dephosphorylation [297]. Intriguingly, AMPK exhibits viral-sensing capability by directly phosphorylating TBK1 and promoting type I interferon gene expression in response to viral nucleic acids detected by the cGAS-STING pathway [279,298]. The genetic ablation of AMPK reduced Sendai virus sensing and damage-induced DNA sensing, underlying a compromised innate immunosurveillance [298]. Thus, during aging, which may be linked to increased circulating bacterial and viral products, an inflammatory environment can distort AMPK signaling, reducing the beneficial effects of AMPK-dependent autophagy, mitochondrial biogenesis, and antiviral immune responses, the reduction of which are all linked to frailty and chronic disease in the elderly.

Moreover, age-associated metabolic inflammation can deplete NAD⁺, leading to age-dependent loss of Sirt activity [88,89,299]. Meanwhile, reduced Sirt1 reduces mitochondrial biogenesis, reduced mitochondrial Sirt3 reduces mitochondrial antioxidant and repair systems, and reduced NAD⁺ may also impair Sirt2, enhancing NLRP3 inflammatory activity [22]. Thus, inflammaging can induce a positive feedback loop through altered Sirt function to drive ROS production, mitochondrial dysfunction, and more inflammation. In addition, as a natural by-product of amino acid metabolism and urea cycle, ammonia levels in

the brain increase with age, contributing to neuroinflammation through p38 MAPK/NF- κ B pathway and blood–brain-barrier breakdown [300]. In human and murine skeletal muscle/myotubes, hyperammonemia can inhibit the mitochondrial electron transport chain complex I that oxidizes NADH to NAD⁺, leading to a lower redox ratio, Sirt3 dysfunction, and postmitotic senescence in myotubes, accompanied by increased acetyl–NF- κ B p65 expression, an activating post-translational modification [301]. Thus, multiple metabolic mechanisms contribute to Sirt decline with age.

Collectively, the major nutrient-sensing pathways, including the IIS, mTOR, Sirts, and AMPK pathways, orchestrate the physiology of longevity. Their deregulation, as a hallmark of aging, fuels the metabolic inflammation that impacts the healthspan and lifespan of aging populations. Early detection of immunometabolic dysregulation with multi-omics approaches, such as lipidomics, metabolomics, and glycomics [302], permits the early diagnosis of metabolic and ARDs and represents a future direction of personalized geroprotective treatments.

3.2. Mitochondrial dysfunction

Mitochondria developed through the merging of a prokaryotic organism into a eukaryotic cell [303]. From these origins, mitochondria maintain their own DNA (mtDNA) and DNA replication process. They act to supply the cell with ATP and synthesize key molecules in the processes of inflammation, oxidation, and metabolism. In recent years surmounting evidence suggests that mitochondria not only play an influential role in the progression of the inflammaging phenotype but are also heavily impaired by chronic inflammatory states [85]. Indeed, mitochondrial function declines with aging, and this change is associated with increased ROS, reduced ATP production, elevated mitochondrial damage, and age-related epiphenomena (Figure 2) [304–306]. While multiple mechanisms link mitochondrial dysfunction with aging, one such mechanism relates to age-related depletion of NAD⁺, which causes loss of efficient sirtuin activity, resulting in impaired mitochondrial SIRT3. Impaired Sirt3 leads to dysregulation in mitochondrial antioxidant systems, mtDNA repair, and mitochondrial quality control and biogenesis pathways [307]. Here, we examine the reciprocal links between inflammation and mitochondria with age.

3.2.1. mtDNA promotes chronic inflammation

Because of their endosymbiotic origins, mitochondria carry a milieu of potential DAMPs that may be uniquely immuno-stimulating [308]. Some of the most putative mitochondrial DAMPs include the release of mtDNA, N-formyl peptides, and unique mitochondrial lipid species such as cardiolipin [309]. During aging, there is increased mitochondrial dysfunction, reduced genome integrity, and damage [310], allowing for the increased potential of age-related interactions with mitochondrial DAMPs.

The phospholipid cardiolipin is essential for maintaining mitochondrial structure and function and normally siloed to the inner mitochondrial membrane. However, when exposed to the cytoplasm, it stimulates mitophagy, the process in which dysfunctional mitochondria are eliminated and new mitochondria are formed [311]. Apoptosis is precipitated in most severe cases when cardiolipin stimulates the release of cytochrome c [312]. Mitochondrial proteins, including N-Formyl peptides, primarily induce inflammation through binding to formyl peptide receptor-1 [313].

In the case of mtDNA, immune activation can be mediated by TLR signaling. Indeed, mtDNA binds TLR-9, a receptor capable of detecting unmethylated DNA with CpG motifs derived from bacteria and viruses and initiating the innate immune response [314]. Activation of TLR-9

leads to NF- κ B signaling, which induces the expression of several pro-inflammatory cytokine genes [315] and is aberrant in many chronic diseases [316]. In this way, mitochondria not only contribute to the transmission of danger signals within the cell but are also a major source of molecules capable of activating the innate immune system. For reasons not entirely understood, circulating mtDNA appears to increase gradually with age after the fifth decade of life [317], and the abundance of unhoused mtDNA is associated with accelerated aging pathology [318]. Various inflammatory pathways are activated when ROS-damaged mtDNA and cardiolipin are released into the cytosol. Responses that have been best studied involve the NLRP3 inflammasome, the cGAS-STING pathway, and NF- κ B [309,319]. Cytosolic mitochondrial RNA has similar effects, stimulating TBK1/IKK and NF- κ B through RIG-1, MDA5, and mitochondrial antiviral-signaling protein (MAVS) [320]. Mitochondrial damage also activates other inflammasomes such as NLRP10, and AIM2 in keratinocytes and macrophages, respectively [321,322], thus implicating their potential role during mitochondrial damage-induced inflammation in aging. Finally, mitochondrial-derived phosphocreatine and ATP can also instigate inflammation through inflammasome activation or, in the case of ATP, by also binding extracellular P2X purinoceptor 7 on innate immune cells [322,323].

In addition to mtDNA and ROS, mitochondrial dysfunction releases TCA cycle intermediate metabolites into the cytosol with immunomodulatory effects [324]. Fumarate has been shown to impose immunomodulatory effect through controlling chromatin modifications and regulating protein succination [325]. Specifically, pro-inflammatory insults can lead to the accumulation of fumarate through glutamine anaplerosis which has been shown to be necessary for trained immunity and inflammation by inhibiting lysine-specific demethylase 5 A (KDM5) histone demethylase activity; inhibition of KDM5 increases the levels of H3K4me3, a marker of active gene transcription at the promoters of TNF α and IL6 cytokines [326]. Furthermore, elevated levels of succinate, resulting from mitochondrial dysfunction, can potentially contribute to chronic inflammation by activating the HIF-1 α pathway through stabilization of HIF-1 α . Succinate inhibits prolyl hydroxylases, enzymes responsible for the degradation of HIF-1 α . Consequently, stabilized HIF-1 α translocates into the nucleus and initiates the transcription of key inflammatory cytokines, particularly IL-1 β [327]. It remains to be seen if such mechanisms occur during inflammaging. Itaconate has been shown to inhibit the nuclear factor-kappa B (NF- κ B) pathway, a key regulator of inflammation [328]. Itaconate acts by directly modifying specific cysteine residues on key proteins involved in NF- κ B signaling, such as I κ B kinase (IKK) and Kelch-like ECH-associated protein 1 (Keap1). This modification disrupts the activation of NF- κ B and promotes its nuclear export, thereby suppressing the expression of pro-inflammatory genes. Additionally, itaconate can also promote anti-inflammatory responses by activating the anti-inflammatory transcription factor NRF2, which induces the expression of antioxidant and anti-inflammatory genes including GPX1 and SOD1 which decrease ROS [328,329]. Itaconate may be induced during aging inside immune cells like macrophages, where it may act as a compensatory factor to help dampen inflammation or maintain tissue homeostasis [330].

3.2.2. Inflammation impairs mitochondrial function

Conversely, chronic inflammation has been shown to impair mitochondrial function through inflammasome activation and other pro-inflammatory immune signaling engagements, such as the exaggerated generation of ROS, which produce damage in mitochondrial proteins, lipids, and mtDNA [331]. An array of ARDs,

including CVDs, cancer, neurodegenerative disease, and metabolic diseases, has been linked to mitochondrial dysfunction [332–334]. Similar to DNA damage, mitochondrial machinery, and compartments accumulate damage over time and rely on repair and replacement mechanisms to maintain performance. However, instead of repairing damaged entities, dysfunctional mitochondrial proteins and even entire organelles are selectively removed through mitophagy [335,336]. This quality control process selects for healthy mitochondria to replicate and govern homeostasis. However, both excessive chronic inflammation and aging impair autophagic mechanisms. For instance, excessive inflammation due to the NLRP3 inflammasome can trigger CASP1-dependent mitochondrial damage, ROS production, dissipation of mitochondrial membrane potential, permeabilization, and inhibition of mitophagy through cleavage of Parkin [319]. The resultant accumulation of damaged mitochondria leads to uncontrolled ROS and mtDNA leakage and the ensuing IL-1 β - and IL-6-dependent inflammation orchestrated by cGAS-STING signaling (Figure 2) [337].

3.3. Cellular senescence

Cellular senescence is a state of proliferative arrest in response to cell stressors [338]. Senescence can be induced by various stimuli, including DNA damage, oxidative stress, replicative stress, mitochondrial signaling, and altered expression of certain oncogenes [339]. Senescent cells remain mechanically intact, metabolically operational, and involved in intercellular signaling while still arresting growth and replication. Senescence growth and replicative arrest have been shown to be mediated by major tumor-suppressor pathways such as p16^{INK4a}, pRB (retinoblastoma protein), and by p53 [340,341]. Senescent cells and their secretory products are now widely accepted as important contributors to aging and age-related disease. Cell-specific effects of cellular senescence and inflammaging have been recently reviewed elsewhere [342].

3.3.1. Cell senescence and its SASP promotes inflammation

The secretion profile of factors specific to senescent cells has been termed the “senescence-associated secretory phenotype” or SASP [343,344]. The SASP is a bioactive secretome thought to promote the recruitment and activation of immune cells that would clear senescent cells. When clearance fails, the result is the accumulation of senescent cells and SASP factors which leads to diminished tissue function and steadily elevated proinflammatory tone. SASP factors are also thought to be critical mediators of tissue repair [345]. However, they may also potentially facilitate senescence spreading into neighboring cells [346]. Some of the most robustly induced secreted pro-inflammatory SASP proteins include, but are not limited to, IL-6, IL-8, IL-1, granulocyte-macrophage colony-stimulating factor (GM-CSF), growth-regulated oncogene (GRO) α , monocyte chemoattractant protein (MCP)-2, MCP-3, matrix metalloproteinase (MMP)-9, -1, -3 and several IGF-binding proteins. SASP factors that have been proposed to be biomarker candidates of aging include markers such as growth differentiation factor (GDF)-15, stanniocalcin 1, MMP-1, Inhibin Subunit Beta A (INHBA), and serpin family E member 1 (SERPINE1) [343,347].

In addition to pro-senescence mechanisms, such as persistent DDR signaling, GATA4 stabilization, and ensuing NF- κ B and C/EBP β cascades [25,348,349], the secretion of SASP factors can also be initiated or maintained by two microRNAs, mir-146a/b. These microRNAs orchestrate a negative feedback loop dampening the escalation of NF- κ B activity and maintaining a chronic low-grade inflammatory phenotype [350]. Despite the induction of mir-146a/b, the SASPs persist in promoting the low-grade inflammation thought to drive

chronic pathologies associated with aging [351]. Interestingly, the genetic determinants of senescence also leads to unique SASPs and inflammatory profiles. For instance, p21-induced senescence induces a distinct SASP characterized by C-X-C motif chemokine 14 (CXCL14) expression, which puts stressed cells under immunosurveillance [352]. In addition to recruiting immune cells, SASP factors can also induce markers of aging, such as CD38, on immune cells. Macrophages expressing CD38 in aged tissues deplete NAD⁺, which further drives aging phenotypes [87,89,299]. Thus, it will be interesting in the future to map out how underlying instigators of senescence contribute to specific inflammatory profiles across the chronic diseases of aging and to better understand distinct impacts on immune cell populations. Of note, not all SASP immunological factors should be considered pathogenic with age. For instance, senescent cells can also secrete factors that stimulate tissue repair and epithelial regeneration [345]. As a result, the use of senolytics that target senescent cells should be used with caution, as indiscriminate senolysis can interfere with less appreciated benefits of senescent cells.

3.3.2. Inflammation promotes senescence

Senescent cells have been shown to accumulate over the life span of humans and predominantly in renewable tissues and tissues that experience prolonged inflammation [353,354]. Many inflammatory diseases, such as atherosclerosis, osteoarthritis, idiopathic pulmonary fibrosis, and insulin resistance, also show elevations in senescent cells associated with inflammation [22]. As previously mentioned, molecular damage, including DNA and structural damage induced by oxidative stress associated with chronic inflammation, can trigger senescence. The accumulation of oxidative stress, DNA damage, and even reactivation of HERV can exacerbate inflammation in aging individuals by activating the cGAS-STING pathway, resulting in increased interferons or SASP signaling (Figure 2) [125,355]. In addition, as discussed in section 3.1., mTORC1 signaling mediates the production of various pro-inflammatory cytokines including IL-1A, while rapamycin can suppress the translation of the membrane-bound cytokine IL-1A, leading to diminished NF- κ B transcriptional activity and reduced SASP [356]. Moreover, viral infection, such as by SARS-CoV-2, can evoke “cytokine storms” characterized as pro-inflammatory cytokines, particularly in the elderly, and amplify SASP from pre-existing senescent cells. The resulted increase in SASP can further exacerbate tissue damage in addition to the direct cellular damage due to virus itself [357]. The innate immune sensing of viral RNA or cytosolic chromatin fragments associated with aging through cGAS-STING and its downstream mediators, predominantly IRF3, NF- κ B and C/EBP β , may also promote both induction and spreading of senescence [357–359].

4. INTEGRATIVE HALLMARKS

The integrative hallmarks arise when the homeostatic machinery of aging tissues fails to resolve the accumulated damage from the primary and antagonistic hallmarks [6]. Because of the broad, tissue-specific etiology underlying the integrative hallmarks, their initiation, and progression can exacerbate the primary and antagonistic hallmarks, engendering a large set of age-related pathologies at the systemic level [360].

4.1. Altered intercellular communication

Intercellular signaling, as a hallmark of aging, encompasses many modes of cell-to-cell signaling, including endocrine, neuronal, and neuroendocrine pathways, as well as cell-to-cell contact and the

exchange of vesicle-packaged and free soluble factors, including inflammatory signals [6]. Because of the vast role of aberrant intercellular signaling in aging pathology and the degree to which chronic inflammation contributes, these two hallmarks were differentiated in the 2023 Hallmarks of Aging update [6]. Here we briefly highlight links between chronic inflammation and other forms of intercellular communication, including hormones and extracellular vesicles.

4.1.1. Hormones and chronic inflammation

In a similar age-dependent manner, secretory patterns of the hormones produced by the hypothalamic–pituitary axis (HPA) [361] and gut–pancreas–liver axis (GPLA) change [362], as does the sensitivity of the axis response to feedback. The HPA is a master regulator of the body's systemic homeostasis, such as energy balance, body temperature, sleep, blood pressure, and circadian rhythms. Background-level inflammation in the hypothalamus, concisely termed “hypothalamic microinflammation,” is thought to contribute to the deviation from homeostasis, particularly metabolic dysregulation, with age and the tonal systemic inflammation that ensues [363].

In addition to decreased insulin sensitivity and clearance, aging is associated with reductions in fasting glucagon-like peptide-1 (GLP-1) and glucose-dependent insulinotropic polypeptide (GIP), which may predispose to the development of glucose intolerance and T2DM with age [362]. The consequence of dysregulated insulin is the inflammatory reprogramming of immunometabolism and immune cell function (discussed in the section *IIS deregulation induces inflammation*). Furthermore, the age-associated downregulation of sex hormones, estrogen, and testosterone, may also promote inflammation. Estrogen can induce Th2 responses [364], and testosterone receptors in T cells promote forkhead box protein P3 (Foxp3) expression and Treg fates [365].

4.1.2. Extracellular vesicles as mediators of inflammaging

EVs are lipid membrane vesicles derived from multivesicular bodies or plasma membrane containing proteins, lipids, and nucleotide species, sharing a snapshot of the donor cell state with the recipient [366]. Released by most cell types, they are a newly appreciated form of intercellular communication, linking hallmarks of aging through inflammatory mediators [367,368]. In cancer and autoimmunity, EVs promote inflammation [369], for instance, in the delivery of TLR ligands to plasmacytoid dendritic cells [370]. EVs also exhibit regenerative potential as stem cell-derived or “young” EVs can improve healthspan, though they can lose their regenerative potential over time. For example, mesenchymal stem cell (MSC)-derived small EVs reduce endothelial cell senescence via delivery of miR-146a [371], while reduced levels of EVs containing galectin-3 (Gal-3) in aging plasma donors correlate with a reduced ability to induce osteoblasts from MSCs [372] and may also potentiate PAMP-TLR responses [373]. Similarly, EVs from young human donors were found to have higher levels of glutathione-related protein (GSTM2) than old EVs, which promotes antioxidant activity [374] (Figure 2). EV research is a burgeoning field, and the modes by which EVs partake in inflammaging require further study, including their immunosuppressive role in expanding senescence [375] and their ability to serve as a biomarker for age-related conditions [376].

4.2. Stem cell exhaustion

Adult stem cells function as self-renewal pools that facilitate tissue repair by replacing damaged cells in various tissues and organs [377]. During aging, multiple factors, including telomere attrition, DNA damage, epigenetic dysregulation, and chronic inflammation, cause a

decline in both the regenerative and proliferative capacities of stem cells, leading to stem cell exhaustion and accumulation of senescent cells [378,379]. Senescent cells can in turn facilitate additional stem cell dysfunction. For instance, in aged progeroid INK-ATTAC mice, senescent fat progenitors inhibit adipogenesis in non-senescent progenitors through activin A secretion [380]. Chronic inflammation, even early in life, can contribute to the depletion of HSCs, causing phenotypes of premature aging [381], whereas the inhibition of NF- κ B activation demonstrates a functional rejuvenation of aged skeletal stem/progenitor cells (SSPC) [382]. Meanwhile, senescent cells can exacerbate stem cell exhaustion via SASP cytokines that favor the infiltration of immune cells, creating an inflamed niche that further impedes stem cell function [383–385]. For example, aged skeletal stem cells (SSCs) become pro-inflammatory and contribute to an inflamed bone marrow niche, which can promote the dysfunction of HSCs [386], while aged HSCs can also activate NF- κ B, facilitating the myeloid-skewed differentiation and the eventual loss of self-renewal [387,388]. Additionally, the regenerative capacity of differentiated cells in response to injury is also weakened in aged animals, indicating that the injury-induced cellular plasticity also may be impaired during aging [389]. Although acute inflammation can provoke somatic stemness and promote tissue recovery by IL-6-mediated epigenetic modifications [390], chronic inflammation can lock cells in epigenetically plastic states disabled for reparative differentiation, leading to the accumulation of damaged cells and even tumorigenesis [391].

Recent advances have illustrated the importance of stem cell-related factors that rejuvenate tissues by transiently reprogramming somatic cells. Specifically, the Yamakana factors (Oct3/4, Sox2, Klf4, c-Myc, or OSKM) can induce aging somatic cells to transiently de-differentiate into a younger state, after which the somatic identity is regained with age-related epigenetic patterns erased and cellular functions rejuvenated [392]. Multiple *in vivo* and *in vitro* studies have proved the efficacy of OSKM- or OSK-mediated transient reprogramming in murine and human cells, leading to the marked reversal of epigenetic clocks, reduction in inflammatory profiles, and restoration of lost functionality in diseased or aged cells [393–395]. Notably, as transient reprogramming recapitulates features of natural tissue repair, which often declines with age due to accumulated senescent cells, the removal of senescent cells by senolysis extends *Drosophila* lifespan synergistically with OSKM expression [395]. Other means for stem cell renewal include fecal microbiota transplantation from young mice to aged mice, which can enhance hematopoietic repopulation capacity, thus exhibiting the potential to rejuvenate HSCs [396].

4.3. Dysbiosis

The gut microbiome plays a vital role in maintaining human health through nutrient digestion and absorption, protection against pathogens, and the production of important metabolites such as vitamins, as well as amino acid and fatty acid derivatives [397]. Dysbiosis refers to an imbalance in the gut microbiome, which can occur for various reasons and negatively impact health and serve as a catalyst for fueling inflammaging. However, the contribution of dysbiosis in the context of the human microbiome interaction, particularly regarding its impact on systemic immune function or deterioration of this function among the elderly as it relates to ARDs, is an ongoing area of research [398]. A recent study by Galkin and colleagues shows that microbiome profiles can predict the age of healthy individuals, and patients with T1D exhibit age acceleration according to the microbiome clock [399]. Furthermore, a growing body of literature implicates age-related dysbiosis of the gut microbiome as contributing to tissue-specific (such as in the liver, adipose, and intestine) as well as a global inflammatory state in

the elderly [257]. Interestingly, besides the gut–brain axis, oral dysbiosis also plays a role in mediating age-related neurodegenerative diseases, expanding the term into the “oral–gut–brain axis”. Metabolites produced by gut and oral microbes (particularly *Streptococcus mutans* and *Porphyromonas gingivalis*) have been linked to the formation of β -amyloid and its accumulation in the brain, tau protein phosphorylation, and neuroinflammation in individuals with Alzheimer’s disease [400].

Short-chain fatty acids (SCFA), such as butyrate, are considered a primary microbial anti-inflammatory metabolite that is known to inhibit the NF- κ B pathway [401] and regulate Th17/Treg differentiation by inhibiting IL-6/signal transducer and the activator of transcription 3 (STAT3)/IL-17 pathway and promoting Foxp3 [402,403]. *Faecalibacterium prausnitzii* has been identified as a butyrate-producing bacterium with an inverse relationship with different proinflammatory markers [402].

Another feature of the centenarians’ gut ecosystem is the increase in facultative anaerobes, such as bacteria belonging to the *Micrococcaceae* family, the *Fusobacterium*, *Bacillus*, *Staphylococcus*, and *Corynebacterium* genera, and many members of the phylum Proteobacteria. Such opportunistic microbes, especially the *Enterobacteriaceae* family, thrive in an inflamed environment and are known to increase in the elderly and are associated with a decrease in *F. prausnitzii* [404]. Overall, studies with centenarian populations typically show reduced core taxa, such as *Bacteroides* and *Roseburia*, associated with increases in *Akkermansia* and *Bifidobacterium*, which may have pro-longevity effects [405]. *Akkermansia* can be modulated by local levels of IFN- γ [406], a cytokine linked to inflammaging, and has immunomodulatory effects in the host, possibly through its pili structures [407]. Other mechanisms by which changes in the gut

microbiota can intersect with aging or immunity include the production of amino acid metabolites, such as from tryptophan (e.g., indoles that bind the aryl hydrocarbon receptor [408] and phenylalanine/tyrosine [409], and through the production of secondary bile acids [410]. Despite these advances, more work, such as through fecal microbial transplantation studies [411], will undoubtedly improve our knowledge of mechanisms by which the gut microbiota intersects with the immune system to control healthy aging. A full scope of the recent advances in characterizing the role of the gut microbiome in aging and longevity has been systematically reviewed here [412].

5. CONCLUSIONS AND FUTURE PERSPECTIVES

This review highlights the bidirectional relationships between chronic inflammation and the hallmarks of aging by summarizing the underlying molecular mechanisms (Figure 3). Immune dysfunction and changes in inflammatory pathways are transversal contributors to the aging process, propagating a rippling effect through the other drivers of aging. Due to its interconnectedness and ability to accentuate aging, drivers of chronic inflammation may be the ideal target to address aging with high translational potential. While their clinical application and long-term safety remain to be proven in clinical trials, current evidence suggests that immune-targeting approaches have the potential to revolutionize longevity research and treatment, as they have done in cancer therapies. Boosting and targeting the immune system to clear senescent cells is just one example, as we have discussed. Progress in this field will require technological improvements in the measurement and analysis of multi-omics and rapid cycles of translation from model organisms to humans and *vice versa* to identify new promising therapeutic targets.

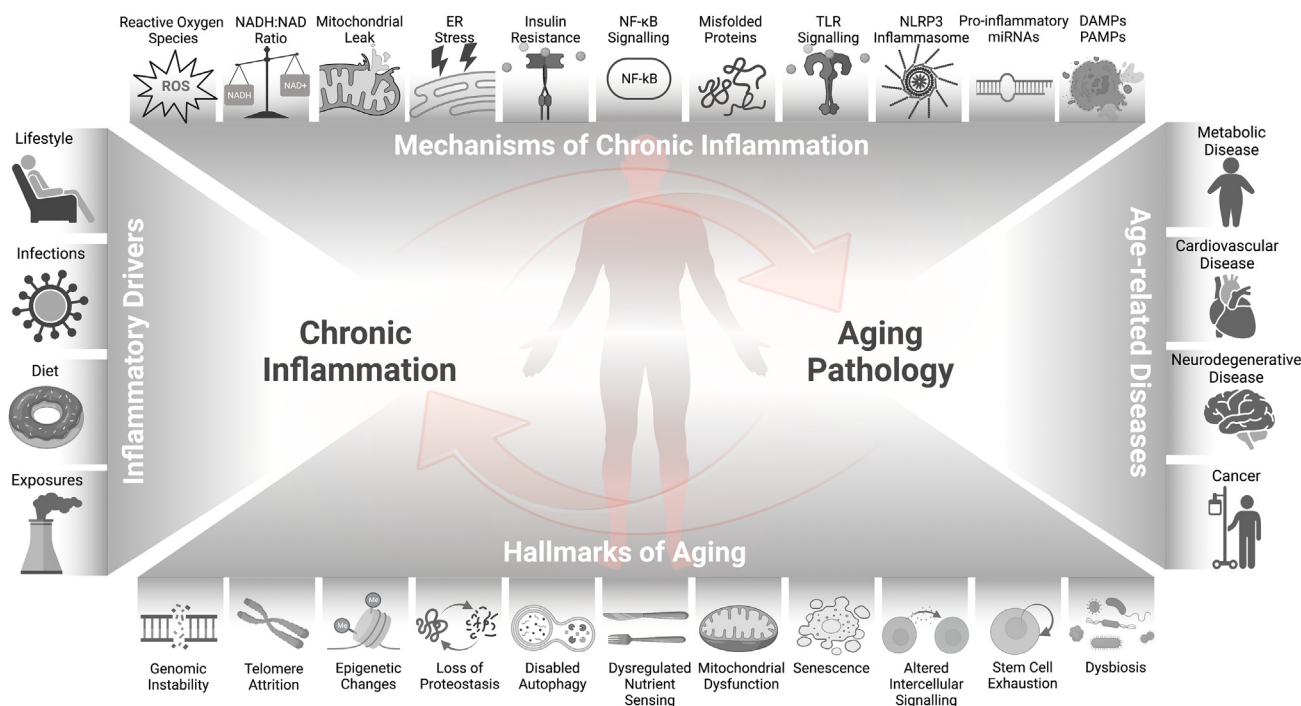


Figure 3: Clinical and Molecular Drivers Underlying the Loop of Chronic Inflammation and the Hallmarks of Aging: Through various molecular mechanisms and environmental triggers, chronic inflammation can drive aging and age-related diseases, which in turn can exacerbate chronic inflammation along with other hallmarks of aging. DAMPs = deoxyribonucleic acid damage-associated molecular patterns, ER = endoplasmic reticulum, NAD/NADH = nicotinamide adenine dinucleotide, NF- κ B = nuclear factor kappa B, NLRP3 = nod-like receptor family pyrin domain-containing 3, PAMPs = pathogen-associated molecular pattern molecules, TLR = toll-like receptor.

As our understanding of aging continues to expand, new hallmarks of aging will inevitably emerge. Foreseeable future hallmarks of aging include those linked to the extracellular matrix (ECM) and mechanobiology, both up-and-coming areas of basic aging science research [413–415]. Indeed, aging and ARDs are associated with changes in the ECM and mechanical cell surroundings, including immune cells. Mechanosensing by immune cells, such as through YAP/TAZ or PIEZO1 signaling [416,417], alters immune cell function, maturation, and metabolism to drive aberrant inflammation [418]. Thus, one future direction of inflammaging research is to link physical cues with chemical cues, thus generating a better understanding of immunological changes with age.

AUTHOR CONTRIBUTIONS

All authors contributed to the writing of this manuscript.

DECLARATION OF COMPETING INTEREST

DF is co-founder and Chairman of the Scientific Advisory Board of Edifice Health, a company that utilizes Inflammatory Age (iAge) to quantify chronic inflammation and immune health. The remaining authors declare no competing interests.

DATA AVAILABILITY

No data was used for the research described in the article.

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ABBREVIATIONS

Abbreviation Definition

ABCs	age-associated B cells
AD	Alzheimer's Disease
AGEs	advanced glycation end-products
AID	activation-induced cytidine deaminase
ALS	autophagy-lysosome system
AMPK	5'-adenosine monophosphate-activated protein kinase
AP-1	activator protein 1
APCs	antigen-presenting cells
ARDs	age-related diseases
ATF6	ATF5 activating transcription factor 6 Autophagy-related 5
ATM	ataxia-telangiectasia mutated
ATR	ataxia telangiectasia and Rad3-related protein
BAX	Bcl-2 Associated X-protein
C/EBP	CCAAT-enhancer-binding proteins
CBP	cyclic adenosine monophosphate response element binding protein binding protein
cfDNA	cell-free DNA
cGAS	cyclic GMP—AMP synthase
CHOP	CCAAT-enhancer-binding proteins homologous protein
CMA	chaperone-mediated autophagy
COPD	chronic obstructive pulmonary disease
COVID-19	coronavirus disease 2019
CRP	c-reactive protein
CVD	cardiovascular disease

DAMPs	damage-associated molecular patterns
DDR	DNA damage response
DNA	deoxyribonucleic acid
DNMT	DNA methyltransferase
eIF2 α	eukaryotic translation initiation factor 2
ER	endoplasmic reticulum
FFAs	Free fatty acids
FoxO	forkhead box protein O
GDF	growth differentiation factor
GM-CSF	granulocyte-macrophage colony-stimulating factor
GRO	growth-regulated oncogene
H3K	H3 lysine
HATs	histone acetyltransferases
HDACs	histone deacetylases
HERV-K	human endogenous retrovirus-K
HSC	hematopoietic stem cells
hTERT	human telomerase reverse transcriptase
IFN	interferon
IGF-1	insulin-like growth factor 1
IGFR	insulin-like growth factor receptor
IKK β	inhibitor of nuclear factor kappa-B kinase subunit beta
IL	interleukin
INHBA	inhibin Subunit Beta A
IP	immunoproteasome
IR	insulin resistance
IRE1 α	inositol-requiring transmembrane kinase/endoribonuclease 1 α
IRF3	interferon regulatory factor 3
IRS	insulin receptor substrate
IIS	insulin and insulin-like growth factor 1 signaling
JNK	jun n-terminal kinase
LC3B	microtubule-associated protein 1 A/1 B-light chain 3
LINE1	long interspersed nuclear element 1
LKB-1	liver kinase B1
LPS	lipopolysaccharide
MAVS	mitochondrial antiviral-signaling protein
MCP	monocyte chemotactic protein
MDA5	anti-melanoma differentiation-associated gene 5
miRNAs	microribonucleic acid
MLH1	MutL homolog 1
MMP	matrix metalloproteinase
MMR	mismatch repair
mRNA	messenger ribonucleic acid
mtDNA	mitochondrial DNA
mTOR	mammalian target of rapamycin
mTORC1	mammalian target of rapamycin complex 1
mTORC2	mammalian target of rapamycin complex 2
MuSCs	muscle stem cells
NAD	nicotinamide adenine dinucleotide
NADPH	nicotinamide adenine dinucleotide phosphate hydrogen
NAFLD	nonalcoholic fatty liver disease
ncRNA	non-coding RNA
NF- κ B	nuclear factor kappa B
NLR	nod-like receptor
NLRP3	nod-like receptor family pyrin domain-containing 3
NOS2	nitric oxide synthase
PAMPs	pathogen-associated molecular pattern molecules
PARP1	poly(adenosine diphosphate-ribose) polymerase 1
PD	Parkinson's Disease
PERK	protein kinase R-like endoplasmic reticulum kinase
PGC	peroxisome proliferator-activated receptor gamma coactivator
Phox	phagocyte oxidase
PI3K	phosphatidylinositol-3-kinase
PRAAS	proteasome-associated autoinflammatory syndrome
PRRs	pattern-recognition receptors

RIDD	regulated IRE1 α -dependent decay
RIG-I	retinoic acid-inducible gene I
RNS	reactive nitrogen species
ROS	reactive oxygen species
RTPs	retrotransposons
SARS-CoV-2	severe acute respiratory syndrome coronavirus 2
SASP	senescence-associated secretory phenotype
SERPINE1	serpin family E member 1
SESNs	sestrins
Sirt	sirtuin
SSPC	skeletal stem/progenitor cells
STING	stimulator of interferon genes
T2D	type 2 diabetes
TBK1	tank-binding kinase 1
TCR	T cell receptor
TGF β	transforming growth factor-beta
TLR	toll-like receptor
TNF	tumor necrosis factor
TRAF6	tumor necrosis factor receptor-associated factor 6
UPR	unfolded protein response
UPS	ubiquitin-proteolytic system
UV	ultraviolet light
VAT	visceral adipose tissue
YAP1	yes-associated protein 1

REFERENCES

- Keilich SR, Bartley JM, Haynes L. Diminished immune responses with aging predispose older adults to common and uncommon influenza complications. *Cell Immunol* 2019;345(103992):103992. <https://doi.org/10.1016/j.cellimm.2019.103992>.
- Bartleson JM, Radenkovic D, Covarrubias AJ, Furman D, Winer DA, Verdin E. SARS-CoV-2, COVID-19 and the ageing immune system. *Nat Aging* 2021;1(9):769–82. <https://doi.org/10.1038/s43587-021-00114-7>.
- Niccoli T, Partridge L. Ageing as a risk factor for disease. *Curr Biol* 2012;22(17):R741–52. <https://doi.org/10.1016/j.cub.2012.07.024>.
- Jiang X, Holmes C, McVean G. The impact of age on genetic risk for common diseases. *PLoS Genet* 2021;17(8):e1009723. <https://doi.org/10.1371/journal.pgen.1009723>.
- López-Otín C, Blasco MA, Partridge L, Serrano M, Kroemer G. The hallmarks of aging. *Cell* 2013;153(6):1194–217. <https://doi.org/10.1016/j.cell.2013.05.039>.
- López-Otín C, Blasco MA, Partridge L, Serrano M, Kroemer G. Hallmarks of aging: an expanding universe. *Cell* 2023;186(2):243–78. <https://doi.org/10.1016/j.cell.2022.11.001>.
- Furman D, Campisi J, Verdin E, Carrera-Bastos P, Targ S, Franceschi C, et al. Chronic inflammation in the etiology of disease across the life span. *Nat Med* 2019;25(12):1822–32. <https://doi.org/10.1038/s41591-019-0675-0>.
- Melov S. Mitochondrial oxidative stress. Physiologic consequences and potential for a role in aging. *Ann N Y Acad Sci* 2000;908(1):219–25. <https://doi.org/10.1111/j.1749-6632.2000.tb06649.x>.
- Fulop T, Larbi A, Pawelec G, Khalil A, Cohen AA, Hirokawa K, et al. Immunology of aging: the birth of inflammaging. *Clin Rev Allergy Immunol* 2023;64(2):109–22. <https://doi.org/10.1007/s12016-021-08899-6>.
- Saxena S, Zou L. Hallmarks of DNA replication stress. *Mol Cell* 2022;82(12):2298–314. <https://doi.org/10.1016/j.molcel.2022.05.004>.
- Lindahl T. Instability and decay of the primary structure of DNA. *Nature* 1993;362(6422):709–15. <https://doi.org/10.1038/362709a0>.
- Giglia-Mari G, Zotter A, Vermeulen W. DNA damage response. *Cold Spring Harb Perspect Biol* 2011;3(1):a000745. <https://doi.org/10.1101/cshperspect.a000745>.
- Chen Y, Geng A, Zhang W, Qian Z, Wan X, Jiang Y, et al. Fight to the bitter end: DNA repair and aging. *Ageing Res Rev* 2020;64(101154):101154. <https://doi.org/10.1016/j.arr.2020.101154>.
- Crusz SM, Balkwill FR. Inflammation and cancer: advances and new agents. *Nat Rev Clin Oncol* 2015;12(10):584–96. <https://doi.org/10.1038/nrclinonc.2015.105>.
- Guo S, Zhu X, Huang Z, Wei C, Yu J, Zhang L, et al. Genomic instability drives tumorigenesis and metastasis and its implications for cancer therapy. *BioMed Pharmacother* 2023;157(114036):114036. <https://doi.org/10.1016/j.biopha.2022.114036>.
- Hanahan D. Hallmarks of cancer: new dimensions. *Cancer Discov* 2022;12(1):31–46. <https://doi.org/10.1158/2159-8290.CD-21-1059>.
- Abbotts R, Wilson 3rd DM. Coordination of DNA single strand break repair. *Free Radic Biol Med* 2017;107:228–44. <https://doi.org/10.1016/j.freeradbiomed.2016.11.039>.
- Verdin E. NAD⁺ in aging, metabolism, and neurodegeneration. *Science* 2015;350(6265):1208–13. <https://doi.org/10.1126/science.aac4854>.
- Lee JH, Hussain M, Kim EW, Cheng SJ, Leung AKL, Fakouri NB, et al. Mitochondrial PARP1 regulates NAD⁺-dependent poly ADP-ribosylation of mitochondrial nucleoids. *Exp Mol Med* 2022;54(12):2135–47. <https://doi.org/10.1038/s12276-022-00894-x>.
- Wiley CD, Velarde MC, Lecot P, Liu S, Sarnoski EA, Freund A, et al. Mitochondrial dysfunction induces senescence with a distinct secretory phenotype. *Cell Metabol* 2016;23(2):303–14. <https://doi.org/10.1016/j.cmet.2015.11.011>.
- Andina N, de Meuron L, Schnegg-Kaufmann AS, Sarangdhar MA, Ansermet C, Bombaci G, et al. Increased inflammasome activation is associated with aging and chronic myelomonocytic leukemia disease severity. *J Immunol* 2023;210(5):580–9. <https://doi.org/10.4049/jimmunol.2200412>.
- Walker KA, Basisty N, Wilson 3rd DM, Ferrucci L. Connecting aging biology and inflammation in the omics era. *J Clin Invest* 2022;132(14). <https://doi.org/10.1172/JCI158448>.
- Erdal E, Haider S, Rehwinkel J, Harris AL, McHugh PJ. A prosurvival DNA damage-induced cytoplasmic interferon response is mediated by end resection factors and is limited by Trex1. *Genes Dev* 2017;31(4):353–69. <https://doi.org/10.1101/gad.289769.116>.
- Zhao J, Zhang L, Lu A, Han Y, Colangelo D, Bukata C, et al. ATM is a key driver of NF- κ B-dependent DNA-damage-induced senescence, stem cell dysfunction and aging. *Aging (Albany NY)* 2020;12(6):4688–710. <https://doi.org/10.18632/aging.102863>.
- Kang C, Xu Q, Martin TD, Li MZ, Demara M, Aron L, et al. The DNA damage response induces inflammation and senescence by inhibiting autophagy of GATA4. *Science* 2015;349(6255):aaa5612. <https://doi.org/10.1126/science.aaa5612>.
- Dunphy G, Flannery SM, Almine JF, Connolly DJ, Paulus C, Jonsson KL, et al. Non-canonical activation of the DNA sensing adaptor STING by ATM and IFI16 mediates NF- κ B signaling after nuclear DNA damage. *Mol Cell* 2018;71(5):745–60. <https://doi.org/10.1016/j.molcel.2018.07.034>. e5.
- Hinz M, Stilmann M, Arslan SÇ, Khanna KK, Dittmar G, Scheidereit C. A cytoplasmic ATM-TRAF6-cIAP1 module links nuclear DNA damage signaling to ubiquitin-mediated NF- κ B activation. *Mol Cell* 2010;40(1):63–74. <https://doi.org/10.1016/j.molcel.2010.09.008>.
- Zhao Y, Simon M, Seluanov A, Gorbunova V. DNA damage and repair in age-related inflammation. *Nat Rev Immunol* 2023;23(2):75–89. <https://doi.org/10.1038/s41577-022-00751-y>.
- Cercek A, Lumish M, Sinopoli J, Weiss J, Shia J, Lamendola-Essel M, et al. PD-1 blockade in mismatch repair-deficient, locally advanced rectal cancer. *N Engl J Med* 2022;386(25):2363–76. <https://doi.org/10.1056/NEJMoa2201445>.
- St Sauver J, Rocca W, LeBrasseur N, Chamberlain A, Olson J, Jacobson D, et al. Inflammatory biomarkers, multi-morbidity, and biologic aging. *J Int Med Res* 2022;50(7). <https://doi.org/10.1177/03000605221109393>.

- [31] Chen X, Andresen BT, Hill M, Zhang J, Booth F, Zhang C. Role of reactive oxygen species in tumor necrosis factor- α induced endothelial dysfunction. *Curr Hypertens Rev* 2008;4(4):245–55. <https://doi.org/10.2174/157340208786241336>.
- [32] Blaser H, Dostert C, Mak TW, Brenner D. TNF and ROS crosstalk in inflammation. *Trends Cell Biol* 2016;26(4):249–61. <https://doi.org/10.1016/j.tcb.2015.12.002>.
- [33] Dizdaroglu M, Jaruga P. Mechanisms of free radical-induced damage to DNA. *Free Radic Res* 2012;46(4):382–419. <https://doi.org/10.3109/10715762.2011.653969>.
- [34] Ionescu-Tucker A, Cotman CW. Emerging roles of oxidative stress in brain aging and Alzheimer's disease. *Neurobiol Aging* 2021;107:86–95. <https://doi.org/10.1016/j.neurobiolaging.2021.07.014>.
- [35] Kolodkin A, Sharma RP, Colangelo AM, Ignatenko A, Martorana F, Jennen D, et al. ROS networks: designs, aging, Parkinson's disease and precision therapies. *NPJ Syst Biol Appl* 2020;6(1):34. <https://doi.org/10.1038/s41540-020-00150-w>.
- [36] Pignatelli P, Menichelli D, Pastori D, Violi F. Oxidative stress and cardiovascular disease: new insights. *Kardiol Pol* 2018;76(4):713–22. <https://doi.org/10.5603/KP.a2018.0071>.
- [37] Otoupalova E, Smith S, Cheng G, Thannickal VJ. Oxidative stress in pulmonary fibrosis. *Compr Physiol* 2020;10(2):509–47. <https://doi.org/10.1002/cphy.c190017>.
- [38] Tang SP, Mao XL, Chen YH, Yan LL, Ye LP, Li SW. Reactive oxygen species induce fatty liver and ischemia-reperfusion injury by promoting inflammation and cell death. *Front Immunol* 2022;13:870239. <https://doi.org/10.3389/fimmu.2022.870239>.
- [39] Wang L, Tang J, Wang L, Tan F, Song H, Zhou J, et al. Oxidative stress in oocyte aging and female reproduction. *J Cell Physiol* 2021;236(12):7966–83. <https://doi.org/10.1002/jcp.30468>.
- [40] Nago M, Arichi A, Omura N, Iwashita Y, Kawamura T, Yumura Y. Aging increases oxidative stress in semen. *Investig Clin Urol* 2021;62(2):233–8. <https://doi.org/10.4111/icu.20200066>.
- [41] Kay J, Thadhani E, Samson L, Engelward B. Inflammation-induced DNA damage, mutations and cancer. *DNA Repair* 2019;83(102673):102673. <https://doi.org/10.1016/j.dnarep.2019.102673>.
- [42] Koi M, Tseng-Rogenski SS, Carethers JM. Inflammation-associated microsatellite alterations: mechanisms and significance in the prognosis of patients with colorectal cancer. *World J Gastrointest Oncol* 2018;10(1):1–14. <https://doi.org/10.4251/wjgo.v10.i1.1>.
- [43] Neri S, Gardini A, Facchini A, Oliveri F, Franceschi C, Ravaglia G, et al. Mismatch repair system and aging: microsatellite instability in peripheral blood cells from differently aged participants. *J Gerontol A Biol Sci Med Sci* 2005;60(3):285–92. <https://doi.org/10.1093/gerona/60.3.285>.
- [44] Mayer-Hamblett N, Kronmal RA, Gibson RL, Rosenfeld M, Retsch-Bogart G, Treggiari MM, et al. Initial *Pseudomonas aeruginosa* treatment failure is associated with exacerbations in cystic fibrosis. *Pediatr Pulmonol* 2012;47(2):125–34. <https://doi.org/10.1002/ppul.21525>.
- [45] Mechtcheriakova D, Svoboda M, Meshcheryakova A, Jensen-Jarolim E. Activation-induced cytidine deaminase (AID) linking immunity, chronic inflammation, and cancer. *Cancer Immunol Immunother* 2012;61(9):1591–8. <https://doi.org/10.1007/s00262-012-1255-z>.
- [46] Shimizu T, Marusawa H, Endo Y, Chiba T. Inflammation-mediated genomic instability: roles of activation-induced cytidine deaminase in carcinogenesis. *Cancer Sci* 2012;103(7):1201–6. <https://doi.org/10.1111/j.1349-7006.2012.02293.x>.
- [47] Çakan E, Gunaydin G. Activation induced cytidine deaminase: an old friend with new faces. *Front Immunol* 2022;13:965312. <https://doi.org/10.3389/fimmu.2022.965312>.
- [48] Qiao Y, Han X, Guan G, Wu N, Sun J, Pak V, et al. TGF- β triggers HBV cccDNA degradation through AID-dependent deamination. *FEBS Lett* 2016;590(3):419–27. <https://doi.org/10.1002/1873-3468.12058>.
- [49] Chiba T, Marusawa H, Ushijima T. Inflammation-associated cancer development in digestive organs: mechanisms and roles for genetic and epigenetic modulation. *Gastroenterology* 2012;143(3):550–63. <https://doi.org/10.1053/j.gastro.2012.07.009>.
- [50] Gushima M, Hirahashi M, Matsumoto T, Fujita K, Ohuchida K, Oda Y, et al. Expression of activation-induced cytidine deaminase in ulcerative colitis-associated carcinogenesis. *Histopathology* 2011;59(3):460–9. <https://doi.org/10.1111/j.1365-2559.2011.03965.x>.
- [51] Sawai Y, Kodama Y, Shimizu T, Ota Y, Maruna T, Eso Y, et al. Activation-induced cytidine deaminase contributes to pancreatic tumorigenesis by inducing tumor-related gene mutations. *Cancer Res* 2015;75(16):3292–301. <https://doi.org/10.1158/0008-5472.CAN-14-3028>.
- [52] Switzer CH, Cho HJ, Eykyn TR, Lavender P, Eaton P. NOS2 and S-nitrosothiol signaling induces DNA hypomethylation and LINE-1 retrotransposon expression. *Proc Natl Acad Sci U S A* 2022;119(21):e2200022119. <https://doi.org/10.1073/pnas.2200022119>.
- [53] De Cecco M, Ito T, Petrashen AP, Elias AE, Skvir NJ, Criscione SW, et al. L1 drives IFN in senescent cells and promotes age-associated inflammation. *Nature* 2019;566(7742):73–8. <https://doi.org/10.1038/s41586-018-0784-9>.
- [54] Bellon M, Nicot C. Telomere dynamics in immune senescence and exhaustion triggered by chronic viral infection. *Viruses* 2017;9(10). <https://doi.org/10.3390/v9100289>.
- [55] Hayflick L. The limited in vitro lifetime of human diploid cell strains. *Exp Cell Res* 1965;37(3):614–36. [https://doi.org/10.1016/0014-4827\(65\)90211-9](https://doi.org/10.1016/0014-4827(65)90211-9).
- [56] Sławińska N, Krupa R. Molecular aspects of senescence and organismal ageing-DNA damage response, telomeres, inflammation and chromatin. *Int J Mol Sci* 2021;22(2):590. <https://doi.org/10.3390/ijms22020590>.
- [57] Rossiello F, Jurk D, Passos JF, d'Adda di Fagagna F. Telomere dysfunction in ageing and age-related diseases. *Nat Cell Biol* 2022;24(2):135–47. <https://doi.org/10.1038/s41556-022-00842-x>.
- [58] Lagnado A, Leslie J, Ruchaud-Sparagano MH, Victorelli S, Hirsova P, Ogrodnik M, et al. Neutrophils induce paracrine telomere dysfunction and senescence in ROS-dependent manner. *EMBO J* 2021;40(9):e106048. <https://doi.org/10.15252/embj.202106048>.
- [59] Bloom SI, Tucker JR, Lim J, Thomas TJ, Stoddard GJ, Lesniewski LA, et al. Aging results in DNA damage and telomere dysfunction that is greater in endothelial versus vascular smooth muscle cells and is exacerbated in atheroprone regions. *GeroScience* 2022;44(6):2741–55. <https://doi.org/10.1007/s11357-022-00681-6>.
- [60] Wang J, Thomas HR, Li Z, Yeo NCF, Scott HE, Dang N, et al. Puma, noxa, p53, and p63 differentially mediate stress pathway induced apoptosis. *Cell Death Dis* 2021;12(7):659. <https://doi.org/10.1038/s41419-021-03902-6>.
- [61] Kumari R, Jat P. Mechanisms of cellular senescence: cell cycle arrest and senescence associated secretory phenotype. *Front Cell Dev Biol* 2021;9:645593. <https://doi.org/10.3389/fcell.2021.645593>.
- [62] Chakravarti D, LaBella KA, DePinho RA. Telomeres: history, health, and hallmarks of aging. *Cell* 2021;184(2):306–22. <https://doi.org/10.1016/j.cell.2020.12.028>.
- [63] Maciejowski J, de Lange T. Telomeres in cancer: tumour suppression and genome instability. *Nat Rev Mol Cell Biol* 2017;18(3):175–86. <https://doi.org/10.1038/nrm.2016.171>.
- [64] Nassour J, Aguiar LG, Correia A, Schmidt TT, Mainz L, Przetocka S, et al. Telomere-to-mitochondria signalling by ZBP1 mediates replicative crisis. *Nature* 2023;614(7949):767–73. <https://doi.org/10.1038/s41586-023-05710-8>.

- [65] Amano H, Chaudhury A, Rodriguez-Aguayo C, Lu L, Akhanov V, Catic A, et al. Telomere dysfunction induces sirtuin repression that drives telomere-dependent disease. *Cell Metabol* 2019;29(6):1274–90. <https://doi.org/10.1016/j.cmet.2019.03.001>. e9.
- [66] Amano H, Sahin E. Telomeres and sirtuins: at the end we meet again. *Mol Cell Oncol* 2019;6(5):e1632613. <https://doi.org/10.1080/23723556.2019.1632613>.
- [67] Heba AC, Toupance S, Arnone D, Peyrin-Biroulet L, Benetos A, Ndiaye NC. Telomeres: new players in immune-mediated inflammatory diseases? *J Autoimmun* 2021;123(102699):102699. <https://doi.org/10.1016/j.jaut.2021.102699>.
- [68] Van Der Stukken C, Nawrot TS, Alfano R, Wang C, Langie SAS, Plusquin M, et al. The telomere-mitochondrial axis of aging in newborns. *Aging (Albany NY)* 2022;14(4):1627–50. <https://doi.org/10.18632/aging.203897>.
- [69] Kang Y, Zhang H, Zhao Y, Wang Y, Wang W, He Y, et al. Telomere dysfunction disturbs macrophage mitochondrial metabolism and the NLRP3 inflammasome through the PGC-1 α /TNFAIP3 axis. *Cell Rep* 2018;22(13):3493–506. <https://doi.org/10.1016/j.celrep.2018.02.071>.
- [70] Nassour J, Radford R, Correia A, Fuste JM, Schoelle B, Jauch A, et al. Autophagic cell death restricts chromosomal instability during replicative crisis. *Nature* 2019;565(7741):659–63. <https://doi.org/10.1038/s41586-019-0885-0>.
- [71] Mender I, Zhang A, Ren Z, Han C, Deng Y, Siteni S, et al. Telomere stress potentiates STING-dependent anti-tumor immunity. *Cancer Cell* 2020;38(3):400–11. <https://doi.org/10.1016/j.ccell.2020.05.020.e6>.
- [72] Mittelbrunn M, Kroemer G. Hallmarks of T cell aging. *Nat Immunol* 2021;22(6):687–98. <https://doi.org/10.1038/s41590-021-00927-z>.
- [73] Laphanuwat P, Gomes DCO, Akbar AN. Senescent T cells: beneficial and detrimental roles. *Immunol Rev* 2023. <https://doi.org/10.1111/imr.13206>. Published online 25 April.
- [74] Marrella V, Facoetti A, Cassani B. Cellular senescence in immunity against infections. *Int J Mol Sci* 2022;23(19):11845. <https://doi.org/10.3390/ijms231911845>.
- [75] Matthe DM, Thoma OM, Sperka T, Neurath MF, Waldner MJ. Telomerase deficiency reflects age-associated changes in CD4⁺ T cells. *Immun Ageing* 2022;19(1):16. <https://doi.org/10.1186/s12979-022-00273-0>.
- [76] Ruiz A, Flores-Gonzalez J, Buendia-Roldan I, Chavez-Galan L. Telomere shortening and its association with cell dysfunction in lung diseases. *Int J Mol Sci* 2021;23(1):425. <https://doi.org/10.3390/ijms23010425>.
- [77] Aviv A. The bullwhip effect, T-cell telomeres, and SARS-CoV-2. *Lancet Healthy Longev* 2022;3(10):e715–21. [https://doi.org/10.1016/S2666-7568\(22\)00190-8](https://doi.org/10.1016/S2666-7568(22)00190-8).
- [78] Lanna A, Vaz B, D'Ambra C, Valvo S, Vuotto C, Chiruchiù V, et al. An intercellular transfer of telomeres rescues T cells from senescence and promotes long-term immunological memory. *Nat Cell Biol* 2022;24(10):1461–74. <https://doi.org/10.1038/s41556-022-00991-z>.
- [79] Tahara T, Shibata T, Kawamura T, Ishizuka T, Okubo M, Nagasaka M, et al. Telomere length in non-neoplastic gastric mucosa and its relationship to *H. pylori* infection, degree of gastritis, and NSAID use. *Clin Exp Med* 2016;16(1):65–71. <https://doi.org/10.1007/s10238-014-0335-0>.
- [80] Laish I, Mannasse-Green B, Hadary R, Biron-Shental T, Konikoff FM, Amiel A, et al. Telomere dysfunction in nonalcoholic fatty liver disease and cryptogenic cirrhosis. *Cytogenet Genome Res* 2016;150(2):93–9. <https://doi.org/10.1159/000454654>.
- [81] Lim HF, Tan NS, Dehghan R, Shen M, Liew MF, Bee SWL, et al. Shortened telomere length in sputum cells of bronchiectasis patients is associated with dysfunctional inflammatory pathways. *Lung* 2022;200(3):401–7. <https://doi.org/10.1007/s00408-022-00535-0>.
- [82] Slusher AL, Zúñiga TM, Acevedo EO. Inflamm-aging is associated with lower plasma PTX3 concentrations and an impaired capacity of PBMCs to express hTERT following LPS stimulation. *Mediat Inflamm* 2019;2019:2324193. <https://doi.org/10.1155/2019/2324193>.
- [83] Chung SS, Wu Y, Okobi Q, Adekoya D, Atefi M, Clarke O, et al. Proinflammatory cytokines IL-6 and TNF- α increased telomerase activity through NF- κ B/STAT1/STAT3 activation, and withaferin A inhibited the signaling in colorectal cancer cells. *Mediat Inflamm* 2017;2017:5958429. <https://doi.org/10.1155/2017/5958429>.
- [84] Maekawa T, Liu B, Nakai D, Yoshida K, Nakamura KI, Yasukawa M, et al. ATF7 mediates TNF- α -induced telomere shortening. *Nucleic Acids Res* 2018;46(9):4487–504. <https://doi.org/10.1093/nar/gky155>.
- [85] Conte M, Martucci M, Chiariello A, Franceschi C, Salvio S. Mitochondria, immunosenescence and inflammaging: a role for mitokines? *Semin Immunopathol* 2020;42(5):607–17. <https://doi.org/10.1007/s00281-020-00813-0>.
- [86] Jacome Burbano MS, Gilson E. The power of stress: the Telo-hormesis hypothesis. *Cells* 2021;10(5):1156. <https://doi.org/10.3390/cells10051156>.
- [87] Covarrubias AJ, Perrone R, Grozio A, Verdin E. NAD⁺ metabolism and its roles in cellular processes during ageing. *Nat Rev Mol Cell Biol* 2021;22(2):119–41. <https://doi.org/10.1038/s41580-020-00313-x>.
- [88] Zeidler JD, Kashyap S, Hogan KA, Chini EN. Implications of the NADase CD38 in COVID pathophysiology. *Physiol Rev* 2022;102(1):339–41. <https://doi.org/10.1152/physrev.00007.2021>.
- [89] Zeidler JD, Hogan KA, Agorrodoy G, Peclat TR, Kashyap S, Kanamori KS, et al. The CD38 glycohydrolase and the NAD sink: implications for pathological conditions. *Am J Physiol Cell Physiol* 2022;322(3):C521–45. <https://doi.org/10.1152/ajpcell.00451.2021>.
- [90] Guo Z, Li P, Ge J, Li H. SIRT6 in aging, metabolism, inflammation and cardiovascular diseases. *Aging Dis* 2022;13(6):1787–822. <https://doi.org/10.14336/AD.2022.0413>.
- [91] Abbas AA, Akbar AN. Induction of T cell senescence by cytokine induced bystander activation. *Front Aging* 2021;2:714239. <https://doi.org/10.3389/fragi.2021.714239>.
- [92] Lanna A, Coutavas E, Levati L, Seidel J, Rustin MH, Henson SM, et al. IFN- α inhibits telomerase in human CD8⁺ T cells by both hTERT downregulation and induction of p38 MAPK signaling. *J Immunol* 2013;191(7):3744–52. <https://doi.org/10.4049/jimmunol.1301409>.
- [93] Reed JR, Vukmanovic-Stejic M, Fletcher JM, Soares MV, Cook JE, Orteu CH, et al. Telomere erosion in memory T cells induced by telomerase inhibition at the site of antigenic challenge in vivo. *J Exp Med* 2004;199(10):1433–43. <https://doi.org/10.1084/jem.20040178>.
- [94] Tichy ED, Ma N, Sidibe D, Loro E, Kocan J, Chen DZ, et al. Persistent NF- κ B activation in muscle stem cells induces proliferation-independent telomere shortening. *Cell Rep* 2021;35(6):109098. <https://doi.org/10.1016/j.celrep.2021.109098>.
- [95] Jurk D, Wilson C, Passos JF, Oakley F, Correia-Melo C, Greaves L, et al. Chronic inflammation induces telomere dysfunction and accelerates ageing in mice. *Nat Commun* 2014;2(1):4172. <https://doi.org/10.1038/ncomms5172>.
- [96] Nardini C, Moreau JF, Gensous N, Ravaoli F, Garagnani P, Bacalini MG. The epigenetics of inflammaging: the contribution of age-related heterochromatin loss and locus-specific remodelling and the modulation by environmental stimuli. *Semin Immunol* 2018;40:49–60. <https://doi.org/10.1016/j.smim.2018.10.009>.
- [97] Gorbunova V, Seluanov A, Mita P, McKerron W, Fenyó D, Boeke JD, et al. The role of retrotransposable elements in ageing and age-associated diseases. *Nature* 2021;596(7870):43–53. <https://doi.org/10.1038/s41586-021-03542-y>.
- [98] Zhang L, Lu Q, Chang C. Epigenetics in health and disease. *Adv Exp Med Biol* 2020;1253:3–55. https://doi.org/10.1007/978-981-15-3449-2_1.
- [99] Zhang J, Wang S, Liu B. New insights into the genetics and epigenetics of aging plasticity. *Genes* 2023;14(2):329. <https://doi.org/10.3390/genes14020329>.

- [100] Saul D, Kosinsky RL. Epigenetics of aging and aging-associated diseases. *Int J Mol Sci* 2021;22(1):401. <https://doi.org/10.3390/ijms22010401>.
- [101] Sarkar A, Liu NQ, Magallanes J, Tassej J, Lee S, Shkhyan R, et al. STAT3 promotes a youthful epigenetic state in articular chondrocytes. *Aging Cell* 2023;22(2):e13773. <https://doi.org/10.1111/accel.13773>.
- [102] Teo YV, Capri M, Morsiani C, Pizza G, Faria AMC, Franceschi C, et al. Cell-free DNA as a biomarker of aging. *Aging Cell* 2019;18(1):e12890. <https://doi.org/10.1111/accel.12890>.
- [103] Urban LA, Trinh A, Pearlman E, Siryaporn A, Downing TL. The impact of age-related hypomethylated DNA on immune signaling upon cellular demise. *Trends Immunol* 2021;42(6):464–8. <https://doi.org/10.1016/j.it.2021.04.005>.
- [104] Wielscher M, Mandaviya PR, Kuehnel B, Joehanes R, Mustafa R, Robinson O, et al. DNA methylation signature of chronic low-grade inflammation and its role in cardio-respiratory diseases. *Nat Commun* 2022;13(1):2408. <https://doi.org/10.1038/s41467-022-29792-6>.
- [105] Zhao J, Huai J. Role of primary aging hallmarks in Alzheimer's disease. *Theranostics* 2023;13(1):197–230. <https://doi.org/10.7150/thno.79535>.
- [106] Khalil H, Tazi M, Cauton K, Ahmed A, Kanneganti A, Assani K, et al. Aging is associated with hypermethylation of autophagy genes in macrophages. *Epigenetics* 2016;11(5):381–8. <https://doi.org/10.1080/15592294.2016.1144007>.
- [107] Dyer ML, Khouja JN, Jackson AR, Havill MA, Dockrell MJ, Munafò MR, et al. Effects of electronic cigarette e-liquid flavouring on cigarette craving. *Tobac Control* 2023;32(e1):e3–9. <https://doi.org/10.1136/tobaccocontrol-2021-056769>.
- [108] Issa JP, Ahuja N, Toyota M, Bronner MP, Brentnall TA. Accelerated age-related CpG island methylation in ulcerative colitis. *Cancer Res* 2001;61(9):3573–7. <https://www.ncbi.nlm.nih.gov/pubmed/11325821>.
- [109] Niwa T, Tsukamoto T, Toyoda T, Mori A, Tanaka H, Maekita T, et al. Inflammatory processes triggered by *Helicobacter pylori* infection cause aberrant DNA methylation in gastric epithelial cells. *Cancer Res* 2010;70(4):1430–40. <https://doi.org/10.1158/0008-5472.CAN-09-2755>.
- [110] Nishida N, Nagasaka T, Nishimura T, Ikai I, Boland CR, Goel A. Aberrant methylation of multiple tumor suppressor genes in aging liver, chronic hepatitis, and hepatocellular carcinoma. *Hepatology* 2008;47(3):908–18. <https://doi.org/10.1002/hep.22110>.
- [111] Vakoc CR, Sachdeva MM, Wang H, Blobel GA. Profile of histone lysine methylation across transcribed mammalian chromatin. *Mol Cell Biol* 2006;26(24):9185–95. <https://doi.org/10.1128/MCB.01529-06>.
- [112] Yang N, O'cean JR, Melters DP, Shi C, Wang L, Stransky S, et al. A hyper-quiet chromatin state formed during aging is reversed by regeneration. *bioRxiv* 2023. <https://doi.org/10.1101/2023.02.14.528512>. Published online 15 February.
- [113] Hyun K, Jeon J, Park K, Kim J. Writing, erasing and reading histone lysine methylations. *Exp Mol Med* 2017;49(4):e324. <https://doi.org/10.1038/emmm.2017.11>.
- [114] Zhu X, Chen Z, Shen W, Huang G, Sedivy JM, Wang H, et al. Inflammation, epigenetics, and metabolism converge to cell senescence and ageing: the regulation and intervention. *Signal Transduct Targeted Ther* 2021;6(1):245. <https://doi.org/10.1038/s41392-021-00646-9>.
- [115] Soriano-Cantón R, Perez-Villalba A, Morante-Redolat JM, Marqués-Torrejón MÁ, Pallás M, Pérez-Sánchez F, et al. Regulation of the p19(Arf)/p53 pathway by histone acetylation underlies neural stem cell behavior in senescence-prone SAMP8 mice. *Aging Cell* 2015;14(3):453–62. <https://doi.org/10.1111/accel.12328>.
- [116] Sen P, Lan Y, Li CY, Sidoli S, Donahue G, Dou Z, et al. Histone acetyltransferase p300 induces DE Novo super-enhancers to drive cellular senescence. *Mol Cell* 2019;73(4):684–98. <https://doi.org/10.1016/j.molcel.2019.01.021>. e8.
- [117] Andrenacci D, Cavaliere V, Lattanzi G. The role of transposable elements activity in aging and their possible involvement in laminopathic diseases. *Ageing Res Rev* 2020;57(100995):100995. <https://doi.org/10.1016/j.arr.2019.100995>.
- [118] Pappalardo XG, Barra V. Losing DNA methylation at repetitive elements and breaking bad. *Epigenet Chromatin* 2021;14(1):25. <https://doi.org/10.1186/s13072-021-00400-z>.
- [119] Yushkova E, Moskalev A. Transposable elements and their role in aging. *Ageing Res Rev* 2023;86:101881. <https://doi.org/10.1016/j.arr.2023.101881>.
- [120] Simon M, Van Meter M, Ablaeva J, Ke Z, Gonzalez RS, Taguchi T, et al. LINE1 derepression in aged wild-type and SIRT6-deficient mice drives inflammation. *Cell Metabol* 2019;29(4):871–85. <https://doi.org/10.1016/j.cmet.2019.02.014>. e5.
- [121] Geis FK, Goff SP. Silencing and transcriptional regulation of endogenous retroviruses: an overview. *Viruses* 2020;12(8):884. <https://doi.org/10.3390/v12080884>.
- [122] Wang K, Liu H, Hu Q, Wang L, Liu J, Zheng Z, et al. Epigenetic regulation of aging: implications for interventions of aging and diseases. *Signal Transduct Targeted Ther* 2022;7(1):374. <https://doi.org/10.1038/s41392-022-01211-8>.
- [123] Lee MY, Lee J, Hyeon SJ, Cho H, Hwang YJ, Shin JY, et al. Epigenome signatures landscaped by histone H3K9me3 are associated with the synaptic dysfunction in Alzheimer's disease. *Aging Cell* 2020;19(6):e13153. <https://doi.org/10.1111/accel.13153>.
- [124] Dembny P, Newman AG, Singh M, Hinz M, Szczepek M, Krüger C, et al. Human endogenous retrovirus HERV-K(HML-2) RNA causes neurodegeneration through Toll-like receptors. *JCI Insight* 2020;5(7). <https://doi.org/10.1172/jci.insight.131093>.
- [125] Liu X, Liu Z, Wu Z, Ren J, Fan Y, Sun L, et al. Resurrection of endogenous retroviruses during aging reinforces senescence. *Cell* 2023;186(2):287–304. <https://doi.org/10.1016/j.cell.2022.12.017>. e26.
- [126] Kinser HE, Pincus Z. MicroRNAs as modulators of longevity and the aging process. *Hum Genet* 2020;139(3):291–308. <https://doi.org/10.1007/s00439-019-02046-0>.
- [127] Olivieri F, Praticchizzo F, Giuliani A, Maticchione G, Rippon MR, Sabbatinelli J, et al. miR-21 and miR-146a: the microRNAs of inflammaging and age-related diseases. *Ageing Res Rev* 2021;70(101374):101374. <https://doi.org/10.1016/j.arr.2021.101374>.
- [128] Zhang HQ, Wang JY, Li ZF, Cui L, Huang SS, Zhu LB, et al. DNA methyltransferase 1 is dysregulated in Parkinson's disease via mediation of miR-17. *Mol Neurobiol* 2021;58(6):2620–33. <https://doi.org/10.1007/s12035-021-02298-w>.
- [129] Wang X, Zhou Y, Gao Q, Ping D, Wang Y, Wu W, et al. The role of exosomal microRNAs and oxidative stress in neurodegenerative diseases. *Oxid Med Cell Longev* 2020;2020:3232869. <https://doi.org/10.1155/2020/3232869>.
- [130] Jusic A, Thomas PB, Wettinger SB, Dogan S, Farrugia R, Gaetano C, et al. Noncoding RNAs in age-related cardiovascular diseases. *Ageing Res Rev* 2022;77(101610):101610. <https://doi.org/10.1016/j.arr.2022.101610>.
- [131] Zheng YL, Song G, Guo JB, Su X, Chen YM, Yang Z, et al. Interactions among lncRNA/circRNA, miRNA, and mRNA in musculoskeletal degenerative diseases. *Front Cell Dev Biol* 2021;9:753931. <https://doi.org/10.3389/fcell.2021.753931>.
- [132] Li C, Ni YQ, Xu H, Xiang QY, Zhao Y, Zhan JK, et al. Roles and mechanisms of exosomal non-coding RNAs in human health and diseases. *Signal Transduct Targeted Ther* 2021;6(1):383. <https://doi.org/10.1038/s41392-021-00779-x>.
- [133] Colella M, Cuomo D, Peluso T, Falanga I, Mallardo M, De Felice M, et al. Ovarian aging: role of pituitary-ovarian axis hormones and ncRNAs in regulating ovarian mitochondrial activity. *Front Endocrinol* 2021;12:791071. <https://doi.org/10.3389/fendo.2021.791071>.
- [134] Frankowska N, Lisowska K, Witkowski JM. Proteolysis dysfunction in the process of aging and age-related diseases. *Front Aging* 2022;3:927630. <https://doi.org/10.3389/fragi.2022.927630>.
- [135] Rai M, Curley M, Coleman Z, Demontis F. Contribution of proteases to the hallmarks of aging and to age-related neurodegeneration. *Aging Cell* 2022;21(5):e13603. <https://doi.org/10.1111/accel.13603>.

- [136] Miles J, Scherz-Shouval R, van Oosten-Hawle P. Expanding the organismal proteostasis network: linking systemic stress signaling with the innate immune response. *Trends Biochem Sci* 2019;44(11):927–42. <https://doi.org/10.1016/j.tibs.2019.06.009>.
- [137] Ruano D. Proteostasis dysfunction in aged mammalian cells. The stressful role of inflammation. *Front Mol Biosci* 2021;8:658742. <https://doi.org/10.3389/fmolb.2021.658742>.
- [138] Franceschi C, Garagnani P, Vitale G, Capri M, Salvioli S. Inflammaging and 'garb-aging'. *Trends Endocrinol Metabol* 2017;28(3):199–212. <https://doi.org/10.1016/j.tem.2016.09.005>.
- [139] Cuanalo-Contreras K, Schulz J, Mukherjee A, Park KW, Armijo E, Soto C. Extensive accumulation of misfolded protein aggregates during natural aging and senescence. *Front Aging Neurosci* 2022;14:1090109. <https://doi.org/10.3389/fnagi.2022.1090109>.
- [140] Chipurupalli S, Samavedam U, Robinson N. Crosstalk between ER stress, autophagy and inflammation. *Front Med* 2021;8:758311. <https://doi.org/10.3389/fmed.2021.758311>.
- [141] Salminen A, Kaarniranta K, Kauppinen A. ER stress activates immunosuppressive network: implications for aging and Alzheimer's disease. *J Mol Med* 2020;98(5):633–50. <https://doi.org/10.1007/s00109-020-01904-z>.
- [142] Di Conza G, Ho PC, Cubillos-Ruiz JR, Huang SCC. Control of immune cell function by the unfolded protein response. *Nat Rev Immunol* 2023. <https://doi.org/10.1038/s41577-023-00838-0>. Published online 8 February.
- [143] Li D, Wu M. Pattern recognition receptors in health and diseases. *Signal Transduct Targeted Ther* 2021;6(1):291. <https://doi.org/10.1038/s41392-021-00687-0>.
- [144] Tan Y, Chen Q, Li X, Zeng Z, Xiong W, Li G, et al. Pyroptosis: a new paradigm of cell death for fighting against cancer. *J Exp Clin Cancer Res* 2021;40(1):153. <https://doi.org/10.1186/s13046-021-01959-x>.
- [145] Tabibzadeh S. Cell-centric hypotheses of aging. *Front Biosci* 2021;26(1):1–49. <https://doi.org/10.2741/4888>.
- [146] Kulkarni A, Preeti K, Tryphena KP, Srivastava S, Singh SB, Khatri DK. Proteostasis in Parkinson's disease: recent development and possible implication in diagnosis and therapeutics. *Ageing Res Rev* 2023;84(101816):101816. <https://doi.org/10.1016/j.arr.2022.101816>.
- [147] Abbadie C, Pluquet A. Unfolded protein response (UPR) controls major senescence hallmarks. *Trends Biochem Sci* 2020;45(5):371–4. <https://doi.org/10.1016/j.tibs.2020.02.005>.
- [148] Goetzke CC, Ebstein F, Kallinich T. Role of proteasomes in inflammation. *J Clin Med* 2021;10(8):1783. <https://doi.org/10.3390/jcm10081783>.
- [149] Arata Y, Watanabe A, Motosugi R, Murakami R, Goto T, Hori S, et al. Defective induction of the proteasome associated with T-cell receptor signaling underlies T-cell senescence. *Gene Cell* 2019;24(12):801–13. <https://doi.org/10.1111/gtc.12728>.
- [150] Xu F, Zhang C, Zou Z, Fan EKY, Chen L, Li Y, et al. Aging-related Atg5 defect impairs neutrophil extracellular traps formation. *Immunology* 2017;151(4):417–32. <https://doi.org/10.1111/imm.12740>.
- [151] Colombini B, Dinu M, Murgo E, Lotti S, Tarquini R, Sofi F, et al. Ageing and low-level chronic inflammation: the role of the biological clock. *Antioxidants* 2022;11(11). <https://doi.org/10.3390/antiox11112228>.
- [152] Zuo L, Prather ER, Stetskiy M, Garrison DE, Meade JR, Peace TI, et al. Inflammaging and oxidative stress in human diseases: from molecular mechanisms to novel treatments. *Int J Mol Sci* 2019;20(18):4472. <https://doi.org/10.3390/ijms20184472>.
- [153] Lévy E, El Banna N, Baille D, Heneman-Masurel A, Truchet S, Rezaei H, et al. Causative links between protein aggregation and oxidative stress: a review. *Int J Mol Sci* 2019;20(16):3896. <https://doi.org/10.3390/ijms20163896>.
- [154] Fani G, La Torre CE, Cascella R, Cecchi C, Vendruscolo M, Chiti F. Misfolded protein oligomers induce an increase of intracellular Ca²⁺ causing an escalation of reactive oxidative species. *Cell Mol Life Sci* 2022;79(9):500. <https://doi.org/10.1007/s00018-022-04513-w>.
- [155] Martínez de Toda I, Ceprián N, Díaz-Del Cerro E, De la Fuente M. The role of immune cells in oxi-inflamm-aging. *Cells* 2021;10(11):2974. <https://doi.org/10.3390/cells10112974>.
- [156] Li WW, Wang HJ, Tan YZ, Wang YL, Yu SN, Li ZH. Reducing lipofuscin accumulation and cardiomyocytic senescence of aging heart by enhancing autophagy. *Exp Cell Res* 2021;403(1):112585. <https://doi.org/10.1016/j.yexcr.2021.112585>.
- [157] Tubío-Santamaría N, Ebstein F, Heidel FH, Krüger E. Immunoproteasome function in normal and malignant hematopoiesis. *Cells* 2021;10(7):1577. <https://doi.org/10.3390/cells10071577>.
- [158] Papendorf JJ, Krüger E, Ebstein F. Proteostasis perturbations and their roles in causing sterile inflammation and autoinflammatory diseases. *Cells* 2022;11(9):1422. <https://doi.org/10.3390/cells11091422>.
- [159] Sarrabay G, Méchin D, Salhi A, Boursier G, Rittore C, Crow Y, et al. PSMB10, the last immunoproteasome gene missing for PRAAS. *J Allergy Clin Immunol* 2020;145(3):1015–7. <https://doi.org/10.1016/j.jaci.2019.11.024>. e6.
- [160] Pickering AM, Lehr M, Miller RA. Lifespan of mice and primates correlates with immunoproteasome expression. *J Clin Invest* 2015;125(5):2059–68. <https://doi.org/10.1172/JCI80514>.
- [161] Tripodi L, Molinaro D, Fortunato F, Mella C, Cassani B, Torrente Y, et al. Immunoproteasome inhibition ameliorates aged dystrophic mouse muscle environment. *Int J Mol Sci* 2022;23(23):14657. <https://doi.org/10.3390/ijms232314657>.
- [162] Levine B, Kroemer G. Biological functions of autophagy genes: a disease perspective. *Cell* 2019;176(1–2):11–42. <https://doi.org/10.1016/j.cell.2018.09.048>.
- [163] Lőrincz P, Juhász G. Autophagosome-lysosome fusion. *J Mol Biol* 2020;432(8):2462–82. <https://doi.org/10.1016/j.jmb.2019.10.028>.
- [164] Tabibzadeh S. Role of autophagy in aging: the good, the bad, and the ugly. *Aging Cell* 2023;22(1):e13753. <https://doi.org/10.1111/ace1.13753>.
- [165] Aman Y, Schmauck-Medina T, Hansen M, Morimoto RI, Simon AK, Bjedov I, et al. Autophagy in healthy aging and disease. *Nat Aging* 2021;1(8):634–50. <https://doi.org/10.1038/s43587-021-00098-4>.
- [166] Deretic V, Levine B. Autophagy balances inflammation in innate immunity. *Autophagy* 2018;14(2):243–51. <https://doi.org/10.1080/15548627.2017.1402992>.
- [167] Noda NN, Wang Z, Zhang H. Liquid-liquid phase separation in autophagy. *J Cell Biol* 2020;219(8). <https://doi.org/10.1083/jcb.202004062>.
- [168] Rubinsztein DC, Bento CF, Deretic V. Therapeutic targeting of autophagy in neurodegenerative and infectious diseases. *J Exp Med* 2015;212(7):979–90. <https://doi.org/10.1084/jem.20150956>.
- [169] Wang L, Cai J, Zhao X, Ma L, Zeng P, Zhou L, et al. Palmitoylation prevents sustained inflammation by limiting NLRP3 inflammasome activation through chaperone-mediated autophagy. *Mol Cell* 2023;83(2):281–97. <https://doi.org/10.1016/j.molcel.2022.12.002>. e10.
- [170] Qiao L, Ma J, Zhang Z, Sui W, Zhai C, Xu D, et al. Deficient chaperone-mediated autophagy promotes inflammation and atherosclerosis. *Circ Res* 2021;129(12):1141–57. <https://doi.org/10.1161/CIRCRESAHA.121.318908>.
- [171] Bakula D, Scheibye-Knudsen M. Mitophagy: mitophagy in aging and disease. *Front Cell Dev Biol* 2020;8:239. <https://doi.org/10.3389/fcell.2020.00239>.
- [172] Bektas A, Schurman SH, Gonzalez-Freire M, Dunn CA, Singh AK, Macian F, et al. Age-associated changes in human CD4+ T cells point to mitochondrial dysfunction consequent to impaired autophagy. *Aging (Albany NY)* 2019;11(21):9234–63. <https://doi.org/10.18632/aging.102438>.
- [173] Zhang H, Alsaleh G, Feltham J, Sun Y, Napolitano G, Riffelmacher T, et al. Polyamines control eIF5A hypusination, TFE3 translation, and autophagy to reverse B cell senescence. *Mol Cell* 2019;76(1):110–25. <https://doi.org/10.1016/j.molcel.2019.08.005>. e9.
- [174] Abdullah Marzoug B. Autophagy as an anti-senescent in aging neurocytes. *Curr Mol Med* 2023;23. <https://doi.org/10.2174/1566524023666230120102718>.

- [175] Carosi JM, Fourrier C, Bensalem J, Sargeant TJ. The mTOR-lysosome axis at the centre of ageing. *FEBS Open Bio* 2022;12(4):739–57. <https://doi.org/10.1002/2211-5463.13347>.
- [176] Liu JK. Antiaging agents: safe interventions to slow aging and healthy life span extension. *Nat Products Bioprospect* 2022;12(1):18. <https://doi.org/10.1007/s13659-022-00339-y>.
- [177] Bennett DF, Goyal A, Statzer C, Beckett CW, Tyshkovskiy A, Gladyshev VN, et al. Rilmenidine extends lifespan and healthspan in *Caenorhabditis elegans* via a nischarin I1-imidazoline receptor. *Aging Cell* 2023;22(2):e13774. <https://doi.org/10.1111/acel.13774>.
- [178] Hodzic Kuerec A, Maier AB. Why is rapamycin not a rapalog? *Gerontology* 2023. <https://doi.org/10.1159/000528985>. Published online 7 January.
- [179] Reed R, Miwa S. Cellular senescence and ageing. *Subcell Biochem* 2023;102:139–73. https://doi.org/10.1007/978-3-031-21410-3_7.
- [180] Chaib S, Tchkonja T, Kirkland JL. Cellular senescence and senolytics: the path to the clinic. *Nat Med* 2022;28(8):1556–68. <https://doi.org/10.1038/s41591-022-01923-y>.
- [181] Zhuang XX, Wang SF, Tan Y, Song JX, Zhu Z, Wang ZY, et al. Pharmacological enhancement of TFEB-mediated autophagy alleviated neuronal death in oxidative stress-induced Parkinson's disease models. *Cell Death Dis* 2020;11(2):128. <https://doi.org/10.1038/s41419-020-2322-6>.
- [182] Zhang Z, Yang X, Song YQ, Tu J. Autophagy in Alzheimer's disease pathogenesis: therapeutic potential and future perspectives. *Ageing Res Rev* 2021;72(101464):101464. <https://doi.org/10.1016/j.arr.2021.101464>.
- [183] Croce KR, Yamamoto A. A role for autophagy in Huntington's disease. *Neurobiol Dis* 2019;122:16–22. <https://doi.org/10.1016/j.nbd.2018.08.010>.
- [184] Amin A, Perera ND, Beart PM, Turner BJ, Shabanpoor F. Amyotrophic lateral sclerosis and autophagy: dysfunction and therapeutic targeting. *Cells* 2020;9(11):2413. <https://doi.org/10.3390/cells9112413>.
- [185] Zhang L, Dai L, Li D. Mitophagy in neurological disorders. *J Neuroinflammation* 2021;18(1):297. <https://doi.org/10.1186/s12974-021-02334-5>.
- [186] Zhu R, Luo Y, Li S, Wang Z. The role of microglial autophagy in Parkinson's disease. *Front Aging Neurosci* 2022;14:1039780. <https://doi.org/10.3389/fnagi.2022.1039780>.
- [187] Wang Z, Wang Q, Li S, Li XJ, Yang W, He D. Microglial autophagy in Alzheimer's disease and Parkinson's disease. *Front Aging Neurosci* 2022;14:1065183. <https://doi.org/10.3389/fnagi.2022.1065183>.
- [188] Niu H, Wang Q, Zhao W, Liu J, Wang D, Muhammad B, et al. IL-1 β /IL-1R1 signaling induced by intranasal lipopolysaccharide infusion regulates alpha-Synuclein pathology in the olfactory bulb, substantia nigra and striatum. *Brain Pathol* 2020;30(6):1102–18. <https://doi.org/10.1111/bpa.12886>.
- [189] Lv QK, Tao KX, Wang XB, Yao XY, Pang MZ, Liu JY, et al. Role of α -synuclein in microglia: autophagy and phagocytosis balance neuroinflammation in Parkinson's disease. *Inflamm Res* 2023;72(3):443–62. <https://doi.org/10.1007/s00011-022-01676-x>.
- [190] Karpowicz Jr RJ, Trojanowski JQ, Lee VMY. Transmission of α -synuclein seeds in neurodegenerative disease: recent developments. *Lab Invest* 2019;99(7):971–81. <https://doi.org/10.1038/s41374-019-0195-z>.
- [191] Wang C, Fan L, Khawaja RR, Liu B, Zhan L, Kodama L, et al. Microglial NF- κ B drives tau spreading and toxicity in a mouse model of tauopathy. *Nat Commun* 2022;13(1):1969. <https://doi.org/10.1038/s41467-022-29552-6>.
- [192] López-Otín C, Galluzzi L, Freije JMP, Madeo F, Kroemer G. Metabolic control of longevity. *Cell* 2016;166(4):802–21. <https://doi.org/10.1016/j.cell.2016.07.031>.
- [193] Marmontini C, Soares GM, Bronczek GA, Piovani S, Mareze-Costa CE, Carneiro EM, et al. Aging reduces insulin clearance in mice. *Front Endocrinol* 2021;12:679492. <https://doi.org/10.3389/fendo.2021.679492>.
- [194] Evans JL, Goldfine ID. Aging and insulin resistance: just say iNOS. *Diabetes* 2013;62(2):346–8. <https://doi.org/10.2337/db12-1239>.
- [195] Ormazabal V, Nair S, Efkedy O, Aguayo C, Salomon C, Zúñiga FA. Association between insulin resistance and the development of cardiovascular disease. *Cardiovasc Diabetol* 2018;17(1):122. <https://doi.org/10.1186/s12933-018-0762-4>.
- [196] Ieronymaki E, Theodorakis EM, Lyroni K, Vergadi E, Lagoudaki E, Al-Qahtani A, et al. Insulin resistance in macrophages alters their metabolism and promotes an M2-like phenotype. *J Immunol* 2019;202(6):1786–97. <https://doi.org/10.4049/jimmunol.1800065>.
- [197] Guillén C, Benito M. mTORC1 overactivation as a key aging factor in the progression to type 2 diabetes mellitus. *Front Endocrinol* 2018;9:621. <https://doi.org/10.3389/fendo.2018.00621>. Published 2018 Oct 16.
- [198] Cheng SC, Quintin J, Cramer RA, Shepardson KM, Saeed S, Kumar V, et al. mTOR- and HIF-1 α -mediated aerobic glycolysis as metabolic basis for trained immunity. *Science* 2014;345(6204):1250684. <https://doi.org/10.1126/science.1250684>.
- [199] Viola A, Munari F, Sánchez-Rodríguez R, Scolaro T, Castegna A. The metabolic signature of macrophage responses. *Front Immunol* 2019;10:1462. <https://doi.org/10.3389/fimmu.2019.01462>.
- [200] Jin J, Li X, Hu B, Kim C, Cao W, Zhang H, et al. FOXO1 deficiency impairs proteostasis in aged T cells. *Sci Adv* 2020;6(17):eaba1808. <https://doi.org/10.1126/sciadv.aba1808>.
- [201] Tsai S, Clemente-Casares X, Zhou AC, Lei H, Ahn JJ, Chan YT, et al. Insulin receptor-mediated stimulation boosts T cell immunity during inflammation and infection. *Cell Metabol* 2018;28(6):922–34. <https://doi.org/10.1016/j.cmet.2018.08.003>. e4.
- [202] Makhijani P, Basso PJ, Chan YT, Chen N, Baechele J, Khan S, et al. Regulation of the immune system by the insulin receptor in health and disease. *Front Endocrinol* 2023;14:1128622. <https://doi.org/10.3389/fendo.2023.1128622>.
- [203] Wu D, Wong CK, Han JM, Orban PC, Huang Q, Gillies J, et al. T reg-specific insulin receptor deletion prevents diet-induced and age-associated metabolic syndrome. *J Exp Med* 2020;217(8). <https://doi.org/10.1084/jem.20191542>.
- [204] Ma S, Wang C, Mao X, Hao Y. B cell dysfunction associated with aging and autoimmune diseases. *Front Immunol* 2019;10:318. <https://doi.org/10.3389/fimmu.2019.00318>.
- [205] Phalke S, Rivera-Correa J, Jenkins D, Flores Castro D, Giannopoulou E, Pernis AB. Molecular mechanisms controlling age-associated B cells in autoimmunity. *Immunol Rev* 2022;307(1):79–100. <https://doi.org/10.1111/immr.13068>.
- [206] Li K, Romero M, Cañardo M, García D, Díaz A, Blomberg BB, et al. B cells from old mice induce the generation of inflammatory T cells through metabolic pathways. *Mech Ageing Dev* 2023;209(111742):111742. <https://doi.org/10.1016/j.mad.2022.111742>.
- [207] Frasca D, Romero M, García D, Díaz A, Blomberg BB. Hyper-metabolic B cells in the spleens of old mice make antibodies with autoimmune specificities. *Immun Ageing* 2021;18(1):9. <https://doi.org/10.1186/s12979-021-00222-3>.
- [208] Shirakawa K, Sano M. T cell immunosenescence in aging, obesity, and cardiovascular disease. *Cells* 2021;10(9):2435. <https://doi.org/10.3390/cells10092435>.
- [209] Szwed A, Kim E, Jacinto E. Regulation and metabolic functions of mTORC1 and mTORC2. *Physiol Rev* 2021;101(3):1371–426. <https://doi.org/10.1152/physrev.00026.2020>.
- [210] Laplante M, Sabatini DM. mTOR signaling in growth control and disease. *Cell* 2012;149(2):274–93. <https://doi.org/10.1016/j.cell.2012.03.017>.
- [211] Walters HE, Cox LS. mTORC inhibitors as broad-spectrum therapeutics for age-related diseases. *Int J Mol Sci* 2018;19(8). <https://doi.org/10.3390/ijms19082325>.
- [212] Haythorne E, Lloyd M, Walsby-Tickle J, Tarasov AI, Sandbrink J, Portillo I, et al. Altered glycolysis triggers impaired mitochondrial metabolism and mTORC1 activation in diabetic β -cells. *Nat Commun* 2022;13(1):6754. <https://doi.org/10.1038/s41467-022-34095-x>.

- [213] Chrienova Z, Nepovimova E, Kuca K. The role of mTOR in age-related diseases. *J Enzym Inhib Med Chem* 2021;36(1):1679–93. <https://doi.org/10.1080/14756366.2021.1955873>.
- [214] Mota-Martorell N, Jové M, Pamplona R. mTOR complex 1 content and regulation is adapted to animal longevity. *Int J Mol Sci* 2022;23(15):8747. <https://doi.org/10.3390/ijms23158747>.
- [215] Xu T, Liu J, Li XR, Yu Y, Luo X, Zheng X, et al. The mTOR/NF- κ B pathway mediates neuroinflammation and synaptic plasticity in diabetic encephalopathy. *Mol Neurobiol* 2021;58(8):3848–62. <https://doi.org/10.1007/s12035-021-02390-1>.
- [216] Li Y, Yang L, Dong L, Yang ZW, Zhang J, Zhang SL, et al. Crosstalk between the Akt/mTORC1 and NF- κ B signaling pathways promotes hypoxia-induced pulmonary hypertension by increasing DPP4 expression in PASCs. *Acta Pharmacol Sin* 2019;40(10):1322–33. <https://doi.org/10.1038/s41401-019-0272-2>.
- [217] Liu GY, Sabatini DM. mTOR at the nexus of nutrition, growth, ageing and disease. *Nat Rev Mol Cell Biol* 2020;21(4):183–203. <https://doi.org/10.1038/s41580-019-0199-y>.
- [218] Ham DJ, Börsch A, Chojnowska K, Lin S, Leuchtmann AB, Ham AS, et al. Distinct and additive effects of calorie restriction and rapamycin in aging skeletal muscle. *Nat Commun* 2022;13(1):2025. <https://doi.org/10.1038/s41467-022-29714-6>.
- [219] Correia-Melo C, Birch J, Fielder E, Rahmatika D, Taylor J, Chapman J, et al. Rapamycin improves healthspan but not inflammaging in nfkb1-/- mice. *Aging Cell* 2019;18(1):e12882. <https://doi.org/10.1111/ace1.12882>.
- [220] Collins SL, Oh MH, Sun IH, Chan-Li Y, Zhao L, Powell JD, et al. mTORC1 signaling regulates proinflammatory macrophage function and metabolism. *J Immunol* 2021;207(3):913–22. <https://doi.org/10.4049/jimmunol.2100230>.
- [221] Lesniewski LA, Seals DR, Walker AE, Henson GD, Blimline MW, Trott DW, et al. Dietary rapamycin supplementation reverses age-related vascular dysfunction and oxidative stress, while modulating nutrient-sensing, cell cycle, and senescence pathways. *Aging Cell* 2017;16(1):17–26. <https://doi.org/10.1111/ace1.12524>.
- [222] Pan H, Finkel T. Key proteins and pathways that regulate lifespan. *J Biol Chem* 2017;292(16):6452–60. <https://doi.org/10.1074/jbc.R116.771915>.
- [223] Zhu L, Yang T, Li L, Sun L, Hou Y, Hu X, et al. TSC1 controls macrophage polarization to prevent inflammatory disease. *Nat Commun* 2014;5(1):4696. <https://doi.org/10.1038/ncomms5696>.
- [224] Jiang H, Westertep M, Wang C, Zhu Y, Ai D. Macrophage mTORC1 disruption reduces inflammation and insulin resistance in obese mice. *Diabetologia* 2014;57(11):2393–404. <https://doi.org/10.1007/s00125-014-3350-5>.
- [225] Mannick JB, Morris M, Hockey HUP, Roma G, Beibel M, Kulmatycki K, et al. TORC1 inhibition enhances immune function and reduces infections in the elderly. *Sci Transl Med* 2018;10(449):eaq1564. <https://doi.org/10.1126/scitranslmed.aq1564>.
- [226] Mullen PJ, Garcia Jr G, Purkayastha A, Matulionis N, Schmid EW, Momcilovic M, et al. SARS-CoV-2 infection rewires host cell metabolism and is potentially susceptible to mTORC1 inhibition. *Nat Commun* 2021;12(1):1876. <https://doi.org/10.1038/s41467-021-22166-4>.
- [227] Geier C, Perl A. Therapeutic mTOR blockade in systemic autoimmunity: implications for antiviral immunity and extension of lifespan. *Autoimmun Rev* 2021;20(12):102984. <https://doi.org/10.1016/j.autrev.2021.102984>.
- [228] Moldakozhayev A, Gladyshev VN. Metabolism, homeostasis, and aging. *Trends Endocrinol Metabol* 2023;34(3):158–69. <https://doi.org/10.1016/j.tem.2023.01.003>.
- [229] Sharma A, Chabloz S, Lapidus RA, Roeder E, Ewald CY. Potential synergistic supplementation of NAD⁺ promoting compounds as a strategy for increasing healthspan. *Nutrients* 2023;15(2):445. <https://doi.org/10.3390/nu15020445>.
- [230] Chen Y, Liu Y, Jiang K, Wen Z, Cao X, Wu S. Linear ubiquitination of LKB1 activates AMPK pathway to inhibit NLRP3 inflammasome response and reduce chondrocyte pyroptosis in osteoarthritis. *J Orthop Translat* 2023;39:1–11. <https://doi.org/10.1016/j.jot.2022.11.002>.
- [231] Sadria M, Layton AT. Interactions among mTORC, AMPK and SIRT: a computational model for cell energy balance and metabolism. *Cell Commun Signal* 2021;19(1):57. <https://doi.org/10.1186/s12964-021-00706-1>.
- [232] Kim JK, Silwal P, Jo EK. Sirtuin 1 in host defense during infection. *Cells* 2022;11(18):2921. <https://doi.org/10.3390/cells11182921>.
- [233] Desjardins EM, Smith BK, Day EA, Ducommun S, Sanders MJ, Nederveen JP, et al. The phosphorylation of AMPK β 1 is critical for increasing autophagy and maintaining mitochondrial homeostasis in response to fatty acids. *Proc Natl Acad Sci U S A* 2022;119(48):e2119824119. <https://doi.org/10.1073/pnas.2119824119>.
- [234] Lee IH. Mechanisms and disease implications of sirtuin-mediated autophagic regulation. *Exp Mol Med* 2019;51(9):1–11. <https://doi.org/10.1038/s12276-019-0302-7>.
- [235] Ziętara P, Dziewięcka M, Augustyniak M. Why is longevity still a scientific mystery? Sirtuins—past, present and future. *Int J Mol Sci* 2022;24(1):728. <https://doi.org/10.3390/ijms24010728>.
- [236] Malik N, Ferreira BI, Hollstein PE, Curtis SD, Trefts E, Weiser Novak S, et al. Induction of lysosomal and mitochondrial biogenesis by AMPK phosphorylation of FNIP1. *Science* 2023;380(6642):eabj5559. <https://doi.org/10.1126/science.abj5559>.
- [237] Bharath LP, Agrawal M, McCambridge G, Nicholas DA, Hasturk H, Liu J, et al. Metformin enhances autophagy and normalizes mitochondrial function to alleviate aging-associated inflammation. *Cell Metabol* 2020;32(1):44–55. <https://doi.org/10.1016/j.cmet.2020.04.015>.
- [238] Salminen A, Kaarniranta K. AMP-activated protein kinase (AMPK) controls the aging process via an integrated signaling network. *Ageing Res Rev* 2012;11(2):230–41. <https://doi.org/10.1016/j.arr.2011.12.005>.
- [239] Oka SI, Titus AS, Zablocki D, Sadoshima J. Molecular properties and regulation of NAD⁺ kinase (NADK). *Redox Biol* 2023;59(102561):102561. <https://doi.org/10.1016/j.redox.2022.102561>.
- [240] Zhou H, Liu S, Zhang N, Fang K, Zong J, An Y, et al. Downregulation of Sirt6 by CD38 promotes cell senescence and aging. *Aging (Albany NY)* 2022;14(23):9730–57. <https://doi.org/10.18632/aging.204425>.
- [241] de Candia P, Procaccini C, Russo C, Lepore MT, Matarese G. Regulatory T cells as metabolic sensors. *Immunity* 2022;55(11):1981–92. <https://doi.org/10.1016/j.immuni.2022.10.006>.
- [242] Deng B, Zhang W, Zhu Y, Li Y, Li D, Li B. FOXP3⁺ regulatory T cells and age-related diseases. *FEBS J* 2022;289(2):319–35. <https://doi.org/10.1111/febs.15743>.
- [243] Zhu H, Liu Z, An J, Zhang M, Qiu Y, Zou MH. Activation of AMPK α 1 is essential for regulatory T cell function and autoimmune liver disease prevention. *Cell Mol Immunol* 2021;18(12):2609–17. <https://doi.org/10.1038/s41423-021-00790-w>.
- [244] Phair IR, Nisir RB, Howden AJM, Sovakova M, Alqurashi N, Foretz M, et al. AMPK integrates metabolite and kinase-based immunometabolic control in macrophages. *Mol Metabol* 2023;68(101661):101661. <https://doi.org/10.1016/j.molmet.2022.101661>.
- [245] Yan C, Tian X, Li J, Liu D, Ye D, Xie Z, et al. A high-fat diet attenuates AMPK α 1 in adipocytes to induce exosome shedding and nonalcoholic fatty liver development in vivo. *Diabetes* 2021;70(2):577–88. <https://doi.org/10.2337/db20-0146>.
- [246] Ma H, Guo X, Cui S, Wu Y, Zhang Y, Shen X, et al. Dephosphorylation of AMP-activated protein kinase exacerbates ischemia/reperfusion-induced acute kidney injury via mitochondrial dysfunction. *Kidney Int* 2022;101(2):315–30. <https://doi.org/10.1016/j.kint.2021.10.028>.
- [247] Smirnov D, Eremenko E, Stein D, Kaluski S, Jasinska W, Cosentino C, et al. SIRT6 is a key regulator of mitochondrial function in the brain. *Cell Death Dis* 2023;14(1):35. <https://doi.org/10.1038/s41419-022-05542-w>.

- [248] He H, Xu P, Zhang X, Liao M, Dong Q, Cong T, et al. Aging-induced IL27Ra signaling impairs hematopoietic stem cells. *Blood* 2020;136(2):183–98. <https://doi.org/10.1182/blood.2019003910>.
- [249] Li YQ, Jiao Y, Liu YN, Fu JY, Sun LK, Su J. PGC-1 α protects from myocardial ischaemia-reperfusion injury by regulating mitonuclear communication. *J Cell Mol Med* 2022;26(3):593–600. <https://doi.org/10.1111/jcmm.16236>.
- [250] Nam BY, Jhee JH, Park J, Kim S, Kim G, Park JT, et al. PGC-1 α inhibits the NLRP3 inflammasome via preserving mitochondrial viability to protect kidney fibrosis. *Cell Death Dis* 2022;13(1):31. <https://doi.org/10.1038/s41419-021-04480-3>.
- [251] Fang Y, An N, Zhu L, Gu Y, Qian J, Jiang G, et al. Autophagy-Sirt3 axis decelerates hematopoietic aging. *Aging Cell* 2020;19(10):e13232. <https://doi.org/10.1111/ace1.13232>.
- [252] Salvetrini V, Sell C, Lorenzini A. Obesity may accelerate the aging process. *Front Endocrinol* 2019;10:266. <https://doi.org/10.3389/fendo.2019.00266>.
- [253] Khan S, Chan YT, Revelo XS, Winer DA. The immune landscape of visceral adipose tissue during obesity and aging. *Front Endocrinol* 2020;11:267. <https://doi.org/10.3389/fendo.2020.00267>.
- [254] Song J, Farris D, Ariza P, Moorjani S, Varghese M, Blin M, et al. Age-associated adipose tissue inflammation promotes monocyte chemotaxis and enhances atherosclerosis. *Aging Cell* 2023;22(2):e13783. <https://doi.org/10.1111/ace1.13783>.
- [255] Shou J, Chen PJ, Xiao WH. Mechanism of increased risk of insulin resistance in aging skeletal muscle. *Diabetol Metab Syndr* 2020;12(1):14. <https://doi.org/10.1186/s13098-020-0523-x>.
- [256] Ragonnaud E, Biragyn A. Gut microbiota as the key controllers of 'healthy' aging of elderly people. *Immun Ageing* 2021;18(1):2. <https://doi.org/10.1186/s12979-020-00213-w>.
- [257] Shemtov SJ, Emani R, Bielska O, Covarrubias AJ, Verdin E, Andersen JK, et al. The intestinal immune system and gut barrier function in obesity and ageing. *FEBS J*. Published online June 2022;21. <https://doi.org/10.1111/febs.16558>.
- [258] Yung JHM, Giacca A. Role of c-jun N-terminal kinase (JNK) in obesity and type 2 diabetes. *Cells* 2020;9(3):706. <https://doi.org/10.3390/cells9030706>.
- [259] Rivas DA, McDonald DJ, Rice NP, Haran PH, Dolnikowski GG, Fielding RA. Diminished anabolic signaling response to insulin induced by intramuscular lipid accumulation is associated with inflammation in aging but not obesity. *Am J Physiol Regul Integr Comp Physiol* 2016;310(7):R561–9. <https://doi.org/10.1152/ajpregu.00198.2015>.
- [260] Tilg H, Moschen AR. Inflammatory mechanisms in the regulation of insulin resistance. *Mol Med* 2008;14(3–4):222–31. <https://doi.org/10.2119/2007-00119.Tilg>.
- [261] Bárcena C, Quirós PM, Durand S, Mayoral P, Rodríguez F, Caravia XM, et al. Methionine restriction extends lifespan in progeroid mice and alters lipid and bile acid metabolism. *Cell Rep* 2018;24(9):2392–403. <https://doi.org/10.1016/j.celrep.2018.07.089>.
- [262] Rajabian N, Ikhapoh I, Shahini S, Choudhury D, Thiyagarajan R, Shahini A, et al. Methionine adenosyltransferase2A inhibition restores metabolism to improve regenerative capacity and strength of aged skeletal muscle. *Nat Commun* 2023;14(1):886. <https://doi.org/10.1038/s41467-023-36483-3>.
- [263] Xu C, Zhu H, Qiu P. Aging progression of human gut microbiota. *BMC Microbiol* 2019;19(1):236. <https://doi.org/10.1186/s12866-019-1616-2>.
- [264] Martino C, Dilmore AH, Burcham ZM, Metcalf JL, Jeste D, Knight R. Microbiota succession throughout life from the cradle to the grave. *Nat Rev Microbiol* 2022;20(12):707–20. <https://doi.org/10.1038/s41579-022-00768-z>.
- [265] Luck H, Tsai S, Chung J, Clemente-Casares X, Ghazarian M, Revelo XS, et al. Regulation of obesity-related insulin resistance with gut anti-inflammatory agents. *Cell Metabol* 2015;21(4):527–42. <https://doi.org/10.1016/j.cmet.2015.03.001>.
- [266] Thevaranjan N, Puchta A, Schulz C, Naidoo A, Szamosi JC, Verschoor CP, et al. Age-associated microbial dysbiosis promotes intestinal permeability, systemic inflammation, and macrophage dysfunction. *Cell Host Microbe* 2017;21(4):455–66. <https://doi.org/10.1016/j.chom.2017.03.002>. e4.
- [267] Winer DA, Winer S, Dranse HJ, Lam TKT. Immunologic impact of the intestine in metabolic disease. *J Clin Invest* 2017;127(1):33–42. <https://doi.org/10.1172/JCI88879>.
- [268] Luck H, Khan S, Kim JH, Copeland JK, Revelo XS, Tsai S, et al. Gut-associated IgA+ immune cells regulate obesity-related insulin resistance. *Nat Commun* 2019;10(1):3650. <https://doi.org/10.1038/s41467-019-11370-y>.
- [269] Ciocca M, Zaffina S, Fernandez Salinas A, Bocci C, Palomba P, Conti MG, et al. Evolution of human memory B cells from childhood to old age. *Front Immunol* 2021;12:690534. <https://doi.org/10.3389/fimmu.2021.690534>.
- [270] Hancock ML, Meyer RC, Mistry M, Khetani RS, Wagschal A, Shin T, et al. Insulin receptor associates with promoters genome-wide and regulates gene expression. *Cell* 2019;177(3):722–36. <https://doi.org/10.1016/j.cell.2019.02.030>. e22.
- [271] Nagao H, Jayavelu AK, Cai W, Pan H, Dreyfuss JM, Batista TM, et al. Unique ligand and kinase-independent roles of the insulin receptor in regulation of cell cycle, senescence and apoptosis. *Nat Commun* 2023;14(1):57. <https://doi.org/10.1038/s41467-022-35693-5>.
- [272] Dall'Agnese A, Platt JM, Zheng MM, Friesen M, Dall'Agnese G, Blaise AM, et al. The dynamic clustering of insulin receptor underlies its signaling and is disrupted in insulin resistance. *Nat Commun* 2022;13(1):7522. <https://doi.org/10.1038/s41467-022-35176-7>.
- [273] Blevie A, Motta F, Durante B, Pandolfo C, Selmi C, Sica A. Immunosenescence, inflammaging, and frailty: role of myeloid cells in age-related diseases. *Clin Rev Allergy Immunol* 2023;64(2):123–44. <https://doi.org/10.1007/s12016-021-08909-7>.
- [274] Davison-Castillo P, McMahon B, Aguila S, Bark D, Ashworth K, Allowazi A, et al. TNF- α -driven inflammation and mitochondrial dysfunction define the platelet hyperreactivity of aging. *Blood* 2019;134(9):727–40. <https://doi.org/10.1182/blood.2019000200>.
- [275] Lee DF, Kuo HP, Chen CT, Wei Y, Chou CK, Hung JY, et al. IKK β suppression of TSC1 function links the mTOR pathway with insulin resistance. *Int J Mol Med* 2008;22(5):633–8. <https://doi.org/10.3892/ijmm.00000065>.
- [276] Tzatsos A, Kandror KV. Nutrients suppress phosphatidylinositol 3-kinase/Akt signaling via raptor-dependent mTOR-mediated insulin receptor substrate 1 phosphorylation. *Mol Cell Biol* 2006;26(1):63–76. <https://doi.org/10.1128/MCB.26.1.63-76.2006>.
- [277] Yoneyama Y, Inamitsu T, Chida K, Iemura SI, Natsume T, Maeda T, et al. Serine phosphorylation by mTORC1 promotes IRS-1 degradation through SCF β -TRCP E3 ubiquitin ligase. *iScience* 2018;5:1–18. <https://doi.org/10.1016/j.isci.2018.06.006>.
- [278] Brisse M, Ly H. Comparative structure and function analysis of the RIG-I-like receptors: RIG-I and MDA5. *Front Immunol* 2019;10:1586. <https://doi.org/10.3389/fimmu.2019.01586>.
- [279] Decout A, Katz JD, Venkatraman S, Ablasser A. The cGAS-STING pathway as a therapeutic target in inflammatory diseases. *Nat Rev Immunol* 2021;21(9):548–69. <https://doi.org/10.1038/s41577-021-00524-z>.
- [280] Di Giorgio E, Xodo LE. Endogenous retroviruses (ERVs): does RLR (RIG-I-like receptors)-MAVS pathway directly control senescence and aging as a consequence of ERV DE-repression? *Front Immunol* 2022;13:917998. <https://doi.org/10.3389/fimmu.2022.917998>.
- [281] Bodur C, Kazyken D, Huang K, Ekim Ustunel B, Siroky KA, Tooley AS, et al. The IKK-related kinase TBK1 activates mTORC1 directly in response to growth factors and innate immune agonists. *EMBO J* 2018;37(1):19–38. <https://doi.org/10.15252/emboj.201696164>.
- [282] Tooley AS, Kazyken D, Bodur C, Gonzalez IE, Fingar DC. The innate immune kinase TBK1 directly increases mTORC2 activity and downstream signaling to

- Akt. *J Biol Chem* 2021;297(2):100942. <https://doi.org/10.1016/j.jbc.2021.100942>.
- [283] Schädel P, Czapka A, Gebert N, Jacobsen ID, Ori A, Wenz O. Metabololipidomic and proteomic profiling reveals aberrant macrophage activation and interrelated immunomodulatory mediator release during aging. *Aging Cell* 2023; e13856. <https://doi.org/10.1111/ace1.13856>. Published online 26 April.
- [284] Fan X, Zeng Y, Song W, Li J, Ai S, Yang D, et al. The role of Sestrins in the regulation of the aging process. *Mech Ageing Dev* 2020;188(111251): 111251. <https://doi.org/10.1016/j.mad.2020.111251>.
- [285] Chen Y, Huang T, Yu Z, Yu Q, Wang Y, Hu J, et al. The functions and roles of sestrins in regulating human diseases. *Cell Mol Biol Lett* 2022;27(1):2. <https://doi.org/10.1186/s11658-021-00302-8>.
- [286] Cangelosi AL, Puzyszynska AM, Roberts JM, Armani A, Nguyen TP, Spinelli JB, et al. Zonated leucine sensing by Sestrin-mTORC1 in the liver controls the response to dietary leucine. *Science* 2022;377(6601):47–56. <https://doi.org/10.1126/science.abi9547>.
- [287] Gu X, Jouandin P, Lalgudi PV, Binari R, Valenstein ML, Reid MA, et al. Sestrin mediates detection of and adaptation to low-leucine diets in *Drosophila*. *Nature* 2022;608(7921):209–16. <https://doi.org/10.1038/s41586-022-04960-2>.
- [288] Wang LX, Zhu XM, Yao YM. Sestrin2: its potential role and regulatory mechanism in host immune response in diseases. *Front Immunol* 2019;10: 2797. <https://doi.org/10.3389/fimmu.2019.02797>.
- [289] Ben-Sahra I, Dirat B, Laurent K, Puissant A, Auberger P, Budanov A, et al. Sestrin2 integrates Akt and mTOR signaling to protect cells against energetic stress-induced death. *Cell Death Differ* 2013;20(4):611–9. <https://doi.org/10.1038/cdd.2012.157>.
- [290] Zeng N, D'Souza RF, Mitchell CJ, Cameron-Smith D. Sestrins are differentially expressed with age in the skeletal muscle of men: a cross-sectional analysis. *Exp Gerontol* 2018;110:23–34. <https://doi.org/10.1016/j.exger.2018.05.006>.
- [291] Yang BA, Castor-Macias J, Fraczek P, Cornett A, Brown LA, Kim M, et al. Sestrins regulate muscle stem cell metabolic homeostasis. *Stem Cell Rep* 2021;16(9):2078–88. <https://doi.org/10.1016/j.stemcr.2021.07.014>.
- [292] Muller FL, Lustgarten MS, Jang Y, Richardson A, Van Remmen H. Trends in oxidative aging theories. *Free Radic Biol Med* 2007;43(4):477–503. <https://doi.org/10.1016/j.freeradbiomed.2007.03.034>.
- [293] Naidoo N. ER and aging-Protein folding and the ER stress response. *Ageing Res Rev* 2009;8(3):150–9. <https://doi.org/10.1016/j.arr.2009.03.001>.
- [294] Sag D, Carling D, Stout RD, Suttles J. Adenosine 5'-monophosphate-activated protein kinase promotes macrophage polarization to an anti-inflammatory functional phenotype. *J Immunol* 2008;181(12):8633–41. <https://doi.org/10.4049/jimmunol.181.12.8633>.
- [295] Nath N, Khan M, Rattan R, Mangalam A, Makkar RS, de Meester C, et al. Loss of AMPK exacerbates experimental autoimmune encephalomyelitis disease severity. *Biochem Biophys Res Commun* 2009;386(1):16–20. <https://doi.org/10.1016/j.bbrc.2009.05.106>.
- [296] Viollet B, Horman S, Leclerc J, Lantier L, Foretz M, Billaud M, et al. AMPK inhibition in health and disease. *Crit Rev Biochem Mol Biol* 2010;45(4):276–95. <https://doi.org/10.3109/10409238.2010.488215>.
- [297] Fan K, Lin L, Ai Q, Wan J, Dai J, Liu G, et al. Lipopolysaccharide-induced dephosphorylation of AMPK-activated protein kinase potentiates inflammatory injury via repression of ULK1-dependent autophagy. *Front Immunol* 2018;9:1464. <https://doi.org/10.3389/fimmu.2018.01464>.
- [298] Zhang Q, Liu S, Zhang CS, Wu Q, Yu X, Zhou R, et al. AMPK directly phosphorylates TBK1 to integrate glucose sensing into innate immunity. *Mol Cell* 2022;82(23):4519–36. <https://doi.org/10.1016/j.molcel.2022.10.026>. e7.
- [299] Covarrubias AJ, Kale A, Perrone R, Lopez-Dominguez JA, Pisco AO, Kasler HG, et al. Senescent cells promote tissue NAD⁺ decline during ageing via the activation of CD38⁺ macrophages. *Nat Metab* 2020;2(11):1265–83. <https://doi.org/10.1038/s42255-020-00305-3>.
- [300] Bobermin LD, Roppa RHA, Gonçalves CA, Quincozes-Santos A. Ammonia-induced glial-inflammation. *Mol Neurobiol* 2020;57(8):3552–67. <https://doi.org/10.1007/s12035-020-01985-4>.
- [301] Mishra S, Welch N, Karthikeyan M, Bellar A, Musich R, Singh SS, et al. Dysregulated cellular redox status during hyperammonemia causes mitochondrial dysfunction and senescence by inhibiting sirtuin-mediated deacetylation. *Aging Cell* 2023:e13852. <https://doi.org/10.1111/ace1.13852>.
- [302] Franceschi C, Garagnani P, Parini P, Giuliani C, Santoro A. Inflammaging: a new immune-metabolic viewpoint for age-related diseases. *Nat Rev Endocrinol* 2018;14(10):576–90. <https://doi.org/10.1038/s41574-018-0059-4>.
- [303] Dyal SD, Brown MT, Johnson PJ. Ancient invasions: from endosymbionts to organelles. *Science* 2004;304(5668):253–7. <https://doi.org/10.1126/science.1094884>.
- [304] Zampino M, Brennan NA, Kuo PL, Spencer RG, Fishbein KW, Simonsick EM, et al. Poor mitochondrial health and systemic inflammation? Test of a classic hypothesis in the Baltimore Longitudinal Study of Aging. *GeroScience* 2020;42(4):1175–82. <https://doi.org/10.1007/s11357-020-00208-x>.
- [305] Tian Q, Mitchell BA, Zampino M, Fishbein KW, Spencer RG, Ferrucci L. Muscle mitochondrial energetics predicts mobility decline in well-functioning older adults: the baltimore longitudinal study of aging. *Aging Cell* 2022;21(2): e13552. <https://doi.org/10.1111/ace1.13552>.
- [306] Bencivenga L, Strumia M, Rolland Y, Martinez L, Cestac P, Guyonnet S, et al. Biomarkers of mitochondrial dysfunction and inflammaging in older adults and blood pressure variability. *GeroScience* 2023;45(2):797–809. <https://doi.org/10.1007/s11357-022-00697-y>.
- [307] Bugga P, Alam MJ, Kumar R, Pal S, Chattopadhyay N, Banerjee SK. Sirt3 ameliorates mitochondrial dysfunction and oxidative stress through regulating mitochondrial biogenesis and dynamics in cardiomyoblast. *Cell Signal* 2022;94(110309):110309. <https://doi.org/10.1016/j.cellsig.2022.110309>.
- [308] Kroemer G, Galassi C, Zitvogel L, Galluzzi L. Immunogenic cell stress and death. *Nat Immunol* 2022;23(4):487–500. <https://doi.org/10.1038/s41590-022-01132-2>.
- [309] Marchi S, Guilbaud E, Tait SWG, Yamazaki T, Galluzzi L. Mitochondrial control of inflammation. *Nat Rev Immunol* 2023;23(3):159–73. <https://doi.org/10.1038/s41577-022-00760-x>.
- [310] Picca A, Lezza AMS, Leeuwenburgh C, Pesce V, Calvani R, Landi F, et al. Fueling inflamm-aging through mitochondrial dysfunction: mechanisms and molecular targets. *Int J Mol Sci* 2017;18(5). <https://doi.org/10.3390/ijms18050933>.
- [311] Chu CT, Ji J, Dagda RK, Jiang JF, Tyurina YY, Kapralov AA, et al. Cardiolipin externalization to the outer mitochondrial membrane acts as an elimination signal for mitophagy in neuronal cells. *Nat Cell Biol* 2013;15(10):1197–205. <https://doi.org/10.1038/ncb2837>.
- [312] Li M, Mandal A, Tyurin VA, DeLucia M, Ahn J, Kagan VE, et al. Surface-binding to cardiolipin nanodomains triggers cytochrome c pro-apoptotic peroxidase activity via localized dynamics. *Structure* 2019;27(5):806–15. <https://doi.org/10.1016/j.str.2019.02.007>. e4.
- [313] Rabiet MJ, Huet E, Boulay F. Human mitochondria-derived N-formylated peptides are novel agonists equally active on FPR and FPRL1, while *Listeria monocytogenes*-derived peptides preferentially activate FPR. *Eur J Immunol* 2005;35(8):2486–95. <https://doi.org/10.1002/eji.200526338>.
- [314] Mallavia B, Liu F, Lefrançois E, Cleary SJ, Kwaan N, Tian JJ, et al. Mitochondrial DNA stimulates TLR9-dependent neutrophil extracellular trap formation in primary graft dysfunction. *Am J Respir Cell Mol Biol* 2020;62(3): 364–72. <https://doi.org/10.1165/rcmb.2019-01400C>.
- [315] Shen J, Dai Z, Li Y, Zhu H, Zhao L. TLR9 regulates NLRP3 inflammasome activation via the NF-κB signaling pathway in diabetic nephropathy. *Diabetol Metab Syndr* 2022;14(1):26. <https://doi.org/10.1186/s13098-021-00780-y>.
- [316] Revelo XS, Ghazarian M, Chng MHY, Luck H, Kim JH, Zeng K, et al. Nucleic acid-targeting pathways promote inflammation in obesity-related insulin

- resistance. *Cell Rep* 2016;16(3):717–30. <https://doi.org/10.1016/j.celrep.2016.06.024>.
- [317] Pinti M, Cevenini E, Nasi M, De Biasi S, Salvioli S, Monti D, et al. Circulating mitochondrial DNA increases with age and is a familiar trait: implications for ‘inflamm-aging’. *Eur J Immunol* 2014;44(5):1552–62. <https://doi.org/10.1002/eji.201343921>.
- [318] Picca A, Guerra F, Calvani R, Romano R, Coelho-Junior HJ, Damiano FP, et al. Circulating mitochondrial DNA and inter-organelle contact sites in aging and associated conditions. *Cells* 2022;11(4):675. <https://doi.org/10.3390/cells11040675>.
- [319] Yu J, Nagasu H, Murakami T, Hoang H, Broderick L, Hoffman HM, et al. Inflammasome activation leads to Caspase-1-dependent mitochondrial damage and block of mitophagy. *Proc Natl Acad Sci U S A* 2014;111(43):15514–9. <https://doi.org/10.1073/pnas.1414859111>.
- [320] Tigano M, Vargas DC, Tremblay-Belzile S, Fu Y, Sfeir A. Nuclear sensing of breaks in mitochondrial DNA enhances immune surveillance. *Nature* 2021;591(7850):477–81. <https://doi.org/10.1038/s41586-021-03269-w>.
- [321] Próchnicki T, Vasconcelos MB, Robinson KS, Mangan MSJ, De Graaf D, Shkarina K, et al. Mitochondrial damage activates the NLRP10 inflammasome. *Nat Immunol* 2023;24(4):595–603. <https://doi.org/10.1038/s41590-023-01451-y>.
- [322] Ghiringhelli F, Apetoh L, Tesniere A, Aymeric L, Ma Y, Ortiz C, et al. Activation of the NLRP3 inflammasome in dendritic cells induces IL-1 β -dependent adaptive immunity against tumors. *Nat Med* 2009;15(10):1170–8. <https://doi.org/10.1038/nm.2028>.
- [323] Billingham LK, Stoolman JS, Vasan K, Rodriguez AE, Poor TA, Szibor M, et al. Mitochondrial electron transport chain is necessary for NLRP3 inflammasome activation. *Nat Immunol* 2022;23(5):692–704. <https://doi.org/10.1038/s41590-022-01185-3>.
- [324] Martínez-Reyes I, Chandel NS. Mitochondrial TCA cycle metabolites control physiology and disease. *Nat Commun* 2020;11(1):102. <https://doi.org/10.1038/s41467-019-13668-3>.
- [325] Jové M, Pradas I, Mota-Martorell N, Cabré R, Ayala V, Ferrer I, et al. Succination of protein thiols in human brain aging. *Front Aging Neurosci* 2020;12:52. <https://doi.org/10.3389/fnagi.2020.00052>.
- [326] Arts RJW, Novakovic B, Ter Horst R, Carvalho A, Bekkering S, Lachmandas E, et al. Glutaminolysis and fumarate accumulation integrate immunometabolic and epigenetic programs in trained immunity. *Cell Metab* 2016;24(6):807–19. <https://doi.org/10.1016/j.cmet.2016.10.008>.
- [327] Tannahill GM, Curtis AM, Adamik J, Palsson-McDermott EM, McGettrick AF, Goel G, et al. Succinate is an inflammatory signal that induces IL-1 β through HIF-1 α . *Nature* 2013;496(7444):238–42. <https://doi.org/10.1038/nature11986>.
- [328] Wang L, He C. Nrf2-mediated anti-inflammatory polarization of macrophages as therapeutic targets for osteoarthritis. *Front Immunol* 2022;13:967193. <https://doi.org/10.3389/fimmu.2022.967193>.
- [329] Mills EL, Ryan DG, Prag HA, Dikovskaya D, Menon D, Zaslon Z, et al. *Nature* 2018 Apr;556(7699):113–7. <https://doi.org/10.1038/nature25986>.
- [330] Wang Y, Li S, Zhao L, Cheng P, Liu J, Guo F, et al. Aging relevant metabolite itaconate inhibits inflammatory bone loss. *Front Endocrinol* 2022;13:885879. <https://doi.org/10.3389/fendo.2022.885879>.
- [331] Nakahira K, Haspel JA, Rathinam VA, Lee SJ, Dolinay T, Lam HC, et al. Autophagy proteins regulate innate immune responses by inhibiting the release of mitochondrial DNA mediated by the NALP3 inflammasome. *Nat Immunol* 2011;12(3):222–30. <https://doi.org/10.1038/ni.1980>.
- [332] Cloonan SM, Choi AMK. Mitochondria in lung disease. *J Clin Invest* 2016;126(3):809–20. <https://doi.org/10.1172/JCI81113>.
- [333] Dromparis P, Michelakis ED. Mitochondria in vascular health and disease. *Annu Rev Physiol* 2013;75(1):95–126. <https://doi.org/10.1146/annurev-physiol-030212-183804>.
- [334] Nunnari J, Suomalainen A. Mitochondria: in sickness and in health. *Cell* 2012;148(6):1145–59. <https://doi.org/10.1016/j.cell.2012.02.035>.
- [335] Fischer F, Hamann A, Osiewacz HD. Mitochondrial quality control: an integrated network of pathways. *Trends Biochem Sci* 2012;37(7):284–92. <https://doi.org/10.1016/j.tibs.2012.02.004>.
- [336] Szklarczyk R, Nooteboom M, Osiewacz HD. Control of mitochondrial integrity in ageing and disease. *Philos Trans R Soc Lond B Biol Sci* 2014;369(1646):20130439. <https://doi.org/10.1098/rstb.2013.0439>.
- [337] Zhong W, Rao Z, Xu J, Sun Y, Hu H, Wang P, et al. Defective mitophagy in aged macrophages promotes mitochondrial DNA cytosolic leakage to activate STING signaling during liver sterile inflammation. *Aging Cell* 2022;21(6):e13622. <https://doi.org/10.1111/ace1.13622>.
- [338] Palmer AK, Tchonia T, Kirkland JL. Targeting cellular senescence in metabolic disease. *Mol Metabol* 2022;66(101601):101601. <https://doi.org/10.1016/j.molmet.2022.101601>.
- [339] Gorgoulis V, Adams PD, Alimonti A, Bennett DC, Bischof O, Bishop C, et al. Cellular senescence: defining a path forward. *Cell* 2019;179(4):813–27. <https://doi.org/10.1016/j.cell.2019.10.005>.
- [340] Rayess H, Wang MB, Srivatsan ES. Cellular senescence and tumor suppressor gene p16. *Int J Cancer* 2012;130(8):1715–25. <https://doi.org/10.1002/ijc.27316>.
- [341] Kastenhuber ER, Lowe SW. Putting p53 in context. *Cell* 2017;170(6):1062–78. <https://doi.org/10.1016/j.cell.2017.08.028>.
- [342] Uyar B, Palmer D, Kowald A, Murua Escobar H, Barrantes I, et al. Single-cell analyses of aging, inflammation and senescence. *Ageing Res Rev* 2020;64(101156):101156. <https://doi.org/10.1016/j.arr.2020.101156>.
- [343] Basisty N, Kale A, Jeon OH, Kuehnemann C, Payne T, Rao C, et al. A proteomic atlas of senescence-associated secretomes for aging biomarker development. *PLoS Biol* 2020;18(1):e3000599. <https://doi.org/10.1371/journal.pbio.3000599>.
- [344] Saul D, Kosinsky RL, Atkinson EJ, Doolittle ML, Zhang X, LeBrasseur NK, et al. A new gene set identifies senescent cells and predicts senescence-associated pathways across tissues. *Nat Commun* 2022;13(1):4827. <https://doi.org/10.1038/s41467-022-32552-1>.
- [345] Reyes NS, Krasilnikov M, Allen NC, Lee JY, Hyams B, Zhou M, et al. Sentinel p16INK4a+ cells in the basement membrane form a reparative niche in the lung. *Science* 2022;378(6616):192–201. <https://doi.org/10.1126/science.abf3326>.
- [346] Jeon OH, Mehdipour M, Gil TH, Kang M, Aguirre NW, Robinson ZR, et al. Systemic induction of senescence in young mice after single heterochronic blood exchange. *Nat Metab* 2022;4(8):995–1006. <https://doi.org/10.1038/s42255-022-00609-6>.
- [347] Schafer MJ, Zhang X, Kumar A, Atkinson EJ, Zhu Y, Jachim S, et al. The senescence-associated secretome as an indicator of age and medical risk. *JCI Insight* 2020;5(12). <https://doi.org/10.1172/jci.insight.133668>.
- [348] Pezone A, Olivieri F, Napoli MV, Procopio A, Avedimento EV, Gabrielli A. Inflammation and DNA damage: cause, effect or both. *Nat Rev Rheumatol* 2023;19(4):200–11. <https://doi.org/10.1038/s41584-022-00905-1>.
- [349] Rodier F, Coppé JP, Patil CK, Hoeijmakers WA, Muñoz DP, Raza SR, et al. Persistent DNA damage signalling triggers senescence-associated inflammatory cytokine secretion. *Nat Cell Biol* 2009;11(8):973–9. <https://doi.org/10.1038/ncb1909>.
- [350] Olivieri F, Albertini MC, Orciani M, Ceka A, Cricca M, Procopio AD, et al. DNA damage response (DDR) and senescence: shuttled inflamma-miRNAs on the stage of inflamm-aging. *Oncotarget* 2015;6(34):35509–21. <https://doi.org/10.18632/oncotarget.5899>.
- [351] Suh N. MicroRNA controls of cellular senescence. *BMB Rep* 2018;51(10):493–9. <https://doi.org/10.5483/bmbrep.2018.51.10.209>.
- [352] Sturmlechner I, Zhang C, Sine CC, van Deursen EJ, Jeganathan KB, Hamada N, et al. P21 produces a bioactive secretome that places stressed cells under immunosurveillance. *Science* 2021;374(6567):eabb3420. <https://doi.org/10.1126/science.abb3420>.
- [353] Xu P, Wang M, Song WM, Wang Q, Yuan GC, Sudmant PH, et al. The landscape of human tissue and cell type specific expression and co-

- regulation of senescence genes. *Mol Neurodegener* 2022;17(1):5. <https://doi.org/10.1186/s13024-021-00507-7>.
- [354] Sayed N, Huang Y, Nguyen K, Krejciowa-Rajaniemi Z, Grawe AP, et al. An inflammatory aging clock (iAge) based on deep learning tracks multimorbidity, immunosenescence, frailty and cardiovascular aging. *Nat Aging* 2021;1(7):598–615. <https://doi.org/10.1038/s43587-021-00082-y>.
- [355] Skopelja-Gardner S, An J, Elkon KB. Role of the cGAS-STING pathway in systemic and organ-specific diseases. *Nat Rev Nephrol* 2022;18(9):558–72. <https://doi.org/10.1038/s41581-022-00589-6>.
- [356] Laberge RM, Sun Y, Orjalo AV, Patil CK, Freund A, Zhou L, et al. MTOR regulates the pro-tumorigenic senescence-associated secretory phenotype by promoting IL1A translation. *Nat Cell Biol* 2015;17(8):1049–61. <https://doi.org/10.1038/ncb3195>.
- [357] Schmitt CA, Tchkonja T, Niedernhofer LJ, Robbins PD, Kirkland JL, Lee S. COVID-19 and cellular senescence. *Nat Rev Immunol* 2023;23(4):251–63. <https://doi.org/10.1038/s41577-022-00785-2>.
- [358] Li T, Chen ZJ. The cGAS-cGAMP-STING pathway connects DNA damage to inflammation, senescence, and cancer. *J Exp Med* 2018;215(5):1287–99. <https://doi.org/10.1084/jem.20180139>.
- [359] Yang H, Wang H, Ren J, Chen Q, Chen ZJ. cGAS is essential for cellular senescence. *Proc Natl Acad Sci U S A* 2017;114(23):E4612–20. <https://doi.org/10.1073/pnas.1705499114>.
- [360] Alle Q, Le Borgne E, Milhavel O, Lemaître JM. Reprogramming: emerging strategies to rejuvenate aging cells and tissues. *Int J Mol Sci* 2021;22(8):3990. <https://doi.org/10.3390/ijms22083990>.
- [361] Cai D, Khor S. 'hypothalamic microinflammation' paradigm in aging and metabolic diseases. *Cell Metabol* 2019;30(1):19–35. <https://doi.org/10.1016/j.cmet.2019.05.021>.
- [362] Pham H, Marathe CS, Phillips LK, Trahair LG, Hatzinikolas S, Huynh L, et al. Longitudinal changes in fasting and glucose-stimulated GLP-1 and GIP in healthy older subjects. *J Clin Endocrinol Metab* 2019;104(12):6201–6. <https://doi.org/10.1210/je.2019-01262>.
- [363] Tang Y, Purkayastha S, Cai D. Hypothalamic microinflammation: a common basis of metabolic syndrome and aging. *Trends Neurosci* 2015;38(1):36–44. <https://doi.org/10.1016/j.tins.2014.10.002>.
- [364] Priyanka HP, Nair RS. Neuroimmunomodulation by estrogen in health and disease. *AIMS Neurosci* 2020;7(4):401–17. <https://doi.org/10.3934/Neuroscience.2020025>.
- [365] Walecki M, Eisel F, Klug J, Baal N, Paradowska-Dogan A, Wahle E, et al. Androgen receptor modulates Foxp3 expression in CD4+CD25+Foxp3+ regulatory T-cells. *Mol Biol Cell* 2015;26(15):2845–57. <https://doi.org/10.1091/mbc.E14-08-1323>.
- [366] Kalluri R, LeBleu VS. The biology, function, and biomedical applications of exosomes. *Science* 2020;367(6478):eaau6977. <https://doi.org/10.1126/science.aau6977>.
- [367] Romero-García N, Huete-Acevedo J, Mas-Bargues C, Sanz-Ros J, Dromant M, Borrás C. The double-edged role of extracellular vesicles in the hallmarks of aging. *Biomolecules* 2023;13(1):165. <https://doi.org/10.3390/biom13010165>.
- [368] van Niel G, D'Angelo G, Raposo G. Shedding light on the cell biology of extracellular vesicles. *Nat Rev Mol Cell Biol* 2018;19(4):213–28. <https://doi.org/10.1038/nrm.2017.125>.
- [369] Makhijani P, McGaha TL. Myeloid responses to extracellular vesicles in health and disease. *Front Immunol* 2022;13:818538. <https://doi.org/10.3389/fimmu.2022.818538>.
- [370] Salvi V, Gianello V, Busatto S, Bergese P, Andreoli L, D'Oro U, et al. Exosome-delivered microRNAs promote IFN- α secretion by human plasmacytoid DCs via TLR7. *JCI Insight* 2018;3(10). <https://doi.org/10.1172/jci.insight.98204>.
- [371] Xiao X, Xu M, Yu H, Wang L, Li X, Rak J, et al. Mesenchymal stem cell-derived small extracellular vesicles mitigate oxidative stress-induced senescence in endothelial cells via regulation of miR-146a/Src. *Signal Transduct Targeted Ther* 2021;6(1):354. <https://doi.org/10.1038/s41392-021-00765-3>.
- [372] Weilner S, Keider V, Winter M, Harreither E, Salzer B, Weiss F, et al. Vesicular Galectin-3 levels decrease with donor age and contribute to the reduced osteo-inductive potential of human plasma derived extracellular vesicles. *Aging (Albany NY)* 2016;8(1):16–33. <https://doi.org/10.18632/aging.100865>.
- [373] Díaz-Alvarez L, Ortega E. The many roles of galectin-3, a multifaceted molecule, in innate immune responses against pathogens. *Mediat Inflamm* 2017;2017:9247574. <https://doi.org/10.1155/2017/9247574>.
- [374] Fafián-Labora JA, Rodríguez-Navarro JA, O'Loghlen A. Small extracellular vesicles have GST activity and ameliorate senescence-related tissue damage. *Cell Metabol* 2020;32(1):71–86. <https://doi.org/10.1016/j.cmet.2020.06.004>.
- [375] Salminen A, Kaarniranta K, Kauppinen A. Exosomal vesicles enhance immunosuppression in chronic inflammation: impact in cellular senescence and the aging process. *Cell Signal* 2020;75(109771):109771. <https://doi.org/10.1016/j.cellsig.2020.109771>.
- [376] Ludwig N, Whiteside TL, Reichert TE. Challenges in exosome isolation and analysis in health and disease. *Int J Mol Sci* 2019;20(19):4684. <https://doi.org/10.3390/ijms20194684>.
- [377] Spehar K, Pan A, Beerman I. Restoring aged stem cell functionality: current progress and future directions. *Stem Cell* 2020;38(9):1060–77. <https://doi.org/10.1002/stem.3234>.
- [378] Guo J, Huang X, Dou L, Yan M, Shen T, Tang W, et al. Aging and aging-related diseases: from molecular mechanisms to interventions and treatments. *Signal Transduct Targeted Ther* 2022;7(1):391. <https://doi.org/10.1038/s41392-022-01251-0>.
- [379] Mejia-Ramirez E, Florian MC. Understanding intrinsic hematopoietic stem cell aging. *Haematologica* 2020;105(1):22–37. <https://doi.org/10.3324/haematol.2018.211342>.
- [380] Xu M, Palmer AK, Ding H, Weivoda MM, Pirtskhalava T, White TA, et al. Targeting senescent cells enhances adipogenesis and metabolic function in old age. *Elife* 2015;4:e12997. <https://doi.org/10.7554/eLife.12997>.
- [381] Bogeska R, Mikecin AM, Kaschnitig P, Fawaz M, Büchler-Schäff M, Le D, et al. Inflammatory exposure drives long-lived impairment of hematopoietic stem cell self-renewal activity and accelerated aging. *Cell Stem Cell* 2022;29(8):1273–84. <https://doi.org/10.1016/j.stem.2022.06.012>.
- [382] Josephson AM, Bradaschia-Correa V, Lee S, Leclerc K, Patel KS, Muinos Lopez E, et al. Age-related inflammation triggers skeletal stem/progenitor cell dysfunction. *Proc Natl Acad Sci U S A* 2019;116(14):6995–7004. <https://doi.org/10.1073/pnas.1810692116>.
- [383] Brunet A, Goodell MA, Rando TA. Ageing and rejuvenation of tissue stem cells and their niches. *Nat Rev Mol Cell Biol* 2023;24(1):45–62. <https://doi.org/10.1038/s41580-022-00510-w>.
- [384] Moiseeva V, Cisneros A, Sica V, Deryagin O, Lai Y, Jung S, et al. Senescence atlas reveals an aged-like inflamed niche that blunts muscle regeneration. *Nature* 2023;613(7942):169–78. <https://doi.org/10.1038/s41586-022-05535-x>.
- [385] Cai Y, Song W, Li J, Jing Y, Liang C, Zhang L, et al. The landscape of aging. *Sci China Life Sci* 2022;65(12):2354–454. <https://doi.org/10.1007/s11427-022-2161-3>.
- [386] Ambrosi TH, Marecic O, McArdle A, Sinha R, Gulati GS, Tong X, et al. Aged skeletal stem cells generate an inflammatory degenerative niche. *Nature* 2021;597(7875):256–62. <https://doi.org/10.1038/s41586-021-03795-7>.
- [387] Yang D, de Haan G. Inflammation and aging of hematopoietic stem cells in their niche. *Cells* 2021;10(8):1849. <https://doi.org/10.3390/cells10081849>.
- [388] Chen Z, Amro EM, Becker F, Hölzer M, Rasa SMM, Njeru SN, et al. Cohesin-mediated NF- κ B signaling limits hematopoietic stem cell self-renewal in aging and inflammation. *J Exp Med* 2019;216(1):152–75. <https://doi.org/10.1084/jem.20181505>.

- [389] Zhang B, Gladyshev VN. How can aging be reversed? Exploring rejuvenation from a damage-based perspective. *Adv Genet* 2020;1(1):e10025. <https://doi.org/10.1002/ggn2.10025>.
- [390] Zhu H, Liu X, Ding Y, Tan K, Ni W, Ouyang W, et al. IL-6 coaxes cellular dedifferentiation as a pro-regenerative intermediate that contributes to pericardial ADSC-induced cardiac repair. *Stem. Cell Res Ther* 2022;13(1):44. <https://doi.org/10.1186/s13287-021-02675-1>.
- [391] Menendez JA, Alarcón T. Senescence-inflammatory regulation of reparative cellular reprogramming in aging and cancer. *Front Cell Dev Biol* 2017;5:49. <https://doi.org/10.3389/fcell.2017.00049>.
- [392] Simpson DJ, Olova NN, Chandra T. Cellular reprogramming and epigenetic rejuvenation. *Clin Epigenet* 2021;13(1):170. <https://doi.org/10.1186/s13148-021-01158-7>.
- [393] Yang JH, Hayano M, Griffin PT, Amorim JA, Bonkowski MS, Apostolides JK, et al. Loss of epigenetic information as a cause of mammalian aging. *Cell* 2023;186(2):305–26. <https://doi.org/10.1016/j.cell.2022.12.027>. e27.
- [394] Sarkar TJ, Quarta M, Mukherjee S, Colville A, Paine P, Doan L, et al. Transient non-integrative expression of nuclear reprogramming factors promotes multifaceted amelioration of aging in human cells. *Nat Commun* 2020;11(1):1545. <https://doi.org/10.1038/s41467-020-15174-3>.
- [395] Gill D, Parry A, Santos F, Okkenhaug H, Todd CD, Hernando-Herraez I, et al. Multi-omic rejuvenation of human cells by maturation phase transient reprogramming. *Elife* 2022;11. <https://doi.org/10.7554/eLife.71624>.
- [396] Zeng X, Li X, Li X, Wei C, Shi C, Hu K, et al. Fecal microbiota transplantation from young mice rejuvenates aged hematopoietic stem cells by suppressing inflammation. *Blood* 2023;141(14):1691–707. <https://doi.org/10.1182/blood.2022017514>.
- [397] Ghosh TS, Shanahan F, O'Toole PW. The gut microbiome as a modulator of healthy ageing. *Nat Rev Gastroenterol Hepatol* 2022;19(9):565–84. <https://doi.org/10.1038/s41575-022-00605-x>.
- [398] Amsterdam D, Ostrov BE. The impact of the microbiome on immunosenescence. *Immunol Invest* 2018;47(8):801–11. <https://doi.org/10.1080/08820139.2018.1537570>.
- [399] Galkin F, Mamoshina P, Aliper A, Putin E, Moskalev V, Gladyshev VN, et al. Human gut microbiome aging clock based on taxonomic profiling and deep learning. *iScience* 2020;23(6):101199. <https://doi.org/10.1016/j.isci.2020.101199>.
- [400] Molinero N, Antón-Fernández A, Hernández F, Ávila J, Bartolomé B, Moreno-Arribas MV. Gut Microbiota, an additional hallmark of human aging and neurodegeneration. *Neuroscience* 2023;518:141–61. <https://doi.org/10.1016/j.neuroscience.2023.02.014>.
- [401] Segain JP, Raingeard de la Blétière D, Bourrille A, Leray V, Gervois N, Rosales C, et al. Butyrate inhibits inflammatory responses through NFκB inhibition: implications for Crohn's disease. *Gut* 2000;47(3):397–403. <https://doi.org/10.1136/gut.47.3.397>.
- [402] Zhou L, Zhang M, Wang Y, Dorfman RG, Liu H, Yu T, et al. Faecalibacterium prausnitzii produces butyrate to maintain Th17/Treg balance and to ameliorate colorectal colitis by inhibiting histone deacetylase 1. *Inflamm Bowel Dis* 2018;24(9):1926–40. <https://doi.org/10.1093/ibd/izy182>.
- [403] Ang QY, Alexander M, Newman JC, Tian Y, Cai J, Upadhyay V, et al. Ketogenic diets alter the gut microbiome resulting in decreased intestinal Th17 cells. *Cell* 2020;181(6):1263–75. <https://doi.org/10.1016/j.cell.2020.04.027>. e16.
- [404] Biagi E, Nylund L, Candela M, Ostan R, Bucci L, Pini E, et al. Through ageing, and beyond: gut microbiota and inflammatory status in seniors and centenarians. *PLoS One* 2010;5(5):e10667. <https://doi.org/10.1371/journal.pone.0010667>.
- [405] Biagi E, Franceschi C, Rampelli S, Severgnini M, Ostan R, Turroni S, et al. Gut Microbiota and extreme longevity. *Curr Biol* 2016;26(11):1480–5. <https://doi.org/10.1016/j.cub.2016.04.016>.
- [406] Greer RL, Dong X, Moraes AC, Zielke RA, Fernandes GR, Peremyslova E, et al. Akkermansia muciniphila mediates negative effects of IFNγ on glucose metabolism. *Nat Commun* 2016;7:13329. <https://doi.org/10.1038/ncomms13329>.
- [407] Ottman N, Reunanen J, Meijerink M, Pietilä TE, Kainulainen V, Klivink J, et al. Pili-like proteins of Akkermansia muciniphila modulate host immune responses and gut barrier function. *PLoS One* 2017;12(3):e0173004. <https://doi.org/10.1371/journal.pone.0173004>.
- [408] Lin YH, Luck H, Khan S, Schneeberger PHH, Tsai S, Clemente-Casares X, et al. Aryl hydrocarbon receptor agonist indigo protects against obesity-related insulin resistance through modulation of intestinal and metabolic tissue immunity. *Int J Obes* 2019;43(12):2407–21. <https://doi.org/10.1038/s41366-019-0340-1>.
- [409] Ghosh TS, Das M, Jeffery IB, O'Toole PW. Adjusting for age improves identification of gut microbiome alterations in multiple diseases. *Elife* 2020;9. <https://doi.org/10.7554/eLife.50240>.
- [410] Campbell C, McKenney PT, Konstantinovskiy D, Isaeva OI, Schizas M, Verter J, et al. Bacterial metabolism of bile acids promotes generation of peripheral regulatory T cells. *Nature* 2020;581(7809):475–9. <https://doi.org/10.1038/s41586-020-2193-0>.
- [411] Franssen F, van Beek AA, Borghuis T, Aidy SE, Hugenholtz F, van der Gaast-de Jongh C, et al. Aged gut microbiota contributes to systemical inflammaging after transfer to germ-free mice. *Front Immunol* 2017;8:1385. <https://doi.org/10.3389/fimmu.2017.01385>.
- [412] Badal VD, Vaccariello ED, Murray ER, Yu KE, Knight R, Jeste DV, et al. The gut microbiome, aging, and longevity: a systematic review. *Nutrients* 2020;12(12):3759. <https://doi.org/10.3390/nu12123759>.
- [413] Du H, Bartleson JM, Butenko S, Alonso V, Liu WF, Winer DA, et al. Tuning immunity through tissue mechanotransduction. *Nat Rev Immunol* 2023;23(3):174–88. <https://doi.org/10.1038/s41577-022-00761-w>.
- [414] Park S, Kim B. Aging-related structural change in 3D extracellular matrix affects its mechanics. *Med Eng Phys* 2022;106(103843):103843. <https://doi.org/10.1016/j.medengphy.2022.103843>.
- [415] Schmauck-Medina T, Molière A, Lautrup S, Zhang J, Chlopicki S, Madsen HB, et al. New hallmarks of ageing: a 2022 Copenhagen ageing meeting summary, 14. NY: Aging (Albany); 2022. p. 6829–39. <https://doi.org/10.18632/aging.204248>.
- [416] Lee M, Du H, Winer DA, Clemente-Casares X, Tsai S. Mechanosensing in macrophages and dendritic cells in steady-state and disease. *Front Cell Dev Biol* 2022;10:1044729. <https://doi.org/10.3389/fcell.2022.1044729>.
- [417] Sładitschek-Martens HL, Guarnieri A, Brumana G, Zanconato F, Battilana G, Xiccato RL, et al. YAP/TAZ activity in stromal cells prevents ageing by controlling cGAS-STING. *Nature* 2022;607(7920):790–8. <https://doi.org/10.1038/s41586-022-04924-6>.
- [418] Chakraborty M, Chu K, Shrestha A, Revelo XS, Zhang X, Gold MJ, et al. Mechanical stiffness controls dendritic cell metabolism and function. *Cell Rep* 2021;34(2):108609. <https://doi.org/10.1016/j.celrep.2020.108609>.